Interleukin-1 in the pathogenesis and treatment of inflammatory diseases

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Abstract

More than any other cytokine family, the interleukin-1 (IL-1) family of ligands and receptors is primarily associated with acute and chronic inflammation. The cytosolic segment of each member of the IL-1 receptor family contains the Toll-IL-1-Receptor (TIR) domain. This domain is also present in each member of the Toll-like Receptors (TLR), the receptors that respond to the wide spectrum of microbial products and viruses. Since TIR is the functional domain of both receptor families, responses to IL-1 are fundamental to the innate immunity. Of the 11 members of the IL-1 family, IL-1β has emerged as a therapeutic target for an expanding number of systemic and local inflammatory conditions termed “auto-inflammatory” diseases. In these diseases, neutralization of IL-1β results in a rapid and sustained reduction in disease severity. Auto-inflammatory diseases are distinct from “autoimmune” diseases. Whereas treatments for autoimmune diseases are immunosuppressive and target T-or B-cells, treatment of auto-inflammatory diseases often target IL-1β. Although some auto-inflammatory diseases are due to mutations in the control of caspase-1 activity, common diseases such as gouty arthritis, type 2 diabetes, post-myocardial heart failure, recurrent pericarditis, osteoarthritis and smoldering multiple myeloma also are responsive to IL-1β neutralization. This review summarizes acute and chronic inflammatory diseases that are treated by reducing IL-1β activity and proposes that disease severity is affected by the anti-inflammatory members of the IL-1 family of ligands and receptors.
Introduction

Since the 1996 publication in *Blood* “Biological Basis for Interleukin-1 in Disease”\(^1\), there have been several major advances in understanding a role for IL-1 in the pathogenesis of disease. Because of its property as a hematopoietic factor, IL-1 was administered to patients in order to improve recovery following bone marrow transplantation (human responses to IL-1 were reviewed in detail in 1996).\(^1\) Effective in reducing the duration of thrombocytopenia and leukopenia, recipients of IL-1 therapy experienced unacceptable signs and symptoms of systemic inflammation, including hypotension. Therefore, attention was initially focused on blocking IL-1 activity in sepsis using the naturally occurring IL-1 receptor antagonist (IL-1Ra), now known by its generic name anakinra. There were three controlled trials of anakinra in human sepsis. Although in each trial there was a reduction in 28-day all cause mortality compared to placebo-treated patients, the reductions did not reach statistical significance.\(^2\) The failure of blocking IL-1 to significantly reduce mortality in septic shock is not unusual, as most anti-cytokines and anti-inflammatory agents have also failed in sepsis trials (reviewed in Eichacker).\(^3\)

Subsequently, attention focused on blocking IL-1 in non-infectious, chronic inflammatory conditions, such as rheumatoid arthritis. Anakinra is approved for reducing the signs and symptoms of rheumatoid arthritis and slows the progressive joint destructive characteristics of the disease. Anakinra has also been administered to patients with smoldering/indolent myeloma at high risk for progression to multiple myeloma. In combination with a weekly low dose of dexamethasone, anakinra treatment provided a significant increase in the number of years of progression free disease.\(^4\) Consistently, there are no organ toxicities or gastrointestinal disturbances with anakinra or other parenteral therapies to reduce IL-1 activity. Blocking IL-1, particularly IL-1\(\beta\), is now the standard of therapy for a class of inflammatory syndromes termed “auto-inflammatory” diseases (reviewed by Kastner and Simon).\(^5,6\) Auto-inflammatory syndromes are distinct from autoimmune diseases. In autoimmune diseases, the T-cell is associated with pathogenesis as the dysfunctional cell or “driver” of inflammation. Immunosuppressive therapies targeting T-cell function as well as antibodies that deplete T and B cells are effective in treating autoimmune diseases. In contrast, in auto-inflammatory diseases, the monocyte-macrophage is the dysfunctional cell, which directly promotes
inflammation. Auto-inflammatory conditions are characterized by recurrent bouts of fever with debilitating local and systemic inflammation; they are often responsive to IL-1β blockade (Table 1). In general, these diseases are poorly controlled with immunosuppressive therapies and responses to blocking TNFα, if any, are modest. In this review, a growing number of unrelated diseases often responsive to blocking IL-1β are discussed.

**From IL-1α and IL-1β to the IL-1 superfamily of ligands and receptors**

*Similarities in the IL-1 family.* Although the original IL-1 family was comprised of only IL-1α and IL-1β, the IL-1 family has expanded considerably. As shown in Table 2, there are eleven members. The IL-1 receptor family has also expanded to nine distinct genes and includes co-receptors, decoy receptors, binding proteins and inhibitory receptors. In terms of human disease, the properties of IL-1 itself still remain the model for mediating inflammation. As shown in Figure 1, IL-1α or IL-1β bind first to the ligand binding chain, termed type I (IL-1RI). This is followed by recruitment of the co-receptor chain, termed the accessory protein (IL-1RAcP). A complex is formed of IL-1RI plus IL-1 plus the co-receptor. The signal is initiated with recruitment of the adaptor protein MyD88 to the Toll-IL-1- Receptor (TIR) domain (see below). Several kinases are phosphorylated, NFκB translocates to the nucleus and the expression of a large portfolio of inflammatory genes takes place. Signal transduction in IL-1-stimulated cells has been reviewed in detail by Kracht and colleagues.

Similarly, IL-18 first binds to its ligand binding chain, IL-18Rα, recruits its co-receptor the IL-18Rβ chain and a pro-inflammatory signal is initiated (Figure 1 and Table 2). Family members IL-36α, β and γ (formerly termed IL-1F6, IL-1F8 and IL-1F9) bind to the IL-1Rp2, recruit the same IL-1RAcP and a pro-inflammatory signal is initiated. IL-36Ra (formerly IL-1F5) also binds to the IL-1Rp2, but does not recruit IL-1RAcP and acts as the natural occurring receptor antagonist for IL-36α, β and γ. IL-33 binds to ST2, also recruits the IL-1RAcP, but Th2-like properties characterize IL-33. Thus, the six pro-inflammatory members of the IL-1 family each recruit the IL-1RAcP co-receptor with the TIR domain and MyD88 docks to each.
Active precursors. With the sole exception of IL-1Ra, each member of the IL-1 family is first synthesized as a precursor without a clear signal peptide for processing and secretion and none are found in the Golgi. IL-1α and IL-33 are similar in that their precursor forms can bind to their respective receptor and trigger signal transduction. Although recombinant mature forms of IL-1α and IL-33 are active, it remains unclear whether the mature forms are representative of the naturally occurring IL-1α and IL-33 produced in vivo. The importance of the biological activity of the IL-1α precursor is discussed below. The precursor forms of IL-18 and IL-1β do not bind their respective receptors, are not active and require cleavage by either intracellular caspase-1 or extracellular neutrophilic proteases (see below).

Dual function cytokines. IL-1α and IL-33 are “dual function” cytokines in that in addition to binding to their respective cell surface receptors, the intracellular precursor forms translocate to the nucleus and influence transcription. IL-33 is a bona fide transcription factor and the IL-1α N-terminal amino acids contain a nuclear localization site (reviewed in 7). In general, the nuclear function of IL-1α or IL-33 is transcription of proinflammatory genes. For example, expression of the N-terminal amino acids of IL-1α stimulates IL-8 production in the presence of complete blockade of the IL-1RI on the cell surface. IL-37, formerly IL-1F7, also translocates to the nucleus but in the case of IL-37, there is a net decrease in inflammation. As such, IL-37 is a unique ligand in the IL-1 family as it functions as an anti-inflammatory cytokine (see below).

IL-1 and Toll-like receptor (TLR) share similar functions. The IL-1 family of receptors is unique in its class of Ig-like receptors due to the presence of the TIR domain in cytoplasmic segment of each member (reviewed in O’Neill). The Toll protein was originally studied for its role in embryogenesis of the fruit fly Drosophila. After John Sims reported the cDNA of the IL-1RI, it was reported that the cytosolic segment of the IL-1RI shared a 50 amino acid homologous domain with the Toll protein in Drosophila. The significance of this observation was unclear until a few months later Heguy and co-workers demonstrated that the 50 amino acid domain in the IL-1RI was essential for IL-1 activities. Was the same domain in the Toll protein also functional for properties that were similar to IL-1 properties? It was not until 1996 that a role for the Toll protein in host defense of the fly was demonstrated. But in 1988 van der Meer had already reported that a low dose of IL-1 protected the host against live infections. Thus, the link of the Toll protein to IL-1 and to non-specific host defense against invading
microorganisms had already been made. In 1992, Charles Janeway replaced the term “non-specific resistance to infection” with “innate immune response”. Still later, the existence a mammalian cell surface receptor for microbial products was demonstrated and as reviewed by O’Neill, these receptors are now called Toll-like Receptors (TLR). All TLR contain share the TIR domain with the IL-1RI. With the new term “innate immunity” came the term “acquired immunity” and the concept that innate immunity affects acquired immunity. However, looking back, IL-1 is the best example of the non-specific nature of an “innate” cytokine. That innate immunity affects acquired immunity had already been reported in 1979, with the demonstration that human IL-1β enhanced antigen-specific responses of murine T-cells.

Humans have eleven TLRs. Each has a unique extracellular domain comprised of leucine rich repeats, which recognize a diverse number of bacterial products, nucleic acids and possibly some endogenous lipoproteins. Intracellularly, however, each TLR has a functional TIR domain as does the IL-1RI. Similar to IL-1 receptors, TLRs recruit the intracellular adaptor protein MyD88 to the TIR domain as the first step in signal transduction. The TIR domain is also present in two IL-1 family receptors that function to inhibit inflammation (see Figure 1): the single Ig IL-1 related receptor (SIGIRR) and the newly discovered IL-1R accessory protein, IL-1RAcP. The TIR of these inhibitory receptors have amino acid substitutions, which alter charge distribution of the TIR domain and prevent the recruitment of MyD88.

Since deletion of the TIR domain from IL-1RI prevented IL-1 signaling, the presence of a functional TIR domain in both IL-1 and Toll-like receptors suggested that microbial products such as endotoxin could be ligands for TLRs. However, it was not until 1998-99 that different groups reported microbial products as ligands for TLRs. The TIR domain is perhaps one of the most conserved sequences in both plants and animals. The near ubiquitous presence of the TIR domain reveals an unexpected duplication in evolution. Receptor signaling by the IL-1 family as well as for a large number of bacterial products and viral nucleic acids depend on the same TIR domain and therefore, the inflammatory manifestations are similar.

Are endogenous mammalian proteins ligands for TLRs? There is considerable specificity of each IL-1 family ligand for its respective receptor. There appears to be less specificity in the TLRs for the diversity of molecular structures in binding ligands, particularly for TLR2. For example, several products of Gram-positive bacterial and mycobacterial organisms are ligands...
for TLR2. However, there are reports demonstrating a reduction in disease severity with blocking TLR2 in models that are IL-1-dependent but independent of microbial products.\textsuperscript{19,20} A role for TLRs in such models raises the issue whether the extracellular segment of the TLRs recognize a wide variety of endogenous proteins. Although there is some evidence for this possibility particularly in TLR2, an alternative explanation for disease modification in mice deficient in specific TLRs may relate to a non-specific co-activation or recruitment of the TIR domain of a particular TLR during signal transduction of a specific cell surface or intracellular receptor. For example, during signal transduction following the binding of IL-1 to its receptor, the TIR domain on TLR2 recruits MyD88 and contributes to the activation of NFκB. This intracellular co-activation is often termed “cross-talk”.

**Limiting inflammation in the IL-1 family of cytokines and receptors.**

*Two receptor antagonists.* As listed in Table 2, IL-1Ra binds to IL-1RI and is specific for preventing the activity of IL-1α and IL-1β. As shown in Figure 1, IL-36α, IL-36β and IL-36γ each bind to IL-1Rrp2, recruit IL-1RAcP and initiate a pro-inflammatory signal. For example, IL-36α inhibits the differentiation of adipocytes and induces insulin resistance.\textsuperscript{21} However, IL-36Ra also binds to IL-1Rrp2, but similar to IL-1Ra, does not recruit IL-1RAcP. Therefore, IL-36Ra is the specific receptor antagonist for IL-1Rrp2 and prevents the activity of IL-36α, IL-36β and IL-36γ.\textsuperscript{22} In the skin, overexpression of IL-36α produces an inflammatory phenotype similar to that of psoriasis and is antagonized by IL-36Ra.\textsuperscript{22}

**IL-1 Receptor type II.** IL-1 receptor type II (IL-1RII) does not signal because it lacks a cytoplasmic domain and without a TIR domain, docking of MyD88 cannot take place (Figure 1). This receptor is expressed mostly on macrophages and B-cells. IL-1RII binds IL-1β with a greater affinity than that of IL-1RI, thereby sequestering IL-1β. Hence cell surface IL-1RII functions as decoy receptor for IL-1β.\textsuperscript{23} Since IL-1RAcP is recruited to the IL-1RII-IL-1β complex, the decoy receptor also serves to sequester the accessory receptor from participating in IL-1 signaling from the IL-1RI. The type II receptor is highly conserved and is found in bony fish where it functions to inhibit inflammation due to IL-1β. The soluble (extracellular) domain binds both mature as well as the IL-1β precursor. Although the soluble type II receptor
is an effective anti-inflammatory agent, for optimal neutralization of IL-1β, the soluble type II receptor requires the soluble form of the IL-1RAcP. A splice variant of this receptor codes for the extracellular domains and functions as an acute phase protein synthesized and released from the liver.

From a clinical perspective, expression of surface IL-1RII or circulating levels of soluble IL-1RII appear to affect the degree of inflammation in several diseases. Glucocorticoids induce expression of surface IL-1RII and this property may contribute to their anti-inflammatory effects. For instance, in patients with an acute loss in hearing due to an unknown autoimmune process, a robust induction of IL-1RII by dexamethasone on blood monocytes correlated with clinical response to oral prednisone. In contrast, cells from patients unresponsive to prednisone did not increase expression of the decoy receptor. The production of IL-1β appears to play a role in endometriosis and related infertility. Peritoneal fluid from fertile and infertile women with endometriosis had lower soluble IL-1RII and higher IL-1β concentrations than healthy controls and the increase in IL-1β was significant in women with endometriosis reporting pelvic pain. These and several clinical studies support the concept that like the balance of IL-1 to IL-1Ra, the amount of surface or soluble IL-1RII affects disease severity.

**SIGIRR.** The single IgG IL-1 related receptor (SIGIRR) (also known as TIR8) is a unique receptor in the IL-1 receptor family and functions to inhibit TLR- as well as cytokine-mediated signals. As shown in Figure 1, the extracellular domain of SIGIRR is comprised of a single IgG-like domain; the cytosolic portion does contain the TIR domain but the two amino acids required for a functional TIR domain are absent. In addition, the C-terminal contains an extra 95 amino acid extension, which may contribute to the anti-inflammatory properties of this receptor. Although a ligand for SIGIRR has not been identified, the single Ig-like domain is required for its inhibitory function. SIGIRR also forms a complex with the precursor form of IL-33, which may explain some of the anti-inflammatory properties of IL-33. For example, IL-33 reduces the development of atherosclerosis as well as cardiomyocyte hypertrophy. Intracellularly, SIGIRR reduces NFκB activation by members of the IL-1 family (IL-1, IL-18 and IL-33) as well as by the family of TLRs. Compared to wild-type mice, mice deficient in SIGIRR exhibit a severe inflammatory response to colitis and more severe responses to acute renal injury, which is due to a greater infiltration of myeloid cells into the renal parenchyma. Using the same model of acute renal injury, caspase-1
deficient mice are protected, most likely due to decreased IL-18 activity rather than decreased IL-1 effects.\textsuperscript{31} Thus SIGIRR may keep IL-18 activity in the kidney at bay.

\textit{IL-1RAcPb}. The IL-1RAcP serves as the co-receptor for IL-1\(\alpha\), IL-1\(\beta\), IL-36\(\alpha\), IL-36\(\beta\), IL-36\(\gamma\) and IL-33, each a pro-inflammatory cytokine. However, a unique form of the IL-1RAcP has been found in the brain where it serves to decrease IL-1 activities.\textsuperscript{18} Termed IL-1RAcPb, this co-receptor forms the expected complex with IL-1 and IL-1RI but does not recruit MyD88 or phosphorylate IRAK4. Therefore, most of IL-1 signaling is arrested. But since some genes increase by the IL-1RI/IL-1RAcPb complex, partial IL-1 signaling takes place. Nevertheless, IL-1RAcPb functions as an inhibitory receptor chain only in the brain. Mice deficient in IL-1RAcPb exhibit a normal inflammatory response in the periphery but greater neuro-degeneration in the brain. As such, IL-1RAcPb could play a role in chronic inflammatory responses in the brain by “buffering” IL-1-mediated neuro-degeneration.

Like SIGIRR, IL-1RAcPb contains the amino acids differences in its TIR domain that likely reduce binding of MyD88.\textsuperscript{18} In addition to an altered TIR domain, IL-1RAcPb has a carboxyl extension of 140 amino acids. Carboxyl extensions are also present in SIGIRR as well as two other members of the IL-1 receptor family, TIGIRR-1 and TIGIRR-2. TIGIRR-2 is associated with an X-linked cognitive deficiency, which is apparently independent of IL-1 functions. Little is known whether these C-terminal segments contribute to the inhibitory properties of these receptors.

\textit{A compelling case for the importance of the naturally occurring IL-1 receptor antagonist.} Rabbits passively immunized against their own endogenous IL-1Ra exhibit a worsening of colitis\textsuperscript{32} and mice deficient in IL-1Ra develop spontaneous diseases such as a destructive arthritis\textsuperscript{33}, an arteritis \textsuperscript{34} and a psoriatic-like skin eruption. Furthermore, mice deficient in endogenous IL-1Ra develop aggressive tumors following exposure to carcinogens.\textsuperscript{35} These data support the concept that the concentrations of IL-1\(\beta\) versus IL-1Ra affect the severity of some diseases. The single nucleotide polymorphism (SNP) rs4251961 C allele is associated with lower circulating levels of IL-1Ra (reviewed in \textsuperscript{36,37}) and is common in type 2 diabetes.\textsuperscript{37} The same polymorphism is associated with reduced survival in patients with colon carcinoma compared to those with the wild-type allele. The concept of an imbalance between IL-1\(\beta\) and IL-1Ra gained considerable legitimacy with reports of infants born with non-functional IL-1Ra.\textsuperscript{38,39} Soon after birth, the affected infants exhibited impressive systemic and local
inflammation. Multiple neutrophil-laden pustular skin eruptions, vasculitis, bone abnormalities with large numbers of osteoclasts, osteolytic lesions and sterile osteomyelitis were observed. The inflammation resembled infection with sepsis-like multiorgan failure, but all cultures were sterile. Treatment with anakinra was life-saving; the inflammation abated and the bone lesions disappeared. IL-17 was prominently expressed in cells from these patients (see below). These findings are an extreme example that without functional IL-1Ra, the activity of endogenous IL-1 is “unopposed” and IL-1-driven inflammation can run rampant.

One can conclude that low levels of IL-1β can induce inflammation but the presence of natural IL-1Ra is sufficient to limit the inflammation. IL-1Ra is found in the circulation of healthy subjects in the range of 100-300 ng/mL, whereas IL-1β is in the picogram per milliliter range and is not easily detectable by standard ELISA methods in the same individuals. In healthy humans, daily total constitutive production of IL-1β has been calculated at 6 ng; in patients with cryopyrin-associated periodic syndromes (CAPS), a rare auto-inflammatory diseases associated with increased IL-1β secretion, the calculated amount of IL-1β was 31 ng/day.40 These low amounts are consistent with reports of increased release of IL-1β from blood monocytes of patients with auto-inflammatory diseases. The increase is usually five to ten-fold more than that from blood monocytes of healthy subjects.41-43

IL-37, a fundamental inhibitor of innate immune responses. The function of interleukin-37 (IL-37; formerly IL-1 family member 7) has remained elusive. Studies with the recombinant form of the cytokine have been inconsistent due to aggregation, although some reports show that recombinant IL-37 binds to the IL-18Rα chain. Nevertheless, a role for natural occurring IL-37 in inflammation was demonstrated by suppressing endogenous IL-37 in human blood monocytes. In those studies, both LPS- and IL-1β-induced cytokine production increased significantly.12 Consistent with this observation, overexpression of IL-37 in macrophages or epithelial cells nearly completely suppressed production of pro-inflammatory cytokines. Mice with transgenic expression of IL-37 were protected from LPS-induced shock, and showed markedly improved lung and kidney function with reduced liver damage.12 IL-37 transgenic mice also exhibited lower concentrations of circulating and tissue cytokines (72-95% less) than wild-type mice and showed less dendritic cell activation. Previous studies demonstrated that IL-37 translocates to the nucleus via a caspase-1-dependent fashion.11 IL-37 interacts intracellularly with Smad3 and IL-37-expressing cells and transgenic mice showed less
cytokine suppression when endogenous Smad3 was depleted.\textsuperscript{12} It appears that IL-37 co-localizes to the nucleus with Smad3 and suppresses transcription of inflammatory genes induced by microbial products as well as cytokines themselves. Thus, IL-37 emerges as a natural suppressor of innate inflammatory and immune responses. Whether disease severity in humans is affected by the level of IL-37 expression remains to be determined.

**IL-1β and the spectrum of auto-inflammatory diseases**

*The distinction between auto-inflammatory and autoimmune diseases.* Nearly all autoimmune diseases have an inflammatory component. However, auto-inflammatory diseases appear to be primarily inflammatory, with systemic as well as local manifestations that are often periodic rather than progressive. Autoimmune diseases are primarily caused by dysregulation of adaptive immune responses and due to pathologic antibodies or auto-reactive T-cells. In contrast, some classic auto-inflammatory diseases are caused by genetic defects in innate inflammatory pathways and usually are manifested early in life. Typically, auto-inflammatory diseases have no associations with HLA or MHC Class II haplotypes and there is absence of auto-reactive T-cells or auto-antibodies. However, some complex systemic inflammatory diseases are driven by auto-inflammatory mechanisms but adaptive pathways also seem to play a role. For example, in Behçet’s disease, there is a strong association with HLA-B51, which likely contributes an autoimmune component of the disease.

Familial Mediterranean Fever, Familial Cold-induced Autoinflammatory Syndrome Cryopyrin-Associated Periodic Syndrome (CAPS) are classic examples of auto-inflammatory diseases.\textsuperscript{6,44} TNF Receptor Associated Periodic Syndrome (TRAPS) was the first disease to be labeled as auto-inflammatory. As shown in Table 1, there is a growing number of diseases that fall under the category of being primarily auto-inflammatory. One of the most consistent characteristics of these auto-inflammatory diseases is a rapid and sustained resolution of disease severity upon treatment with IL-1β blockade, although there are exceptions. In the case of systemic onset juvenile idiopathic arthritis, there are often but not always dramatic responses to IL-1β blockade.\textsuperscript{41,45} Although antibodies to the IL-6 receptor are also effective in this disease, a reduction in IL-1β activity likely reduces IL-6 production. Auto-inflammatory diseases are weakly responsive to TNFα neutralization and in some reports, disease worsens with TNFα.
blockade. Whereas autoimmune diseases respond to anti-TNFα, CTLA4-Ig, anti-IL-6 receptor, depletion of CD20 B-cells, depletion of CD3 T-cells, anti-IL-17, or anti-IL-12/IL-23 antibodies, these agents are often ineffective in patients with auto-inflammatory diseases. For example, in the case of Schnitzler’s syndrome with a monoclonal gammopathy, depletion of B-cells resulted in a dramatic reduction in the paraprotein but was of no benefit on systemic symptoms. In contrast, anakinra resulted in a rapid improvement sometimes within hours and a complete remission in disease activity within days. Table 1 lists auto-inflammatory diseases that for the most part respond to blocking IL-1 activity.

**Clinical characteristics of systemic auto-inflammatory diseases.** Systemic auto-inflammatory diseases are syndromes. As listed in Table 1, although many are rare, the clinical manifestations are common. Fever, neutrophilia and elevated hepatic acute phase proteins are characteristically present with painful joint, myalgias and debilitating fatigue. These manifestations are consistent with the responses observed in humans following administration of intravenous IL-1; fever, neutrophilia and elevated IL-6 and ACTH levels were observed at doses of IL-1α or IL-1β as low as 1-3 ng/kg.1

CAPS patients can present with a spectrum of disease severity. In the mildest form, Familial Cold-induced Autoinflammatory Syndrome, the episodes are triggered by cold exposure. A more severe form is Muckle-Wells Syndrome in which the inflammation is persistent and apparently not related to cold but associated with the development of hearing loss and amyloidosis. The most severe form of CAPS is Neonatal-Onset Multisystem Inflammatory Disease, which in addition to systemic inflammation is also associated with joint deformities and developmental disabilities in early childhood. Often, upon blocking IL-1β, patients with CAPS and other auto-inflammatory diseases experience a rapid and sustained cessation of symptoms as well as reductions in biochemical, hematological and functional markers of their disease. Since treatment with the soluble IL-1RI rilonacept or a monoclonal anti-IL-1β antibody (canakinumab) are equally effective in treating auto-inflammatory diseases, the culprit in these diseases is IL-1β and not IL-1α.
Caspase-1 dependent and independent processing of the IL-1β precursor into an active cytokine.

*Increased release of IL-1β in auto-inflammatory diseases.* Fundamental to many auto-inflammatory diseases is an increased release of active IL-1β. Caspase-1 is the intracellular cysteine protease that cleaves the N-terminal 116 amino acids from the IL-1β precursor thus converting the inactive precursor to the active “mature” cytokine. Caspase-1 exists in tissue macrophages and dendritic cells as an inactive zymogen and requires conversion to an active enzyme by autocatalysis. However, in circulating human blood monocytes, caspase-1 is present in an active state. Caspase-1 is also constitutively active in highly metastatic human melanoma cells. In general, the release of active IL-1β from blood monocytes is tightly controlled with less than 20% of the total IL-1β precursor being processed and released. Although the release of active IL-1β from blood monocytes of healthy subjects takes place over several hours, the process can be accelerated by increased levels of extracellular ATP, which triggers the P2X7 purinergic receptor (Figure 2).

*Activation of caspase-1.* As shown in Figure 2, following ATP binding to the P2X7 receptor, there is a rapid exit of potassium from the cell and intracellular potassium falls. With inhibition of ATP binding or in cells deficient in the P2X7 receptor, the amount of secreted IL-1β is low or absent. The fall in intracellular potassium is thought to bring about the oligomerization of a highly specialize group of intracellular proteins termed the “inflammasome”, which convert procaspase-1 to an active enzyme. Active caspase-1 then cleaves the IL-1β precursor in specialize secretory lysosomes or in the cytosol, followed by secretion of “mature” IL-1β (Figure 2). Blood monocytes from patients with auto-inflammatory syndromes release more processed IL-1β than cells for healthy subjects and thus likely accounts for the inflammation in these diseases. An increase in the secretion of active IL-1β is observed in monocytes from patients with a gain of function mutation in a gene originally called Cold-Induced Auto-inflammatory Syndrome-1. The mutations in this gene result in a single amino acid change in the intracellular protein named cryopyrin, since following the exposure to cold, the patients develop fever and other manifestations of systemic inflammation. Cryopyrin...
is now termed nucleotide-binding domain and leucine-rich repeat containing protein 3 (NLRP3).

The gene Cold-Induced Auto-inflammatory Syndrome-1 was first reported by Hoffman in 2001\textsuperscript{57} and a historical perspective of this important discovery has been published.\textsuperscript{7} In 2004, NLRP3 was reported to associate with procaspase-1 and other intracellular proteins to form complex, which was termed the “inflammasome”.\textsuperscript{56} As shown in Figure 2, these components activate procaspase-1, resulting in active caspase-1 followed by the processing and secretion of active IL-1\textbeta. However, about half the patients with classic symptoms and biochemical markers of CAPS, FMF and other auto-inflammatory diseases do not have mutations and the basis for increased secretion of IL-1\textbeta remains unknown. As noted above, the increase in IL-1\textbeta secretion in monocytes from patients with auto-inflammatory diseases is modest compared to that of monocytes from healthy subjects. Despite severe systemic inflammation, only a five-fold more IL-1\textbeta is produced each day in patients with CAPS compared to healthy controls.\textsuperscript{40}

\textit{Secretion of IL-1\textbeta}. As shown in Figure 2, multiple mechanisms have been reported for the secretion of the processed mature IL-1\textbeta; these include the loss of membrane integrity, a requirement for phospholipase C and secretory lysosomes\textsuperscript{53}, the shedding of plasma membrane microvesicles or multivesicular bodies containing exosomes. In general, the release of active IL-1\textbeta takes place before there is significant release of lactate dehydrogenase, although some report that the release of IL-1\textbeta parallels that of lactate dehydrogenase. Pyroptosis is a caspase-1-dependent mechanism for cell death with the release of IL-1\textbeta\textsuperscript{58} and may account for the release of active IL-1\textbeta in ischemic disease.\textsuperscript{58}

Since caspase-1 is present in an active form in freshly obtained blood monocytes from healthy subjects,\textsuperscript{51} the rate-limiting step in the secretion of IL-1\textbeta is at the transcriptional and translational level. But in monocytes from patients with single amino acid mutation in NLRP3, the release of active IL-1\textbeta occurs without a requirement for a rapid fall in the level of intracellular potassium.\textsuperscript{43} Although often studied using endotoxin-induced synthesis of the IL-1\textbeta precursor, inflammation in auto-inflammatory diseases is sterile. Continued treatment with anakinra results in a progressive reduction in the secretion of IL-1\textbeta from monocytes compared to secretion before treatment.\textsuperscript{42,43} A reduction in steady-state caspase-1 gene
expression also takes place.\textsuperscript{42} Thus, IL-1 itself increases the synthesis of caspase-1 as well as its own IL-1\(\beta\) precursor\textsuperscript{59} and may account for the auto-inflammation of these diseases.

\textit{Caspase-1 independent processing of the IL-1\(\beta\) precursor into an active cytokine.} In the arthritic joint and following blunt trauma, hypoxia, hemorrhage or exposure to irritants, there is a brisk myeloid response into the affected tissues dominated by neutrophils. Although IL-1\(\beta\) often plays a pivotal role in these conditions, caspase-1 may not be required. For example, IL-1\(\beta\) is required for irritant-induced inflammation in muscle tissue, but is caspase-1 independent.\textsuperscript{60} Similarly, cartilage destruction in joints\textsuperscript{61} and urate crystal-induced inflammation \textsuperscript{62} are also IL-1\(\beta\)-dependent, but caspase-1 independent. In many models of sterile inflammation, cell death and release of the IL-1\(\beta\) precursor from resident macrophages or infiltrating monocytes takes place. With the infiltrating neutrophils, there is also a role for neutrophil proteases. As shown in Figure 3, these proteases include elastase, chymases, granzyme A, cathepsin G and proteinase-3. In particular, proteinase-3 cleaves the inactive IL-1\(\beta\) precursor close to the caspase-1 site resulting in active IL-1\(\beta\).\textsuperscript{60,61} Moreover, inhibition of neutrophil proteases prevented urate crystal-induced peritonitis.\textsuperscript{62} These findings question whether caspase-1 is responsible for IL-1\(\beta\)-dependent inflammation in patients with gout or any inflammatory process where there is a robust infiltration of neutrophils in the release of the IL-1\(\beta\) precursor into the extracellular space.

\textit{Role of reactive oxygen species on caspase-1-dependent secretion of IL-1\(\beta\).} Inflammation and carcinogenesis are associated with the generation of reactive oxygen species (ROS) and ROS have been proposed to activate the caspase-1 inflammasome.\textsuperscript{63} Since IL-1\(\beta\) appears to play a role in carcinogenesis\textsuperscript{35}, it is an attractive hypothesis that ROS directly activates the inflammasome to induce release of IL-1\(\beta\) and trigger inflammation. However, in cells deficient in deficient in gp91phox ROS, levels are low and IL-1\(\beta\) levels are high and in cells deficient in super oxide dismutase-1, ROS levels are high but caspase-1 is suppressed and IL-1\(\beta\) levels are low.\textsuperscript{64}

Comparable to murine cells deficient in gp91phox, humans with chronic granulomatous disease (CGD) have a mutation in p47-phox, with defective NADPH activity and are unable to generate ROS. Blood monocytes from these patients release IL-1\(\beta\) following stimulation in vitro.\textsuperscript{65,66} Although the well-known inhibitor of ROS, diphenylene iodonium (DPI), reduces the
secretion of IL-1β in all studies, DPI inhibits IL-1β and also TNFα gene expression rather than caspase-1 activity. Thus, it appears that ROS may dampen the inflammasome and this may explain why CGD patients, who are unable to generate ROS, have increased inflammation with granulomatous lesions and a form of colitis indistinguishable from that found in Crohn’s disease. It has also been shown that ROS induces intracellular NAC, a potent anti-oxidant, which facilitates the secretion of active IL-1β. Although ROS generation from mitochondria may contribute to caspase-1 activation, there is the conundrum that regardless of the source of ROS, the concept is not supported by decades of failed placebo controlled anti-oxidant trials.

Role of IL-1α in ischemia-induced inflammation. Cells of mesenchymal origin constitutively contain preformed IL-1α precursor, for example, keratinocytes, endothelial cells and epithelial cells of the lung, kidney and gastrointestinal tract. If released from its intracellular pool in healthy cells, the IL-1α precursor is inflammatory. Thus it was not unexpected that a robust inflammatory responses were observed when extracts of cellular contents from healthy tissues containing the IL-1α precursor upon injection into the peritoneal cavity of mice. Importantly, inflammation was shown to be MyD88 and IL-1RI dependent but TLR independent. Intracellular IL-1α is found diffusely distributed in the cytosol as well as in the nucleus where it associates with chromatin. The N-terminal domain of the IL-1α precursor contains a nuclear localization sequence, which actively participates in transcription and affects the senescence of cells.

As shown in Figure 4, during the hypoxia of an ischemic event, there is loss of membrane integrity in dying cells with the release of intracellular contents, which include the IL-1α precursor. Subsequently, resident tissue macrophages respond to IL-1α and produce IL-1β (Figure 4). Although ischemia-induced inflammation is often initiated by the IL-1α precursor, it rapidly becomes caspase-1 and IL-1β dependent. Adenoviruses infection induces a neutrophil infiltration which is independent of TLR9 and NLRP3 and the inflammation is due to IL-1α activation of the IL-1RI with a brisk production of chemokines. Cell death by a caspase-1-dependent process called pyroptosis may also take place in ischemic tissues with in the release of cell contents.

In contrast, the same mesenchymally-derived cells undergo apoptosis as part of normal tissue renewal. During apoptosis, intracellular IL-1α concentrates in dense nuclear foci and
chromatin binding is not reversed by histone deacetylases or affected by inhibitors of reactive oxygen species.\textsuperscript{70} As illustrated in Figure 4, in apoptotic cells, IL-1\(\alpha\) is retained within the chromatin fraction and is not present in the cytoplasmic contents. As such, lysates of cells undergoing apoptotic death are biologically inactive.\textsuperscript{70} Thus, nuclear trafficking and differential release during necrosis versus apoptosis (Figure 4) demonstrates that inflammation by IL-1\(\alpha\) is tightly controlled.

\textit{IL-1\(\beta\), periodontitis and bone loss.} Epidemiologic studies have examined polymorphisms in the IL-1 gene cluster (IL-1\(\alpha\), IL-1\(\beta\) and IL-1Ra) with increased periodontitis and tooth loss as well as with coronary heart disease (CHD) and type 2 diabetes. In a study in Germany, over 1,500 subjects with increased levels of HbA1c were found to have significant periodontal disease compared to normoglycemic subjects. Diabetics with IL-1\(\beta\) promoter genotype C/T or T/T exhibited a more severe form of periodontitis compared to wild-type IL-1\(\beta\) counterparts suggesting that carrying these polymorphisms increases the inflammation. In another study, mean alveolar bone levels were measured in patients with CHD. The same polymorphisms were associated with bone loss. An IL-1Ra polymorphism was also associated with both CHD and the highest level of bone loss. In patients with the IL-1\(\alpha\) +4845T polymorphism, there was a statistically significant correlation with acute coronary syndrome and severe periodontitis.

The mechanism of IL-1\(\beta\) production in periodontitis has been extensively studied using bacteria isolated from patients with long-standing disease. The most persistent bacterial infection in periodontitis is \textit{A. actinomyctemcomitans}, which infects the periodontal space in many throughout the world and is the most common cause of tooth loss.\textsuperscript{74} These organisms produce a leukotoxin, which induces degranulation and lysis in human neutrophils. The leukotoxin also activates caspase-1 and the secretion of large amounts of IL-1\(\beta\) from human macrophages.\textsuperscript{74} IL-1\(\beta\) is known a player in bone loss in patients with rheumatoid arthritis and the amino acid sequence of purified osteoclast activating factor is identical to that of IL-1\(\beta\). Thus, the induction of IL-1\(\beta\) in the periodontal space by the leukotoxin likely explains alveolar bone tooth loss with periodontitis. Since the inflammation in periodontitis appears to be IL-1\(\beta\) mediated, IL-1\(\beta\) entering the circulation from the local inflammation in the periodontal space
may affect the distant insulin-producing beta-cell in the pancreatic islet and account for the association of periodontitis with type 2 diabetes.

**IL-1 in B- and T-cell Responses**

*IL-1β as an adjuvant.* A role for IL-1β in antibody production has been repeatedly reported (reviewed in). Not unexpectedly, the most widely used adjuvant, aluminum hydroxide (alum), induces IL-1β via a caspase-1 dependent mechanism. Previous studies revealed that mice deficient in IL-1β do not produce anti-sheep red blood cell antibodies, a T-dependent response. In wild-type mice, IL-1β causes a marked increase in the expansion of naive and memory CD4 T cells in response to antigen and particularly when used with LPS as a co-stimulant. In fact, when LPS is used as an adjuvant approximately 55% of the antibody response is due to IL-1. There are also striking increases in serum IgE and IgG1 levels. The role of IL-1β on the expansion and differentiation of CD4 T-cells is independent of IL-6 or CD28 but rather due to IL-17- and IL-4-producing cells. In human T-cells, Th17 polarization induced by *M. tuberculosis* is IL-1β dependent via dectin-1 and TLR4 stimulation. This observation has relevance to models of experimental autoimmune encephalomyelitis (EAE) and collagen-induced arthritis, both of which require the adjuvant complete Freund’s, a mixture of killed *M. tuberculosis* and the irritant croton oil. These results indicate that IL-1β signaling in T cells induces durable primary and secondary CD4 responses, most likely due to the induction of IL-17.

*A role for IL-1 in the generation of Th17 polarization.* Increasing clinical studies indicate that T-cell differentiation into IL-17 producing cells plays a major role in some autoimmune diseases and IL-1 appears to play a pivotal role in the polarization of T-cells towards Th17. The role of IL-1β in the generation of Th17 cells is also consistent with the inflammation in mice deficient in IL-1Ra. In mice deficient in IL-1Ra, the activity of endogenous IL-1 is unopposed and spontaneous rheumatoid arthritis-like disease develops, but not in mice also deficient in IL-17. In mice specifically deficient in endogenous IL-1Ra in myeloid cells, collagen-induced arthritis appears to be due to unopposed IL-1 activity, which drives IL-17. IL-1-dependent IL-17 production may be relevant to EAE, the rodent model for multiple
sclerosis. Mice deficient in both IL-1α and IL-1β do not develop EAE, which is thought to be a failure to mount a Th17 response. In mice deficient in caspase-1, EAE is markedly attenuated. Thus, IL-1 rather than TNFα blockade reduces disease severity in mice subjected to EAE; in fact, blocking TNFα worsens the outcome of the EAE model. In rheumatoid arthritis, depleting CD20 bearing B-cells results in decreased IL-17 production, which may be due to a reduction in B-cell IL-1β.

Although there is no dearth of reports that adding IL-1 to cultures induces IL-17, an essential role for IL-1 was shown when mice deficient in the IL-1R fail to induce IL-17 upon antigen challenge. Moreover, IL-23 fails to sustain IL-17 in IL-1R deficient T-cells and TNFα and IL-6 enhancement of IL-23-induced IL-17 is also IL-1 dependent. In a subsequent study of EAE, γδ CD4 positive T-cells were the source of IL-17 but independent of T-cell receptor engagement. γδ CD4 positive T-cells contain the RORγ transcription factor and thus a direct induction of IL-17 by IL-1 is fundamental to the Th17 paradigm. Therefore, it is likely that there is a cascade of IL-1β-induced IL-23 as well as IL-1-induced IL-6 for Th17 differentiation.

In human T-cells, IL-1β dependent Th17 differentiation is due to the intermediate production of PGE2 from macrophages. This observation is consistent with ability of IL-1β to induce COX-2 (reviewed in) and PGE2 to induce IL-6. Thus, an innate inflammatory response that is IL-1β-driven appears to play a pivotal role in the outcome of EAE. In order to maintain Th1-driven responses, IFNγ suppresses the differentiation into the Th17 phenotype. One possible mechanism by which IFNγ suppresses Th17 is via the suppression of IL-1. IFNγ reduces the induction of IL-1 by IL-1 and also of IL-1-induced PGE2 (reviewed in Dinarello). Thus, IFNγ suppression of IL-1-driven PGE2 as well as IL-1 itself may explain the reduction in Th17 differentiation.

**Blocking IL-1β-mediated Disease**

*Therapeutic agents.* Initially, anakinra was used to treat several unrelated chronic inflammatory diseases as listed in Table 1. Today, these diseases are also successfully treated with neutralization by human anti-IL-1β monoclonal antibodies. For example in type 2
diabetes, either anakinra or monoclonal antibodies to IL-1β improve glycemic control. At this writing, one anti-IL-1β antibody (canakinumab) has been approved for treating CAPS, whereas others are presently in clinical trials. Rilonacept is a construct of the two extracellular chains of the IL-1 receptor complex (IL-1RI plus IL-1RAcP) fused to the Fc segment of IgG and has also been approved for use in CAPS.

Targeting IL-1β in smoldering/indolent myeloma. MGUS and smoldering myeloma present a challenge to medicine as the population ages. Several years of research has focused on the role of IL-1β and IL-6 in the pathogenesis of multiple myeloma. Similar to mature B-cells, the myeloma plasma cell produces IL-1β. In the microenvironment of the bone marrow, stromal cells respond to low concentrations IL-1β and release large amounts of IL-6, which in turn promotes the survival and expansion of the myeloma cells. Although IL-6 is an essential growth factor for myeloma cells, antibodies to IL-6 have not been effective in treating the disease. Lust, Donovan and co-workers reasoned that in the indolent stages of multiple myeloma, blocking IL-1β would provide better reduction of IL-6 activity. Bone marrow cells from patients with smoldering myeloma were co-cultured with a myeloma cell line actively secreting IL-1β. Although the addition of dexamethasone reduced stromal cell IL-6 production, the amount of IL-6 remained sufficiently high enough to protect the plasma cell against dexamethasone-mediated apoptosis. However, anakinra added to these co-cultures significantly reduced IL-6 by nearly 90% and the combination of anakinra plus dexamethasone induced myeloma cell death.

Patients with smoldering or indolent myeloma were selected with the clinical objective of slowing or preventing progression to active disease. Based on in vitro data, 47 patients with smoldering/indolent myeloma at high risk for progression to multiple myeloma were treated with daily anakinra for six months. During the 6 months, there were decreases in CRP in most but not all patients, which paralleled a decrease in the plasma cell labeling index, a measure of myeloma cell proliferation in unfractionated bone marrow cells. After 6 months of anakinra, a low dose of dexamethasone (20 mg per week) was added. Of the 47 patients that received anakinra (25 with dexamethasone), progression-free disease was over three years and in 8 patients over 4 years. Compared to historical experience, the findings indicate a significant failure to progress to active disease and a modest fall in CRP predicted responders who continued with stable disease. Patients with a decrease in serum CRP of 15% or greater after 6
months of anakinra monotherapy resulted in progression-free disease greater than 3 years compared to 6 months in patients with less than a 15% fall during anakinra therapy (p<0.002). Thus, an effective reduction in IL-1β activity using CRP as the marker for IL-1β-induced IL-6 halts progression to active myeloma. With the use of anakinra in over 200,000 patients with rheumatoid arthritis or auto-inflammatory diseases, no opportunistic infections, including *Mycobacterium tuberculosis* reactivation, have been reported.

**Angiogenesis.** Angiogenesis plays a pathological role in multiple myeloma. In vitro and in animal models, there are a growing number of reports on IL-1β as a key cytokine in angiogenesis. Approved by the NIH, an ongoing trial of anakinra as an anti-angiogenic therapy in patients with cutaneous melanoma. Because blocking IL-1β reduces IL-6 as well as the pro-angiogenic chemokine IL-8, the use of IL-1β blocking strategies may result in therapy in high risk patients with smoldering/indolent myeloma or metastatic melanoma. There is also a role for IL-1β in the angiogenic process of macula degeneration and anakinra treatment in rheumatoid arthritis reduces the vascularization of the pannus.

**Role for IL-1 in graft-versus-host disease.** Reducing cytokine activity in the development of graft-versus-host disease (GvHD) is based on pre-clinical studies. Of the three phases of GvHD, immunization, proliferation and targeting of tissue, IL-1 is least important in the primary phases of the disease, in part, because IL-1 is not dominant in the Th1 paradigm of T-cell and natural killer cell activation compared to IL-2, IL-15 and IL-18. In a placebo-controlled trial, patients were treated with anakinra four days prior to and ten days following allogeneic stem cell transplantation added to standards of therapy. Moderate to severe GvHD developed in both anakinra and placebo-treated patients. One can conclude that during conditioning and 10 days after transplantation, IL-1 plays no significant role in the development of the disease. In contrast, in patients with full-blown GvHD, blocking IL-1 reduced the severity of the disease. Using a continuous intravenous infusion of anakinra, there was improvement in the clinical score in 16 of 17 treated patients. Moreover, a decrease in the steady state mRNA for TNF-α in peripheral blood mononuclear cells correlated with improvement (p=0.001). It is likely that there is role for IL-1 in the inflammatory manifestations of GvHD rather than the immunological development of the disease.
Treating gout with IL-1β neutralization. There is a long association of IL-1 production with urate crystals and a causal relationship of IL-1 with gout. Some patients with recurrent attacks of gouty arthritis resistant to colchicine and other standards of therapy often require steroids to control disease flares. When treated with anakinra, rilonacept or canakinumab, a rapid, sustained and remarkable reduction in pain as well as objective signs of reduced inflammation have been observed. The effect of IL-1 blockade appears to be superior to that of steroids and result in prolonged periods without flares. Although some studies report that urate crystals directly activate the NLRP3 for processing of the IL-1β precursor, it is likely that two signals are required; urate crystals plus free fatty acids account for IL-1β activity in flares of gout. Given the characteristic neutrophilic infiltration in gouty joints, it is also likely that the IL-1β precursor is processed extracellularly by neutrophilic enzymes as shown in Figure 3.

Effects of IL-1 receptor blockade in stroke. Although infection, particularly septic shock, evokes a “cytokine storm” as part of a systemic inflammatory response, in many ischemic diseases such as acute lung injury, thrombotic stroke, acute renal failure, myocardial infarction and hepatic failure, inflammation is characterized by cell death and infiltration of neutrophils into the ischemic area. The induction of IL-1β in human monocytes exposed to hypoxia was first reported in 1991. Subsequently, many studies have demonstrated an essential role for IL-1 and specifically for IL-1β and caspase-1 in various models of ischemic injury without and with reperfusion. A randomized, double blind, placebo controlled trial of anakinra was carried-out in patients with acute stroke. Three days of a high dose of intravenous anakinra was administered, the same dose used in the sepsis trials. Peripheral white blood cells, neutrophil counts, CRP and IL-6 concentrations were lower in anakinra treated compared to placebo-treated patients. After 3 months, the anakinra-treated patients exhibited less loss of cognitive function compared to the placebo group.

Reducing IL-1β in type-2 diabetes. For over 25 years, the cytotoxic effects of IL-1β for the insulin-producing pancreatic beta cell have been studied. Subsequently, it was shown that high concentrations of glucose stimulated IL-1β production from the beta-cell itself implicating a role for IL-1β in type 2 diabetes. In addition, free fatty acids act together with glucose to stimulate IL-1β from the beta cell. The production of IL-1β also increases to the
deposition of amyloid via activation of NLRP3. In support of these studies, gene expression for IL-1β was over one hundred-fold higher in beta cells from type 2 patients compared to non-diabetic patients. The clinical proof of a role for IL-1β in the pathogenesis of type 2 diabetes came from a randomized, placebo controlled study of anakinra for 13 weeks, in which improved insulin production and glycemic control associated with decreased CRP and IL-6 levels was reported. In the 39 weeks following the 13 week course of anakinra, patients who responded to anakinra used 66% less insulin to obtain the same glycemic control compared to baseline requirements. This observation suggests that blocking IL-1β even for a short period of time restores the function of the beta cells or possibly allows for partial regeneration of beta cells.

Several trials blocking TNFα in type-2 diabetes have succeeded in reducing CRP but did not improve glycemic control. The findings of anakinra therapy in type 2 diabetic patients have been confirmed using canakinumab as well as another neutralizing monoclonal antibody to IL-1β. The data also provides evidence that short-term blockade of IL-1β restores the function of the beta cells or possibly regeneration.

The above studies are consistent with the concept that type-2 diabetes is a chronic IL-1-mediated disease and could be classified as an auto-inflammatory disease in which IL-1β-mediated inflammation progressively destroys the insulin-producing beta cells. The IL-1β can come from the beta-cell itself, from infiltrating macrophages into the islet or from the adipose tissue stores. A requirement for caspase-1 in regulating IL-1β production from the insulin-producing beta cell as well as the adipocyte provides a molecular mechanism. Differentiation of the adipocyte is caspase-1 dependent. In diabetic mice, administration of a caspase-1 inhibitor reduces insulin resistance; in mice deficient in caspase-1, there is improved sensitivity to insulin. Because type 2 diabetes increases risk of cardiovascular events, blocking IL-1β activity in these patients may also reduce the incidence in myocardial infarction and stroke as reviewed in Dinarello.

IL-1β as a mediator of cartilage destruction in osteoarthritis. In the joint, IL-1β is the mediator of reduced chondrocyte proteoglycan synthesis, increased synthesis of matrix metalloproteinases and the release of nitric oxide. Mice deficient in IL-1β are protected from inflammation-induced arthritis. The role of IL-1β in the destructive processes of osteoarthritis has also been studied in rabbits, pigs, dogs and horses. In a placebo-controlled trial of
intraarticular anakinra in patients with knee osteoarthritis, there was a clear dose-dependent
(50 mg versus 150 mg) reduction in pain and stiffness scores, but the benefit did not extend
beyond one month. The modest reduction may be due to the heterogeneity of the
osteoarthritis population in general but also to the short duration of IL-1RI blockade by
anakinra.

Does IL-1 contribute to heart failure after myocardial infarction? ST elevation
myocardial infarction (STEMI) has a high risk of death but patients who survive the acute event
progress to chronic heart failure due to loss of viable myocardium. In a randomized, placebo-
controlled trial of patients with ST elevation myocardial infarction, daily anakinra was added to
standard of therapy the day after angioplasty and continued for 14 days. Serial imaging and
echocardiographic studies were performed over the following 10-14 weeks. Left ventricular
remodeling was significantly reduced with anakinra treatment and consistent with the
reductions in CRP compared to patients receiving 14 days of placebo. After 18 months, 60% of
placebo-treated patients had developed stage IV heart failure whereas there were none in the
anakinra treated patients. These findings are consistent with experimental myocardial infarction
in mice where blocking IL-1β results is a reduction in post-infarction remodeling. It remains
to be tested whether chronic heart failure in non-post myocardial infarction patients is reduced
by IL-1β blockade.

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References


Table 1  Blocking IL-1β in Treatment of Acute and Chronic Inflammatory Diseases

**Classic Auto-inflammatory Diseases**
- familial Mediterranean fever (FMF)
- pyogenic arthritis, pyoderma gangrenosum, acne (PAPA)\(^a,b\)
- cryopyrin associated periodic syndromes (CAPS)
- hyper IgD syndrome (HIDS)
- adult and juvenile Still’s Disease
- Schnitzler’s syndrome
- TNF receptor-associated periodic syndrome (TRAPS)
- Blau’s Syndrome; Sweet’s Syndrome
- deficiency in IL-1 receptor antagonist (DIRA)

**Probable Autoinflammatory Diseases**
- Recurrent idiopathic pericarditis
- macrophage activation syndrome (MAS)
- urticarial vasculitis
- anti-synthetase syndrome
- relapsing chondritis
- Behçet’s disease
- Erdheim-Chester syndrome (histiocytosis)
- synovitis, acne, pustulosis, hyperostosis, osteitis (SAPHO)

**Common Diseases Mediated by IL-1β**
- rheumatoid arthritis\(^c\)
- periodic fever, aphthous stomatitis, pharyngitis, adenitidis syndrome (PFAPA)
- urate crystal arthritis (gout)
- type 2 diabetes
- smoldering multiple myeloma
- post-myocardial infarction heart failure
- osteoarthritis

\(^1\)See Supplement for complete list of citations
\(^a\) Incomplete responses to IL-1β blockade have been reported.
\(^b\) The combination of IL-1 plus TNFα have been used in some disorders.
\(^c\) Rheumatoid arthritis is also treatable with other anti-cytokines, anti-receptors, immunomodulating agents and B-cell depleting antibodies
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Figure 1. Signaling and inhibition of signaling by IL-1 receptors. A. IL-1α (either precursor or mature) or mature IL-1β bind to the IL-1RI and with the IL-1RAcP forms the receptor heterodimeric complex. The TIR domain of each receptor chain approximate, recruit MyD88, followed by phosphorylation of IRAKs and IKKβ resulting in a signal to the nucleus. Details on IL-1 signal transduction have been reviewed.8  B. In the brain and spinal cord, the variant IL-1RAcPb can form the heterodimeric complex with IL-1α or IL-1β and IL-1RI, but this complex fails to recruit MyD88 and there is inhibition of the IL-1 signal. The failure to recruit MyD88 may be due to an altered TIR domain (indicated as TIRb).  C. IL-1Ra binds to IL-1RI but there is no signal since there is failure to form a complex with IL-1RAcP.  D. IL-1β binds to the IL-1RII, but lacking a cytoplasmic segment, there is no signal.  E. Due to an altered TIR domain (indicated as TIRb), SIGIRR inhibits IL-1 and TLR signaling. SIGIRR can form a complex with IL-33 (not shown) and inhibit IL-33 signaling.29  F. IL-1Rrp2 binds IL-36α, IL-36β or IL-36γ and forms a complex with IL-1RAcP. The TIR domain of each receptor chain approximates and recruits MyD88 similar to that shown in A.  G. IL-36 Receptor antagonist (IL-36Rα) binds to IL-1Rrp2 but fails to form a complex with IL-1R AcP. Thus, IL-36Ra prevents the binding of IL-36α, IL-36β or IL-36γ to IL-1Rrp2 and IL-36Ra is the natural receptor antagonist IL-36.
Figure 2. Steps in the processing and release of IL-1 induced by IL-1. 1. Primary blood monocytes, tissue macrophages or dendritic cells are activated by either mature IL-1β or the IL-1α precursor with the formation of the IL-1 receptor complex heterodimer comprised of the IL-1RI with IL-1RAcP. 2. Approximation of the intracellular TIR domains. 3. Recruitment of MyD88 and phosphorylation of IRAKs and IKKβ. 4. Transcription of IL-1β. 5. Translation into the IL-1β mRNA takes place on polysomes. IL-1β mRNA is not bound to actin microfilaments but rather intermediate filaments. 6. ATP released from the activated monocyte/macrophage accumulates extracellularly. 7. Activation of the P2X7 receptor by ATP. 8. Efflux of potassium from the cell following ATP binding to P2X7 receptor. 9. Fall in intracellular levels of potassium. 10. The fall in intracellular potassium levels triggers the assembly of the components of the caspase-1 inflammasome with the conversion of procaspase-1 to active caspase-1. 11. Caspase-1 is found in the secretory lysosome together with the IL-1β precursor and lysosomal enzymes. Active caspase-1 cleaves the IL-1β precursor in the secretory lysosome, generating the active, carboxyl-terminal mature IL-1β. 12. An influx of calcium with an increase in intracellular calcium levels. The rise in intracellular calcium activates phosphatidylcholine-specific phospholipase C and calcium-dependent phospholipase A. 13. The release of mature IL-1β, the IL-1β precursor and the contents of the secretory lysosomes by exocytosis of the in the absence of cell death. 14. Processing of the IL-1β precursor in the cytosol. Rab39a, a member of the GTPase family, contributes to the secretion of by helping traffic IL-1β from the cytosol into a vesicle compartment. 15. Mature IL-1β exists the cells via loss in membrane integrity, associated with the release of lactic dehydrogenase or microvesicles.
Figure 3. Non-caspase-1 extracellular processing of the IL-1β precursor. The 269 amino acid long IL-1β precursor is shown with the caspase-1 site at aspartic acid (D) at position 116. Extracellular protease sites are indicated by their amino acid recognition sites. The likely proteinase-3 site was derived combinatorial methods of the specific substrate.113.
Figure 4. Role of IL-1α in necrosis versus apoptosis. 1, left and right. In healthy cells of mesenchymal origin, the IL-1α precursor is found diffusely in the cytoplasm but also in the nucleus where it binds to chromatin. 2, right. During normal cell turnover, an apoptotic signal drives cytoplasmic IL-1α into the nucleus and is no longer a dynamic in the cell. The cell shrinks. 2, left. Cells exposed to hypoxia begin to die and nuclear IL-1α moves out of the nucleus into the cytoplasm. Taking on water, the cell swells as the necrotic process begins. 3, left. As the necrotic process continues, there is loss of membrane integrity and cytoplasmic contents containing the IL-1α precursor leak out. 3, right. Tissue macrophages take-up the apoptotic cell into endocytotic vesicles. 4. In the vesicles, the apoptotic cell is digested and there is no inflammatory response from the macrophage. 5, left. The IL-1α precursor is released into the extracellular compartment and binds to IL-1RI expressed on adjacent cells or to resident tissue macrophages. The tissue macrophage responds with synthesis of the IL-1β precursor as well as increased in caspase-1. From step 3, left, ATP is also released upon cell death and activates the P2X7 receptor for activation of caspase-1. 6. Caspase-1 is activated by the inflammasome, cleaves the IL-1β precursor and mature, active IL-1β is released. Alternatively, IL-1β is released via pyroptosis. 7. Once released, IL-1β induces chemokine production, resulting in a chemoattractant gradient. 8. As the endothelium of the microcirculation expresses adhesion molecule, blood neutrophils adhere and cross into the ischemic area. 9. With the infiltration of myeloid cells (monocytes and neutrophils), there is expanded inflammation, which extends beyond the initial area of ischemia.
Interleukin-1 in the pathogenesis and treatment of inflammatory diseases

Charles A. Dinarello

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