Relative telomere length (RTL) was determined by a PCR-based method in CD56\textsuperscript{bright}, CD56\textsuperscript{dim} 1KIR\(^{-}\)CD57\(^{-}\), 1KIR\(^{-}\)CD57\(^{+}\), and 3-4KIR\(^{-}\)CD57\(^{+}\) NK cell subsets \((n = 7; \text{ mean})\). Telomere length was measured by quantitative PCR as previously described (Cawthon RM., Nucelic Acids Research. 2002;30:e47). The following amendments were made to the original protocol: total DNA concentration of 20 ng per aliquot and SYBR Green/ROX PCR master mix (SABiosciences) were used.
Figure S2. Gating scheme used to identify human NK cells in BALB/c Rag2$^{-/-}$γcR$^{-/-}$ mice

Gating of lymphocytes was based on size (FSC vs. SSC) followed by exclusion of dead cells (not shown). CD45$^+$ human cells were identified followed by exclusion of CD19$^+$ B cells and CD3$^+$ T cells. Thereafter, NK cells were identified as CD56$^-$NKG2A$^{+/+}$ cells.
Figure S3. Differentiation and education are uncoupled processes

(A) Expression of CD57 on singleKIR⁻NKG2A⁻ educated and uneducated NK cells stratified based on individual KIRs and donor HLA type. (B) Degranulation (CD107a expression) by educated and uneducated singleKIR⁻CD57⁺/⁻NKG2A⁻ NK cell subsets stratified based on individual KIRs and donor HLA type after co-culture with K562 cells for six hours. (C) Expression of IFN-γ in educated and uneducated
singleKIR^+CD57^+/−NKG2A^− NK cell subsets stratified based on individual KIRs and donor HLA type after 24 hours stimulation with IL-12 + IL-15. (D) Expression of IFN-γ in educated and uneducated singleKIR^+CD57^+/−NKG2A^− NK cell subsets stratified based on individual KIRs and donor HLA type after 18 hours of stimulation with IL-12 + IL-15 and six hours of co-culture with K562 cells.