Residual platelet factor V ensures thrombin generation in patients with severe congenital factor V deficiency and mild bleeding symptoms

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Abstract

Coagulation factor V (FV), present in plasma and platelets, is indispensable to thrombin formation, yet patients with undetectable plasma FV seldom experience major bleeding. We used thrombin generation assays to explore the role of platelet FV in four patients with severe congenital FV deficiency (three with plasma FV:C <1%). When triggered with tissue factor (TF) concentrations up to 50 pM, platelet-poor plasma (PPP) from the patients with undetectable plasma FV showed no thrombin generation, whereas platelet-rich plasma (PRP) formed thrombin already at 1-5 pM TF. Thrombin generation in PRP from the FV-deficient patients was enhanced to near-normal levels by platelet activators (collagen or Ca^{2+}-ionophore) and could be completely suppressed by specific FV inhibitors, suggesting FV-dependence. Accordingly, platelet FV antigen and activity were measurable in all FV-deficient patients and platelet FVa could be visualized by Western blotting. Normalization of the tissue factor pathway inhibitor (TFPI) level, which is physiologically low in FV-deficient plasma, almost completely abolished thrombin generation in PRP from the FV-deficient patients. In conclusion, patients with undetectable plasma FV may contain functional FV in their platelets. In combination with the low TFPI level, residual platelet FV allows sufficient thrombin generation to rescue these patients from fatal bleeding.
Coagulation factor V (FV) is a 330-kDa glycoprotein which is present in plasma (80%) and platelets (20%). Plasma FV (20-25 nM) is synthesized in the liver and is released in the blood stream as a single-chain inactive procofactor. Following limited proteolysis by thrombin or activated factor X (FXa), FV is converted to its activated form (FVa), consisting of a heavy chain (105 kDa) and a light chain (71/74 kDa) that are non-covalently linked via a single Ca$^{2+}$ ion. FVa acts as a non-enzymatic cofactor of FXa in the conversion of prothrombin to thrombin and increases the rate of prothrombin activation more than 1000-fold. FVa activity is down-regulated by activated protein C (APC), which inactivates FVa by cleaving the heavy chain at Arg$^{306}$, Arg$^{506}$ and Arg$^{679}$. Although megakaryocytes are capable of FV synthesis, platelet FV originates from the plasma pool. Bone-marrow megakaryocytes internalize plasma FV via a specific receptor-mediated process and store it in secretory α-granules. After endocytosis, FV gains several unique modifications that make platelet FV structurally and functionally different from plasma FV. In particular, platelet FV is stored in a partially activated form which, after exposure on the platelet surface, is further activated by FXa or thrombin and is resistant to APC-catalysed inactivation.

Congenital FV deficiency (Owren parahaemophilia), caused by loss-of-function mutations in the F5 gene, is a rare (1:10$^6$) bleeding disorder inherited as an autosomal recessive trait. Homozygous and compound heterozygous individuals have FV levels <10% and present with a life-long haemorrhagic diathesis whose severity is poorly correlated with the plasma FV level. Many patients with undetectable FV experience only...
mild/moderate bleeding\textsuperscript{21} and do not require routine prophylaxis. The reason for the relatively mild clinical presentation of many FV-deficient patients is presently unknown.

Given the uniform lethality of FV-null mice\textsuperscript{25} it has been argued that FV-deficient individuals who survive to post-natal life always have some residual FV expression.\textsuperscript{19,25,26} Since the FV requirement for minimal thrombin generation is well below 1\%,\textsuperscript{26-29} traces of FV would already be sufficient to guarantee thrombin formation and to rescue FV-deficient individuals from fatal bleeding. However, \textit{in vitro} experiments have failed to detect any thrombin generation in plasma from patients with undetectable FV,\textsuperscript{29,30} despite the fact that FV-deficient patients have low plasma levels of the anticoagulant protein tissue factor pathway inhibitor (TFPI),\textsuperscript{29} which considerably reduces the FV requirement for thrombin generation.

Some 30 years ago, Miletich \textit{et al.}\textsuperscript{31} showed that, in patients with severe FV deficiency, the FXa-binding capacity of platelets (which is a measure of platelet FV) is a better predictor of bleeding tendency than the plasma FV level. Despite this important observation, platelet FV is not routinely evaluated in FV-deficient patients and only three other studies report platelet FV levels in patients with severe FV deficiency.\textsuperscript{32-34} No platelet FV antigen or activity could be demonstrated in two patients with undetectable plasma FV.\textsuperscript{32,33} In another FV-deficient patient platelet FV could be visualised by Western blotting, but its activity was not determined.\textsuperscript{34} To get more insight into the role of platelet FV in FV deficiency, we have measured thrombin generation and platelet FV levels in four patients with severe congenital FV deficiency.
Materials and Methods

Patients

Four unrelated female patients with severe congenital FV deficiency were enrolled. All four were referred to Padua Academic Hospital from district hospitals in North-Eastern Italy and their demographic and clinical characteristics have been previously described. Briefly, patient PD I (age 64 years, FV clotting activity (FV:C) <1%) was diagnosed at the age of 5 years because of recurrent epistaxis and gum bleeding. Immediately after the menarche at the age of 8 years, she presented with abundant menses. During the post-partum period of her only pregnancy, she developed three severe haemorrhages, which were treated by transfusion with fresh frozen plasma, platelets and red blood cell concentrates. However, she never experienced major spontaneous bleeding. Patient PD II (age 44 years, FV:C <1%) suffered from recurrent epistaxis in childhood, which led to the diagnosis of severe FV deficiency at the age of 8 years. At the age of 15 years, she presented with excessive bleeding after a tooth extraction. She has always had abundant menses and in her thirties she was admitted to hospital twice for severe metrorrhagia. On both occasions, administration of fresh frozen plasma was effective in controlling the haemorrhage. Her parents are first degree cousins. Patient PD III (age 35 years, FV:C <1%) experienced only very mild episodes of epistaxis and gum bleeding and was diagnosed at the age of 8 years. Her occasional abundant menses have worsened lately because of the presence of uterine fibromatosis. In order to reduce menstrual bleeding, she was prescribed oral contraceptives, but the treatment had to be stopped after a few cycles because of her intolerance to the drug. Fresh frozen plasma and/or antifibrinolytic agents were given during all risk situations and bleeding complications never ensued. Patient PD VII (age 62 years, FV:C 6%) has experienced only mild menorrhagia. She was diagnosed
at the age of 23 years, when she presented with a post-partum haemorrhage requiring transfusion with fresh frozen plasma. She also experienced one uncomplicated spontaneous abortion. None of the patients received substitutive therapy in the three months preceding blood sampling. All patients were homozygous or compound heterozygous for missense mutations in the F5 gene (Table 1).35-37

The study was approved by the Ethics Committee of Padua Academic Hospital and conducted in accordance with the Declaration of Helsinki. All patients gave informed consent to participation. Eight healthy volunteers recruited among hospital employees served as normal controls.

**Blood collection and plasma preparation**

Blood was collected from each FV-deficient patient on two separate occasions, with an interval of ~6 months. On each occasion, 40 mL blood were drawn in 129 mM sodium citrate (1/10 vol/vol) for thrombin generation experiments, and 20 mL blood were drawn in 80 mM sodium citrate, 52 mM citric acid, 183 mM glucose (ACD, 1/7 vol/vol) for platelet isolation (see below). For each patient, blood from two normal controls was also collected on the same day and handled in the same way.

Citrated blood was centrifuged at 250×g for 15 min to obtain platelet-rich plasma (PRP). Part of the PRP was further centrifuged at 14,000×g for 5 min to yield platelet-poor plasma (PPP).

The FV-deficient plasma employed in the experiment shown in Figure 1 was purchased from George King Biochemical Inc. (Overland Park KS, USA).

**Thrombin generation assays**
Thrombin generation in PPP and PRP was determined using the Calibrated Automated Thrombogram method. Coagulation was triggered by recalcification (16 mM added CaCl₂) in the presence of recombinant tissue factor (Innovin, Dade-Behring, Marburg, Germany). The TF concentration in the Innovin stock solution (prepared according to the manufacturer’s instructions) was measured with the Imubind TF-ELISA (American Diagnostica, Stamford CT, USA) and found to be 331 ng/ml (7.36 nM). For the measurement in PPP, synthetic phospholipids composed of phosphatidylcholine, phosphatidylerine and sphingomyelin (Phospholipid-TGT, Rossix, Mölndal, Sweden) were added to a final concentration of 20 μM. PRP was adjusted to 1.5*10⁸ platelets/mL. In some experiments, PRP was pre-incubated with collagen (10 μg/mL, Dynamyte Medical, München, Germany) or Ca²⁺-ionophore A23187 (20 μM, Sigma, Buchs, Switzerland) for 10 min at 37 °C to activate the platelets before triggering coagulation. To prevent thrombin formation via the intrinsic pathway, all thrombin generation experiments were performed in the presence of 32 μg/mL corn trypsin inhibitor (Haematologic Technologies Inc., Essex Junction VT, USA). Thrombin generation curves were determined in duplicate and were calibrated against the fluorescence signal obtained in the same plasma with 100 nM Thrombin Calibrator (Thrombinscope BV, Maastricht, The Netherlands). Fluorescence was read in a Fluoroskan Ascent reader (Thermo Labsystems, Helsinki, Finland) and thrombin generation curves were analysed for lag time, peak height and endogenous thrombin potential (ETP) using the Thrombinscope software (Thrombinscope BV).

In some thrombin generation experiments, an anti-FV polyclonal antibody (SAFV-IG, Affinity Biologicals Inc., Ancaster ON, Canada), plasma-purified human APC (Innovative Research, Southfield MI, USA), in-house plasma-purified FV or recombinant
human full-length TFPI (kind gift of Prof. W. Buurman, Maastricht University) were added to the plasma.

**Platelet isolation and preparation**

Washed platelet suspensions were prepared as described previously\(^1\) and divided in two aliquots: one, with a concentration of \(0.7 \times 10^9\) platelets/mL, was frozen for the preparation of platelet lysates; the other was diluted to \(0.5 \times 10^9\) platelets/mL in platelet buffer (PB, 10 mM Hapes, 136 mM NaCl, 2.7 mM KCl, 2 mM MgCl\(_2\), pH 7.5, containing 2 mg/mL bovine serum albumin (BSA) and 2 mg/mL glucose) and used for the preparation of activated platelet suspensions. Platelet lysates were used for the measurement of FV and TFPI antigen levels, whereas activated platelet suspensions represented the starting material for platelet FV\(a\) activity determination and platelet FV\(a\) immunoprecipitation.

For the preparation of platelet lysates, 0.5% (vol/vol) Triton X100 (Fluka, Buchs, Switzerland) was added to the frozen platelet suspension in the presence of the following protease inhibitors: 5 mM EDTA (Merck, Darmstadt, Germany), 2 mM 4-(2-aminoethyl)benzenesulfonyl fluoride hydrochloride (AEBSF, Fluka), 5 mM N-ethylmaleimide (NEM, Merck), 10 \(\mu\)g/mL leupeptin (Sigma), 10 mM benzamidine (Sigma) and 25 \(\mu\)g/mL soybean trypsin inhibitor (SBTI, Sigma). After 2 hours lysis on ice, platelet lysates were analyzed immediately for FV and TFPI antigen levels.

For the preparation of activated platelet suspensions, Arg-Gly-Asp-Ser-peptide (H-1155, BACHEM AG, Bubendorf, Switzerland) was added to the washed platelet suspensions at a final concentration of 0.3 mg/mL to inhibit platelet aggregation, and platelets \((0.5 \times 10^9)/\text{mL in PB pH 7.5}) were activated with 34 nM thrombin and 20 \(\mu\)M Ca\(^{2+}\)-ionophore (A23187) in the presence of 5 mM CaCl\(_2\) for 30 min at 37 °C. Added thrombin
was subsequently neutralized with hirudin (Kordia Life Sciences, Leiden, The Netherlands). Activated platelet suspensions were frozen at -80 °C for later determination of FV activity.

Measurement of FV antigen levels in plasma and platelets

Plasma and platelet FV antigen levels were quantified with an ELISA. Microtiter plates were coated overnight at 4 °C with 2.2 μg/well polyclonal anti-FV antibody (The Binding Site Ltd, Birmingham, UK) in carbonate buffer. After rinsing 5 times with washing buffer (0.05 M Tris/0.15 M NaCl/5 mM EDTA, pH 7.5, 0.1% Tween 20), 100 μL 1:200-diluted plasma or platelet lysate (2.5*10^7 or 5*10^7 platelets/mL in washing buffer) was added to the wells and incubated at room temperature (RT) for 2 hours. After washing, bound FV was detected by incubation with an HRP-conjugated polyclonal anti-FV antibody (The Binding Site Ltd, 0.5 μg/well) for 1.5 hours, and plates were developed with tetramethylbenzidine/peroxide (KPL, Gaithersburg MD, USA).

Measurement of FV activity levels in plasma and platelets

FV activity levels of plasma and washed platelets were determined with prothrombinase-based assays. Plasma FV was determined as previously described.\(^{29}\) For platelet FV determination, activated platelet suspensions were thawed at 37 °C and diluted with Hepes-buffered saline (HBS, 25 mM Hepes, 175 mM NaCl, pH 7.7) containing 0.5 mg/mL ovalbumin and 2.7 mM CaCl\(_2\) to obtain a final concentration of 12*10^6 platelets/mL in the assay. Platelet FVa was quantified with 5 nM FXa, 1 μM prothrombin, 40 μM di-oleoyl phosphatidylserine/di-oleoyl phosphatidylcholine (10/90 mol/mol) phospholipid vesicles and 2.5 mM CaCl\(_2\). Pooled normal plasma and an activated platelet
pool prepared from platelets from 20 healthy individuals were used as a reference for plasma and platelet FV measurements, respectively.

**Immunoprecipitation of FVa from plasma and platelets**

Activated platelet suspensions (individual patient samples and a pool of activated platelets from 20 controls) were solubilized by the addition of 0.5% (final concentration) Triton X100 in the presence of protease inhibitors (2 mM AEBSF, 5 mM NEM, 10 μg/mL leupeptin, 10 mM benzamidine, 25 μg/mL SBTI) and 5 mM CaCl₂ for 30 min at RT.

FV-deficient or normal pooled plasma was defibrinated with 1 U/mL Ancrod (NIBSC, Potters Bar, UK) for 10 min at 37 °C. Plasma FV was activated with 34 nM thrombin and 16 mM added CaCl₂ in the presence of a polyclonal sheep anti-protein C antibody (0.16 mg/mL, Innovative Research) for 10 min at 37 °C, after which the same cocktail of protease inhibitors as used for the platelet preparation was added.

Platelet lysates and plasma were pre-cleared of endogenous IgG with protein A-Sepharose beads (rProtein A Sepharose Fast Flow; GE Healthcare, Uppsala, Sweden), that were subsequently removed by centrifugation. 200 μL pre-cleared platelet lysate (1.5*10⁸ platelets/mL) or pre-cleared plasma was added to protein A-Sepharose beads (15 μL drained volume) bearing monoclonal anti-FV heavy chain antibodies (3B1, a kind gift from Prof B. N. Bouma, Utrecht University Hospital, Utrecht, The Netherlands) and FVa was immunoprecipitated for 30 min at RT under rotation. The beads were collected by centrifugation, washed 3 times with HBS containing 5 mM CaCl₂ and 10 mM benzamidine and resuspended in HBS containing 1/2 volume sample buffer (40 mM Tris, pH 6.7, 3.33% SDS, 6.25% mercaptoethanol, 50% glycerol, 0.01% bromophenol blue). Plasma or platelet samples from FV-deficient patients were resuspended in 33.3 μL (to achieve a 6-fold concentration of the FVa), while control samples were resuspended in
200 μL (no concentration). After 5 min incubation at 96 °C, samples were subjected to SDS-PAGE on 6% polyacrylamide gels according to Laemmli under reducing conditions. Proteins were subsequently transferred to PVDF membranes by semi-dry blotting. Membranes were blocked with 5% (wt/vol) skim milk (Merck) in HBS/0.5% Tween 20 for 1 h at RT and the primary antibody (monoclonal anti-FV heavy chain AHV-5146 (Haematologic Technologies Inc.), 5 μg/mL in blocking buffer) was incubated overnight at 4 °C under shaking. Membranes were washed with HBS/0.3% Tween 20 and incubated with a secondary antibody (HRP-conjugated rabbit anti-mouse immunoglobulins (DAKO), 1:2000 in blocking buffer) for 1 hour at RT. HRP activity was detected using chemiluminescence (Supersignal West Pico Chemiluminescent Substrate, Pierce, Rockford, IL) and the LAS-3000 image analyzer (Fujifilm, Tokyo, Japan).

**TFPI ELISA**

TFPI levels were determined by a full-length TFPI ELISA in plasma as well as in platelet lysates containing 10^8 platelets/mL. The ELISA was calibrated against normal pooled plasma or a pool of lysed platelets from 20 healthy individuals.

**Statistical analysis**

Data are expressed as mean ± standard deviation (SD). Correlations are expressed as Pearson coefficients (r). Statistical analysis was performed using SPSS 15.0 (SPSS, Chicago, IL).
Results

Contributions of plasma and platelet FV to thrombin generation

To explore the relative contributions of plasma and platelet FV to thrombin generation, FV-deficient plasma with undetectable FV was supplemented with normal platelets and with increasing concentrations (0-100%) of purified plasma FV. After pre-activating the platelets with collagen, coagulation was triggered with 5 pM TF. Thrombin generation was already optimal in the absence of plasma FV and did not increase further when the concentration of plasma FV was increased up to 100% (Figure 1). This experiment shows that in vitro thrombin generation in the presence of normal platelets does not require plasma FV, in line with the in vivo observation that platelet FV could maintain haemostasis in a patient with an acquired antibody that fully neutralised plasma FV.39

Thrombin generation in PPP and PRP from FV-deficient patients

To investigate the role of platelet FV in congenital FV deficiency, four patients with severe congenital FV deficiency were enrolled. All were homozygous or compound heterozygous for missense mutations in the F5 gene (Table 1). Plasma FV clotting activity (FV:C) was undetectable in all patients, except PD VII (FV:C 6%).

Thrombin generation was determined in PPP and PRP from these patients after triggering coagulation with 1, 5, 10 or 50 pM TF. Thrombin generation curves obtained in plasma from the four patients and one representative control are shown in Figure 2. Thrombin generation parameters obtained at 5 pM TF are presented in Table 2.

In control PPP and PRP, thrombin generation was already appreciable at 1 pM TF and increased at increasing TF concentrations. The lag time of thrombin generation
decreased at increasing TF concentrations and at 50 pM TF it became too short to be measurable. At each TF concentration, thrombin generation was lower in PRP than in PPP, most probably because activated platelets provide a less favourable phospholipid surface for coagulation reactions than the synthetic phospholipid vesicles added to PPP.

No thrombin generation was observed in PPP from patients PD I, PD II, and PD III at any of the TF concentrations tested. PPP from patient PD VII showed thrombin generation at TF concentrations ≥5 pM, although this was lower and started later in time than in control plasma. In contrast to PPP, PRP from all four FV-deficient patients showed thrombin generation, suggesting the presence of some residual FV in the patients’ platelets. Thrombin generation was already measurable at 1 pM TF in PRP from patients PD II and PD VII and at 5 pM TF in PRP from patients PD I and PD III.

When PRP was pre-incubated with collagen to activate platelets before initiating coagulation, thrombin generation was higher and in the FV-deficient patients it also started earlier than in the absence of platelet agonists. In fact, thrombin generation was already observed at 1 pM TF in PRP from all four FV-deficient patients. Increasing TF concentrations progressively decreased the lag time, but had no effect on the peak height or ETP.

Pre-incubation of PRP with Ca$^{2+}$-ionophore yielded higher and earlier thrombin formation than pre-incubation with collagen. Also under these conditions, the lag time decreased at increasing TF concentrations, while the amount of thrombin formed (peak height and ETP) hardly changed.

Remarkably, in the presence of collagen or Ca$^{2+}$-ionophore, thrombin generation in PRP from the FV-deficient patients was almost as high as or even higher (PD VII) than in control PRP.
FV-dependence of thrombin generation in PRP from the FV-deficient patients

To investigate whether thrombin generation observed in PRP from the FV-deficient patients was FV-dependent, thrombin generation was determined in the patients’ PRP in the presence of specific inhibitors of FV(a), *i.e.* an anti-FV antibody or APC.

When coagulation was triggered with 50 pM TF in the presence of Ca$^{2+}$-ionophore, the addition of an anti-FV antibody completely inhibited thrombin generation in FV-deficient PRP (Figure 3A), whereas it had no effect on thrombin generation in control PRP (data not shown).

Since APC concentrations up to 200 nM had no effect on thrombin generation in PRP triggered with 50 pM TF in the presence of Ca$^{2+}$-ionophore, the APC-sensitivity of thrombin generation in PRP was tested at 5 pM TF in the presence of Ca$^{2+}$-ionophore. Under these conditions, thrombin generation in FV-deficient PRP was completely inhibited by 50 nM APC (Figure 3B). Thrombin generation in control PRP could also be fully inhibited by APC, but a considerably higher APC concentration (200 nM) was required (data not shown).

FV antigen and activity levels in plasma and platelets

The fact that thrombin generation was observed in PRP, but not in PPP, from the FV-deficient patients suggested that their platelets contained FV. To verify this, FV antigen and activity levels were measured in washed platelets from the FV-deficient patients and eight healthy controls, and compared to plasma FV levels (Table 3).

Although FV antigen was detectable in plasma from all four FV-deficient patients, plasma FV activity was below detection limit in all FV-deficient patients except PD VII (4.4%). In platelets, not only FV antigen but also FV activity was measurable in all four FV-deficient patients, confirming the presence of functional platelet FV even in the
patients with undetectable plasma FV activity. Although platelet FV antigen and activity levels did not show a correlation in the FV-deficient patients, they were well correlated in the normal controls \((r = 0.94, p = .002)\). Moreover, in spite of the very small sample size, platelet FV activity levels in FV-deficient patients correlated \((r = 0.91, p = .09)\) with the peak height of thrombin generation determined in the respective PRPs in the presence of Ca\(^{2+}\)-ionophore, a condition where all platelet FV(a) is exposed.

### Immunoprecipitation of plasma and platelet FVa

To visualize any residual FV present in plasma and/or platelets from the FV-deficient patients, FV was concentrated from plasma and platelets by immunoprecipitation and detected by Western blotting (Figure 4). Since plasma FV is structurally different from platelet FV, whereas the corresponding activated forms are identical,\(^{40}\) activated plasma and platelets (containing FVa) were used as a starting material for the immunoprecipitation. As shown in Figure 4A, no FVa was detectable in PPP from the FV-deficient patients, except for PD VII. In contrast, all patients showed FVa on the Western blot of activated platelet lysates (Figure 4B). Since an anti-heavy chain antibody was used for the detection of FVa, only the heavy chain is visible on the Western blot. This appears as a single band of 105 kDa in plasma FVa, and as multiple bands in platelet FVa. The additional bands of 75/80 kDa observed in platelet FVa represent degradation products of the FVa heavy chain which are probably generated during platelet activation and/or immunoprecipitation.\(^{41,42}\)

### Platelet TFPI level

We have recently shown that full-length TFPI is markedly reduced in FV-deficient plasma,\(^{29}\) a finding that was confirmed in this study (Table 4). Since platelets also contain...
full-length TFPI\textsuperscript{43,44} we measured platelet TFPI antigen in the FV-deficient patients (Table 4). Platelet TFPI levels showed a large inter-individual variation both in controls and in FV-deficient patients. On average, platelet TFPI levels were slightly but not significantly lower in FV-deficient patients (58.3±19.9\%) than in normal controls (77.4±31.7\%).

Effect of TFPI on thrombin generation in PRP
To verify whether the low plasma levels of full-length TFPI also improve thrombin generation in FV-deficient PRP (as they do in PPP),\textsuperscript{29} increasing concentrations of TFPI (0.8 to 14.8 ng/mL) were added to PRP from patient PD II and to control PRP, and thrombin generation was initiated with 50 pM TF in the presence of collagen. In control plasma, addition of TFPI increased the lag time of thrombin generation from 4.2 min to 8.0 min, but did not affect peak height (Figure 5). Differently, in FV-deficient PRP, increasing the TFPI concentration not only prolonged the lag time (from 14.3 min to 29.5 min), but it also profoundly reduced the amount of thrombin formed (Figure 5). The addition of 7.8 ng/mL TFPI, a concentration which normalises the plasma TFPI level in FV-deficient PRP, almost completely abolished thrombin generation in FV-deficient PRP, which confirms the beneficial effect of low plasma levels of TFPI for thrombin generation in FV-deficient patients.
Discussion

Despite the essential role of FV in prothrombin activation, many individuals with severe congenital FV deficiency experience only mild-to-moderate bleeding.\textsuperscript{21} The present study provides an explanation for this paradox by showing that patients with undetectable plasma FV may contain enough functional FV in their platelets to guarantee thrombin generation and protect them against major bleeding.

Four patients with severe congenital FV deficiency and relatively mild bleeding symptoms were investigated. While no thrombin generation was observed in PPP from the three patients with undetectable plasma FV even when plasma was triggered with 50 pM TF, the corresponding PRP showed appreciable thrombin generation already at 5 pM TF. Thrombin generation in PRP from the FV-deficient patients was already observed in the absence of platelet agonists and was greatly stimulated by pre-activating the platelets with collagen or Ca\textsuperscript{2+}-ionophore. The progressive increase in the amount of thrombin formed going from no platelet agonist to collagen and Ca\textsuperscript{2+}-ionophore probably reflects the extent of FV secretion and the parallel exposure of procoagulant phospholipids (phosphatidylserine) on the outer platelet membrane surface.\textsuperscript{45,46} Remarkably, the total amount of thrombin formed in PRP from FV-deficient patients stimulated with collagen or Ca\textsuperscript{2+}-ionophore was similar to that of controls, although the lag time of thrombin generation was considerably prolonged. Moreover, when PRP from the FV-deficient patients was activated with collagen or Ca\textsuperscript{2+}-ionophore, the amount of thrombin formed was independent of the TF concentration, suggesting that a fixed prothrombinase activity was generated at all TF concentrations. A possible explanation for this observation is that the amount of platelet FV(a) that is released is the limiting factor for prothrombinase
activity in FV-deficient PRP, although more factor X (FX) is activated at increasing TF concentrations.

To prove that thrombin generation in PRP from the FV-deficient patients was dependent on platelet FV and not caused by an unknown platelet protein capable of stimulating prothrombin activation, we showed that thrombin generation in PRP from FV-deficient patients could be completely abolished by two specific inhibitors of FV, an anti-FV antibody and APC. Thrombin generation in control plasma was much less sensitive to the FV antibody and to APC, which is likely caused by the excessive amount of FVa in combination with the reduced sensitivity of platelet FVa for APC.14-16

The presence of FV in platelets from the FV-deficient patients was confirmed in several ways. First, platelet preparations from all four patients showed FV activity in a FVa activity assay, i.e. they were able to stimulate FXa-catalysed prothrombin activation in a model system. Second, platelet FVa from the FV-deficient patients could be visualised on Western blot. In addition to the 105-kDa band, corresponding to the full-length heavy chain, platelet FVa showed several smaller fragments on the Western blot. These fragments represent degradation products of the FVa heavy chain that are probably generated during platelet activation47 and/or FVa immunoprecipitation, as many platelet proteases, including calpain41 and lysosomal enzymes,42 can cleave FV(a). Finally, FV antigen was also detectable in the patients’ platelets by an ELISA. Although FV antigen was also measurable in the patients’ plasma, this is most likely non-functional as the patients’ PPP showed no FV activity in the prothrombinase-based and thrombin generation assays (except PD VII). The rather poor correlation between the amount of platelet FV antigen measured by ELISA and that visible on western blot (e.g. for PD III) might be due to interference of the F5 mutations with the recognition of FV by the different antibodies used in the ELISA, immunoprecipitation and western blot.
The presence of functional FV in platelets from patients with congenital FV deficiency can only be explained if the underlying F5 gene mutations are compatible with some residual FV expression. Remarkably, all patients under study carried F5 missense mutations as the underlying cause of FV deficiency. Although all amino acid substitutions involved charge changes at highly conserved residues, it should be noted that missense mutations do not per se prevent protein synthesis, but rather impair the folding, secretion and/or stability of the mutant protein. Moreover, being point mutations, their effects can be abolished by rare somatic reversion events or by ribosome slippage during mRNA translation. Therefore, they can hardly be considered truly “null” mutations. Although it has been argued that all patients with severe FV deficiency actually retain some minimal FV expression, as suggested by the uniform lethality of FV-knock out mice and by the absence of gross deletions from the F5 mutational spectrum, further studies are needed to verify the presence of functional platelet FV in patients with F5 molecular defects other than missense mutations. As a matter of fact, no functional FV could be demonstrated in plasma or platelets from two young patients with severe FV deficiency caused by short out-of-frame deletions/insertions and from two other patients with unknown F5 mutations. Patients with undetectable platelet FV tend to have more severe clinical manifestations than the patients in our study and their PRP would be expected to generate less or no thrombin.

The mechanism underlying the preferential localization of FV in platelets rather than in plasma from patients with severe FV deficiency remains unclear. One possibility is that megakaryocytes take up all FV available in plasma, simultaneously depleting the plasma FV pool. This would imply a high-affinity receptor for FV endocytosis. Alternatively, minimal FV synthesis might occur in megakaryocytes as well as in the liver, but plasma FV might be more rapidly cleared than the FV stored in platelets. This
hypothesis is supported by the observation that platelet concentrates confer FV-deficient patients a longer-lasting protection from bleeding than fresh-frozen plasma, suggesting that platelet FV has a longer half-life than plasma FV.48

Since the FV requirement for minimal haemostasis is <1%, the small amount of FV found in platelets may well be sufficient to explain the relatively mild clinical phenotype of our patients. In fact, the specific characteristics of platelet FV (rapid activation by FXa and resistance to APC-catalysed inactivation)14,40 and its targeted release at the site of injury, resulting in high local concentrations,39 make platelet FV a very efficient procoagulant, able to maintain haemostasis even in the absence of plasma FV. This concept is illustrated by the in vitro observation that, in the presence of normal activated platelets (and thus of platelet FVa), plasma FV does not contribute to thrombin generation (Figure 1). Moreover, in a patient with a circulating FV inhibitor and undetectable plasma FV, platelet FV proved sufficient to ensure adequate haemostasis even during surgery.39 The pivotal role of platelet FV is further supported by the bleeding diathesis associated with FV New York,49 a selective deficiency of platelet FV with normal levels of plasma FV.

As we have recently reported,29 FV deficiency is associated with markedly reduced plasma levels of full-length TFPI, a condition which reduces the FV requirement for minimal thrombin generation. In the present study we show that normalisation of the plasma TFPI level almost completely abolishes thrombin generation in PRP from FV-deficient patients. This effect of TFPI is likely due to inhibition of the initiation phase of coagulation, since physiological concentrations TFPI do not inhibit prothrombin activation.50 The low TFPI level contributes to enhance thrombin generation in patients with severe FV deficiency and may explain the near-normal thrombin generation observed in collagen- and Ca²⁺-ionophore-stimulated PRP from these patients (Figure 2, Table 2).
In conclusion, this study demonstrates that patients with congenital FV deficiency and undetectable plasma FV may contain functional FV in their platelets. In combination with the low TFPI level, residual platelet FV supports enough thrombin generation to rescue patients with undetectable plasma FV from fatal haemorrhage. These findings further suggest that differences in residual platelet FV level, possibly determined by the nature of the underlying molecular defect, may be responsible for the variation in clinical phenotype observed among patients with equally undetectable plasma FV levels. Therefore, platelet FV should be routinely evaluated for a more accurate estimate of bleeding tendency in these patients.
Acknowledgements

We are grateful to the FV-deficient patients for their enthusiastic participation in this study. We wish to thank Dr. E. Zanon from the Hemophilia Center of Padua University Hospital and Dr. G. Barillari from the Institute of Immuno-Hematology and Transfusion Medicine of Udine General and University Hospital (Italy) for their help in contacting patients and collecting blood samples. Dr. V. Tchaikovski is gratefully acknowledged for his expert assistance and helpful suggestions for Western blot analysis.

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Authors’ contributions

C.D. performed research, analyzed data and wrote the paper; P.S. recruited the patients, collected clinical histories and critically reviewed the manuscript; L.S. recruited the patients and collected clinical histories; C.R. performed research; P.D. performed research; S.G. performed research; J.R. provided major intellectual input and critically reviewed the manuscript; E.C. designed and performed research, analyzed data and wrote the paper. None of the authors have a conflict of interest.
References


47. Magdeleyns EJ, Keuren JF, Curvers J. Factor Va, bound to microparticles released during platelet storage, is resistant to inactivation by activated protein C. Transfusion. 2007;47(10):1880-1888.


**Table 1. F5 gene mutations in FV-deficient patients.**

<table>
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<tr>
<th>F5 gene mutations</th>
<th>Nucleotide change</th>
<th>Amino acid change</th>
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<tbody>
<tr>
<td>PD I</td>
<td>1744 G&gt;C</td>
<td>524 Asp/His</td>
</tr>
<tr>
<td></td>
<td>(homozygous)</td>
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</tr>
<tr>
<td>PD II</td>
<td>1744 G&gt;C</td>
<td>524 Asp/His</td>
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<td>(homozygous)</td>
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<tr>
<td>PD III</td>
<td>853 T&gt;C</td>
<td>227 Trp/Arg</td>
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<tr>
<td></td>
<td>4957 G&gt;C</td>
<td>1595 Tyr/Asp</td>
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<tr>
<td>PD VII</td>
<td>6509 G&gt;A</td>
<td>2112 Gly/Asp</td>
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<tr>
<td></td>
<td>(homozygous)</td>
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Table 2. Thrombin generation parameters in PRP triggered with 5 pM TF.

<table>
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<th>No platelet agonist</th>
<th>Collagen (10 μg/mL)</th>
<th>Ionophore (20 μM)</th>
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<tr>
<td></td>
<td>Lag time (min)</td>
<td>Peak height (nM)</td>
<td>ETP (nM.min)</td>
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<td>PD I</td>
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<td>107</td>
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<td>PD III</td>
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<td>PD VII</td>
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<tr>
<td>Controls (n = 8)</td>
<td>3.0±0.5</td>
<td>48±49</td>
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<td>86±55</td>
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<td>681±69</td>
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<td>191±44</td>
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<td></td>
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<td>725±94</td>
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</table>

ETP, endogenous thrombin potential.

Thrombin generation parameters were obtained from the thrombin generation curves shown in Figure 2.
Table 3. Plasma and platelet FV antigen and activity levels in FV-deficient patients.

<table>
<thead>
<tr>
<th></th>
<th>Plasma FV</th>
<th>Platelet FV</th>
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<tbody>
<tr>
<td></td>
<td>Antigen (%)</td>
<td>Activity (%)*</td>
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<tr>
<td>PD I</td>
<td>1.8</td>
<td>&lt; 0.5</td>
</tr>
<tr>
<td>PD II</td>
<td>5.7</td>
<td>&lt; 0.5</td>
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<tr>
<td>PD III</td>
<td>4.2</td>
<td>&lt; 0.5</td>
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<tr>
<td>PD VII</td>
<td>9.6</td>
<td>4.4</td>
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<tr>
<td>Controls (n = 8)</td>
<td>98.7±20.9</td>
<td>106.5±26.8</td>
</tr>
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</table>

* FV activity as determined with a prothrombinase-based assay (see Materials and Methods section).
**Table 4.** Plasma and platelet TFPI antigen levels.

<table>
<thead>
<tr>
<th>Full-length TFPI level</th>
<th>Plasma (%)</th>
<th>Platelet (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>PD I</td>
<td>36.9</td>
<td>81.6</td>
</tr>
<tr>
<td>PD II</td>
<td>55.8</td>
<td>58.7</td>
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<tr>
<td>PD III</td>
<td>22.0</td>
<td>32.9</td>
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<tr>
<td>PD VII</td>
<td>51.0</td>
<td>60.1</td>
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<tr>
<td>Controls (n = 8)</td>
<td>96.7±22.0</td>
<td>77.4±31.7</td>
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Figure legends

Figure 1

**Contribution of plasma and platelet FV to thrombin generation.** FV-deficient plasma was reconstituted with 1.5*10^8 normal platelets/mL and increasing amounts of purified plasma FV (0%, 5%, 25%, and 100% = 23 nM). Platelets were activated with collagen and thrombin generation was triggered with 5 pM TF.

Figure 2

**Thrombin generation in PPP and PRP from patients with severe congenital FV deficiency.** PPP and PRP from patients PD I, PD II, PD III and PD VII were triggered with 1 (light gray), 5 (middle gray), 10 (dark gray), or 50 (black) pM TF, and thrombin generation was determined as described under Methods. Thrombin generation in PRP was measured without the addition of any platelet agonist and after pre-activation of platelets with collagen (10 µg/mL) or Ca^{2+}-ionophore (20 µM). A representative control is shown for comparison.

Figure 3

**FV-dependence of thrombin generation in PRP from a FV-deficient patient.** (A) Effect of an anti-FV antibody on thrombin generation in FV-deficient PRP. PRP from patient PD III was triggered with 50 pM TF in the presence of Ca^{2+}-ionophore and thrombin generation was
measured in the absence of an anti-FV antibody (●) and in the presence of 8 μg/mL (○), 28 μg/mL (▲), 60 μg/mL (△) and 108 μg/mL (◇) anti-FV antibody. (B) Effect of APC on thrombin generation in FV-deficient PRP. PRP from patient PD III was triggered with 5 pM TF in the presence of calcium ionophore and thrombin generation was measured in the absence of APC (●) and in the presence of 50 nM APC (△).

Figure 4

**Immunoprecipitation of FVa from plasma and platelets.** FVa was immunoprecipitated from 200 μL plasma (A) or 200 μL 1.5*10⁸ activated platelets/mL (B), concentrated 6 times (FV-deficient patients PD I, PD II, PD III, PD VII) or not concentrated (control), and subjected to SDS-PAGE and Western blotting. Control plasma and platelet FVa preparations were run at different dilutions (100%, 50%, 25% or 5%). FVa was detected with a monoclonal anti-FV heavy chain antibody and chemiluminescence. The FVa heavy chain has a molecular weight of 105 kDa, smaller fragments (only visible in platelet FVa) are degradation products of the FVa heavy chain.

Figure 5

**TFPI-titration of thrombin generation in PRP.** Control PRP (●) and FV-deficient PRP (patient PD II, ▲) were supplemented with increasing amounts of recombinant full-length TFPI and coagulation was initiated with 50 pM TF in the presence of collagen (10 μg/mL). The peak height of thrombin generation is plotted as a function of the concentration of added TFPI.
Duckers et al, Figure 1
Duckers et al, Figure 3
A  

Plasma FVa

B  

Platelet FVa

Duckers et al, Figure 4
Residual platelet factor V ensures thrombin generation in patients with severe congenital factor V deficiency and mild bleeding symptoms

Connie Duckers, Paolo Simioni, Luca Spiezia, Claudia Radu, Paolo Dabrilli, Sabrina Gavasso, Jan Rosing and Elisabetta Castoldi