Title: XMEN disease: a new primary immunodeficiency affecting Mg\(^{2+}\) regulation of immunity against Epstein-Barr virus

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Abstract

Epstein Barr virus (EBV) is an oncogenic gammaherpesvirus that infects and persists in 95% of adults worldwide and has the potential to cause fatal disease, especially lymphoma, in immunocompromised hosts. Primary immunodeficiencies (PIDs) that predispose to EBV-associated malignancies have provided novel insights into the molecular mechanisms of immune defense against EBV. We have recently characterized a novel PID now named “X-linked immunodeficiency with magnesium defect, Epstein-Barr virus (EBV) infection, and neoplasia” (XMEN) disease characterized by loss-of-function mutations in the gene encoding magnesium transporter 1 (MAGT1), chronic high level EBV with increased EBV-infected B cells, and heightened susceptibility to EBV-associated lymphomas. The genetic etiology of XMEN disease has revealed an unexpected quantitative role for intracellular free magnesium in immune functions and led to novel diagnostic and therapeutic strategies. Here, we review the clinical presentation, genetic mutation spectrum, molecular mechanisms of pathogenesis, and diagnostic and therapeutic considerations for this previously unrecognized disease.

Introduction

Epstein-Barr virus (EBV) is a B cell-tropic gammaherpesvirus present in latent form is widespread and affects the majority of adults and children worldwide. While most infections are asymptomatic or cause nonspecific symptoms, about 75% of adolescents and young adults with primary EBV infection develop mononucleosis. Moreover, hosts with acquired immunodeficiencies secondary to post-transplant immunosuppression or HIV are at increased risk of developing EBV+ B cell lymphomas and other opportunistic infections. Failure to control EBV along with the potential lethal sequelae associated with persistent active EBV infection, such as EBV+ B cell lymphomas, fulminant infectious mononucleosis (FIM), chronic active EBV infections (CAEBV) and/or hemophagocytic lymphohistiocytosis (HLH), are key pathological hallmarks of primary immunodeficiencies (PIDs) such as X-linked lymphoproliferative disease type 1 (XLP1), interleukin-2 inducible tyrosine kinase (ITK) deficiency, or CD27 deficiency. These PIDs illustrate key proteins in T cells and natural killer (NK) cells that are important for EBV control. We recently identified a new PID associated with chronic high level EBV and susceptibility to EBV+ B cell lymphomas in a cohort of 7 patients, which has now been named
X-linked immunodeficiency with magnesium defect, EBV infection, and neoplasia (XMEN) disease\textsuperscript{4,5}. Identification of the gene mutated in XMEN, \textit{MAGT1}, has revealed an unexpected but essential role of free intracellular magnesium in the control of EBV by T and NK cells\textsuperscript{4,5}. Here, we review the phenotypic and genotypic features of this disease and how its novel pathogenetic mechanism demonstrates a fundamental role of magnesium transport for immunoregulation and provided new unconventional diagnostic and therapeutic strategies.

\textbf{Clinical manifestations}

We initially identified 3 patients with XMEN disease based on persistent high EBV levels and inverted CD4/CD8 ratio\textsuperscript{4}. Subsequently, we screened 61 patients for CD4 lymphopenia, chronic high EBV levels comparable to those in chronic active EBV (CAEBV) patients (ranging from 1000’s to 100,000’s), and/or EBV-associated B cell lymphomas. Although EBV levels detected in X-Men patients vary widely, uncontrolled EBV should be detectable and extremely high elevations of EBV in a male child should raise the index of suspicion of this disease. While we did not find any additional XMEN patients with only inverted CD4/CD8 ratios, we identified 5 additional unrelated XMEN patients with chronic EBV infection and persistent EBV levels in the blood (Table 1)\textsuperscript{5}. All patients also had splenomegaly. The patients were developmentally normal and had no signs of end organ dysfunction except for patient B.1, who presented with CAEBV disease including hepatitis, pancytopenia, and hemophagocytosis\textsuperscript{6}. We found that except for the youngest individuals (A.1, A.2, C.1), all post-pubertal XMEN patients developed various EBV-associated B cell lymphoproliferative disorders (Table 1). The most consistent clinical feature among the seven XMEN patients characterized thus far is persistent high level of EBV along with increased susceptibility for developing EBV\textsuperscript{+} lymphomas\textsuperscript{4}. All human subjects in this study provided written informed consent in accordance with Helsinki principles, for enrollment in research protocols that were approved by the institutional review board of the National Institute of Allergy and Infectious Diseases, NIH. This study was conducted in accordance with the Declaration of Helsinki.

Two XMEN patients (E.1 and F.1) apparently developed two sequential EBV\textsuperscript{+} tumors. Two patients (B.1 and F.1) underwent allogeneic hematopoietic stem cell transplant and died from transplant-related complications shortly post transplant. Patient F.1 received a 7/8 matched unrelated donor peripheral blood stem cell transplant from a female donor after Campath,
fludarabine, and melphalan conditioning. He had 99.8% engraftment by day +30 but died on day +60 after developing Staphylcoccal bacteremia, hemorrhagic shock, and renal failure. Patient B.1 received a 6/6 matched sibling transplant with cytoxan and fludarabine conditioning but died on day +20 with multiorgan failure, hemophagocytic syndrome, and central pontine demyelination. However, his lymphoma was well-controlled and was largely necrotic at the time of death ⁶.

In addition to elevated EBV levels, two XMEN patients also had excessive childhood infections consistent with an underlying PID. Our index patients (A.1 and A.2) had a history of recurrent otitis media, sinusitis, and diarrhea along with repeated hospitalizations for viral pneumonia. While two XMEN patients had recurrent virus infections, including two episodes of molluscum contagiosum (patient E.1) and severe varicella followed by recurrent zoster (patient F.1), other upper respiratory infections (patient D.1, F.1), epiglottitis (patient D.1), and pertussis (patient F.1) were observed. None of the patients were noted to have failure to thrive. Autoimmune features were not prominent among XMEN patients, but two patients did have autoimmune cytopenias (patients A.1 and D.1). Remarkably, many of the patients did not come to medical attention until they developed EBV-associated malignancies, sometimes as old as age 45.

Although the absolute CD⁴⁺ T cell counts were often above the CD4 lymphopenic threshold (<300 cells/μL or <20% of T cells), we observed that all XMEN patients had CD4:CD8 T cell ratios decreased to ~1 or less, except for patient B.1 who presented with pancytopenia due to malignancy. This is consistent with our finding that CD31⁺ cells amongst the CD4⁺CD27⁻CD45RO⁻ naïve T cells were reduced in the patients compared to normal controls, suggesting reduced thymic output of CD4⁺ T cells ⁴. The patients also had moderately high B cell counts, with increased transitional B cells, and mild neutropenia, which may be secondary to chronic EBV. Variable deficiencies in immunoglobulin levels (IgG, IgA, and IgM) and vaccination responses were found in some patients (Table 1), which might be attributable to deficiency in T follicular helper cells or an underlying B cell dysfunction. T cell proliferation in response to mitogen (phytohaemagglutinin, Candida, concanavalin A, or tetanus) stimulation was inconsistently defective. The clinical features of XMEN disease are summarized in Figure 1A.
Genetic mutation spectrum

XMEN disease is caused by loss of function mutations in \textit{MAGT1}, a 70 kb gene with 10 exons encoding for a 335 amino acid protein. We initially detected a 10 base pair (bp) deletion in the index probands (A.1 and A.2) at the exon-intron junction of exon 7. This mutation led to altered splicing, frame shift, early termination, and loss of protein expression by nonsense-mediated decay of the mRNA\textsuperscript{4}. Subsequent XMEN patients had different \textit{MAGT1} mutations also leading to premature translational termination with loss of mRNA expression. In all cases, there was premature termination of the MAGT1 protein and nonsense-mediated decay of the mRNA leading to deficient gene expression. The mothers of XMEN patients have preferential X chromosome inactivation of the chromosome harboring the mutant allele and appear to be asymptomatic\textsuperscript{4}. The fact that all patients had mutations leading to the same functional deficiency of MAGT1 but had somewhat different clinical presentations at highly variable ages (3-45), suggests that the disease is phenotypically heterogeneous. MAGT1 genotype alone does not seem to be predictive for the time to the onset of lymphoma, which may depend on environmental variables.

Mechanisms of pathogenesis

MAGT1 is an evolutionarily conserved, magnesium-specific transporter expressed ubiquitously in all mammalian cells with higher expression in certain tissues including the hematopoietic lineages\textsuperscript{7}. While it was previously shown to play an essential role in magnesium homeostasis, its function in the immune system was uncharacterized until our discovery of XMEN disease\textsuperscript{4}. We found unexpected immunoregulatory roles of free (unbound) intracellular \textit{Mg}\textsuperscript{2+} which constitutes about 5\% of total intracellular \textit{Mg}\textsuperscript{2+}, most of which is tightly bound to ATP, other polyphosphates, and other intracellular molecules\textsuperscript{4,5}. While loss of MAGT1 does not alter the bound or total pool of intracellular magnesium, it has two consequences on the intracellular free \textit{Mg}\textsuperscript{2+}: (1) the abolition of a rapidly-induced, transient TCR-induced free \textit{Mg}\textsuperscript{2+} flux required for optimal T cell activation (Figure 1B) and (2) the chronic decrease in the basal level of free \textit{Mg}\textsuperscript{2+} (Figure 1C). The second defect leads to loss of expression of NKG2D, a receptor on natural killer (NK) and cytotoxic T lymphocytes (CTLs) involved in antiviral and antitumor cytotoxicity\textsuperscript{4,5}. Remarkably, addition of extracellular \textit{Mg}\textsuperscript{2+} can increase intracellular
free Mg$^{2+}$ and restore NKG2D and cytolytic functions that are abnormal in XMEN lymphocytes. This has therapeutic implications as detailed below.

The transient MAGT1-mediated Mg$^{2+}$ flux induced by TCR stimulation is required to coordinate T cell signaling during T cell activation. Loss of this flux leads to delayed phospholipase γ-1 (PLCγ-1) phosphorylation and impaired downstream signaling events resulting in a reduced TCR-gated calcium flux$^4$ (Figure 1B). Thus, MAGT1 is upstream in the same pathway of calcium regulation that is targeted by conventional immunosuppressants such as cyclosporin A and sirolimus, making it an attractive extracellular therapeutic target. Besides gating the transient Mg$^{2+}$ flux during T cell activation, MAGT1 also sets the basal level of free intracellular Mg$^{2+}$, which is required to maintain the expression of the cytolytic activating receptor NKG2D (Figure 1C). Loss of this activating receptor debilitates specific NKG2D ligand-mediated cytotoxicity functions in NK cells and CTLs, which play essential roles in anti-EBV and anti-tumor immunity$^8$. Consequently, while XMEN patients are capable of generating EBV-specific CTLs in vivo, their cells exhibit a profound killing defect for autologous EBV-immortalized lymphoblastoid B-cell lines (LCLs) and other tumor cell line targets$^5$. XMEN disease is the first PID associated with a specific loss of NKG2D expression. The compromised EBV control in XMEN disease together with the restoration of EBV suppression as NKG2D expression is rescued with Mg$^{2+}$ supplementation indicating that there is a secondary, albeit less efficient, Mg$^{2+}$ transporter that can substitute for MAGT1 in restoring the intracellular free Mg$^{2+}$ level (Figure 1C). These results also established that NKG2D is important for anti-EBV immunity. EBV, unlike CMV, lacks extensive mechanisms of evading NKG2D recognition. Lytic induction in EBV-infected cells can lead to downregulation of HLA class I and upregulation of the NKG2D ligand ULBP1, both of which trigger NK cytotoxicity via NKG2D$^9$. NKG2D is implicated in tumor immunosurveillance, which may also account for the tumorigenic susceptibility in XMEN disease.

The immune cell cytotoxicity defect in XMEN disease also sheds light on the clinical similarities between XMEN disease and XLP1, another PID manifested by inability to control EBV infection and a high incidence of lymphomas. The unusual susceptibility to EBV in XLP1 has been attributed to the loss of 2B4-mediated cytotoxicity against EBV-infected B cells, which requires SAP-associated receptors for recognition$^{10-13}$. Interestingly, NKG2D ligation can synergize with many NK activating receptors including 2B4$^{14}$. The convergence of the disease
pathogenesis of XMEN and XLP1 suggests that the NKG2D-2B4 synergistic cytotoxic pathway may be particularly important for control of EBV and preventing lymphoma development. Thus far, we have only observed EBV+ tumors in XMEN disease. Whether XMEN patients are also susceptible to developing EBV− tumors as in XLP1 remains to be determined.

**Diagnostic and Therapeutic considerations**

The significant mortality associated with XMEN disease due to lymphoma raises the necessity of early diagnosis and close follow-up. Males with unexplained chronic elevated EBV level in the blood and splenomegaly, and/or a past history or family history of either EBV+ B cell lymphoproliferative disease or EBV lymphoma, should be considered for XMEN screening. Patients may have EBV serologies indicative of past infection with borderline transaminase elevations (Table 1), but EBV PCR reveals persistently elevated levels of EBV DNA indicative of poor control of EBV replication. Other clinical features that may suggest the diagnosis of XMEN include a reduced CD4:CD8 ratio (< 1.3), elevated levels of B cells, dysgammaglobulinemia, or defective T lymphocyte stimulation responses.

Reduced NKG2D expression can be detected by flow cytometry on whole blood samples in a research laboratory. As NKG2D can also be down-regulated in the setting of malignancies, a final diagnosis needs to be confirmed by examining MAGT1 mRNA expression or MAGT1 exon sequencing. If no mutations are found, quantitative assessment of MAGT1 mRNA expression is necessary to rule out potential non-coding mutations affecting MAGT1 expression. Although flow cytometry for NKG2D is not currently available in commercial laboratories, MAGT1 sequencing can be performed through the Cincinnati Children’s Molecular Genetics Laboratory or through the NIH Clinical Center.

Once the diagnosis is made, treatment depends on the patient’s clinical condition. Rituximab (anti-CD20 antibody) will likely reduce the level of EBV-infected cells in the blood, but does not completely deplete CD20+ B cells in the tissues. EBV+CD20+ lymphoproliferative lesions have developed after treatment of EBV+CD20+ lesions in patients with chronic active EBV; therefore, it is unlikely that rituximab will cure XMEN. Because T cells from patients with XMEN have impaired EBV-specific killing, infusions of additional autologous virus-specific T cells would be unlikely to be effective. While using closely matched 3rd party EBV specific T cells is another potential therapeutic option, it might complicate future transplantation.
due to allo-sensitization. Chemotherapy is useful to treat lymphomas, but does not prevent future
development of other lymphomas. Two patients with XMEN and lymphoma underwent
allogeneic hematopoietic stem cell transplant (HSCT) but did not survive due transplant related
complications (Table 1). Nonetheless, HSCT might still be curative for patients with a good
match. Patients with XMEN disease who have lymphoproliferative disease on biopsy or who are
in remission after treatment for lymphoma should be considered for HSCT, especially since these
patients have a high incidence of primary and secondary lymphomas. An alternative therapeutic
strategy that clinicians can consider is magnesium supplementation. We have observed that oral
Mg\(^{2+}\) supplementation restored basal intracellular free Mg\(^{2+}\) along with NKG2D expression and
decreased the number of EBV-infected cells in two XMEN patients in vivo\(^5\). The high levels of
EBV in XMEN disease, together with the fact that the percentage of EBV-infected cells is
suppressed as NKG2D expression is reinstated with Mg\(^{2+}\) supplementation, establish the
importance of NKG2D for anti-EBV immunity. Thus far, we have found that continuous oral
magnesium threonate supplementation is a safe and well tolerated treatment\(^5\). Whether this
dietary supplement is sufficient to avert the development of lymphoma or promote lymphoma
clearance remains to be determined.

**Perspectives**

The discovery of XMEN disease has provided a novel understanding of magnesium
biology for immunoregulation. The pathogenesis of XMEN is consistent with its clinical features
given that it highlights the importance of T cell activation signaling and immune cell cytotoxicity
for controlling EBV pathogenesis similar to other PIDs with poor control of EBV\(^3\). As the T cell
activation defect is weak and can be overcome with strong or prolonged stimulation, XMEN
patients do not present with severe, life-threatening infections like many T cell
immunodeficiencies. XMEN disease also has some important differences from XLP1 and ITK
deficiency. Specifically, XMEN patients do not develop FIM, rarely present with HLH, and have
normal levels of NKT cells or iNKT cells, which have also been implicated for EBV control \(^3\).
The lack of these fatal, dysregulated immune responses against EBV in XMEN disease probably
also explains why XLP1 and ITK deficient patients generally present at a younger age with
greater mortality (70% by age 10). While the pathology of XLP1 results from an uncontrolled,
overwhelming CD8+ T cell response triggered by primary EBV infection, XMEN mainly
results from the lack of a sufficient immune response to clear EBV. The morbidity and mortality of XMEN disease mainly stems from EBV-associated malignancies, which may not develop until the second decade. This also makes XMEN disease difficult to recognize early in life. Nonetheless, there could be signs of immunodeficiency prior to acquiring EBV, including recurrent infections and decreased CD4 T cells. The long-term disease course of XMEN patients who never acquires EBV remains uncertain at this point, but one may speculate that they may develop lymphoma regardless as in XLP1 or other types of cancers due to the loss of NKG2D-mediated tumor immunosurveillance. The CD4 lymphopenia and T cell activation defect is mild in XMEN patients, but they may worsen with age and make XMEN patients susceptible to opportunistic infections. As Mg\(^{2+}\) supplementation may reduce the number of EBV infected cells and potentially reduce the risk of developing lymphoma, early screening of XMEN disease in males may be beneficial. Besides providing this new therapeutic strategy for XMEN disease, our revelation of the pathologic mechanism of XMEN disease also suggests MAGT1 is an attractive extracellular therapeutic target for immunomodulation in autoimmune diseases and allogeneic transplantations given that it plays a specific role in regulating T cell activation and immune cell cytotoxicity without affecting B cell activation.
Authorship

F.Y.L., B.C.D. and M.J.L wrote the manuscript. H.S, G.U. and H.M. collected clinical data. All authors reviewed and edit the manuscript. The authors declare no conflict of interest.

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References

**Figure 1: XMEN disease**

(A) XMEN disease clinical features. Major features are present in almost all XMEN patients, while minor features are only found in some. (B) Model of the role of MAGT1 in T Cell Receptor (TCR) activation. In normal T cells, stimulation of the TCR triggers a Mg\(^{2+}\) flux through MAGT1. This transient increase of intracellular Mg\(^{2+}\) is required for the activation of phospholipase C \(\gamma 1\) (PLC\(\gamma 1\)) required for the downstream generation of Ca\(^{2+}\) flux through ORAI by promoting the release of Ca\(^{2+}\) from the endoplasmic reticulum via Inositol 1,4,5-Triphosphate (IP\(_3\)) and its receptor. (C) Model of role of MAGT1 in Mg\(^{2+}\) homeostasis and NKG2D expression. In XMEN patients, the absence of MAGT1 leads to the chronic reduction of intracellular free Mg\(^{2+}\), which is required to maintain the expression of NKG2D.
Table 1: Clinical phenotype of XMEN patients

<table>
<thead>
<tr>
<th>Patient</th>
<th>A.2</th>
<th>C.1</th>
<th>A.1</th>
<th>D.1</th>
<th>E.1</th>
<th>F.1</th>
<th>B.1</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age, yo</td>
<td>3</td>
<td>4</td>
<td>7</td>
<td>16</td>
<td>16</td>
<td>23</td>
<td>45</td>
</tr>
<tr>
<td>Age of death, yo</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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</table>

**MAGT1 mutation**

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<tr>
<th>Genomic</th>
<th>g.46668_46677del10</th>
<th>g.25009G&gt;A</th>
<th>g.46668_46677del10</th>
<th>g.29684C&gt;T</th>
<th>g.43183delC</th>
<th>g.46604G&gt;T</th>
<th>g.29684C&gt;T</th>
</tr>
</thead>
<tbody>
<tr>
<td>cDNA</td>
<td>c.859_997del139</td>
<td>c.172G&gt;A</td>
<td>c.859_997del139</td>
<td>c.409C&gt;T</td>
<td>c.598delC</td>
<td>c.859_997del139</td>
<td>c.409C&gt;T</td>
</tr>
</tbody>
</table>

**Recurents infections**

- Epstein-Barr Virus: +, +, +, +, +, +, +
- Herpes simplex virus: +, - , +, - , - , - , -
- Viral pneumonia: +, - , +, - , - , - , -
- Otitis Media: +, - , +, - , +, +, -
- Sinusitis: +, + , +, +, - , - , -
- Streptococcal pharyngitis: - , - , - , +, - , +, -
- Epiglottitis: - , - , - , +, - , - , -
- Molluscum contagiosum: - , - , - , - , +, - , -
- Varicella + Recurrent zoster: - , - , - , - , - , +, -
- Pertussis: - , - , - , - , - , - , +

**Cancers**

- Lymphoma (age of onset, yo): None, None, None, None, B cell LPD (12), Burkitt’s (7, 14), Hodgkin’s (17, 22), Lymphoma (45)

**Vaccinations Titer**

<table>
<thead>
<tr>
<th>Tetanus Toxoid</th>
<th>+</th>
<th>+</th>
<th>-</th>
<th>ND</th>
<th>+</th>
<th>+/-</th>
<th>ND</th>
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<tbody>
<tr>
<td>Haemophilus influenzae type B</td>
<td>+</td>
<td>ND</td>
<td>+</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
<td>ND</td>
</tr>
<tr>
<td>Diphtheria</td>
<td>ND</td>
<td>ND</td>
<td>+</td>
<td>ND</td>
<td>+</td>
<td>+/-</td>
<td>ND</td>
</tr>
<tr>
<td>Pneumococcal</td>
<td>+/-</td>
<td>+</td>
<td>+</td>
<td>ND</td>
<td>-</td>
<td>-</td>
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**Peripheral blood cell numbers**

<table>
<thead>
<tr>
<th>T cells, %</th>
<th>61.2 (53-75)</th>
<th>ND</th>
<th>54.3 (53-75)</th>
<th>48.6 (53-75)</th>
<th>53.6 (53-75)</th>
<th>69 (55-83)</th>
<th>83.9 (57.3-86.4)</th>
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<tbody>
<tr>
<td>CD4 T cells, %</td>
<td>27.8 (32-51)</td>
<td>ND</td>
<td>13.5 (28-47)</td>
<td>19.1 (31-47)</td>
<td>17 (31-47)</td>
<td>40 (28-57)</td>
<td>74.4 (28.6-57.2)</td>
</tr>
<tr>
<td>CD8 T cells, %</td>
<td>20.8 (14-30)</td>
<td>ND</td>
<td>22.4 (16-30)</td>
<td>43 (18-35)</td>
<td>34 (18-35)</td>
<td>34 (10-39)</td>
<td>8.6 (12.9-46.9)</td>
</tr>
<tr>
<td>CD4/CD8 Ratio</td>
<td>0.7 (0.9-3.7)</td>
<td>0.6 (0.9-3.4)</td>
<td>0.6 (1.4-1.7)</td>
<td>0.55 (0.9-3.4)</td>
<td>0.5 (0.9-3.4)</td>
<td>1.1 (1.0-3.6)</td>
<td>8.5 (1.0-3.6)</td>
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<tr>
<td>B cells, %</td>
<td>26.9 (16-35)</td>
<td>ND</td>
<td>37.1 (14-33)</td>
<td>46 (13-27)</td>
<td>0 (RITX)</td>
<td>44 (6-23)</td>
<td>14.1 (6-23)</td>
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<tr>
<td>NK cells, %</td>
<td>15 (3-15)</td>
<td>ND</td>
<td>5.4 (4-17)</td>
<td>5 (3-22)</td>
<td>7 (3-22)</td>
<td>11 (3-22)</td>
<td>1.7 (4.6-29.8)</td>
</tr>
<tr>
<td>Eosinophils, %</td>
<td>1.85 (0.4-1)</td>
<td>ND</td>
<td>1.2 (0.4-7)</td>
<td>1.5 (0.8-7)</td>
<td>1.2 (0.8-7)</td>
<td>1 (0.8-7)</td>
<td>0.2 (0.8-7)</td>
</tr>
<tr>
<td>Neutrophils, %</td>
<td>8 (22.4-69)</td>
<td>ND</td>
<td>20.4 (28.6-74.5)</td>
<td>37.9 (28.6-74.5)</td>
<td>55 (28.6-74.5)</td>
<td>34 (34-67.9)</td>
<td>88.2 (34-67.9)</td>
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<tr>
<td>Monocytes, %</td>
<td>8.1 (4.2-12.2)</td>
<td>ND</td>
<td>9.7 (4.2-12.3)</td>
<td>13 (4.2-12.3)</td>
<td>9.9 (4.2-12.3)</td>
<td>7 (4.2-12.3)</td>
<td>3.4 (4.2-12.3)</td>
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**Immunoglobulin’s levels (normal range)**

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<thead>
<tr>
<th>IgG, mg/dL</th>
<th>286 (424-1051)</th>
<th>1030 (620-1300)</th>
<th>1160 (633-1280)</th>
<th>1690 (639-1349)</th>
<th>611 (639-1349)</th>
<th>619 (639-1349)</th>
<th>734 (642-1730)</th>
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<tbody>
<tr>
<td>IgA, mg/dL</td>
<td>7 (14-23)</td>
<td>56 (50-200)</td>
<td>87 (25-154)</td>
<td>14.8 (45-236)</td>
<td>35.6 (45-236)</td>
<td>29.9 (70-312)</td>
<td>128 (91-499)</td>
</tr>
<tr>
<td>IgM, mg/dL</td>
<td>55 (48-1680)</td>
<td>115 (60-200)</td>
<td>92 (43-1960)</td>
<td>29 (56-352)</td>
<td>87 (56-352)</td>
<td>38 (56-352)</td>
<td>14 (34-342)</td>
</tr>
<tr>
<td>IgE, IU/mL</td>
<td>2000 (310-2950)</td>
<td>ND</td>
<td>1750 (1070-6890)</td>
<td>2100 (206-1952)</td>
<td>1500 (206-1952)</td>
<td>5 (1.53-114)</td>
<td>5 (0.90)</td>
</tr>
</tbody>
</table>

yo indicates years old; LPD, lymphoproliferation disorder; ND, not determined and +/-, positive for some serotypes, negative for others. Adapted from

* Lymphocyte numbers in peripheral blood were measured a few months prior to death but before chemotherapy and transplant

† EBV+ B LPD in the central nervous system.

‡ B cell lymphomas of two different restrictions: first lambda light chain and then kappa light chain; no MYC/IGH gene rearrangements were found.

§ Presumably two independent lymphomas based on the timing of onset.
XMEN disease: a new primary immunodeficiency affecting Mg$^{2+}$ regulation of immunity against Epstein-Barr virus

Feng-Yen Li, Benjamin Chaigne-Delalande, Helen Su, Gulbu Uzel, Helen Matthews and Michael J. Lenardo