Interaction between platelet factor 4 and heparins: thermodynamics determines conformational changes required for binding of anti-platelet factor 4/heparin antibodies

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Short title: Characterization of the antigen in HIT

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Abstract

The chemokine platelet factor 4 (PF4) undergoes conformational changes when complexing with polyanions. This can induce the antibody-mediated adverse drug effect heparin-induced thrombocytopenia (HIT). Understanding why the endogenous protein PF4 becomes immunogenic when complexing with heparin is important for the development of other negatively charged drugs and may also hint towards more general mechanisms underlying the induction of autoantibodies to other proteins. By circular dichroism spectroscopy, atomic force microscopy and isothermal titration calorimetry we characterized the interaction of PF4 with unfractionated heparin (UFH), its 16-, 8-, and 6-mer subfractions, low molecular weight heparin (LMWH), and the pentasaccharide fondaparinux. To bind anti-PF4/heparin antibodies, PF4/heparin complexes require: 1) an increase in PF4 anti-parallel β-sheets exceeding ~30% (achieved by UFH, LMWH, 16-, 8-, 6-mer), 2) formation of multimolecular complexes (UFH, 16-, 8-mer), and 3) energy (needed for a conformational change) which is released by binding of ≥11-mer heparins to PF4, but not by smaller heparins. These findings may help to synthesize safer heparins. Beyond PF4 and HIT, the methods applied in the current study may be relevant to unravel mechanisms making other endogenous proteins more vulnerable, to undergo conformational changes with little energy requirement (e.g. point mutations; post-translational modifications) and thereby prone them to become immunogenic.

Key Points:

1. Besides clustering platelet factor 4/polyanion complexes require input of energy to become immunogenic.
Introduction

Heparin, clinically widely used as a parenteral anticoagulant\textsuperscript{1-3} is a polyanion consisting of iduronic acid (IdoA), glucuronic acid (GlcA), and glucosamine (GlcN) residues carrying sulfate groups. Pharmaceutical heparin is obtained from porcine gut mucosa and, as biological material, composed of polysaccharide chains with variable length. Degradation of unfractionated heparin (UFH) results in less polydisperse and smaller low molecular weight heparins (LMWH). The pentasaccharide fondaparinux consists of the shortest sequence able to catalyze the activity of antithrombin. It was the first synthesized heparin approved for clinical use.\textsuperscript{4} Currently, several other heparin-based, synthesized polysaccharides are in preclinical development.\textsuperscript{5,6} Beside their anticoagulant activity, heparins have other biological effects including potential anti-tumor activity,\textsuperscript{7} which, however, differ depending on their chain length.\textsuperscript{7}

Clinically, beside bleeding, heparin-induced thrombocytopenia (HIT) is the most important adverse effect of heparin.\textsuperscript{8} HIT is a life-threatening immune-driven adverse effect, which occurs in up to 3\% of patients receiving UFH after major surgery.\textsuperscript{9} HIT is caused by antibodies that recognize platelet factor 4 (PF4) - a CXC chemokine family protein - in ultralarge multimolecular complexes (ULC) with heparin.\textsuperscript{10} Several of these pathogenic antibodies can bind to the multimolecular complexes of PF4 and heparin, forming immunocomplexes. When these PF4/heparin-IgG immunocomplexes bind to platelets, the Fc-parts of the antibodies crosslink FcγIIa-receptors on platelets, which induces platelet activation and aggregation.\textsuperscript{11} This results in a prothrombotic state and an increased risk for new thrombosis.\textsuperscript{12} Heparin-induced antibodies recognize an antigen exposed on PF4 at a certain PF4/heparin ratio\textsuperscript{10} at which PF4 tetramers\textsuperscript{13} are forced into close approximation\textsuperscript{14} accompanied by charge neutralization.\textsuperscript{14-16}
Previously, we have shown, using circular dichroism (CD) spectroscopy in combination with an enzyme-linked immunosorbent assay (EIA), that the antigenic site is a composite surface formed by at least two (or three) PF4 monomers in a PF4 tetramer. Exposure of this antigenic site occurs, when polyanions induce changes in the structure of PF4 resulting in an increase of the antiparallel β-sheet content in the PF4 secondary structure to more than approximately 30%.17

Here we report the physicochemical characterization of complexes formed between PF4 and UFH, LMWH, or subfractions produced from unfractionated heparin with defined chain lengths (16- (HO16), 8- (HO08), 6- (HO06) mer), as well as with synthetic fondaparinux (5-mer).4 We found that beside the conformational change of PF4 exposing >30% antiparallel β-sheets and formation of large PF4/heparin complexes, the enthalpy of binding (released heat) has to exceed a threshold value to provide the energy for the conformational change of PF4 required to expose the antigenic epitope. These findings may provide relevant aspects to understand the structure-function relationship for other biological functions of heparins derived drugs18 and may also underlay mechanisms making other endogenous proteins immunogenic.

Methods

Ethics

The use of human sera containing anti-PF4/heparin antibodies and human platelets and obtaining whole blood from healthy volunteers has been approved by the Greifswald ethics board.
Reagents: Lyophilized human PF4 isolated from platelets: Chromatec (Greifswald, Germany); unfractionated heparin (UFH) Heparin-Natrium-25000 (Ratiopharm, Ulm, Germany), fondaparinux (Arixtra, GlaxoSmithKline, London, UK); LMWH reviparin (Clivarin 1750, Abbott GmbH, Wiesbaden Germany): hospital pharmacy; heparin oligosaccharides 6-mer (HO06), 8-mer (HO08) and 16-mer (HO16): Iduron Ltd. (Manchester, UK). These heparin fractions are obtained by partial heparin lyase digestion followed by high resolution gel filtration and show a defined length as determined by the manufacturer. However, their interaction with antithrombin, i.e. their anticoagulant capacity, is not defined. The smaller the oligosaccharides become, the more disruption on the pentasaccharide structure required for antithrombin binding occurs.

Enzyme-linked Immunosorbent Assay (EIA)

PF4/heparin EIA was performed as described\textsuperscript{19} with three human sera of patients known to contain anti-PF4/heparin IgG antibodies verified by PF4/heparin EIA and heparin-induced platelet activation (HIPA) test.\textsuperscript{19} UFH, LMWH and the defined length heparins were added in rising concentrations to PF4 to form the complexes before these were coated on the microtiter plate (as indicated in the figures; UFH: 0-14µg/mL, HO16 and HO08: 0-11.7µg/mL, HO06: 0-15µg/mL, fondaparinux: 0-30µg/mL). To verify the reactivity pattern, we then coated the PF4/defined length heparin complexes at the optimal concentration determined by the titration experiment and tested them with an additional panel of 14 characterized sera from patients with serologically confirmed HIT. LMWH had been assessed by the same method.\textsuperscript{17}

Circular Dichroism (CD) Spectroscopy

Changes in the secondary structure of PF4 upon interaction with heparins were studied by recording far-UV CD spectra (200–260nm) using a Chirascan CD spectrometer (Applied Photophysics, Leatherhead, UK) as previously described.\textsuperscript{17} PF4 was dissolved in phosphate
buffered saline (PBS; 155mM NaCl, 1.54mM KH2PO4, 2.71mM Na2HPO4-7H2O, pH 7.2) to a final concentration of 80 µg/mL (2.5µM). Complex formation was carried out at 20°C directly in the CD cuvette (Hellma, Müllheim, Germany) with 5 mm path length. Each measurement started with a pure PF4 solution. Subsequently, increasing amounts of the heparins (UFH, HO16, HO08, HO06 and fondaparinux) were added to the cuvette and CD spectra were recorded for each PF4/heparin ratio. Buffer baselines and baselines of each heparin concentration step (i.e., without PF4 in the solution) were recorded. In the data analysis, the spectra of PF4 alone and of PF4/heparin complexes were corrected for the baselines, path length, number of amino acids, and concentration to obtain the wavelength dependent mean residue delta epsilon (MRDE) values of the PF4/heparin complex. To estimate the secondary structure content of PF4, deconvolution of CD-spectra was carried out with CDNN (software, circular dichroism neural network) using a database of 33 reference proteins.20 LMWH had been assessed by the same method.17

Atomic Force Microscopy (AFM)

To characterize the structural features of the PF4/heparin complexes, AFM imaging in liquid was carried out using a BioScope II scanning probe microscope from Digital Instruments Inc. (Santa Barbara, CA). NanoScope® 7.3 software was used to control the AFM, to set the imaging parameters and for flattening of the images. Pre-formed PF4/heparin complexes (at the optimal ratios found by isothermal titration calorimetry (ITC)) were incubated on freshly cleaved mica for 10s, followed by washing the mica surface with ultrapure water. For imaging, a few drops of ultrapure water were added on the mica with adsorbed samples. All samples were imaged in tapping mode at room temperature using silicon nitride cantilevers DNP-S (Veeco, Camarillo, CA) with a drive frequency of 8-14 kHz in water and a nominal curvature radius of 10nm. AFM images of the PF4/heparin complexes and of their constituents were recorded. All experiments were repeated at least three times.
The height distribution of the AFM imaged features was analyzed using a MatLab script (MathWorks, Natick, MA). Briefly, the script scans over the images identifying the highest points within a moving 17x17 rectangle (see Figure SI1), giving the position and maximum height of the samples adsorbed to mica. To avoid crosstalk with measurement noise, a 0.9 nm threshold (the standard deviation of the background noise in the AFM images) was used in this study. Height values were then merged and plotted into a semi-logarithmic point histogram. As the typically observed height values were on the order of the tip curvature radius, tip convolution led to a strong lateral broadening of the observed structures. We therefore did not further analyze the lateral dimensions of the detected structures.

**Isothermal Titration Calorimetry (ITC)**

The different heparins and PF4 were separately dialyzed against PBS buffer at pH 7.4. ITC measurements were carried out using an iTC200 calorimeter (GE Healthcare Life Sciences). A PF4 solution (500-2470µg/mL [15.6-70µM] in PBS) was added to the sample cell, and a solution of heparin (450-900µg/mL [37-500µM]) was loaded into the injection syringe. For each experiment, a 60 s delay was followed by 19 injections of 1µL of the titrant solution, spaced 240 s apart. The sample cell was stirred at 1,000 rpm throughout and maintained at 25°C. Control titration was performed by injecting heparin into PBS buffer and subtracted prior to data analysis. The area under each peak of the resulting heat profile was integrated, normalized by the concentrations, and plotted against the molar ratio of heparin to PF4 using an Origin script supplied with the instrument (Origin 7, OriginLab Corporation, Northampton, USA). The resulting binding isotherms were fitted by non-linear regression using the single-site model. The stoichiometry of the interaction \( n = c_{\text{Heparin}} / c_{\text{PF4}} \), where \( c \) is the concentration in mol/L, the equilibrium constant \( K_A \), and the change in enthalpy \( \Delta H \) were obtained during the fitting of all titration data. Equilibrium dissociation constants \( K_D \)
were calculated as the reciprocal of $K_A$. The Gibbs free energy change ($\Delta G$) was calculated with the equation $\Delta G = -RT \ln K_A$. All titrations were replicated to determine the experimental standard deviation for each parameter.

**Results**

**Changes in the PF4 secondary structure by UFH, LMWH and defined length heparins and binding of anti-PF4/heparin antibodies**

We compared the structural changes of PF4 (quantified by CD spectroscopy) with anti-PF4/heparin antibody binding to PF4 (quantified by optical density [OD] changes in an EIA at the wavelength 450 nm), when PF4 was incubated with increasing concentrations of UFH, 16-, 8-, 6-mer heparins and fondaparinux. The CD data are given in the left panels and the corresponding EIA results in the right panels of Figure 1; data obtained with LMWH are given in Suppl Figure SI2. We always normalized the heparin concentration by the PF4 tetramer concentration (number of heparin monomers per PF4 tetramer), taking into account that the PF4 concentration slightly changed by adding heparin in the titrating experiments. Moreover, we plotted in Figure 1 only the antiparallel $\beta$-sheet content of PF4, which is indicative for the overall changes of the PF4 structure\(^ {17}\) (Figure SI3).

Figure 1 shows three different patterns of the PF4/polyanion complexes: i) reversible increase in antiparallel $\beta$-sheets of PF4, paralleled by a reversible increase in anti-PF4/heparin antibody binding (complexes with UFH and HO16; Figure 1A-B; and LMWH Figure SI2); ii) non-reversible increase in antiparallel $\beta$-sheets of PF4, and minimal binding of anti-PF4/heparin antibodies (complexes with HO08 and HO06; Figure 1C-D); iii) no increase in antiparallel $\beta$-sheets and no anti-PF4/heparin antibody binding (complexes with
pentasaccharide fondaparinux; Figure 1E). We then tested a panel of 14 well defined sera containing anti-PF4/heparin antibodies, with the PF4/defined length heparin complexes at the optimal concentrations determined by titration, which gave the same results (Figure 1F).

The maximum increase in antiparallel β-sheets (34% for UFH; 34% for LMWH; 39% HO16; 40% for HO08; 36% for HO06) was observed at distinct concentrations of each heparin preparation. Due to the polydisperse (i.e., variably-sized) nature of UFH, it is impossible to define this on a molar basis. We therefore used an approach to take the mean MW of UFH 12 kDa, which corresponds to 39 monomers (saccharide monomers) per molecule. For consistency, the same approach was used for the heparin fragments.

In sharp contrast to all other heparins, no structural changes of PF4 and no anti-PF4/heparin antibody binding were observed for fondaparinux (Figure 1E), which is consistent with previous data.23-25

**AFM morphological characterization of the complexes formed by PF4 with heparins of different chain length**

The differences in anti-PF4/heparin antibody binding raised the question, whether the smaller heparins, albeit causing an increase in anti-parallel β-sheets, form large complexes with PF4. We used tapping mode liquid AFM imaging to characterize the dependency between length of heparin and the size of the complexes formed with PF4.

We show the complexes that PF4 forms with 16, 8, 6-mer heparins and fondaparinux in Figure 2. PF4 forms ultralarge complexes (above 20 nm) with the 16-mer (HO16) and the 8-mer (HO08) heparins (Figure 2C and 2D). The complexes formed by PF4 with HO16 and HO08 have a broad height distribution reaching up to 35 nm and 25 nm, respectively (Figure 2G). The 6-mer (HO06) and the 5-mer (fondaparinux) heparins formed small, if any
complexes with PF4 with heights in the range 1-5 nm (Figure 2E, 2F and 2G). The complexes were similar to PF4 alone, which formed structures with a large height distribution (from 1-7 nm, Figure SI 4A and 4G).

When we tried to measure PF4/UFH complexes, we could not stably immobilize them on the mica surface. Most likely they were so large that they detached when we rinsed the surface. Consistently, in additional experiments with 300 s incubation time and gentle dipping instead of washing we found several large PF4/UFH complexes (data not shown).

We show the control experiment with UFH, 16, 8, 6-mer heparins, and fondaparinux adsorbed on mica surface alone in Figure SI4 B-F where we found grains with average height of 1-2 nm.

**Binding interaction between PF4 and UFH, LMWH and defined chain length heparins: isothermal titration calorimetry (ITC) study**

We assessed the energetic characteristics of the interaction of PF4 with UFH, LMWH and defined chain length heparins by ITC, which directly measures changes in heat that occur during complex formation.\(^{26,27}\) The top panels of Figure 3 show the sequence of the titration, with each peak corresponding to the injection of the solution in the syringe, while the bottom panels show the integrated heat plot as a function of heparin/PF4 tetramer ratio. The thermodynamic parameters are given in Table 1.

For all investigated heparins, the reaction with PF4 was exothermic (heat release; Figure 3), but two distinct reaction patterns occurred: i) reactions with large heat release (enthalpy change) and ii) reactions with little heat release. PF4/UFH, PF4/LMWH, and PF4/HO16 complexes showed the largest heat release (Table 1; Figure 3A-C), while PF4/HO08 (Table 1; Figure 3D), PF4/HO06 (Table 1; Figure 3E) and PF4/fondaparinux complexes showed
approximately 60%, 30% and less than 20% of the heat release of PF4/UFH complexes, respectively (Table 1; Figure 3F, please note the different scales in Figure 3).

We calculated relatively low values of equilibrium dissociation constant $K_D$ for the complexes formed by PF4 with HO16 heparin (0.05μM), LMWH (0.09μM) and with UFH (0.14μM), indicating strong binding, compared to the $K_D$ for the complexes formed with short chain length heparins (1.25μM for PF4/fondaparinux complexes; 2.5μM for PF4/HO06 complexes and 0.2μM for PF4/HO08 complexes).

The key finding of the thermodynamic studies is the change in randomness of the system (entropy, $\Delta S$; Table 1) which correlates very well with the binding capacity of the resulting PF4/heparin complexes for anti-PF4/heparin antibodies (as shown by EIA). We give the enthalpy per PF4 molecule as this is the constant reaction partner in our experiments.

The calculated $\Delta S$ for the PF4/UFH complex and PF4/LMWH complex showed a negative value (-15.6±4.1 cal/mol/K and -11.5±3.8, respectively, with respect to one PF4 molecule), i.e., a considerable amount of energy released after PF4 binding was consumed by the conformational changes of the complexes (an alternative explanation is that hydrophobic functional groups move to the surface of PF4 coming into direct contact with water). A similar pattern (negative change in entropy, $\Delta S = -11.4±6.2$ cal/mol/K with respect to one mol PF4) was found for PF4/HO16 complexes. In contrast, complexes formed by PF4 with HO08, HO06 and fondaparinux showed a positive entropy change, i.e., binding of these heparins does not result in conformational changes of the PF4/heparin complexes which require additional energy.
Discussion

Our studies combining physicochemical characterization with assessment of anti-PF4/heparin antibody binding provide new insights into the interaction mechanism between PF4 and heparins. Using heparin fragments with defined length, we found that PF4/heparin complexes require the following characteristics to bind anti-PF4/heparin antibodies: induction of an increase in anti-parallel $\beta$-sheets in PF4 exceeding ~30%; formation of multimolecular complexes; and an amount of energy larger than -4000 cal/molPF4. This energy is required for the conformational changes in PF4 needed to expose the antigenic epitopes and is provided when heparins $\geq$11-mers bind to PF4.

While we confirm in this study that an increase in the antiparallel $\beta$-sheet content of PF4 above 30% is a requirement\textsuperscript{17} for binding of anti-PF4/heparin antibodies, it was a major surprise when we found that in complexes with HO08, and HO06 the antiparallel $\beta$-sheet content of PF4 increased clearly above 34% (\textit{i.e.} even more than in PF4/UFH complexes). It is well known that a critical heparin chain length of approximately 12 saccharide units is required to form PF4/polyanion complexes that express the antigen to which anti-PF4/heparin antibodies bind\textsuperscript{28} and induce subsequent platelet activation.\textsuperscript{24} 10-mer heparin fragments induce only weak recognition, and 8- and 6-mer heparin fragments are even less\textsuperscript{29,30} or non-reactive.\textsuperscript{28} Consistent with these findings, anti-PF4/heparin antibodies did not (or only minimally) bind to PF4/HO08 and PF4/HO06 complexes in our study. Only fondaparinux did neither increase the antiparallel $\beta$-sheet structures of PF4 nor facilitated binding of anti-PF4/heparin antibodies when complexed with PF4 (Figure 1). However, fondaparinux still binds to PF4 as shown by ITC, where interaction of PF4 with fondaparinux results in an exothermic reaction (\textit{i.e.} the enthalpy change is negative; see Table 1 and Figure 3E).
CD spectroscopy already gave a first hint that HO08 and HO06 form with PF4 different complexes than longer heparins. The typical reversible changes in the secondary structure of PF4 at high heparin concentrations were only seen for UFH, LMWH, and HO16, but not for HO08 and HO06. This feature, can, however, not be the only feature differentiating between PF4/polyanion complexes which bind anti-PF4/heparin antibodies and those which do not. Dextran sulfate also induces irreversible changes in the PF4 secondary structure but at the same time, anti-PF4/heparin antibodies bind strongly to PF4/dextran sulfate complexes.17,31

We assumed that formation of multimolecular complexes is the requirement for anti-PF4/heparin antibody binding as already shown by us14,24,32 and others10 and that at minimum a 12-mer is required for formation of such large complexes.28 Accordingly, we found by AFM that the size of PF4/fondaparinux and PF4/HO06 complexes did not differ largely from the size of PF4 alone (Figure 2). However, PF4/HO08 complexes were as large as PF4/HO16 complexes, and still anti-PF4/heparin antibodies did not bind to them. Thus, exposure of the antigen allowing binding of anti-PF4/heparin antibodies must require more than formation of multimolecular complexes between PF4 and a polyanion even if this induces a change in antiparallel β-sheets of PF4 >30%.

These puzzling observations were further clarified by ITC, which measures the thermodynamic changes when PF4/polyanion complexes are formed. For all investigated heparins, heat was released upon binding to PF4. However, longer heparins induced a higher heat release (negative change in enthalpy) compared to shorter heparins. Normalized per mole PF4 tetramers, the largest heat release (the highest negative values for enthalpy) were measured for the complexes formed by PF4 with UFH, LMWH, and HO16, while HO08, HO06 and fondaparinux (see Tables 1 and 2) induced much less heat release when complexed with PF4. This is an unexpected finding for a mainly electrostatically mediated interaction.33 The smaller heparin molecules should have been able to pack closer to the PF4-
tetramers thereby displacing more water molecules consequently leading to an increased heat release.

By comparing the EIA data with the ITC data, it became obvious that the heat release has to be stronger than approximately −4000 cal/molPF4 to allow expression of the binding site for anti-PF4/heparin antibodies. In other words, this energy is needed to drive the structural changes in PF4 required for exposure of the neoantigens. This is hardly fulfilled by HO08, but not by HO06 and fondaparinux (Table 2).

We then extrapolated the change in enthalpy (heat release) of the different heparins, as shown graphically in Figure 4. The required change in enthalpy approaches the values of UFH and HO16 at a chain length of approximately 11 monosaccharide units. This excellently matches with empirically observed interaction patterns of PF4 with different heparins showing that at least 12 monosaccharides are necessary to induce anti-PF4/heparin antibody binding in vitro. 28-30 This critical heparin chain length can therefore also be interpreted by the minimum chain length that is necessary to release enough energy to drive the PF4 conformational changes, which finally lead to epitope exposure.16

The CD experiments showed different patterns of the anti-parallel β-sheet content of PF4 (Figure 1), which was at higher concentrations reversible when UFH, LMWH, and HO16 were added, but irreversible with HO08 and HO06. Likely, shorter heparins (HO08 and HO06, and presumably all heparins shorter <11-mers) pack closely around PF4 tetramers, thereby each negative charge of the heparin chain finds positive binding partners on PF4. Larger heparins, however, are too long to just bind to one PF4 tetramer. Therefore, they likely bridge between two PF4 tetramers to find maximal binding partners to neutralize their negative charges. This requires close approximation and a conformational change which consumes energy. By adding more heparin, the long heparin molecules all compete for the positive binding sites on PF4. As already proposed in earlier studies34 this then results in
disruption of the multimolecular complexes and reversal of the conformational change. In contrast, fondaparinux, HO06 and HO08 already form energetically favorable complexes with PF4 which will not be reversed by the addition of more fondaparinux, HO06, or HO08.

Our study has some limitations. We determined the structural changes of the PF4 molecule associated with exposure of the antibody binding site. Whether these changes are causative or indirectly related can only be determined when the structure of the binding site(s) for PF4/heparin antibodies are identified. This will require crystallization of PF4/heparin complexes together with an anti-PF4/heparin antibody. Although we did employ typically reacting human anti-PF4/heparin antibodies, the panel of 14 human sera may not cover the entire spectrum of binding characteristics of human anti-PF4/heparin antibodies. In addition, the defined heparin fragments that we used are not characterized for their anticoagulatory potency. They are model substances and might show different physical characteristics, especially depending on the number of their sulfate groups. In the AFM experiments, we had technical difficulties to immobilize PF4/UFH complexes on the mica surface in the fluid phase, in contrast to previous experiments, in which we assessed PF4/heparin complexes by AFM on dried mica. Most likely, the PF4/UFH complexes are so large that they had been flushed away in the present fluid phase experiments. Consistent with this hypothesis we found large PF4/UFH complexes in additional experiments (300 s incubation time and gentle dipping instead of washing; data not shown). In addition, the PF4/polyanion complexes we observed in the fluid phase were predominantly globular, similar to the complexes seen in TEM images of Rauova et al. We did not find ridge-like structures as previously described, which had a height of only about 2.9 nm and a length of up to 200 nm. These differences can be explained by the fact that measurements were carried out in liquid and tip convolution is here much larger than in the previous work. We also did not exclude the interaction of several polyanions with PF4, e.g. endothelial cell heparin sulfate and the
pentasaccharide, which may induce together conformational changes which likely happen in vivo and may be the explanation why fondaparinux can induce anti-PF4/heparin antibodies.

Our study shows for the first time that besides clustering, conformational changes of PF4 by a polyanion do not necessarily lead to the expression of the binding site for anti-PF4/heparin antibodies. This only occurs, if binding of a polyanion to PF4 results in a conformational change which requires input of energy. Only then PF4 exposes structures to which the immune system reacts and which can therefore be seen as danger signal, e.g. for labelling bacteria. Thus our biophysical methods may be applied to guide the development of synthetic heparins and other polyanion based drugs, e.g. aptamers, that do not lead to expression of these danger signals, which results in an increased risk for heparin-induced thrombocytopenia.

Beyond HIT, understanding the conformational changes making PF4 immunogenic may be relevant for mechanisms underlying other autoimmune blood disorders and immune reactions to human recombinant proteins used as biotherapeutics. Many proteins which are the target of autoantibodies in hematology tend to cluster, e.g. PF4 clusters with polyanions like heparin; platelet glycoprotein IIbIIIa (target in immune thrombocytopenia) clusters in rafts, and ADAMTS13 (target in thrombotic thrombocytopenic purpura) may cluster on von Willebrand factor. If in addition to clustering, these proteins undergo conformational changes, they may also trigger an immune response. In this regard, it is of interest that GPIIbIIIa and ADAMTS13 have in common that their genes show many polymorphisms. Potentially, certain point mutation or post-translational modifications allow conformational changes critical for the immune system with less energy input.

In summary, biophysical methods allowed us to characterize the conformational changes, which the endogenous protein PF4 undergoes when it forms complexes with well defined polyanions. Our findings may help to synthesize safer heparins. Beyond HIT these methods
may be relevant to unravel mechanisms which prone other endogenous proteins relevant in autoimmunity to become immunogenic.

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Supplementary Material Available

In supplementary material, an AFM image in which PF4/HO16 complexes on a mica surface are identified using a MatLab script, comparison of PF4 structural changes and PF4/reviparin complex antigenicity obtained from CD and EIA, normalized CD data and deconvolution of PF4 in complexes with heparins, AFM images of PF4 and heparins are given.

Authorship

Martin Kreimann: designed and carried out the ITC and AFM experiments, analyzed and interpreted the results, wrote and reviewed the manuscript.

Sven Brandt: designed and carried out the EIA and CD measurements, analyzed and interpreted the results and reviewed the manuscript.

Krystin Krauel: discussed the results and reviewed the manuscript.

Stephan Block: supported the AFM experiments and participated in the thermodynamical interpretation of the data.
Christiane A. Helm: discussed the results critically and reviewed the manuscript

Werner Weitschies: discussed the results and reviewed the manuscript

Andreas Greinacher: conceptual design of the experiments, critical review and discussion of results, writing of the manuscript

Mihaela Delcea: conceptual design of biophysical characterization of PF4-defined heparins complexes, designed the experiments, interpreted the results, wrote and reviewed the manuscript.

All authors reviewed and approved the final version of the manuscript.

Conflict-of-interest disclosure

None of the authors has to declare any conflict of interest.

References


Table 1: Thermodynamic parameters (calculated per mole PF4 tetramer) for the interaction of PF4 with UFH, HO16, LMWH, HO08, HO06 and fondaparinux at 25°C. The stoichiometry $n$ is the molar ratio heparin molecule/PF4.

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<td>-6663±906</td>
<td>0.09</td>
<td>0.36±0.01</td>
<td>-3515±136</td>
<td>-11.5±3.8</td>
</tr>
<tr>
<td>PF4/HO08</td>
<td>-4240±1467</td>
<td>0.20</td>
<td>1.93±0.42</td>
<td>-17148±635</td>
<td>46.1±23.3</td>
</tr>
<tr>
<td>PF4/HO06</td>
<td>-2071±35</td>
<td>2.50</td>
<td>0.42±0.04</td>
<td>-3190±151</td>
<td>3.7±0.5</td>
</tr>
<tr>
<td>PF4/fondaparinux</td>
<td>-1333±57</td>
<td>1.25</td>
<td>1.30±0.15</td>
<td>-10415±404</td>
<td>30.4±1.4</td>
</tr>
</tbody>
</table>
Table 2: Thermodynamic energies for the interaction of PF4 with UFH, LMWH, HO16, HO08, HO06 and fondaparinux at 25°C, calculated per mole of PF4 tetramers. aThe lower limit for the enthalpy required to drive antigenicity inducing conformational changes was estimated using \( \Delta S \) by taking either the measured \( \Delta S \) values (UFH, LMWH, HO16), or assuming \( \Delta S = -12.8 \text{ cal/mol/K} \) (for heparins labeled with \(^b\)), which is the average of the entropy changes for UFH and HO16. \(^c\)A comparison of the minimum required enthalpy with the measured enthalphy indicates, if an antigenicity inducing conformational change (similar to UFH, LMWH and HO16), is thermodynamically allowed (\( \Delta H > \Delta H_{\text{req}} \)), unlikely (\( \Delta H \approx \Delta H_{\text{req}} \)) or forbidden (\( \Delta H < \Delta H_{\text{req}} \)).

<table>
<thead>
<tr>
<th>Complexes</th>
<th>Enthalpy, ( \Delta H ) [cal/mol]</th>
<th>Required Enthalpy(^a), ( \Delta H_{\text{req}} ) [cal/mol]</th>
</tr>
</thead>
<tbody>
<tr>
<td>PF4/UFH</td>
<td>-6682±1150</td>
<td>&gt; (^c) -4673±1238</td>
</tr>
<tr>
<td>PF4/HO16</td>
<td>-7260±1365</td>
<td>&gt; (^c) -3415±1865</td>
</tr>
<tr>
<td>PF4/LMWH</td>
<td>-6663±906</td>
<td>&gt; (^c) -3450±1140</td>
</tr>
<tr>
<td>PF4/HO08</td>
<td>-4240±1467</td>
<td>( \approx ) (^c) -3840±717(^b)</td>
</tr>
<tr>
<td>PF4/HO06</td>
<td>-2071±35</td>
<td>&lt; (^c) -3840±717(^b)</td>
</tr>
<tr>
<td>PF4/fondaparinux</td>
<td>-1333±57</td>
<td>&lt; (^c) -3840±717(^b)</td>
</tr>
</tbody>
</table>
Figure 1: Comparison of PF4 structural changes (antiparallel β-sheet content, determined by CD spectroscopy) and PF4/heparins antigenicity (OD obtained from EIA measurements) for unfractionated heparin (UFH) (A), HO16 (B), HO08 (C), HO06 (D) and fondaparinux (E) as a function of molar ratio monosaccharides/PF4. The open and filled squares represent average values of antiparallel β-sheet content and of OD values, respectively. Error bars correspond to the standard deviation (calculated from the results of \( n = 3 \) experiments). (F) EIA measurements for PF4 alone or in complex with fondaparinux, HO06, HO08, HO16 and UFH using a panel of 14 well characterized sera containing anti-PF4/heparin antibodies. The filled squares represent average values of the maxima of the OD values. Error bars correspond to the standard deviation. Data for LMWH are given in Figure SI 2.

Figure 2: Representative liquid AFM tapping mode images of PF4 alone (A) and of PF4/UFH (B), PF4/HO16 (C), PF4/HO08 (D), PF4/HO06 (E), and PF4/fondaparinux (F) complexes on mica; the corresponding height histograms derived from all experiments are shown in (G).

Figure 3: Representative binding isotherms for the titration of PF4 with defined chain length heparins. Top panel shows the raw titration data while bottom panel shows the integrated heats as a function of the molar ratio of heparin/PF4 for: UFH (A), LMWH (B), HO16 (C), HO08 (D), HO06 (E), and fondaparinux (F).

Figure 4: Dependence between change in enthalpy and heparin chain length. The change in enthalpy (circles, values taken from Table 2) increases with chain length and reaches the values of HO16 and UFH at a heparin chain length around 11 monosaccharides, which is close to the critical heparin chain length (= 12) that has to be exceeded to form antigenic PF4/heparin complexes. The error bars for HO05 and HO06 are so small that they are hidden by the circles.
A
UFH
β-sheets \( \% \)
24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39
0.0 0.5 1.0 1.5 2.0 2.5

B
HO16
β-sheets \( \% \)
24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39
0.0 0.5 1.0 1.5 2.0 2.5

C
HO08
β-sheets \( \% \)
24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39
0.0 0.5 1.0 1.5 2.0 2.5

D
HO06
β-sheets \( \% \)
24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39
0.0 0.5 1.0 1.5 2.0 2.5

c_{\text{saccharides}}/c_{\text{PF4}}
0 20 40 60

E
Fondaparinux
β-sheets \( \% \)
24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39
0.0 0.5 1.0 1.5 2.0 2.5

c_{\text{saccharides}}/c_{\text{PF4}}
0 20 40 60

F
OD_{450}[\text{AU}]
0.0 0.5 1.0 1.5 2.0 2.5
PF4 Fondaparinux HO06 HO08 HO16 UFH
w/o heparin
Interaction between platelet factor 4 and heparins: thermodynamics determines conformational changes required for binding of anti-platelet factor 4/heparin antibodies

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