Heparin modifies the immunogenicity of positively-charged proteins

Running Title: Immunogenicity of heparin-binding proteins

Shalini L. Chudasama1, Benjamin Espinasse2, Fred Hwang1, Rui Qi1, Manali Joglekar1, Galyna Afonina1, Mark R. Wiesner2, Ian J. Welsby3, Thomas Ortel1, Gowthami M. Arepally1

Divisions of ¹Hematology and ²Cardiovascular Anesthesiology and ³Department of Civil and Environmental Engineering, Duke University Medical Center, Durham, NC;

Scientific section designation: Platelets and Thrombopoiesis

Corresponding author: Gowthami M. Arepally, M.D
Duke University Health System
Box 3486
Room 304 Sands Building
Durham, NC 27710
Phone: 919-668-1550
Fax: 919-684-2420
e-mail: arepa001@mc.duke.edu

Keywords: Platelet Factor 4, protamine, lysozyme, heparin, HIT, antibodies and murine model
Abstract

The immune response in Heparin-Induced Thrombocytopenia (HIT) is initiated by and directed to large multimolecular complexes of Platelet Factor 4 (PF4) and heparin (H). We have previously shown that PF4:H multimolecular complexes assemble through electrostatic interactions and, once formed, are highly immunogenic in vivo. Based on these observations, we hypothesized that other positively charged proteins would exhibit similar biological interactions with heparin. To test this hypothesis, we selected two unrelated positively-charged proteins {protamine (PRT) and lysozyme (Lys)} and studied heparin-dependent interactions using in vitro and in vivo techniques. Our studies indicate that PRT/H and Lys/H, like PF4/H, show heparin-dependent binding over a range of heparin concentrations and that formation of complexes occurs at distinct stoichiometric ratios. We show that protein/heparin complexes are capable of eliciting high titer antigen-specific antibodies in a murine immunization model and that PRT/H antibodies occur in patients undergoing cardiopulmonary bypass surgery. Finally, our studies indicate that protein/heparin complexes, but not uncomplexed protein, directly activate dendritic cells in vitro leading to IL-12 release. Taken together, these studies indicate that heparin significantly alters the biophysical and biological properties of positively-charged compounds through formation of multimolecular complexes that lead to dendritic cell activation and trigger immune responses in vivo.
Introduction

Heparin-Induced Thrombocytopenia (HIT) is an immune-mediated disorder caused by antibodies that recognize multimolecular complexes of platelet factor 4 (PF4), a positively charged platelet protein, and heparin (H), a negatively charged carbohydrate. We, and others, have shown that PF4 and heparin complexes assemble primarily through non-specific electrostatic interactions that are governed by principles of colloidal chemistry \(^1\)\(^-\)\(^5\). In colloidal systems, molecules of opposite charge “aggregate” or grow in size due to effects of charge neutralization. Particle interactions are frequently dependent on stoichiometric ratios of the two compounds, with the largest complexes occurring at molar ratios of the compounds leading to charge neutralization. When either compound is in molar excess, charge re-stabilization occurs and repulsive forces predominate, leading to reduced complex size and/or complex disassembly.

Studies, to date, indicate that PF4/heparin multimolecular complex formation is central to the pathogenesis of HIT. The characteristic “bell-shaped” curve seen with HIT antibody binding over a range of heparin concentrations coincides with heparin-dependent formation of multimolecular complexes \(^2\)\(^-\)\(^3\). HIT antibody binding, as gauged by serologic assays or functional studies of platelet activation, is optimal when multimolecular complexes form at or near equimolar ratios of PF4:H. However, antibody binding is markedly reduced with increasing heparin concentrations, a phenomenon that can be directly attributed to loss of complex formation \(^2\)\(^-\)\(^4\). Recent studies from our laboratory indicate that similar heparin-dependent changes affect the immunogenicity of PF4/H complexes \textit{in vivo} \(^5\)\(^,\)\(^6\). Our studies demonstrate that PF4/H complexes are immunogenic over a certain range of heparin concentrations associated with multimolecular complex formation and that the immune response is attenuated when PF4 or H is given alone or when heparin is in molar excess of PF4 \(^5\).

Heparin and heparin-like molecules bind a number of positively charged proteins in addition to PF4 \(^7\). These heparin-binding proteins (HBPs) are structurally and functionally diverse, and include, to name a few, nuclear proteins (protamine), enzymes (C1 esterase, lysozyme), adhesion molecules (fibronectin, vitronectin) growth factors (fibroblast growth factor), and lipid-binding proteins (apolipoprotein E, lipoprotein lipase). To date, it appears that a majority of HBP-heparin interactions are ionic in nature, with limited or no evidence for unique structural requirements, folding patterns or consensus heparin-binding regions in common \(^8\)\(^-\)\(^10\).
Early experimental studies of several heparin-binding proteins, including protamine and lysozyme, indicate that heparin interacts stoichiometrically with these proteins to form complexes and/or aggregates \(^{10-12}\). As noted with PF4/heparin complex formation\(^1\), protamine and lysozyme interactions with heparin bear the hallmark of charge-dependent colloidal interactions, namely sensitivity to changes in pH and ionic strength of the buffer \(^{10,12}\).

The ubiquity of HBPs in organisms, the non-specific nature of electrostatic interactions of HBPs with heparin and their similarity to PF4/H interactions, prompted us to investigate the biologic response to HBP/H complexes \textit{in vivo}. These studies aim to characterize the multimolecular complexes formed between heparin and two structurally and functionally unrelated HBPs (protamine, PRT and lysozyme, Lys). Using in vitro and in vivo studies, we present data to show that heparin significantly enhances the immunogenicity of positively charged molecules through formation of protein-heparin multimolecular complexes that activate dendritic cells and lead to an antigen-specific immune response in the host.
Materials & Methods

Biophysical studies of PRT/H and Lys/H complexes

Unless specified, reagents were purchased from Sigma Aldrich (St. Louis, MO). Solutions of protamine-sulfate (grade “X” amorphous powder from salmon sperm, Mw 5.1 kDa) or lysozyme (chicken egg white, Mw 14.3 kDa) were mixed with varying concentrations of UFH (100 or 1000 Units/mL; Heplock; Elkins-Sinn Inc, Cherry Hill, NJ) in Hanks Balanced Salt Solution (HBSS, Invitrogen Corp, NY) or H2O. Murine PF4 (mPF4) was prepared from recombinant bacteria as previously described 5,6. For stoichiometric calculations involving UFH, we utilized previously published estimates of UFH specific activity at 140 U/mg 3,5 and mean Mw of 15 kDa 3,5.

Light absorbance of PRT/H and Lys/H solutions was measured in a Spectra Max Plus 384 Plate Reader (MDS Technologies, Sunnyvale, CA) and analyzed using proprietary SoftMax Pro v. 5.3. The electrophoretic mobility of HBP/heparin complexes was measured using a Zetasizer Nano ZS (Malvern, Worcestershire, UK) as previously described 5. Zeta potentials (ζ-potential), which are related to the surface charge of particles, were calculated using the Henry equation and analyzed using the accompanying Zetasizer software (Malvern).

Murine immunization model

Immunizations were performed using modifications of a previously described immunization model 6. Protein or HBP/heparin concentrations for immunization were selected based on preliminary studies from biophysical data shown in Figure 1. Unless specified, mice were injected via retro-orbital plexus with protamine, PRT (125 µg/mL) + H (12.5 U/mL) or Lysozyme, Lys (500 µg/mL) + H 12 U/mL in a final volume of 100 µL daily for five days. Blood samples for ELISA were collected in anesthetized mice from the retro-orbital blood plexus in acid-dextrose citrate solution (ACD formula A, Baxter Healthcare Corporation, Deerfield IL) at baseline and at weekly intervals as specified after the start of immunizations. Animals injected with PRT/H or Lys/H were defined as seropositive if A450nm exceeds the mean A450nm + 3 times the standard deviation (SD) of animals injected with the respective protein in the absence of heparin (PRT alone or Lys alone). All studies were performed with the approval of the Institutional Animal Care & Use Committee at Duke University.
**PRT/H and Lys/H ELISAs**

Murine antibodies to PRT/H and Lys/H complexes were measured by ELISA using a 96 well microtiter plate (Nunc Maxisorp, Nalge Nunc International, Rochester, NY) as previously described \(^5,\) \(^6\) with the following exceptions. Antigen was coated overnight with 100 μL of antigen solution: protamine (31 μg/mL) + heparin (4 U/mL), lysozyme (125 μg/mL) + heparin (2.8 U/mL), mPF4 (10 μg/mL) + heparin (0.4 U/mL), or BSA (50 μg/mL) prepared in phosphate buffered saline (PBS, Invitrogen, Grand Island, NY). The protein to heparin ratios used for coating the plate were based on stoichiometric ratios obtained from biophysical data (Figure 1). Murine plasma was used at 1:100 and horseradish peroxidase-labeled goat anti-mouse IgG gamma was used at 1:500 for PRT/H antibodies and 1:2000 dilution for Lys/H antibodies.

**Dendritic cell activation**

Dendritic cells (DC) were isolated from non-immunized C57BL/6 murine splenocytes using previously published protocols \(^13\). Briefly, spleens were minced in Mg\(^{2+}\)- and Ca\(^{2+}\)-free HBSS/5% FCS and digested with 1 mg/ml of collagenase from Clostridium histolyticum and 0.2 mg/ml of DNAse I for 30 min at 37°C. Cells were passed through 70μm nylon mesh, washed with RPMI+10% FCS/10mM EDTA/20mM HEPES and re-suspended in 5 ml of the solution in a Petri dish, cells were harvested from plate and suspension loaded over equal volume of 14.5% Nycodenz gradient. The interface was collected and further purified using CD11c MicroBeads (Miltenyl Biotes, Auburn, CA) following manufacturer’s protocols. This procedure yielded DCs of >94% purity as assessed by flow cytometry. Endotoxin-free buffers and/or reagents that tested low for endotoxin were used for cell culture experiments.

After isolation, DCs were seeded into 96-well plates at 1x10^5 cells/well in RPMI media and incubated with media containing the following antigens: recombinant mPF4(10 μg/mL) + H (0.4 U/ml) PRT (15.6 μg/mL) + H (2 U/ml) and Lys (125 μg/mL) + H (2.8 U/ml). LPS (1 μg/mL) from E.coli 055:B5 (Sigma-Aldrich, St. Louis, MO) was added to the culture as positive control and cellular activation was measured 24 hours after addition of antigen using an IL-12 optiELISA kits from BD Biosciences (San Diego, CA) following manufacturer’s directions.

**Patient samples**

Plasma from normal subjects (n=45) were purchased from George King Biomedical Inc. (Overland Park, Kansas). Blood from patients undergoing cardiopulmonary bypass (CPB) surgery was collected at baseline, 5 days and 30 days after the procedure using informed
consent under an IRB approved protocol. Human antibodies to PRT/H were measured using ELISA conditions described above for murine plasma with the exception of adding 0.05% Tween to buffers to reduce background binding. Antibodies to hPF4/heparin were detected using a commercial immunoassay (PF4 Enhanced®, Genetics Technology Institute, GTI, Waukesha, WI).

Statistical analysis

Antibody levels in murine and human ELISAs were compared using the Student t-test for comparisons of two groups or by a one-way ANOVA analysis for more than 2 groups. ELISA reactivity to various antigens was expressed as mean ± 1 SD and analyzed for significance using the Student t-test. Statistical analyses were performed using Graph Pad Prism (Graphpad Software, San Diego, CA, USA). Differences were considered significant at a p value < 0.05.
Results

Biophysical characterization of PRT/H and Lys/H complexes

We have previously shown that PF4/H multimolecular complex formation occurs through colloidal interactions, which can be measured through changes in light scattering (absorbance) and surface charge or ζ-potential. In our previous studies, we showed that PF4 alone in solution displays minimal light absorption (Fig 1A right side of curve) on spectrophotometry and carries a positive charge as measured by ζ-potential (Figure 1B, top of curve). With increasing concentrations of heparin, there is an accompanying increase in light transmission (peak curve Figure 1A) due to multimolecular assembly. In the ζ-potential, addition of heparin to a solution of PF4 lowers the surface charge of PF4, and leads to a negative or decreased slope (change from positive to negative, Figure 1B). The “peaks” in light transmission (Figure 1A) and the inflection points in the ζ-potential (Figure 1B) correlate closely with formation of large multimolecular complexes due to effects of charge neutralization.

To determine if other positively charged proteins show similar colloidal interactions with heparin, we selected two HBPs, protamine sulfate and lysozyme, with known high affinity to heparin. To demonstrate multimolecular complex formation, we mixed various concentrations of PRT (31-250 µg/mL, Figure 1C, D) or Lys (125-1000 µg/mL, Figure 1E,F) with increasing amounts of heparin (0-100 U/mL) and measured light absorption (A280nm, Figures 1C,E) or ζ-potential (Figure 1D,F). As shown in Figure 1, increasing amounts of heparin altered the biophysical properties of PRT or Lys as indicated by changes in light transmission/absorption and ζ-potential. For a fixed protein concentration of PRT or Lys, we noted that addition of heparin results in complexes of increasing size, until a “peak” size is achieved; further increases in heparin result in decreased absorbance which correlates with reduced size of complexes. As seen with PF4/H interactions, the heparin concentrations resulting in peak complex size for PRT or Lys, as measured by absorbance (A280nm, Figures 1C & E), were concordant with the heparin concentrations at which charge neutralization occurred (ζ-potential inflection points, Figures 1D & F) for each protein. The concentrations of protein and heparin resulting in the peak absorbance and/or neutral charge allowed us to approximate the stoichiometry of the two compounds necessary for multimolecular complex formation. For protamine, we approximated that peak complex size occurs at a molar ratio of ~3:1, whereas for lysozyme, peak complexes occur at molar ratios of ~5:1. The molar ratios for peak complex formation for these individual
proteins remain constant for a given protein/heparin peak, irrespective of protein concentration (e.g., for PRT, the peaks for 250 and 125 µg/mL curves occur at 32 and 16 U/mL heparin respectively, both approximating a PHR molar ratio of 3:1).

Heparin modifies the immunogenicity of positively charged proteins in vivo

We have previously shown that mPF4/heparin complexes, but not mPF4 alone or heparin alone elicit robust immune responses in mice. To determine if heparin modifies the immunogenicity of PRT and Lys similar to mPF4, we injected C57BL/6 mice with PRT + H or Lys + H as described in Methods. Similar to mPF4/H, mice injected with PRT/H or Lys/H, but not PRT alone or Lys alone, develop robust immune responses within 14 days of immunization (Figure 2 A & B). When similar cohorts from replicate experiments were tested concurrently by ELISA (n=30 for each antigen: PRT, PRT/H, Lys or Lys/H), significant differences were seen in the mean absorbance (A450nm + SD) of animals injected with PRT/H as compared to PRT alone (PRT/H = 0.910+1.173 v PRT= 0.1193 + 1.0193, p<0.0001) and Lys/H as compared to Lys alone (Lys/H=1.537+1.5 v. Lys alone= 0.094 + 0.054; p<0.0001, Supplemental Figure S1). In the combined cohort, seroconversions (as defined in Methods) occurred in 30% of animals injected with PRT/H (Supplemental Figure S1A) and 53% of animals injected with Lys/H (Supplemental Figure S1B). Antibody responses to PRT/H or Lys/H were highly specific for the immunizing antigen as antibodies formed to PRT/H (Figure 2C, closed circles) or Lys/H (Figure 2D, closed squares) did not cross-react with other antigens (mPF4 or BSA) or other HBP/H complexes (mPF4/heparin, PRT/H in mice expressing anti- Lys/H or vice versa; Figure 2C &D). Mice injected with PRT or Lys alone did not react to wells coated with PRT alone or Lys alone (Supplemental Figure S2). Consistent with a polyclonal response, antibodies formed after PRT/H or Lys/H showed variable heparin-dependent reactivity by ELISA (Figure 2C&D and Supplemental Figure S3). Whereas PRT/H antibodies showed binding to wells with increasing amounts of heparin (with minimal binding to PRT alone and increased binding to wells coated with PRT and variable amounts of heparin, Supplemental Figure S3A), Lys/H antibodies showed no preferential heparin-dependent binding (Supplemental Figure S3B).

Characterization of the in vivo immune response to PRT/H

We undertook additional studies to determine if the humoral response to PRT/H or Lys/H shares any important biologic features of mPF4/H immune response. As shown in Figure 3, the PRT/H immune response resembles mPF4/H seroconversions in terms of temporal course and stoichiometric requirements for multimolecular complex formation. Seroconversion is
maximal by day 15 (D15) from start of immunization, with antibody levels declining to baseline levels by D30 (Figure 3A). Mice injected with PRT alone show no antibody responses to PRT/H over time (Figure 3A inset). A similar time course of seroconversions were seen with mice injected with Lys/H (Supplemental Figure S4). As with PF4/H seroconversions, the immunogenicity of PRT/H complexes in vivo is linked to the stoichiometry of complex formation. Mice injected with PRT/H complexes at molar ratios of PRT:H of 3:1 showed a trend towards increased seroconversion, whereas mice injected with complexes at higher or lower molar ratios were less likely to seroconvert (Figure 3B, p=ns). Other similarities with the PF4/H immune response were seen with respect to the doses of multimolecular complexes needed to elicit an immune response (Supplemental Figure S5). These latter studies suggest that a critical mass amount of circulating multimolecular antigen is required, even if the complexes are formed at “optimal” stoichiometry.

**PRT/H complexes and PF4/H complexes elicit dendritic cell activation.**

The similarity of the in vivo immune response to PRT/H and mPF4/H and the knowledge that both antigens formed multimolecular complexes with heparin, suggested the possibility that multimolecular complexes were more effective than uncomplexed protein at triggering cellular activation. To determine if multimolecular complexes activated antigen presenting cells (APCs), dendritic cells were harvested from non-immunized C57Bl/6 mice and incubated with heparin or buffer, protein (mPF4, PRT or Lys), or protein/H complexes. Cell supernatants were harvested and assayed for IL-12, a marker of dendritic cell activation. As shown in Figure 4, IL-12 was increased in wells containing HBP/H complexes as compared to wells containing uncomplexed protein, buffer or heparin. These studies show that antigen in the form of multimolecular complexes elicits potent cellular activation of dendritic cells.

**Antibodies to PRT/H can be demonstrated in human subjects undergoing CPB**

Because patients undergoing CPB are routinely exposed to high doses of PRT and heparin, and because our murine studies indicated that PRT/H complexes are immunogenic in vivo, we next asked if patients undergoing CPB develop antibodies to PRT/H. To study the PRT/H immune response in humans, we analyzed samples from normal subjects (n=45) and plasma from patients undergoing CPB (n=15, for each patient, sample from baseline prior to CPB, 5 days and 30 days after CPB was evaluated). As shown in Figure 5A, plasma from normal subjects as well as plasma from CPB patients prior to surgery and 5 days after surgery showed minimal reactivity in the PRT/H ELISA. However, after 30 days, we observed that 4/15 patients (27%)
developed significantly elevated levels of antibodies to PRT/H as compared to normals, or their own plasma collected at baseline or 5D after surgery. Seropositive patients recognized PRT/H preferentially over PRT (p=ns) and did not cross-react with other antigens (Figure 5B, p<0.001 for PRT/H v. BSA, Lys, Lys/H or hPF4/H).
Discussion

In this study, we show that heparin forms multimolecular complexes with positively charged proteins other than PF4 and, in so doing, enhances the immunogenicity of these proteins in a murine model. We demonstrate that heparin-modified complexes are capable of activating dendritic cells directly *in vitro* and that this process may lead to immune activation *in vivo*, as mice and humans undergoing CPB develop antigen-specific antibodies to PRT/H. These findings, and the similarity of the humoral response to varying heparin-modified complexes, suggest that antigen in the form of multimolecular complexes is potently immunizing *in vivo*.

The observation that heparin modifies the biophysical properties of proteins is not new. In 1935, Fischer showed that heparin-binding significantly alters the spectral properties of proteins 14. Subsequent studies have shown that heparin binds to a variety of proteins leading to alterations that affect not only protein function 9,15, but structural properties that enhance a proteins stability 16,17 and/or its localization, as seen with PF4 or other cytokines 18. Crystallographic studies of several heparin-protein complexes do not reveal the presence of any consistent structural or sequential motifs in heparin-binding proteins, other than clusters of positively charged residues responsible for ionic interactions 8. Our earlier studies have shown that the biophysical alteration of PF4 by heparin significantly enhances the immunogenicity of PF4 5,6,19,20. To date, the immunologic consequences of heparin-binding to other positively charged proteins have not been examined.

For our studies, we selected two structurally and functionally dissimilar proteins, protamine and lysozyme, whose heparin-binding features have been partially characterized. Protamine sulfate is a small (Mw 4100 Da), highly cationic DNA binding family of proteins found in sperm nuclei. Protamines replace sperm-specific histones during late spermatogenesis and compact DNA and inhibit its transcription and degradation 21. Protamines from varying species show conservation of arginine rich domains (60-80%), which account for the protein’s high isoelectric point 22 and conserved functional roles as a DNA binding protein across species. The crystal structure of protamine or the protamine-heparin complex has not been solved; however, spectroscopy studies indicate that free protamine is unstructured in solution 21. Upon binding DNA, protamine binds to the phosphate groups in the major groove of the DNA helix allowing the DNA to coil tightly and condense 21.

Lysozyme, on the other hand, is a bactericidal enzyme (14kDa) found in mucosal secretions (saliva and milk). The protein consists of 129 amino acids and its crystal structure reveals the
presence of 5 helical regions consisting of 3 alpha helices and 5 regions of beta pleated sheets. Like protamine, lysozyme has a high pl (10.5) and has a homogenous distribution of positive charges over the protein surface, to which heparin presumably binds. Bacteria use carbohydrate polyanions to bind lysozyme’s cationic sites and interfere with its catalytic activity. We considered the lack of any structural or functional similarities of these proteins to PF4 as proof-of-concept that multimolecular complex formation and immunogenicity of heparin-containing multimolecular complexes is a generalizable phenomena that extends beyond PF4/H multimolecular complexes.

Using techniques to measure changes in absorbance and ζ-potential, we were able to confirm and extend previous observations on the biophysical interactions of both protamine and lysozyme. We demonstrated that both these proteins show heparin-dependent binding (Figure 1), with binding curves similar to that seen with PF4 and heparin. Specifically, these curves reveal that PRT/H and Lys/H interactions are charge dependent with PRT/H or Lys/H complexes increasing in size until a heparin concentration is reached at which charge neutralization occurs. The molar ratio leading to multimolecular complex assembly varies for each protein (mPF4: H ~2:1, PRT:H ~3-4:1, and Lys:H is 5:1) and are comparable to findings reported in the literature employing other techniques. These studies show that absorbance and ζ-potential are reliable, simple tools for assessing the stoichiometry of heparin-dependent complex formation with positively charged proteins.

We show that structural modification of a protein by heparin significantly alters its immunogenicity (Figure 2 and Supplemental Figure S1). Mice injected with PRT or Lys alone do not make antibodies to these proteins (Supplemental Figure S1 and S2), even though these proteins are foreign antigens in the murine system (salmon-derived protamine and chicken egg-white derived lysozyme). However, mice injected with heparin-modified PRT or Lys are more likely to develop high levels of antibodies to each heparin modified protein (Figure 2A&B and Supplemental Figure S1). The similarity of the immune response with regards to the temporal kinetics (Figure 3 and Supplemental Figure S4) and dependence on stoichiometric complex formation (Figure 3B), of disparate antigens as mPF4/H, PRT/H and Lys/H suggest that the immune response is likely triggered by formation of multimolecular complexes.

Despite use of syngeneic animals and fixed doses of antigen, we recognize that rates of seroconversion were not uniform in our experimental model. We noted an overall seroconversion rate of ~30-50% of animals injected with PRT/H or Lys/H (Supplemental Data,
Figure S1). Several variables likely account for non-uniform seroconversions in our murine model. First, we do not use adjuvant in our immunization strategy. It is known that most peptides and proteins, when injected alone, are poor immunogens \(^{24}\). For this reason, adjuvant, such as alum or Freund’s adjuvant, is typically employed in animal models to enhance seroconversions. We purposefully avoided use of adjuvants in our model, because we were interested in demonstrating the “adjuvant” like properties of heparin. A second variable in this animal model is the use of UFH. UFH is a polydisperse compound containing molecules of variable chain length and charge \(^{25}\). When UFH is mixed with PF4 \(^5\), protamine or lysozyme, the polydispersity of UFH species is likely to result in complexes of variable size and/or charge density. Last, it is possible that complex size changes with time, as previously shown for complexes containing PF4 and heparin \(^5\). While we strived to keep the injection conditions uniform, it is likely that size variation invariably occurred during the time required for sequential animal injections.

While PRT/H and Lys/H seroconversions share some essential features of mPF4/H seroconversions, with regards to timing (Figure 3A and Supplemental Figure S4), stoichiometric requirements (Figure 3B) and dose dependency (Supplemental Figure S5), it is important to recognize that the immune response to each antigen is highly specific. Antibodies to PRT/H do not cross-react with Lys/H or PF4/H and vice versa (Figure 2 C&D). As well, there are some notable biologic differences with regards to the fine specificities of the polyclonal response to each immunogen. Whereas antibodies to PRT/H do not recognize PRT alone and show heparin dependent binding by ELISA (Figure 2C and Supplemental Figure S3A), antibodies formed after Lys/H exposure do not show heparin-dependent reactivity (Figure 2D and Supplemental Figure S3B). We attribute these differences in serologic specificities to the polyclonal nature of the immune response to each antigen, possibly owing to the types of neoepitopes that are displayed on the circulating antigen in vivo. Studies have shown that polyclonal antibodies from HIT patients can also show a range of binding specificities, with some antibodies binding to PF4 alone (26% in study by \(^{26}\)) while other antibodies show lack of heparin-dependent reactivity (4-12% of antibodies in \(^{27-29}\)). Taken together, differences in the heparin-dependent reactivity of Lys/H, PRT/H and PF4/H antibodies in an ELISA, which is reflective of a polyclonal immune response, does not contradict our central observation that heparin modifies the immunogenicity of these individual proteins in vivo (Figure 2A&B and Supplemental Figure S1).
The specificity of the immune response to each respective antigen implies an important role for adaptive immunity over innate immunity or T-cell independent immune responses. As adjuvant was not used in this or previous studies of our immunization model, it is presumed that multimolecular complexes exert an “adjuvant-like” effect on dendritic cells. As shown in Figure 4, our studies confirm that protein/heparin complexes are capable of directly activating dendritic cells. To what extent IL-12 production and/or dendritic cell activation contributes to the HIT immune response remains under investigation. Although our murine studies indicate a compelling role for T-cells in PF4/H antibody production, the role of T-cells in human HIT is less clear.

The identification of a serologic response to PRT/H in humans undergoing CPB, in part, validates our in vitro and in vivo observations on the biological consequences of circulating PRT/H multimolecular complexes. In the limited number of patients studied, we noted PRT/H antibodies in 4/15 patients undergoing CPB (Figure 5A). Patients undergoing CPB are routinely exposed to high doses of PRT (5-9 mg/kg, or an average dose of 350-630 mg in a 70 Kg individual) to reverse the anticoagulant effects of heparin (1.4-4.3 U/mL). While there are no studies of protamine levels in patients undergoing CPB, in healthy volunteers, a 250 mg dose or 3.5 mg/kg dose (in a 70kg adult), is associated with a 10-50 µg/mL level of circulating PRT. The stoichiometric ratios seen in Figure 1A can likely be achieved in some CPB patients (Figure 5A) who may receive either higher doses of protamine or heparin and/or have the additional inflammatory stimulus of surgery.

The clinical implications of a PRT/H immune response in humans are not clear. Protamine allergy is well-described in patients undergoing CPB, vasectomized men, and diabetic patients who receive insulin preparations containing protamine (Neutral Protamine Hagedorn or NPH insulin). In most of these cases, the protamine allergy is associated with a type I hypersensitivity or anaphylactic reaction, associated with high titer IgE and in some cases high-titer IgG. A more recent study documents the occurrence of anti-PRT antibodies in patients undergoing CPB, some of whom may be mistaken for HIT due to weak cross-reactivity of anti-PRT antibodies with PF4/H antibodies. Our studies, however, suggest that the PRT/H antibody response in humans is distinct from previously described protamine reactions (Type I hypersensitivity) or delayed type hypersensitivity skin reactions (Type IV) caused by subcutaneous heparin. The PRT/H seropositive patients in our cohort were asymptomatic and manifested a highly specific immune response to PRT/H (Figure 5B). Because samples from patients undergoing CPB were only available at D5 and D30, the exact timing of PRT/H
seroconversions was not known. Because PRT is likely cleared from circulation by the time seroconversions occur, it is likely that any adverse outcomes associated with anti-PRT/H may be limited to patients who are in need of protamine re-exposure, i.e., patients requiring another CPB or diabetic patients receiving NPH insulin. Additional studies in transgenic mice bearing platelet human FcγRIIR will be informative as to whether immune complexes containing anti-PRT/H and antigen are capable of eliciting platelet activation and thrombocytopenia.

In summary, our studies suggest that the immune response to heparin-modified complexes is a biologically conserved response. These findings have implications for understanding the pathogenesis of HIT and may provide insights into the existence of an immune response to other heparin-binding proteins capable of forming heparin-dependent multimolecular complexes.
Acknowledgement: The authors wish to thank Sharon Hall for assistance with obtaining patient samples. The authors have no conflicts of interest.

Supported by the National Institutes of Health HL081395 (GMA), UO1-HL072289 (TLO) and U54-HL077878 (TLO), American Heart Association Grant-in-Aid (GMA), USEPA STAR award and RD83241301(MRW), and the Howard Hughes Medical Institute Grant, 52005871 (SC).

Conception and design: S.Chudasama, B.Espinasse, M.Weisner, G. Arepally
Provision of study materials or patients: I. Welsby, B.Espinasse, M. Weisner, T. Ortel
Collection and assembly of data: S.Chudasama, B.Espinasse, F. Hwang, M. Joglekar, R. Qi, G. Afonina,
Manuscript writing: S. Chudasama, I. Welsby, T. Ortel, M. Weisner, G. Arepally
References


Figure Legends

**Figure 1:** Heparin dependent multimolecular complex formation with PRT and Lys.  
A-B: Schematic representation of PF4 and H interactions as previously reported.  
PF4 in solution, in the absence of heparin, shows minimal light aborption (left side of curve in panel A) and displays positive charge (top part of curve in panel B). With increasing amounts of heparin (x-axis), charge neutralization occurs resulting in complexes of increasing size (peak in panel A) and neutral charge (inflection point in panel B). Addition of heparin beyond the concentration required for peak complexes results in complexes of reduced size (right side of curve in panel A) and increasingly negative charge (bottom of curve in panel B).  
C-D: PRT (250-31 µg/mL) was incubated with increasing concentrations of heparin. Light absorption (panel C) and ζ-potential (panel D) were measured.  
E-F: Lys (1000-125 µg/mL) was incubated with increasing concentrations of heparin. Light absorption (panel E) and ζ-potential (panel F) were measured.  
Data are representative of three or more independent experiments.

**Figure 2:** Immune responses to PRT/H or Lys/H complexes.  
C57Bl/6 mice were immunized with PRT alone (125 µg/mL, n=10) or PRT/H (125µg/mL PRT and 12.5 U/mL H, n=10) and Lys (500 µg/mL, n=5) or Lys/H (500 µg/mL Lys and 12 U/mL, n=10). A-B: Peak antibody responses to PRT/H (panel A) and Lys/H (panel B) occurred within 14 days from start of immunization.  
C-D: Specificity of representative seropositive PRT/H (panel C, closed circles) and Lys/H (panel D, closed squares) and seronegative PRT/H (open circles) and Lys/H (open squares) are shown.  
Data are representative of greater than three independent immunization experiments involving PRT/H or Lys/H.

**Figure 3:** Characterization of murine immune response to PRT/H.  
(A) Time course of murine immune response to PRT/H (n=10).  
C57Bl/6 mice were immunized with PRT/H (PRT 125 µg/ml and Hep 12.5 U/mL) and antibody levels were followed over a time period of 30 days. Each line represents the seroconversion profile of an individual mouse injected with PRT/H.  
(B) Stoichiometry of the immune response to PRT/H: Mice (n=10/cohort) were immunized with a fixed amount of PRT (125µg/mL) and varying H doses (0, 5, 10, 12.5, 25 and
50 U/mL) to yield varying PRT:H molar ratios as shown above (--; 8:1, 4:1, 3:1, 1.5:1 or 1:1.2). Results of antibody levels are shown at D15.

Figure 4: Dendritic cell (DC) activation by protein/H complexes: Murine DCs were isolated from non-immunized C57Bl/6 mice and incubated with PRT+ H, Lys+ H, mPF4 + H, buffer containing media (Buffer), heparin (0.4U/mL) or lipopolysaccharide (LPS). IL-12 was measured from cell supernatants by ELISA. Shaded bars indicate protein/heparin complexes and light shade is uncomplexed protein. One tailed paired t-test was used for determining significance. *p<0.01 for PF4 v. PF4/H **p<0.001 for PRT v. PRT/H; p=ns for Lys v. Lys/H.

Figure 5: Antibodies to PRT/H in patients undergoing CPB. (A) Plasma from healthy subjects (“Normal”; n=45) and patients undergoing CPB (n=15) were screened for antibodies to PRT/H by ELISA at baseline, D5 and D30. (B) Specificity of antibodies to PRT/H. Plasma from 4 patients with antibodies to PRT/H (solid symbols) and three seronegative CPB patients (open symbols) were incubated with wells coated with BSA, hPF4/heparin, Lys, Lys/H, PRT or PRT/H. P values were calculated for panels A and B using a one-way ANOVA. For Panel B, a Bonferroni post-test was applied for calculating differences between conditions. ** p<0.001 for PRT/H v. BSA, Lys, Lys/H, and hPF4/H.
FIGURE 1

A

B

C

D

E

F

For personal use only. on November 12, 2017. by guest www.bloodjournal.org From
Figure 4

IL-12 (pg/mL)

- PRT
- PRT/H
- Lys
- Lys/H
- PF4
- PF4/H
- Buffer
- Hep
- LPS
Figure 5

A

p < 0.001
p < 0.05

B

*p < 0.001

anti-PRT/H (A\textsubscript{450mm})

Normal  Baseline  Day 5  Day 30

BSA  Lys  Lys/Hep  hPF4/Hep  PRT  PRT/Hep
Heparin modifies the immunogenicity of positively-charged proteins

Shalini L. Chudasama, Benjamin Espinasse, Fred Hwang, Rui Qi, Manali Joglekar, Galyna Afonina, Mark R. Wiesner, Ian J. Welsby, Thomas Ortel and Gowthami M. Arepally

Information about reproducing this article in parts or in its entirety may be found online at: http://www.bloodjournal.org/site/misc/rights.xhtml#repub_requests

Information about ordering reprints may be found online at: http://www.bloodjournal.org/site/misc/rights.xhtml#reprints

Information about subscriptions and ASH membership may be found online at: http://www.bloodjournal.org/site/subscriptions/index.xhtml

Advance online articles have been peer reviewed and accepted for publication but have not yet appeared in the paper journal (edited, typeset versions may be posted when available prior to final publication). Advance online articles are citable and establish publication priority; they are indexed by PubMed from initial publication. Citations to Advance online articles must include digital object identifier (DOIs) and date of initial publication.