Inhibition of the intrinsic coagulation pathway factor XI by antisense oligonucleotides: a novel antithrombotic strategy with lowered bleeding risk

Short Title: Antisense Suppression of FXI activity in mice

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ABSTRACT

Existing anticoagulants effectively inhibit the activity of coagulation factors of the extrinsic and common pathway but have substantial limitations and can cause severe bleeding complications. Here we describe a novel therapeutic approach to thrombosis treatment. We have developed and characterized the efficacy and safety of selective second generation antisense oligonucleotides (ASOs) targeting coagulation factor XI (FXI), a member of the intrinsic coagulation pathway. Systemic treatment of mice with FXI ASO led to a potent, specific and dose-dependent reduction of FXI mRNA levels in the liver with corresponding reductions in plasma levels of FXI protein and activity. FXI ASO treatment produced potent, dose-dependent antithrombotic activity in various venous and arterial thrombosis models, comparable to warfarin or enoxaparin. However, unlike warfarin or enoxaparin, FXI inhibition did not cause bleeding. Co-administration of FXI ASO with enoxaparin or the anti-platelet drug clopidogrel produced improved anti-thrombotic activity without increased bleeding. Finally, plasma-derived FXI concentrate was shown to effectively and rapidly reverse the anticoagulant effect of FXI antisense therapy. These results support the concept that inhibition of FXI through antisense therapy might serve as a new and effective strategy for the treatment and prevention of venous thromboembolism with improved specificity and safety.
INTRODUCTION

Precise regulation of blood coagulation is critical for normal hemostasis. Vessel wall injury-induced exposure of extravascular tissue factor to circulating coagulation factor VIIa (FVIIa) activates the extrinsic coagulation cascade which through subsequent activation of coagulation FIX, FX, and FII (prothrombin) ultimately produces a fibrin and platelet cross-linked clot at the site of injury\(^1\)\(^-\)\(^2\). However, in pathogenic conditions such as thrombosis, additional coagulation pathways are activated that mediate both the propagation and the maintenance of the growing thrombi\(^3\).

Several lines of evidence suggest that the intrinsic coagulation pathway, and in particular FXI, contributes to the pathogenesis of thrombosis. In humans, increased plasma levels of FXI have been associated with deep vein thrombosis\(^4\), ischemic stroke\(^5\) and myocardial infarction\(^6\). In addition, the incidence of ischemic stroke seems to be significantly lower in FXI deficiency compared to the general population\(^7\). Furthermore, FXI deficiency has been shown to protect mice from experimentally induced thrombosis or stroke and to reduce occlusive thrombus formation in venous and arterial models of thrombosis\(^8\)\(^-\)\(^11\). In addition pharmacologic inhibition of FXI has also been shown to promote antithrombotic activity in various animal models and under different conditions\(^12\)\(^-\)\(^14\).

In contrast to its important role in the pathogenesis of thrombosis, FXI appears to play a relatively minor role in normal hemostasis. FXI activation is thought to play a principal function in the propagation and stabilization of a growing thrombus by virtue of a positive feedback amplification loop between thrombin and FXI itself and by activation of thrombin-activatable fibrinolysis inhibitor (TAFI) which protects the fibrin clot against fibrinolysis\(^3\)\(^,\)\(^15\)\(^,\)\(^16\). Consistent with this notion, only a minority of subjects with severe FXI deficiency present with bleeding tendencies, which are generally mild to moderate and typically only occur under extreme circumstances\(^17\)\(^-\)\(^19\). In contrast, deficiencies in components of either the extrinsic or common pathways are consistently associated with severe bleeding or even
incompatible with life\textsuperscript{20-23}. Currently available anticoagulant agents, such as vitamin K antagonists (including warfarin or coumadin), (low molecular weight) heparins (e.g. enoxaparin or Lovenox) and small molecule FXa or FIIa inhibitors, all target components of the extrinsic or common pathways and can be associated with serious bleeding complications\textsuperscript{24-27}. Taken together these findings suggest that targeting intrinsic coagulation factor XI may be an attractive therapeutic approach for identifying novel antithrombotic agents with greater safety.

To further define the role of FXI in thrombosis and hemostasis, we evaluated a previously unexplored therapeutic modality for coagulation factor inhibition. Second generation antisense oligonucleotides (ASOs) selectively depleting FXI expression in the liver were developed and the pharmacological profile of FXI ASO treatment in mice, both alone and in combination with existing standard of care antithrombotic agents, was characterized. Antisense therapy is based on base-pair hybridization through which ASOs selectively bind to their complementary mRNA target\textsuperscript{28}. This binding typically results in the selective and catalytic degradation of the targeted mRNA by a mechanism involving the nuclease RNAse H\textsuperscript{28} and leads to a corresponding reduction in target protein levels. Antisense technology is a potentially attractive therapeutic modality for the development of FXI inhibitors for several reasons. First, FXI is synthesized primarily in the liver, which is one of most sensitive tissues for ASO therapy\textsuperscript{28, 29}. Several second generation ASOs designed against hepatic targets are currently in clinical development and have demonstrated significant clinical activity and safety, for example in hyperlipidemia\textsuperscript{28, 30}. Furthermore, second generation ASOs exhibit dose-linear and predictable pharmacokinetics in humans\textsuperscript{30} which can simplify dose selection and help to ensure patient safety. Third, due to prolonged tissue elimination half-lives, second generation ASOs can be administered relatively infrequently (once weekly or even less frequent), which should provide acceptable patient convenience\textsuperscript{30}. Fourth, compared to other anticoagulant modalities, including small molecule inhibitors and the natural product anticoagulants like warfarin, antisense inhibitors offer a high degree of target selectivity which should provide
greater patient safety. Finally, since second generation ASOs are not substrates for cytochrome P450 enzymes, which are involved in the metabolism of several drug classes in humans, and because of their high specificity, antisense drugs avoid common drug-drug interactions which frequently limit therapeutic applications, thereby making ASO drugs particularly well suited for combination therapy. The results of the current study support the conclusion that FXI antisense therapy could serve as an effective, convenient and safe alternative for the treatment and prevention of venous and arterial thrombosis in humans.

MATERIALS AND METHODS

Oligonucleotides

All oligonucleotides were 20 nucleotides in length and chemically modified with phosphorothioate in the backbone and 2'-O-methoxyethyl (MOE) on the wings with a central deoxy gap ("5-10-5" design). Oligonucleotides were synthesized using an Applied Biosystems 380B automated DNA synthesizer (Perkin Elmer-Applied Biosystems, Foster City, CA, USA) and purified as previously described.

To identify a potent FXI ASO for testing in mice, ASOs were designed and tested in primary mouse hepatocytes for their relative ability to suppress FXI mRNA levels. From these experiments an optimized FXI ASO was selected for evaluation in mice.

Animals and Oligonucleotide Dosing

Eight-week-old male BALB/c and C57BL/6 mice (Charles River Laboratories) were treated according to the indicated treatment schedules. The animals were housed in micro-isolator cages on a constant 12 hour light-dark cycle with controlled temperature and humidity and were given access to food and water ad libitum. All animal husbandry and procedures performed at Isis (CA, USA) and AMC (Amsterdam, The Netherlands) were approved by the Institutional Animal Care and Use Committee and the Dutch Animal Experiments Committee, respectively.
Preparation of Platelet Poor Plasma (PPP)

Blood samples were collected under anesthesia by cardiac puncture. Blood was quickly withdrawn from the heart using a 1-ml plastic syringe with a 27G needle and collected into a final ratio of nine parts of whole blood to one part of 3.2% sodium citrate. Mice were euthanized by cervical dislocation. Blood samples were immediately mixed by tapping and inverting the tube 5 times to ensure proper anticoagulation and then centrifuged for 10 minutes at 2000 rcf at room temperature. After removal of the platelet rich plasma the samples were centrifuged a second time for 5 minutes. Platelet poor plasma (PPP) was stored at -80 °C until assays were performed.

FXI Deficient Plasma

The Factor XI deficient plasma is a commercial preparation from a human patient with a congenital deficiency supplied by George King Bio-Medical Inc. Overland Park, USA.

Measurement of Hepatic FXI mRNA

Mouse liver was homogenized in RLT buffer. Total RNA was prepared according to RNAeasy mini lit (Qiagen, Valencia, CA). The quantitative real-time RT-PCR was done by ABI Prism 7700 sequence detector (Applied Biosystems, Foster City, CA). The sequences of primers and probe used for mouse FXI were: ACATGACAGGCGCGATCTCT (forward), TCTAGGTTCACGTACACATCTTTGC (reverse), and TTCCTTCAAGCAATGCCCTCAGCAATX (probe) (Integrated DNA Technologies, Coralville, IA). The amount of each FXI mRNA was normalized to amount of total RNA determined by Ribogreen. RNase protection assay (RPA) was performed using the RPA kit (BD Pharmingen, San Jose, CA) according to the manufacturer’s instructions.

Activated Partial Thromboplastin Time (aPTT) and Prothrombin Time (PT)

APTT and PT were measured using the ACL 1000 coagulation analyzer (IL Instrumentation, Beckman Coulter, Fullerton, CA). APTT test of mouse PPP was initiated using ellagic acid
mixture (APTT-XL, Pacific Hemostasis, Fisher Diagnostics, Middletown, CA, USA) and calcium chloride, while initiation of the PT assay in mouse PPP was performed by adding thromboplastin. APTT and PT measurements of saline treated mice were pooled and served as baseline reference. APTT and PT ratios were calculated by dividing measured values by these baseline values.

**FXI and FXII Activity**

To measure plasma FXI and FXII activity mouse plasma was 40 times diluted in human FXI or FXII deficient plasma followed by aPTT measurement, using pooled values from normal mice as reference.

**Ferric Chloride Induced Inferior Vena Cava Thrombosis**

Antithrombotic activity was studied using a well established ferric chloride (FeCl₃) induced inferior vena cava (IVC) thrombosis model, previously described by others. Total mRNA was purified from liver and vena cava tissue samples, and analyzed by RT-PCR for FXI and Platelet Factor 4 (PF4) mRNA levels, respectively. PF4 mRNA levels were used to determine the effect of treatment on platelet deposition as a measure of thrombus formation. PF4 mRNA levels in the IVC tissue exposed to FeCl₃ was normalized to non-exposed vena cava tissue.

**Ferric Chloride Induced Mesenteric Vein Thrombosis**

Male BALB/c mice were maintained under isoflurane (2.5% inhalant). Fluorescently labeled mouse platelets were injected intravenously followed by a 3 minute exposure of the mesenteric vein to a 10% FeCl₃ solution. Using intravital microscopy, thrombus formation in the mesenteric vein was continuously recorded on video for a maximum of 40 minutes or until total occlusion. Time to formation of 30 μm thrombi and full occlusion was recorded.

**Stenosis Induced Inferior Vena Cava Thrombosis**
For stenosis induced IVC thrombosis a model also known as the “St. Tomas’ model” which uses a combination of reduced blood flow and endothelial damage was applied. In short, the IVC of male BALB/c mice, anaesthetized with 2.5% inhalant isoflurane, was exposed and mobilized via a midline abdominal incision below the left renal vein, and separated from the abdominal aorta by blunt dissection. A 6-0 silk tie (Ethicon, UK) was placed behind the vessel just below the left renal vein and 2 silk sutures (4-0, Ethicon, UK) were placed longitudinally over the IVC and tied over the top. Finally the 4-0 sutures were removed. Next two neurovascular surgical clips (Braun Medical) were applied at two separate positions below the ligation for twenty seconds each. Finally the bowel was placed back into the abdominal cavity and the laparotomy was closed. Twenty-four hours after thrombosis induction thrombi that had formed in the IVC were collected and fixed in 10% formalin for over 24 hours and then photographed and weighed. Liver total mRNA was prepared and analyzed for FXI mRNA levels by RT-PCR.

**Ferric Chloride Induced Aortic Thrombosis**

Arterial antithrombotic activity of FXI ASO treatment in mice was studied using a method similar to the FeCl₃ induced IVC model described above with this difference that in this model not the IVC but the descending aorta was exposed to 10% FeCl₃. Thirty minutes after initial FeCl₃ exposure two small hemostats were used to clamp both the descending aorta and IVC. The descending aorta was cut and peeled away from the IVC and the extracted part was processed similar to the IVC tissue described previously.

**Tail Vein Bleeding Assay**

A mouse tail bleeding assay was employed as described. Bleeding was assessed by total blood loss or weight difference before and after bleeding within a 40-minute period, or by bleeding time, determined as time until the bleeding stopped with a maximum recording time of 10 minutes.
**Hepatectomy Surgical Bleeding Model**

Bleeding risk of FXI antisense treatment after a surgical procedure was assessed using a previously described partial hepatectomy model. FXI or FII ASO was dosed sc at 20 mg/kg twice/week for 2.5 weeks, and low molecular weight heparin (Lovenox) was dosed sc at 50 mg/kg once/day on three consecutive days. Two days following ASO treatment or two hours following Lovenox treatment, male BALB/c mice were anesthetized with ketamine/xylazine (100/10 mg/kg, i.p.) and the liver was exposed via a midline abdominal incision. After ligation and resection of the lower left lobe of the liver the body cavity and incision were closed by suture and wound clips respectively. Four hours later health of the mice was evaluated by recording survival and cage conditions, and blood was drawn for red blood cell count, hemoglobin and hematocrit analysis using a clinical analyzer.

**FXI ASO Reversal Studies**

Plasma-derived FXI (Hemoleven, LFB, Lille) was tested for its ability to reverse FXI ASO anticoagulant activity in mice. Male BALB/c mice were treated with saline or FXI ASO (40 mg/kg twice-weekly for three weeks). Two days after final dosing half of the FXI ASO treated mice received Hemoleven (10 µg) intravenously once daily for two consecutive days. APTT was measured 24 hours following the final dose of Hemoleven.

**Anticoagulant Co-administration Studies**

Male BALB/c mice were treated with different doses of Lovenox or the anti-platelet drug clopidogrel either alone or in combination with FXI ASO. Lovenox was injected subcutaneously once daily for three consecutive days, whereas clopidogrel was administered orally, with two loading doses on day one and one single dose on day two. Measurements were performed two hours following dose. In both treatment groups half of the mice additionally received FXI ASO treatment (20 mg/kg) s.c. twice-weekly for 3 weeks. Antithrombotic activity and bleeding tendency were determined using the FeCl₃ induced IVC thrombosis model and the tail vein bleeding assay described above. In addition, bleeding
potential for clopidogrel was also determined in combination with a small molecule FXa inhibitor\textsuperscript{36} (0.5 mg/kg daily on two consecutive days) four hours after the final dose.

**Statistical Analysis**

Statistical comparisons were made by analysis of variance (ANOVA). Student t-test was used for comparison of single pairs. Correction for multiple comparisons between individual groups was made using Fisher's LSD post-hoc test. Data are expressed as mean +/- SEM. \( P \) values < 0.05 were considered significant. Data were analyzed using SPSS software package for Windows version 14.0 (SPSS Inc, Chicago, IL, USA). Graphics were constructed using GraphPad Prism, version 5 for Windows (GraphPad Software, Inc., CA, USA).

**RESULTS**

**ASO Suppression of FXI Levels in Mice**

For initial characterization of FXI ASO activity in mice we measured the effect of FXI ASO treatment on both hepatic FXI mRNA levels and non-targeted coagulation factor mRNA levels (figure 1a), protein levels (figure 1b) and levels of plasma FXI activity (figure 1c). FXI ASO treatment resulted in a dose-dependent reduction of FXI mRNA levels in liver with a maximal reduction of approximately 98% observed at the highest dose tested (figure 1d,e). FXI ASO was highly specific for FXI as all other measured coagulation factor mRNAs remained unchanged, as did the activity of another intrinsic pathway factor, FXII (not shown). Furthermore, reduction in FXI mRNA levels correlated well with a reduction in circulating FXI plasma protein and activity levels (figure 1b,c). Next, we determined the anticoagulant effect of FXI ASO treatment in mice by measuring aPTT and PT ratios. FXI antisense therapy resulted in a dose-dependent prolongation of aPTT (figure 1d). To characterize the kinetics of FXI ASO activity, the onset and duration of action in mice following a single subcutaneous injection (50 mg/kg) was determined by monitoring both hepatic FXI mRNA levels, aPTT and PT. FXI ASO treatment resulted in a time-dependent reduction in FXI mRNA levels with an onset of action of approximately one day following treatment. This activity was maintained at
maximal inhibition (~90%) for days 2-7 which followed by a gradual return to basal levels between days 14-28 (figure 1e). APTT prolongation with no effect on PT correlated well with the decrease in hepatic FXI mRNA levels. APTT started to increase by day 2 post-treatment. Maximal aPTT prolongation was achieved at day 3 and was maintained until approximately day 7 when levels gradually normalized again returning to baseline by day 28 (figure 1e).

These data are consistent with other reports on the duration of action of second generation ASOs in liver and indicate that an approximate 60% reduction in FXI mRNA levels is needed to promote anticoagulant activity in mice, as measured by aPTT prolongation.

**Effect of FXI ASO Treatment on Thrombogenesis and Hemostasis in Mice**

To study the effect of FXI inhibition on thrombus formation and hemostasis, FXI ASO treatment was evaluated in several mouse models of thrombosis and bleeding respectively.

**Ferric Chloride Induced Inferior Vena Cava Thrombosis.** The antithrombotic activity of FXI ASO therapy was first evaluated in a FeCl₃ induced IVC thrombosis model and compared to the effect of warfarin treatment. Within two days after the final FXI ASO injection, thrombosis was induced and PF4 mRNA levels were determined at the lesion site as a measure of thrombosis. Similar to the dose-dependent effects of FXI ASO treatment on hepatic FXI mRNA protein and FXI activity levels (figure 1a,b), FXI ASO therapy also produced a dose-dependent reduction in thrombus formation with maximal effects observed at doses between 5-10 mg/kg (figure 2a). While the observed maximal antithrombotic activity of FXI ASO treatment was similar to that observed with warfarin at doses of 3 mg/kg and higher, the effects of these two treatment strategies on bleeding were remarkably different. As expected, warfarin treatment resulted in marked blood loss in a tail bleeding assay (figure 2b). Moreover, the increase in bleeding caused by warfarin was observed at sub-efficacious doses starting as low as 1 mg/kg. In contrast, FXI ASO treatment did not affect bleeding in mice even at the highest doses tested. The effect of FXI ASO treatment on bleeding evaluated in the tail vein bleeding assay was also compared to that of low molecular weight
heparin Lovenox (figure 2b insert). Compared to control ASO treatment, FXI ASO treatment had no effect on bleeding, whereas Lovenox treatment resulted in markedly prolonged bleeding times (P value <0.001) and exceeded the maximally recorded time of 10 minutes in 50% of the cases (figure 2b, insert).

**Ferric Chloride Induced Mesenteric Vein Thrombosis.** To examine the mechanisms underlying FXI ASO antithrombotic activity, we evaluated the effects of FXI ASO treatment by intravital microscopy in a FeCl₃ induced mesenteric vein thrombosis model as described in methods (see supplementary video). As depicted in figure 3a, mice treated with saline (“untreated”) or control ASO exhibited complete mesenteric vein occlusion within 30 minutes following FeCl₃ injury. In contrast, FXI ASO treatment completely prevented occlusion of the mesenteric vein up to the end of the observation period (40 minutes after FeCl₃ exposure). Although small thrombi (approximately 30 μm in diameter) did initially form near the injured vessel wall in the FXI ASO treatment group (figure 3b), this partial thrombus formation was markedly delayed in time, reduced in size, density and stability, and eventually cleared (figure 3a-b). Continuous video recording using the intravital microscopy system demonstrated that impairment of thrombus growth in the mesenteric vein of FXI ASO treated mice was due to platelet shedding with a continuous break down of small platelet aggregates from the growing thrombi. The small thrombi that initially had formed appeared to detach easily from the vessel wall, leading to complete clearance of the thrombotic lesion at the end of observation (See video in supplemental materials).

**Stenosis Induced Inferior Vena Cava Thrombosis**

A third model that was used to further profile the antithrombotic activity of FXI ASO treatment in mice was a stenosis induced IVC thrombosis model (“St. Tomas’ model”) which uses a combination of reduced blood flow and endothelial damage to induce thrombosis³⁴ (see Materials and Methods). FXI ASO treatment resulted in a dose-dependent reduction in thrombus formation as was shown by a decrease in thrombus size and weight (figure 3c-d).
Furthermore, dose-dependent reduction in thrombus weight correlated well with a dose-dependent reduction in hepatic FXI mRNA levels in treated animals (figure 3d). Finally, the antithrombotic activity of FXI ASO treatment appeared to be specific as no antithrombotic activity was observed in animals treated with control ASO (figure 3c-d).

**Partial Hepatectomy Surgical Bleeding Model.** To further assess the risk for bleeding following FXI ASO treatment, a surgical hepatectomy bleeding model was employed in mice pretreated with either vehicle, FXI ASO, a prothrombin (FII) ASO, or Lovenox as described in Materials and Methods. Treatment with FXI and FII ASO resulted in reductions of over 90% of hepatic FXI and FII mRNA levels respectively (data not shown). All animals in the various treatment groups survived the surgical procedure except for the Lovenox and FII ASO treatment groups where respectively 4 and 1 animals died. General cage inspection revealed obvious bloody cage conditions in the Lovenox and FII ASO treatment groups, whereas cage conditions appeared normal in animals treated with vehicle or FXI ASO. Hematological analysis demonstrated significantly reduced levels of red blood cell count, hemoglobin and hematocrit levels in surviving mice treated with either Lovenox or FII ASO compared to control animals (figure 4). In contrast, no differences were observed in any of these parameters between FXI ASO and vehicle treated animals.

**Reversal of FXI ASO Activity by Plasma-Derived FXI Concentrate**

Despite the absence of bleeding associated with FXI ASO treatment, availability of an effective and convenient antidote strategy is nonetheless desirable. To address this, we evaluated the ability of plasma-derived FXI concentrate (Hemoleven) to reverse FXI antisense mediated effects on aPTT in mice. As expected FXI ASO treatment produced a significant prolongation of aPTT compared to control treated animals (1.28 ± 0.01 versus 1.00 ± 0.01). Treatment with FXI protein concentrate however, rapidly normalized the FXI ASO mediated aPTT prolongation (1.04 ± 0.02) (data not shown).
Antithrombotic Effect and Safety of FXI ASO in Combination with Enoxaparin (Lovenox) or Clopidogrel (Plavix)

Since combination therapy is a relatively common approach to maximize antithrombotic activity in patients, but may have undesirable effects, particularly relating to exacerbation of bleeding complications, we evaluated the effects of FXI ASO in combination with either Lovenox or clopidogrel treatment on thrombosis and bleeding in mice. The antithrombotic activity of Lovenox alone or in combination with FXI ASO treatment was evaluated in the IVC mouse model of venous thrombosis. Lovenox treatment alone resulted in a dose-dependent reduction in thrombus formation, which was significantly improved when combined with FXI ASO, as was demonstrated by a lower maximal effective dose of Lovenox (60 mg/kg versus 30 mg/kg) (figure 5a). As expected, treatment with Lovenox resulted in a dose-dependent increase in bleeding as demonstrated in the tail vein bleeding assay. However, combining Lovenox with FXI ASO therapy did not exacerbate the increase in bleeding produced by Lovenox treatment alone (figure 5b).

To determine the effects of FXI ASO treatment in combination with clopidogrel, a FeCl₃-induced descending aorta model of arterial thrombosis was employed. Treatment with clopidogrel alone resulted in a reduction in thrombus formation which was markedly improved when combined with FXI ASO treatment, as was demonstrated by enhanced antithrombotic efficacy at both the lower dose and the higher dose of FXI ASO treatment, as compared to FXI or clopidogrel treatment alone (figure 6a).

To determine the effects of clopidogrel on bleeding when administered in combination with FXI ASO or a small molecule FXa inhibitor, mice were treated with increasing doses of clopidogrel, either alone or in combination with FXI ASO (20 mg/kg) or a small molecule FXa inhibitor (0.5 mg/kg) and the tail vein bleeding assay was employed. The dose chosen for the FXa small molecule inhibitor was the lowest dose that produced maximum antithrombotic activity in the mouse FeCl₃ IVC model (data not shown). As expected, clopidogrel alone
produced a dose-dependent increase in bleeding time (figure 6b). Moreover, the effect of clopidogrel on bleeding was significantly increased when combined with a small molecule FXa inhibitor (figure 6c). However, combination of clopidogrel with FXI ASO therapy did not significantly exacerbate the increase in bleeding produced by treatment with clopidogrel alone (figure 6b).

DISCUSSION

In this report, we are the first to describe the use of antisense technology to target hepatic-derived coagulation factor XI as a novel treatment strategy for thromboembolic diseases. FXI antisense treatment in mice produced a dose-dependent and highly specific suppression of hepatic FXI mRNA levels. Even at maximal dose, which resulted in a reduction in FXI mRNA levels of approximately 98%, the expression of other, non-targeted liver-derived coagulation factors remained unaffected. Kinetic studies and dose response analysis demonstrated that suppression of FXI mRNA levels correlated well with reduced plasma FXI activity levels and aPTT prolongation without affecting PT values. Following cessation of FXI ASO treatment, FXI mRNA levels, FXI protein and activity levels, and aPTT values, returned to pre-treatment levels without a “rebound effect”, which is probably due to the slow, steady and gradual loss of activity following treatment termination. The absence of this rebound effect following cessation of FXI ASO dosing is an attractive property for anticoagulant treatment as it avoids potential exposure to a prothrombotic condition.

FXI ASO antithrombotic activity was demonstrated in several mouse models of thrombosis, with maximal antithrombotic effects observed with reductions in circulating FXI activity of approximately 70%. Using a FeCl₃ induced IVC thrombosis model, FXI ASO treatment was shown to have an antithrombotic effect similar to that of warfarin. However, unlike warfarin, FXI ASO treatment did not result in increased bleeding. In a FeCl₃ induced mesenteric vein thrombosis model, intravital microscopy analysis demonstrated that FXI ASO therapy prevented vessel occlusion in contrast to mice treated with control ASO or saline, in which
complete occlusion occurred within 20 minutes following FeCl₃ challenge. Although small thrombi did initially form at the site of injury in FXI ASO treated mice, this partial thrombus formation was substantially delayed in time, reduced in size, density and stability, and eventually cleared. This observation suggests that FXI is not involved in the initiation of thrombus formation, but instead, is mainly involved in the propagation and/or stabilization of thrombus growth. This may also explain why inhibition of FXI is not associated with increased bleeding, as this may not affect the initial formation of small thrombi at the site of injury helping to achieve hemostasis, but may limit the propagation and stabilization of a growing thrombus thereby preventing vessel occlusion.

FXI ASO treatment also produced marked antithrombotic activity in a stenosis-induced venous thrombosis model and in an arterial thrombosis model. The demonstration of antithrombotic activity in both arterial and venous models of thrombosis, and under conditions of either local vascular wall injury (FeCl₃) or reduced blood flow (stenosis), demonstrates that targeting FXI has potential utility for the treatment and prevention of a broad range of thrombotic conditions.

Vitamin K antagonists, like warfarin, have several features that limit their therapeutic use, including a narrow therapeutic window and high inter- and intra-patient pharmacokinetic variability resulting in high rates of serious bleeding complications. Low molecular weight heparins have more predictable pharmacokinetic and pharmacologic profiles but require multiple subcutaneous injections every week, and complications such as bleeding events and heparin induced thrombocytopenia remain an issue. In contrast, recent studies suggest that the intrinsic coagulation pathway, and FXI in particular, plays a relatively minor role in normal hemostasis, whereas it plays a significant role in thrombosis, which is further supported by our data. Despite the strong antithrombotic activity of FXI ASO treatment in mice, FXI ASO treatment had no detectable effect on bleeding as demonstrated in both a tail vein bleeding assay and a surgical hepatectomy model. In contrast, treatment
with warfarin or the low molecular weight heparin Lovenox resulted in marked increases in bleeding, which was observed at sub-efficacious doses.

The long tissue half-life of ASOs potentially represents an important advantage for antithrombotic therapy since it can permit reasonable patient convenience based on once weekly or less frequent subcutaneous injection. However, the extended duration of action of ASO treatment further emphasizes the importance of an antidote strategy that effectively and rapidly reverses ASO activity during a bleeding crisis or during acute surgery. Since antisense treatment produces a reduction in the circulating levels of the targeted coagulation factor, simple protein replacement therapy would be expected to produce immediate reversal of anticoagulant activity. Indeed, replacement therapy by plasma-derived FXI concentrate was shown to rapidly and completely reverse the anticoagulant effect of FXI antisense treatment, as demonstrated by normalization of aPTT. The use of plasma or plasma derived coagulation factors as antidotes may also be more straightforward in the case of antisense therapy as only small increases in coagulation factor levels may be needed to normalize activity.

Combination therapy is a relatively common approach to maximize antithrombotic activity in patients who are at risk for thromboembolic events, but may also have undesirable effects, particularly relating to exacerbation of bleeding complications. Lovenox is often used for treatment and prevention of venous thrombosis. The anti-platelet drug clopidogrel (Plavix) effectively inhibits platelet aggregation and is commonly used to prevent arterial thrombosis, including atherothrombotic complications after myocardial infarction, ischemic cerebrovascular events or in patients with acute coronary syndrome. Since both Lovenox and clopidogrel can cause significant bleeding complications, either alone or in combination with other anticoagulant agents, it is important to understand whether FXI inhibition has the potential to exacerbate bleeding complications when used in combination with one of these important antithrombotic modalities. We evaluated the safety and efficacy of FXI ASO
therapy in combination with Lovenox or with clopidogrel which was compared to the combination of clopidogrel with a small molecule FXa inhibitor. Although co-administration of FXI ASO with either Lovenox or clopidogrel resulted in an increase in antithrombotic activity compared to either drug alone, this combined treatment did not result in any increase in bleeding beyond that produced by Lovenox or clopidogrel alone. In contrast, combination of clopidogrel with a small molecule FXa inhibitor resulted in marked increases in bleeding beyond that of clopidogrel therapy alone.

A positive feedback amplification loop involving thrombin and FXI is generally believed to be responsible for the role of FXI in thrombus formation and stabilization\(^3,15,16\). Furthermore, since polyphosphate release from activated platelets can promote thrombus formation \(^40\) and because thrombin induces platelet activation and polyphosphate release, the mechanism underlying the role of FXI in thrombus formation may involve the promotion of polyphosphate release by activated platelets as a result of the positive thrombin - FXI positive feedback loop. Studies addressing the role of platelet polyphosphates in the antithrombotic mechanism of FXI depletion will be the subject of future studies.

In summary, we have described the use of antisense technology to target the hepatic-derived, intrinsic pathway coagulation factor XI in mice as a potentially novel strategy for the treatment or prevention of thromboembolic disease. We conclude that antisense therapy against FXI is effective and safe, both alone and in combination with standard antithrombotic agents, in preclinical models. Further evaluation is warranted to confirm its potential as a novel, effective, and possibly safer alternative for the treatment and prevention of thrombosis in humans.
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AUTHORSHIP CONTRIBUTIONS

B.P. Monia, M. Levi, E.S. Stroes and J.C.M. Meijers conceived and implemented the project strategy; E.C. Löwenberg, H. Zhang, J.C. Crosby, A.R. MacLeod, C. Zhao, D. Gao and C. Black designed and performed in vitro studies and animal studies; E.C. Löwenberg and H. Zhang analyzed the data and wrote the first draft of the manuscript; M. Levi, B.P. Monia and J.C.M. Meijers supervised the research and edited and finalized the manuscript.

CONFLICTS OF INTEREST DISCLOSURES

For EC Löwenberg, JCM Meijers, ES Stroes and M Levi there are no conflicts of interest to disclose. JC Crosby, R MacLeod, C Zhao, D Gao, C Black and BP Monia are all employees of ISIS Pharmaceuticals.

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REFERENCES


FIGURE LEGENDS

Figure 1. Effect of FXI ASO treatment on hepatic FXI mRNA levels, FXI enzymatic activity and aPTT ratio in mice. FXI ASO activity was initially characterized in normal mice by measuring the effect of FXI ASO treatment on hepatic FXI mRNA levels and plasma FXI activity levels. Because FXI has a relatively long half-life (approximately 52 hours in human plasma), a multi-dose regimen with doses ranging from 5 to 50 mg/kg was initially used. Male BALB/c mice were treated subcutaneously with FXI ASO twice-weekly for three weeks at indicated doses (n=4 per dosing group). Three days following final dosing we measured (a) hepatic mRNA levels of FXI as well as non-targeted coagulation factors by RPA, (b) FXI plasma protein levels (c) FXI enzymatic activity levels, and (d and e) aPTT and PT ratios. Onset and duration of hepatic FXI mRNA reduction and corresponding anticoagulation in mice in response to a single injection of FXI ASO (50 mg/kg). APTT = activated partial thromboplastin time; RPA = RNase protection assay. *P≤0.05 compared to untreated control.

Figure 2. Effect of FXI ASO, warfarin or Levonox therapy on thrombosis and bleeding in mice. (a) Male BALB/c mice were treated with either FXI ASO at doses ranging from 1.25-40 mg/kg administered subcutaneously twice-weekly for three weeks, or warfarin which was injected intraperitoneally once daily for a period of 6 days with the final dose injected approximately 4 hours prior to endpoint measurements (n=10). Thrombosis was induced by 3 minute exposure of the inferior vena cava to a 10% FeCl₃ solution and assessed by RT-PCR measurement of platelet factor 4 (PF4) mRNA levels at the site of injury. (b) Effect of FXI antisense and warfarin treatment at indicated doses on tail bleeding in male BALB/c mice (n=10). Inserted figure: Male C57BL/6 mice were treated with either FXI ASO, control ASOs (25 mg/kg, subcutaneously injected twice/week for three weeks), or Lovenox (single dose of 25 mg/kg injected subcutaneously 2.5 hours prior to bleeding measurements) and the effects on tail bleeding were evaluated (n=8, 8 and 6 mice per treatment group, respectively).
Figure 3. Antithrombotic effect of FXI ASO treatment in mouse models of ferric chloride induced mesenteric vein thrombosis and stenosis induced IVC thrombosis. Thrombus formation in male BALB/c mice treated with FXI ASO or control ASO (50 mg/kg, s.c.) twice-weekly for three weeks or without treatment (“untreated”). For testing in the mesenteric vein model, thrombosis was induced by a 3 minute exposure of the mesenteric vein to a 10% FeCl₃ solution following intravenous injection of fluorescein labeled mouse platelets. (a) Continuous recording of venous thrombus formation by intravital microscopy (IVM) for a total period of 40 minutes or until total occlusion (see supplementary video file). (b) Time to formation of a 30 μm thrombus or to total occlusion. *P≤0.05 for FXI ASO treated group compared to control group.

For testing in the stenosis IVC model, 30 male BALB/c mice were treated with either, FXI ASO (dose ranging from 1.25 mg/kg to 40 mg/kg), control ASO (40 mg/kg), or saline by a twice-weekly subcutaneous injection for a period of three weeks. Vena cava thrombosis was induced using a model that combines reduced blood flow with endothelial damage. 24 hours after thrombosis induction thrombi that had formed in the IVC were collected, fixed in formalin, photographed and weighed. Thrombus weight relative to saline control mice was calculated. Liver total mRNA was prepared and analyzed for FXI mRNA levels by RT-PCR (c) Appearance of formed thrombi in the vena cava in different treatment and dosing groups. Of note: thrombus formation in the first mouse of the FXI ASO 40mg/kg treatment group was completely prevented. (d) Correlation of hepatic FXI mRNA levels and relative thrombus weight in mice treated with FXI ASO at indicated doses.

Figure 4. Effects of anticoagulant treatment on bleeding in a mouse hepatectomy surgical bleeding model. Mice were treated with vehicle (saline), FXI or FII ASO (20 mg/kg twice/week for 3 weeks) or Lovenox (50 mg/kg once/day on 3 consecutive days). Two days following ASO treatment or two hours following Lovenox treatment, animals underwent a surgical hepatectomy procedure. Four hours following incision closure, relative bleeding was
determined by measuring red blood cell count, haemoglobin levels and hematocrit values. *P≤0.05 compared to vehicle group with surgery.

Figure 5. Effect of FXI antisense treatment alone or in combination with Lovenox on thrombosis and bleeding in mice. Mice (n=12/group) were treated with indicated doses of Lovenox alone (once daily for three consecutive days) or in combination with FXI ASO treatment (20 mg/kg twice/week for three weeks). Effects on FeCl₃-induced inferior vena cava thrombosis (a), or tail vein bleeding (b), were determined two hours following the final dose of Lovenox, as described in methods *p<0.05.

Figure 6. Antithrombotic activity and bleeding evaluation of FXI ASO alone or in combination with clopidogrel in an aortic arterial thrombosis model. As described in methods mice were treated with saline ("0") or FXI ASO (dosed 10 mg/kg or 20 mg/kg twice weekly for three weeks), either alone or in combination with clopidogrel (3 mg/kg), and thrombosis was induced 4 hours following the final dose of clopidogrel by exposing the descending aorta to a ferric chloride solution (a). Effects of FXI ASO or FXa small molecule inhibitor in combination with clopidogrel on bleeding in mice. As described in methods mice were treated with increasing dose levels of clopidogrel as indicated, either alone, or in combination with FXI ASO (20 mg/kg) (b), or in combination with a FXa small molecule inhibitor (0.5 mg/kg) (c) and bleeding was determined using a mouse tail vein bleeding assay.
Figure 1

A

FXI ASO (mg/kg)

TPO
FVIII
FXI
FVII
FIX
FXII
FX
FII
FV
TAFI

B

FXI protein (% of saline)

0 5 10 25 50

C

FXI Activity (% of saline)

0 5 10 25 50

D

FXI mRNA (% of saline)

FXI ASO (mg/kg)

E

FXI mRNA (% of saline)

Days

RNA aPTT PT

PT/aPTT INR

0 1 2 3 4 5 6 7 8 9 10 11 12 13 14 15 16 17 18 19 20 21 22 23 24 25 26 27 28 29 30 31 32 33 34 35 36 37 38 39 40 41 42 43 44 45 46 47 48 49 50 51 52 53 54 55 56

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Figure 2

A

Thrombosis (PF4 RNA-% Control)

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<th>Saline H</th>
<th>FeCl₃</th>
<th>5</th>
<th>4</th>
<th>3</th>
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B

Blood loss (g)

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Inset: Bleeding time (min)

Control ASO: 100, FXI ASO: 200, Clexane: 500
Figure 3

A

![Images showing the effects of different treatments on thrombus formation over time.](image)

B

**Time to form 30 µm thrombus**

- Control
- FXI ASO

*P* < 0.05

**Time to total occlusion**

- Control
- FXI ASO

*P* < 0.001

C

<table>
<thead>
<tr>
<th>Treatment</th>
<th>Liver FXI RNA, %</th>
<th>Thrombus Weight, Ratio</th>
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D

![Graph showing the effect of FXI ASO on liver FXI RNA and thrombus weight.](image)
Figure 4

**RBC**

Cells (X10^6/µL)

- Vehicle
- Vehicle
- FXI
- FII
- Lovenox

**Hemoglobin**

g/dL

- Vehicle
- Vehicle
- FXI
- FII
- Lovenox

**Hematocrit**

% 

- Vehicle
- Vehicle
- FXI
- FII
- Lovenox
Inhibition of the intrinsic coagulation pathway factor XI by antisense oligonucleotides: a novel antithrombotic strategy with lowered bleeding risk

Hong Zhang, Ester C. Löwenberg, Jeffrey R. Crosby, A. Robert MacLeod, Chenguang Zhao, Dacao Gao, Chris Black, Alexey S. Revenko, Joost C.M. Meijers, Erik S. Stroes, Marcel Levi and Brett P. Monia