Enrichment of FLI1 and RUNX1 mutations in families with excessive bleeding and platelet dense granule secretion defects

Running title: FLI1 and RUNX1 defects associated with bleeding

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Key points

1. Identification of novel FLI1 and RUNX1 alterations in 6 of 13 patients with excessive bleeding and platelet granule secretion defects.
2. Two FLI1 alterations predicting amino acid substitutions in the DNA binding domain of FLI1 abolish transcriptional activity of FLI1.

Abstract

We analysed candidate platelet function disorder genes in 13 index cases with a history of excessive bleeding in association with a significant reduction in dense granule secretion and impaired aggregation to a panel of platelet agonists. Five of the index cases also had mild thrombocytopenia. Heterozygous alterations in FLI1 and RUNX1, encoding Friend Leukemia Integration-1 and RUNT-related transcription factor 1, respectively, which have a fundamental role in megakaryocytogenesis, were identified in 6 cases, 4 of whom had mild thrombocytopenia. Two FLI1 alterations predicting p.Arg337Trp and p.Tyr343Cys substitutions in the FLI1 DNA binding domain abolished transcriptional activity of FLI1. A 4 bp deletion in FLI1, and 2 splicing alterations and a nonsense variation in RUNX1, which were predicted to cause haploinsufficiency of either FLI1 or RUNX1, were also identified. Our findings suggest that alterations in FLI1 and RUNX1 may be common in patients with platelet dense granule secretion defects and mild thrombocytopenia.
Introduction

Inherited platelet function disorders (PFDs) which cause excessive bleeding are heterogeneous and can seldom be linked to a causative gene using clinical and laboratory phenotype alone. The advent of techniques allowing selective capture and enrichment of target gene sequences, coupled with next generation sequencing (NGS) technology, has facilitated the rapid identification of disease associated variants by enabling the simultaneous analysis of large numbers of genes. We have used NGS to investigate index cases from 13 unrelated families recruited to the UK Genotyping and Phenotyping of Platelets (UK GAPP) study with significantly reduced platelet dense granule secretion using lumi-aggregometry.

Patients and Methods

Recruitment of patients and platelet phenotyping
Index cases and affected relatives from a subgroup of 13 families, recruited to the UK GAPP study on the basis of excessive clinical bleeding and a suspicion of an inherited platelet disorder, were investigated. These individuals were diagnosed with a platelet dense granule secretion defect following platelet phenotyping, but had no other features of Hermansky-Pudlak syndrome. The inclusion and exclusion criteria for recruitment to the study, and the criteria for diagnosis of a dense granule secretion defect, have been described. Platelet aggregation and ATP secretion were assessed in platelet rich plasma using a rationalised panel of platelet agonists and a dual channel Chronolog lumi-aggregometer. This study was approved by the National Research Ethics Service Committee West Midlands–Edgbaston (REC reference: 06/MRE07/36) and conducted in accordance with the Declaration of Helsinki.

Analysis of candidate genes
Genomic DNA was isolated from peripheral blood and NGS of candidate PFD genes was undertaken on an ABI-SOLiD3+ or an Illumina Genome Analyser IIx platform. Novel sequence variations identified by NGS were confirmed by conventional Sanger sequencing. Where possible, patients were selected for NGS analysis on the basis of having an affected relative who was also available for study.

Detection of MYH10 in platelets
Nonmuscle myosin heavy chain IIB (MYH10) was detected in platelets by SDS-PAGE and immunoblotting as described.
Results and Discussion

To date, 366 patients, from 292 unrelated families, have been recruited to the UK GAPP study, of whom 56 (15%) have been classified as having a secretion defect. NGS analysis of candidate PFD genes was undertaken in 13 of the patients with defects in secretion, who were selected, where possible, on the basis of having an affected relative who was also available for study. Novel heterozygous alterations in either the FLI1 or RUNX1 gene were identified in 6 of the families (F1 to F6) (Table 1). Two nonsynonymous FLI1 alterations, c.1009C>T and c.1028A>G, predicting p.R337W and p.Y343C substitutions in the highly conserved DNA binding domain of FLI1 were identified in affected members of F1 and F2, while a 4 bp frameshift deletion in FLI1 (c.992-995del; p.Asn331Thrfs*4) was present in both affected members of F3. Three RUNX1 variations predicted to cause RUNX1 haploinsufficiency were identified; donor splice site transversions, c.508+1G>T and c.351+1G>T, were detected in F4 and F5 respectively, and a nonsense mutation in codon 106 (c.317G>A; p.Trp106Stop) was detected in the index case from F6.

The presence of a FLI1 or RUNX1 alteration was associated with symptoms of excessive bleeding in all index cases, and their affected family members, and with mild thrombocytopenia in 5 of the families (Table 1, supplemental Figure S1). In F4, the splice site alteration in RUNX1 was associated with a normal platelet count in the index case, but with thrombocytopenia in two affected relatives. The missense variations in FLI1 were also associated with alopecia, eczema or psoriasis, and recurrent viral infections in affected individuals in F1, and with mild thrombocytopenia, alopecia and eczema in the affected individuals in F2 (Figure 1A and B).

Examination of platelet secretion and aggregation in all subjects carrying FLI1 or RUNX1 defects consistently revealed the predominant platelet abnormality to be a significant reduction in platelet ATP secretion in response to all agonists tested (Figure 1C, Table 1, supplemental Figure S2 and data not shown). As reported previously for patients with dense granule secretion defects, reductions in platelet aggregation in response to collagen and PAR-1 were observed in some but not all subjects, and were usually more apparent at low or intermediate agonist concentrations (data not shown). 3

FLI1 is a member of the ETS family of transcription factors which is expressed primarily in hematopoietic cells and regulates genes expressed both early and late in megakaryocytopoiesis, including GP6.5,6,8,10 Patients with Paris-Trousseau syndrome (PTS), who are hemizygous for FLI1 as a result of a deletion of chromosome 11q23, have an increased tendency to bleed which is associated with thrombocytopenia, and enlarged
platelets displaying giant α-granules.\textsuperscript{10-12} It is thought that transient monoallelic expression of FLI1 at an early stage during megakaryocytopoiesis, coupled with hemizygous loss of FLI1 in patients with PTS, which results in a complete lack of FLI1 expression, accounts for the generation of two distinct subpopulations of normal and immature megakaryocytes in patients with PTS.\textsuperscript{10} The identification of alterations predicting substitutions in the DNA binding domain of FLI1 and their association with symptoms in two families suggests a role for FLI1 in the pathogenesis of bleeding in these families.

We investigated whether the p.R337W and p.Y343C substitutions could alter the DNA binding capacity of FLI1 by examining the ability of the recombinant FLI1 variants to bind to an ETS site in the GP6 promoter using the dual-luciferase reporter assay to measure GP6 promoter activity. There was a complete loss in the ability of the R337W and Y343C FLI1 variants to induce GP6 promoter activity compared with that of WT-FLI1 (p<0.001; Figure 1E). Furthermore, co-expression of either the R337W or the Y343C variant with WT-FLI1, to mimic heterozygosity, resulted in significant reductions in the transcriptional activity to approximately 60% of that of WT-FLI1 alone (p<0.01) indicating that these mutations disrupt DNA binding of FLI1 and will cause a reduction in activation of megakaryocyte-specific genes (Figure 1E). Of interest is a previous study which showed that R337 is critical for the function of nuclear localisation signal 2 of FLI1 and that an R337A substitution caused a reduction in nuclear accumulation of FLI1.\textsuperscript{13} However, as with the R337W variant, the R337A substitution was shown to disrupt the DNA binding ability of FLI1, and consequently to down regulate expression of FLI1-inducible megakaryocyte-specific genes.\textsuperscript{13}

As a member of the RUNT family of transcription factors, the role of RUNX1 in regulating megakaryopoiesis and platelet formation is well established. RUNX1 has been shown to co-operate with FLI1 in the late stages of megakaryopoiesis.\textsuperscript{14,15} Inherited RUNX1 mutations are recognised to lead to autosomal dominant thrombocytopenia and impaired platelet function, and are associated with a propensity to myelodysplastic syndrome and acute myeloid leukemia (AML).\textsuperscript{1} RUNX1-mediated silencing of the nonmuscle myosin IIb (MYH10) gene is required for the switch from mitosis to endomitosis during megakaryocyte maturation. Persistence of MYH10 in platelets was recently proposed as a biomarker for RUNX1 alterations in patients with familial thrombocytopenia, and for FLI1 deletions in PTS.\textsuperscript{7,16} The detection of MYH10 in platelets from patients carrying the p.Tyr343Cys and the p.Asn331Thrfs*4 FLI1 variations confirms the use of MYH10 detection as a biomarker for FLI1 alterations (Figure 1F). Indeed, immunoblot detection of MYH10 expression in platelets would be a useful initial screening test for inherited platelet disorders caused by abnormalities in RUNX1 or FLI1. Dosage of RUNX1 is thought to contribute to the risk of AML since missense mutations which result in variants that can heterodimerise with normal
RUNX1 and act in a dominant-negative manner to compete with the normal protein have been reported to be associated with a higher risk of haematological malignancy than mutations resulting in haploinsufficiency. The absence of haematological malignancies to date in the three families with RUNX1 mutations identified in this study, which were all predicted to result in haploinsufficiency would support this observation.

The identification of RUNX1 or FLI1 alterations in 6/13 unrelated index cases with dense granule secretion disorder, 4 of whom had thrombocytopenia, suggests that defects in these genes will account for a significant proportion of patients with defects in platelet granule secretion and excessive bleeding.

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Authorship

Contribution: JS and MED wrote the paper which was read and commented on by all authors. JS, NVM, DB, GCL, ML, BD, MAS, KM, KH and VCL contributed to the data collection and laboratory analyses. GCL, ML, KT, JM, JTW, PWC, and MM recruited patients and contributed clinical data to the study. MED and SPW coordinated the study.

Conflict-of-interest disclosure: the authors declare no competing financial interests.
References


Table 1. Genotypic and phenotypic characteristics of subjects with platelet dense granule secretion defects.

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<th>Family (F)/Patient ID◊</th>
<th>FLI1 or RUNX1 alteration ¥</th>
<th>Effect</th>
<th>Platelet count (x10^9/L)^</th>
<th>Mean platelet volume (fL)</th>
<th>ATP Secretion nmol/1x10^8 platelets*</th>
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Heterozygous nucleotide changes present in FLI1 and RUNX1 and their predicted effects on the resulting RNA or protein are shown. ◊ Index cases are indicated in bold font; ¥ Alterations are numbered according to positions in the NM_002017.3 and NM_001754 transcripts for FLI1 and RUNX1 respectively; ^Mean platelet counts are shown, normal reference range is 150-400 x 10^9 platelets/L, thrombocytopenia is defined as platelet count <150 x 10^9 platelets/L; *ATP secreted in response to 100 μM PAR-1 receptor specific peptide SFLLRN, 5th centile in healthy volunteers is 0.82 nmol/1x10^8 platelets; **ATP secretion was measured in response to 1U/ml thrombin in the centre where these subjects were recruited, against a normal reference range of 0.73-1.80 nmol/1x10^8 platelets. ND= not detectable. NA= not available.
Figure Legend

**Figure 1. Excessive bleeding and platelet dense granule secretion defects are associated with heterozygous mutations in FLI1 and RUNX1.**

A and B: Pedigrees showing inheritance of mild bleeding, alopecia totalis and other clinical features in families 1 (A) and 2 (B). Individuals heterozygous for the c.1009C>T and c.1028A>G transitions in FLI1 are indicated. Lines through symbols indicate deceased individuals. In A, individuals with bleeding symptoms and alopecia, and confirmed platelet dense granule secretion defects are indicated by black filled symbols. An asterisk indicates the presence of eczema and a history of recurrent viral infections. The presence of psoriasis is indicated by a ‘+’ sign. In B, individuals with bleeding symptoms and alopecia are indicated by black or grey filled symbols. Black filled symbols indicate individuals with confirmed platelet dense granule secretion defect and mild thrombocytopenia. An asterisk indicates a history of infective endocarditis and the presence of eczema and colitis are indicated by a ‘+’ sign. C. ATP secretion in response to 100μM PAR1 peptide in two members of F1 with the c.1009C>T transition in FLI1, and three members of F4 with the c.508+1G>T transversion in RUNX1 alongside controls. D. Schematic diagram of RUNX1 and FLI1 showing the regions of the proteins affected by mutations identified in this study. Intronic mutations, predicted to interfere with splicing of the RUNX1 RNA, are shown in italics. E. HEK293T cells were co-transfected with WT and variant FLI1 constructs, or combinations thereof, and pGL3.10-GP6-luciferase and pRLnull-Renilla reporters as shown, and firefly and Renilla luciferase expression assessed in cell lysates 48 hours later. Data represents the means (± SEM) of 3 independent experiments; **p<0.01, ***p<0.001. F. MYH10 protein expression in platelets from patients with FLI1 mutations (F2.1 and F2.2 with p.Tyr343Cys FLI1 mutation; F3.1 and F3.2 with p.Asn331Thrfs*4 FLI1 mutation) and a healthy control.
Enrichment of FLI1 and RUNX1 mutations in families with excessive bleeding and platelet dense granule secretion defects