Kinetics of chronic lymphocytic leukemia (CLL) cells in tissues and blood during therapy with the BTK inhibitor ibrutinib

Dominik Wodarz¹, Naveen Garg², Natalia L. Komarova¹, Ohad Benjamini³, Michael J. Keating³, William G. Wierda³, Hagop Kantarjian³, Danelle James⁴, Susan O’Brien³, Jan A. Burger³

¹ Departments of Mathematics and Ecology and Evolutionary Biology, University of California, Irvine, California
² Department of Diagnostic Radiology, MD Anderson Cancer Center, Houston, Texas
³ Department of Leukemia, MD Anderson Cancer Center, Houston, Texas
⁴ Pharmacyclics Inc., Sunnyvale, California

Corresponding author:
Jan A. Burger, M.D. Ph.D., Department of Leukemia, Unit 428
The University of Texas MD Anderson Cancer Center
PO Box 301402, Houston, TX 77230-1402, USA
Phone (713) 563-1487 or (713) 792-1865, FAX (713) 794-4297
e-mail: jaburger@mdanderson.org

Running title: CLL cell kinetics during ibrutinib therapy
Key Points

- During ibrutinib therapy, 1.7% of blood and 2.7% of tissue CLL cells die per day which is 3 and 5 times higher than without treatment
- The fraction of CLL cells that redistribute into the blood during ibrutinib treatment represents 23.3±17% of the tissue disease burden

Abstract

The Bruton tyrosine kinase (BTK) inhibitor ibrutinib has excellent clinical activity in patients with chronic lymphocytic leukemia (CLL). Characteristically, ibrutinib causes CLL cell redistribution from tissue sites into the peripheral blood during the initial weeks of therapy. To better characterize the dynamics of this redistribution phenomenon, we correlated serial lymphocyte counts with volumetric changes in lymph node and spleen sizes during ibrutinib therapy. Kinetic parameters were estimated by applying a mathematical model to the data. We found that during ibrutinib therapy, 1.7±1.1% of blood CLL cells and 2.7 ± 0.99% of tissue CLL cells die per day. The fraction of the tissue CLL cells that was redistributed into the blood during therapy was estimated to be 23.3±17% of the total tissue disease burden. These data indicate that the reduction of tissue disease burden by ibrutinib is due more to CLL cell death and less to egress from nodal compartments.
Introduction

BTK is part of the B cell receptor (BCR) signaling cascade, which plays a central pathogenic role in CLL\(^1\). Ibrutinib is a potent (IC\(_{50}\), 0.5 nM) BTK inhibitor which inactivates BTK through irreversible covalent bonding to Cys-481 in the ATP binding domain of BTK\(^2\). Early stage clinical trials found ibrutinib to be particularly active in patients with chronic lymphocytic leukemia (CLL)\(^3,4\) and mantle cell lymphoma (MCL)\(^5\), and the drug recently has been FDA-approved for patients with relapsed CLL and MCL. In CLL, ibrutinib characteristically causes an early redistribution of tissue-resident CLL cells into the PB, with rapid resolution of enlarged lymph nodes, along with a surge in lymphocytosis. After weeks to months of continuous ibrutinib therapy, normalization of lymphocyte counts and remission is observed in the majority of patients\(^3,4,6\). While well-documented, a quantitative understanding of the redistribution phenomenon is still lacking, and it is debated whether the degree of tissue shrinkage accounts for the magnitude of the lymphocytosis, or whether tissue cell death plays a significant role.

Materials and methods

Data from ten previously treated CLL patients were analyzed, who received single-agent ibrutinib at a dose of 420 MG continuously daily on a Phase 1/2 clinical trial (PCYC-1102-CA) at MD Anderson Cancer Center between 2010 and 2012 after approval and by the rules of the institutional review board (IRB) and in accordance with the Declaration of Helsinki. The clinical details of these patients are summarized in Table 1A. Ten patients were selected for this analysis in which serial computed tomography (CT) scans were available to quantify changes in volumes of lymph nodes...
and spleen prior to therapy and at two time points during treatment. These volume changes were translated into numbers of affected tissue CLL cells per patients and set into relation with changes in serial blood lymphocyte counts, using average CLL cell volumes and individual blood volumes (Supplemental Materials 1&2).

In order to characterize the kinetics of the lymphocytosis, a two-compartment mathematical model, based on work by Messmer and colleagues, was fit simultaneously to the data in blood and tissue (Supplementary Materials 1&2). Denoting the number of CLL lymphocytes in the tissues and blood by \(x\) and \(y\), respectively, the model is given by the following ordinary differential equations, which describe the time evolution of these populations during treatment:

\[
\frac{dx}{dt} = -mx - d_1 (x - c) \\
\frac{dy}{dt} = mx - d_2 y
\]

In the tissue compartment, cells can die with a rate \(d_1\), and redistribute into the blood with a rate \(m\). In the blood, CLL cells die with a rate \(d_2\). Lymphocyte homing to tissues and cell proliferation can be ignored because these processes are effectively inhibited by ibrutinib. This is evidenced by preclinical data demonstrating that ibrutinib inhibits thymidine incorporation, CLL cell proliferation, and CLL cell migration and homing. The parameter \(c\) is included to phenomenologically account for the observation that the majority of ibrutinib-treated patients do not achieve complete remissions (Supplementary Material 2).
Results and Discussion

The treatment responses in blood and tissue were consistent with previous patterns\(^3,4\) and are shown in Figure 1, demonstrating a good fit of the model to the data. Volumetric tissue changes in a representative patient are displayed in supplemental Figure S1.

During ibrutinib therapy, blood CLL cells are estimated to die on average with a rate \(d_2 = 0.017 \pm 0.012 \text{ days}^{-1}\) (average \(\pm\) standard deviation per day). Expressed differently, 1.7 \(\pm\) 1.1\% of the cells die per day, and their average life-span is about 58 days (Supplementary Materials 2). In the tissues, the average death rate of CLL cells was found to be \(d_1 = 0.027 \pm 0.01 \text{ days}^{-1}\), or 2.7\% \(\pm\) 0.99\% of the cells die per day. Thus, they have a shorter life-span of about 37 days. The average rate at which tissue CLL cells redistribute into the blood is calculated to be \(m = 0.008 \pm 0.005 \text{ days}^{-1}\), and is significantly smaller than the cell death rate in tissue (\(p = 0.0002\)). This translates into 0.8\% \(\pm\) 0.5\% of cells redistributing per day. The overall rate of nodal decline (caused by death + redistribution) is thus given by \(\alpha = d_1 + m = 0.035 \pm 0.006 \text{ days}^{-1}\), i.e. 3.5\% \(\pm\) 0.6\% of cells are lost from tissue per day. This means that the average time until the number of tissue CLL cells has been halved during ibrutinib therapy is 20.3 \(\pm\) 3.6 days. The parameter estimates for the patients are given in Table 1B. This is in line with clinical observation of rapid reduction in lymph nodes sizes during the first weeks of ibrutinib therapy, and CT assessment of nodal sites at later time points (Fig. 1 and Fig. 1 in\(^4\)).

A previous study estimated that in the absence of treatment, approximately 0.5\% of cells died per day\(^7\), although a single combined rather than a compartment-specific death rate was provided. Hence, ibrutinib therapy increases the tissue and blood cell
death rate approximately 5- and 3- fold, respectively. We note that although the methodologies used to estimates parameters previously\textsuperscript{7} and here are different, both are valid techniques and the comparison is instructive. These death rates during ibrutinib therapy may seem high, considering that ibrutinib induced only modest levels of apoptosis in vitro when compared to, for example, cytotoxic agents. Nonetheless, in stromal cell co-cultures and suspension cultures ibrutinib consistently caused apoptosis of approximately 10-20% of CLL cells over 48 hours\textsuperscript{8,9}, which is compatible with our patient-based model. We cannot, however, determine whether the different death rates in the compartments are caused by ibrutinib, or whether this is a treatment-independent observation.

Having obtained these parameters (Table 1B), we can calculate the percentage of the total tissue CLL cells before treatment that have re-distributed into the blood during the lymphocytosis phase of ibrutinib therapy. Significant redistribution originating from the pre-therapy tumor load occurs not only during the initial rise of the lymphocyte levels, but also during the subsequent decline until the dynamics start to stabilize. This phase can be defined mathematically, given by the characteristic time $T_c = 1/d_2 + 1/(d_1 + m)$ (Supplementary Material 2). On average, the percentage of the tissue CLL cell population that was re-distributed into the blood (Table 1B) was thus found to be 23.3 ± 17%. Note that there is a fair amount of variation in this percentage among patients, with numbers ranging from 1.9% to 52.6%. These numbers suggest that reduction of tissue disease burden is largely caused by CLL cell death. This is also supported by supplemental Figure S2, showing a significant positive correlation between the rate of
nodal decline and the death rate of tissue cells, while no correlation was found between the rate of nodal decline and the rate of CLL cell redistribution.

CLL cell redistribution appears to be a class effect of kinase inhibitors interfering with the BCR and chemokine receptor signaling pathways\textsuperscript{11}. Similar clinical effects have been reported for the spleen tyrosine kinase (SYK) inhibitor fostamatinib (R406/R788)\textsuperscript{12} and the PI3K\(\delta\) inhibitor idelalisib (GS-1101)\textsuperscript{13,14}. CLL cell redistribution was even observed in the early treatment approaches using glucocorticoids (GC), and it was thought that the amount of tissue shrinkage was not reflected in the degree of lymphocytosis, pointing to a role for lympholysis\textsuperscript{15}. Our calculations suggest that ibrutinib causes a significant amount of cell death in tissue (a larger death rate than in blood), and that a relatively small fraction of the tissue cell burden redistributes to the blood. We likely overestimated the fraction of total tissue cells that redistributed. Our volumetric analysis did not include the bone marrow, which would increase tissue cell burden and lower the estimated fraction. Our study provides a framework to quantitatively examine treatment efficacy of other tyrosine kinase inhibitors or ibrutinib combination therapy in CLL in different disease compartments.
Acknowledgments

The study was supported by a Cancer Prevention and Research Institute of Texas (CPRIT) grant (to J.A.B.), a Leukemia & Lymphoma Society Scholar Award in Clinical Research (to J.A.B.), and MD Anderson’s Moon Shot Program in CLL.

Author Contributions

D.W., N.G., N.L.K., O.B., and D.J. devised and performed the experiments, analyzed the data, and designed the figures. M.K., W.W., H.K., and S.O’B. provided patient samples and reviewed the manuscript, and J.A.B. designed the research, supervised the study, analyzed the data, and wrote the paper with D.W.

Conflict of Interest

D.J. is an employee of Pharmacyclics. J.A.B. and S.O’B. have received research funding from Pharmacyclics.

Abbreviations and parameter explanations

d_2 = death rate of CLL cells in blood
d_1 = death rate of CLL cells in tissue
m = rate of redistribution of tissue cells to blood
α = overall nodal decline rate, i.e. rate at which cells disappear from the tissue due to redistribution + death, i.e. α=m+d1.
x_0 = total body number of CLL cells in tissue
y_0 = total body number of CLL cells in blood

c = parameter that determines the (relatively low) level to which the number of CLL cells converges in the long term during treatment. It is not important for the analysis, but makes for a better fit once cell numbers stabilize.

Percent = % of pre-treatment tissue tumor burden that is redistributed into the blood during treatment.
References

### Table 1A: Patient characteristics.

<table>
<thead>
<tr>
<th>Patient #</th>
<th>1</th>
<th>2</th>
<th>3</th>
<th>4</th>
<th>5</th>
<th>6</th>
<th>7</th>
<th>8</th>
<th>9</th>
<th>10</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (years)</td>
<td>68</td>
<td>70</td>
<td>81</td>
<td>57</td>
<td>51</td>
<td>75</td>
<td>77</td>
<td>55</td>
<td>67</td>
<td>57</td>
</tr>
<tr>
<td>Sex</td>
<td>F</td>
<td>M</td>
<td>M</td>
<td>M</td>
<td>M</td>
<td>M</td>
<td>M</td>
<td>M</td>
<td>M</td>
<td>M</td>
</tr>
<tr>
<td>IgVH mutation status</td>
<td>M</td>
<td>M</td>
<td>M</td>
<td>U</td>
<td>U</td>
<td>U</td>
<td>U</td>
<td>U</td>
<td>M</td>
<td>U</td>
</tr>
<tr>
<td>FlISH cytogenetics</td>
<td>17p-</td>
<td>neg</td>
<td>11q-</td>
<td>11q-</td>
<td>11q-</td>
<td>11q-</td>
<td>11q-</td>
<td>17p-</td>
<td>11q-</td>
<td>17p-</td>
</tr>
<tr>
<td>Karyotype (abnormal)</td>
<td>yes</td>
<td>yes</td>
<td>no</td>
<td>yes</td>
<td>n.a</td>
<td>yes</td>
<td>yes</td>
<td>yes</td>
<td>yes</td>
<td>yes</td>
</tr>
<tr>
<td>ZAP70*</td>
<td>neg</td>
<td>pos</td>
<td>n.a</td>
<td>neg</td>
<td>neg</td>
<td>pos</td>
<td>pos</td>
<td>neg</td>
<td>n.a</td>
<td>pos</td>
</tr>
<tr>
<td>CD38 (&gt; 30%)</td>
<td>no</td>
<td>yes</td>
<td>yes</td>
<td>no</td>
<td>no</td>
<td>yes</td>
<td>yes</td>
<td>yes</td>
<td>no</td>
<td>yes</td>
</tr>
<tr>
<td>ALC (K/µl)</td>
<td>39.8</td>
<td>10.6</td>
<td>138.1</td>
<td>19.4</td>
<td>26.8</td>
<td>21.2</td>
<td>0.92</td>
<td>29.4</td>
<td>52.5</td>
<td>7.8</td>
</tr>
<tr>
<td>Hb (g/dl)</td>
<td>10.8</td>
<td>15.2</td>
<td>9.6</td>
<td>11.2</td>
<td>13.7</td>
<td>13.5</td>
<td>13.8</td>
<td>13.2</td>
<td>12.2</td>
<td>14.8</td>
</tr>
<tr>
<td>PLT (K/µl)</td>
<td>57</td>
<td>141</td>
<td>72</td>
<td>138</td>
<td>115</td>
<td>68</td>
<td>136</td>
<td>79</td>
<td>104</td>
<td>100</td>
</tr>
</tbody>
</table>

**Abbreviations:** n.a., not available; neg, negative; pos, positive; M, mutated; U, unmutated; ALC, absolute lymphocyte count; Hb, hemoglobin; PLT, platelets; *Zap-70 by immunohistochemistry

All values were at baseline values, prior to start of ibrutinib.
Table 1B: Parameter estimates from the model fit. The notation $d^{-1}$ stands for “per day”. In the last column, percent stands for % of pre-treatment tissue tumor burden that is redistributed into the blood during treatment.

<table>
<thead>
<tr>
<th>patient</th>
<th>blood(ml)</th>
<th>$d_2$($d^{-1}$)</th>
<th>$d_1$($d^{-1}$)</th>
<th>$m$($d^{-1}$)</th>
<th>$\alpha$($d^{-1}$)</th>
<th>$x_0$($x10^6$)</th>
<th>$y_0$($x10^6$)</th>
<th>$c$($x10^6$)</th>
<th>percent</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>3484</td>
<td>0.002</td>
<td>0.027</td>
<td>0.0096</td>
<td>0.037</td>
<td>3034</td>
<td>153</td>
<td>0.4</td>
<td>25.9</td>
</tr>
<tr>
<td>2</td>
<td>5363</td>
<td>0.022</td>
<td>0.015</td>
<td>0.0177</td>
<td>0.033</td>
<td>3064</td>
<td>58</td>
<td>47.5</td>
<td>50</td>
</tr>
<tr>
<td>3</td>
<td>5088</td>
<td>0.014</td>
<td>0.012</td>
<td>0.0146</td>
<td>0.026</td>
<td>7044</td>
<td>674</td>
<td>94.7</td>
<td>52.6</td>
</tr>
<tr>
<td>4</td>
<td>6466</td>
<td>0.016</td>
<td>0.047</td>
<td>0.0009</td>
<td>0.047</td>
<td>30209</td>
<td>120</td>
<td>392.1</td>
<td>1.9</td>
</tr>
<tr>
<td>5</td>
<td>5010</td>
<td>0.018</td>
<td>0.022</td>
<td>0.0095</td>
<td>0.032</td>
<td>2143</td>
<td>217</td>
<td>86.3</td>
<td>29.4</td>
</tr>
<tr>
<td>6</td>
<td>4714</td>
<td>0.014</td>
<td>0.027</td>
<td>0.0061</td>
<td>0.033</td>
<td>4083</td>
<td>73</td>
<td>35.7</td>
<td>18.2</td>
</tr>
<tr>
<td>7</td>
<td>4793</td>
<td>0.010</td>
<td>0.022</td>
<td>0.0056</td>
<td>0.028</td>
<td>1294</td>
<td>3</td>
<td>1.0</td>
<td>19.6</td>
</tr>
<tr>
<td>8</td>
<td>6272</td>
<td>0.011</td>
<td>0.032</td>
<td>0.0023</td>
<td>0.034</td>
<td>15452</td>
<td>521</td>
<td>376.4</td>
<td>6.9</td>
</tr>
<tr>
<td>9</td>
<td>6350</td>
<td>0.047</td>
<td>0.033</td>
<td>0.0088</td>
<td>0.042</td>
<td>6156</td>
<td>358</td>
<td>507.8</td>
<td>19.3</td>
</tr>
<tr>
<td>10</td>
<td>5692</td>
<td>0.018</td>
<td>0.035</td>
<td>0.0034</td>
<td>0.039</td>
<td>7711</td>
<td>38</td>
<td>155.9</td>
<td>8.8</td>
</tr>
<tr>
<td>average</td>
<td></td>
<td>0.017</td>
<td>0.027</td>
<td>0.008</td>
<td>0.035</td>
<td>8019</td>
<td>221</td>
<td>169.8</td>
<td>23.3</td>
</tr>
<tr>
<td>st. dev.</td>
<td></td>
<td>0.011</td>
<td>0.010</td>
<td>0.005</td>
<td>0.006</td>
<td>8799</td>
<td>226</td>
<td>185.4</td>
<td>17.0</td>
</tr>
</tbody>
</table>
Figure legend

Figure 1: Dynamics of cell populations over time for each of the ten patients, numbered consecutively. Dots are clinical data, and lines represent the best model fit (see Table 1 for parameters). For each patient, two graphs are presented. The left graph depicts the total number of blood lymphocytes over time. Note that these numbers do not represent the standard absolute lymphocyte counts, which are typically presented as the number of cells per µL blood. Instead, the number of cells per µL blood was multiplied by the blood volume of each patient (Supplementary Material 1), to provide numbers that are commensurate with the total number of cells in tissue, which are shown in the right graph for each patient. Only two of the three tissue volumes were large enough in order to calculate the number of tissue CLL cells (Supplementary Material 1): the measurement before treatment and the first measurement during treatment. Note different scales on the Y-axes. Note that the measured initial number of cells in blood and the initial number predicted by the fitted model can differ, which is explained further in Supplementary Material 2.
Kinetics of chronic lymphocytic leukemia (CLL) cells in tissues and blood during therapy with the BTK inhibitor ibrutinib

Dominik Wodarz, Naveen Garg, Natalia L. Komarova, Ohad Benjamini, Michael J. Keating, William G. Wierda, Hagop Kantarjian, Danelle James, Susan O’Brien and Jan A. Burger