Mitochondrial clearance is regulated by Atg7-dependent and independent mechanisms during reticulocyte maturation

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Running title: Role of autophagy in mitochondrial clearance

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The online version of this article contains a data supplement.

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Abstract

Mitochondrial clearance is a well recognized but poorly understood biological process, and reticulocytes, which undergo programmed mitochondrial clearance, provide a useful model to study this phenomenon. At the ultrastructural level, mitochondrial clearance resembles an autophagy-related process; however, the role of autophagy in mitochondrial clearance has not been established. Here we provide genetic evidence that autophagy pathways, initially identified in yeast, are involved in mitochondrial clearance from reticulocytes. Atg7 is an autophagy protein, and an E1-like enzyme, which is required for the activity of dual ubiquitin-like conjugation pathways. Atg7 is required for the conjugation of Atg12 to Atg5, and Atg8 to PE, and is essential for autophagosome formation. In the absence of Atg7, mitochondrial clearance from reticulocytes is diminished, but not completely blocked. Mammalian homologs of Atg8 are unmodified in Atg7−/− erythroid cells, indicating that canonical autophagy pathways are inactive. Thus, mitochondrial clearance is regulated by both autophagy-dependent and independent mechanisms. Additionally, mitochondria, which depolarize in wild type cells prior to elimination, remain polarized in Atg7−/− reticulocytes in culture. This suggests that mitochondrial depolarization is a consequence rather than a cause of autophagosome formation in reticulocytes.
Introduction

Mitochondria are the site of oxidative phosphorylation and energy production in animal cells, and one of the by-products of this process, reactive oxygen species (ROS), causes mitochondrial damage. Damaged mitochondria generate more ROS than healthy mitochondria, and can participate in a vicious cycle of ROS-mediated damage and ROS production.\(^1\) Mitochondrial damage accumulates with age, and is thought to have a role in aging, degenerative diseases, and cancer.\(^2,3\) To maintain homeostasis, cells replace damaged mitochondria through mitochondrial elimination and biogenesis. Therefore, to develop novel therapies, it would be useful to understand the mechanisms whereby senescent or defective mitochondria are eliminated.

Several types of cells eliminate their organelles during the course of normal development; these include the lens cells of the eye, and newly-formed erythrocytes, also known as reticulocytes.\(^4,5\) Reticulocytes are formed in the bone marrow from orthochromatic erythroblasts by the process of enucleation. Nascent reticulocytes contain mitochondria, ribosomes, endocytic vesicles, golgi cisternae, and rough endoplasmic reticulum (ER).\(^5,6\) Reticulocytes lose surface area and volume as they mature,\(^7\) and clear all of their membrane-bound organelles, including mitochondria, within a few days of generation. This rapid and coordinated clearance of mitochondria makes reticulocytes an ideal physiological model for the study of programmed mitochondrial clearance.

The primary means of eliminating defective mitochondria is thought to be mitochondrial autophagy, also known as mitophagy (reviewed in ref. 8). Mitophagy is observed in starved hepatocytes treated with glucagon,\(^9\) and in serum-starved neurons treated with caspase inhibitor.\(^10\) Similarly, in reticulocytes, ultrastructural studies indicate that mitophagy is involved in mitochondrial clearance.\(^5,11\) Another mechanism, described in reticulocytes, is 15-lipoxygenase-dependent degradation of mitochondria.\(^12-14\) Recent
work from our laboratory, and by Sandoval et al., demonstrated that an atypical BH3-only protein, NIX, is required for programmed mitochondrial clearance during reticulocyte maturation.\textsuperscript{15,16} These studies showed that NIX is required for mitochondrial autophagy, and specifically for targeting mitochondria to autophagosomes.

Autophagy is an evolutionarily conserved process, which maintains cellular homeostasis in changing nutrient conditions. Starvation induces autophagy, which in turn causes the sequestration of cytoplasm and subcellular organelles by double-membraned vesicles, and their subsequent degradation. Genetic studies in yeast have defined several of the key pathways that regulate autophagy.\textsuperscript{17} Autophagy gene 1 (Atg1) is a serine-threonine protein kinase that is repressed by mTOR. mTOR senses nutrient conditions, and starvation or rapamycin inhibit mTOR and activate autophagy. Along with Atg6 (Beclin-1), a component of a class III PI3-kinase-containing complex, Atg1 is essential for the formation of a preautophagosomal structure. Following formation of the preautophagosomal structure, dual ubiquitin-like conjugation pathways are required for the growth of autophagosomal membranes and autophagosome formation.\textsuperscript{18,19} These pathways, which require the E1-like enzyme Atg7, catalyze the conjugation of Atg12 to Atg5, and Atg8 to phosphatidylethanolamine (PE). Consistent with this function, Atg7 is required for the formation of autophagosomes.\textsuperscript{20} The Atg12-Atg5 and Atg8-PE ubiquitin-like conjugation pathways have been implicated in mitophagy in yeast,\textsuperscript{21-23} but the role of these pathways in mitochondrial clearance in mammalian cells remains to be determined.

The mammalian homologs of a number of autophagy genes have been identified. Targeted disruption of \textit{Atg5} or \textit{Atg7} in the germline of mice causes lethality due to an inability to mobilize essential and branched-chain amino acids in the neonatal period.\textsuperscript{20,24} Circulating erythrocytes from \textit{Atg5}\textsuperscript{-/-} neonates appeared normal, raising the possibility that the ubiquitin-like conjugation pathways may be dispensable for mitochondrial
clearance. However, in this study, an essential role of Atg5 in the induction of autophagy in erythroid cells was not confirmed. By contrast, Atg7 is known to be required for both ubiquitin-like conjugation pathways. Therefore, to gain insight into the role of autophagy in mitochondrial clearance, we examined the effect of Atg7 deficiency on mitochondrial clearance during reticulocyte maturation. Our studies indicate that the ubiquitin-like conjugation pathways make an important contribution, but are not strictly required for mitochondrial clearance in reticulocytes.

Materials and methods

Atg7 fetal liver cell transplants and animal studies

Conditional Atg7<sup>flox/flox</sup> mice (gift of Masaaki Komatsu and Keiji Tanaka) were bred with B6.FVB-Tg(EIIa-Cre) mice to generate heterozygous Atg7<sup>+/-</sup> mice on a C57BL/6J background.20,26 Atg7<sup>+/−</sup> mice were intercrossed to obtain E13.5 Atg7 null embryos. PCR primers for genotyping are tggctgctacttgcaatgatgt and ttagcacagggaacagcgctcatgg; the wild type Atg7 allele gives a 3.3 kb product and the null allele gives a 2.2 kb product. 2.5 to 5.0 X 10<sup>6</sup> fetal liver cells were injected into the tail vein of lethally irradiated (1050-1100 rads) B6.FVB-Tg(H2K-GFP) mice.27 Transplant recipients were treated with Enrofloxacin, 45 mg/kg/day, in their drinking water, prophylactically, to prevent infection. Bone marrow reconstitution was monitored by serial determination of the complete blood count, and by monitoring the expression of GFP in the circulating erythrocytes, platelets, and leukocytes. Reticulocytosis was induced by phlebotomy of 0.35 ml of blood daily for 4 days with saline volume replacement through intraperitoneal injection. The regimen was adjusted to achieve a target hematocrit of 20-25% on the final day, which was well tolerated by the mice. All studies were performed under an animal protocol approved by the Institutional Animal Care and Use Committee of St. Jude Children’s Research Hospital.
Flow cytometry

Reticulocyte-enriched blood was cultured in complete medium (30% fetal bovine serum, 1% deionized bovine serum albumin, 0.001% monothioglycerol, 2 mM glutamine, and penicillin-streptomycin in IMDM) for up to 3 days. ABT-737 (gift of Abbott Laboratories), 10 μM, was added in selected experiments, where indicated. Reticulocytes were stained with 200 nM Mitotracker Red CMXRos (MTR) (Molecular Probes, Eugene, OR) in complete medium for 30 minutes at 37°C, followed by 2 washes with phosphate-buffered saline (PBS). Reticulocytes were stained with thiazole orange (TO) in PBS (2 μg/ml) for 45 minutes at room temperature, followed by 2 washes with PBS. Doubly stained reticulocytes were obtained by sequential staining with MTR, then TO. MTR fluorescence emission between 600 to 620 nm was collected after 562 nm laser excitation, using a BD LSR II flow cytometry analyzer. Alternatively, MTR fluorescence emission greater than 610 nm was collected after 568 nm laser excitation, using a BD FACSVantage SE cell sorter. In both cases, TO fluorescence was collected at 500 to 520 nm after excitation with a 488 nm laser.

Immunofluorescence

For mitochondrial imaging by immunofluorescence, reticulocyte-enriched blood was stained with MTR, as described above, and transferred to a glass-bottom dish. Alternatively, to image mitochondrial depolarization, reticulocytes were stained with 500 nM Mitotracker Green (MTG) and 1 μM tetramethylrhodamine methylester (TMRM) in complete medium for 1 hr, followed by 2 washes with PBS. Cells were viewed with a Nikon TE2000-E inverted microscope equipped with a C1Si confocal system and a X60 Plan Apo oil-immersion objective (N.A. 1.45) (Nikon, Melville, NY). To quantify mitochondrial polarization, the fluorescence of every cell containing mitochondria in a representative high-power field (original magnification, X 60) was quantified in the red
and green channels with EZ-C1 software (Nikon, Melville, NY), and the ratio of the signals determined. Approximately 50-100 cells were quantified per sample.

Electron microscopy

Reticulocyte-enriched blood was fixed and stained, and sections examined with a Jeol JEM-1200EX II electron microscope, as previously described. Images were archived with an Advanced Microscopy Techniques XR111 bottom-mount CCD digital camera. Mitochondria and mature vacuoles with degraded contents were quantified, and the ratio determined. Approximately fifty images, containing 100-200 mitochondria and vacuoles, were quantified per sample.

Protein assays and antibodies

E14.5 fetal liver cells were expanded in culture for 6 days in complete medium with erythropoietin, 2 units/ml (Amgen, Thousand Oaks, CA); murine stem cell factor, 100 ng/ml; human IGF-1, 40 ng/ml (Peprotech, Rocky Hill, NJ; cat. nos. 250-03 and 100-11); and dexamethasone, 1 μM (Sigma, St. Louis, MO; cat. no. D4902). Lineage negative cells were obtained by magnetic bead technology and differentiated in culture for 2 days in complete medium with erythropoietin, 4 units/ml, and the protease inhibitors pepstatin A, 10 μg/ml (Calbiochem, San Diego, CA), and E64d, 10 μg/ml (Peptide Institute, Osaka, Japan). Whole-cell extracts were made with RIPA buffer (150 mM NaCl, 1.0% Nonidet P-40, 0.5% sodium deoxycholate, 0.1% SDS, 1 mM EDTA, and 50 mM Tris-HCl, pH 8.0) with protease and phosphatase inhibitors. Immunoblotting was performed as previously described.

The following primary antibodies were used: LC3, mouse IgG1 monoclonal antibody, 1 μg/ml; GABARAP, mouse IgG1 monoclonal antibody, 1 μg/ml; GATE-16, rabbit polyclonal antibody, 1:1,000 (Medical and Biological Laboratories, Woburn, MA; cat. nos. M115-3, M135-3, and PM038); Atg12, rabbit polyclonal antibody, 1:1,000 (Cell
Signaling Technology, Danvers, MA; cat. no. 2011); Atg7, rabbit polyclonal antibody, 1 μg/ml (ProSci, Poway, CA; cat. no. 3617); NIX, rabbit polyclonal antibody, 0.5 μg/ml (Exalpha Biologicals, Watertown, MA; cat. no. X1120P); Beclin-1, rabbit polyclonal antibody, 0.1 μg/ml (Santa Cruz Biotechnology, Santa Cruz, CA; cat. no. sc-11427); Ter119, rat monoclonal antibody, 0.25 μg/ml (BD Biosciences, San Jose, CA; cat. no. 553670); and β-actin, mouse IgG1 monoclonal antibody, 1:10,000 (Sigma, St. Louis, MO; cat. no. A5441).

Results

*Atg7*−/− fetal liver cell transplant recipients are anemic and lymphopenic

To generate mice that were Atg7 deficient in the hematopoietic lineage, *Atg7*<sup>±/±</sup> mice with a conditional null allele of the *Atg7* gene were bred to the B6.FVB-Tg(EIIa-Cre) Cre-deletor strain,<sup>20,26</sup> and the progeny intercrossed. The Cre transgene was removed by breeding, and heterozygous *Atg7*<sup>±/±</sup> mice were bred to obtain homozygous *Atg7*<sup>−/−</sup> embryos. 2.5 to 5.0 X 10<sup>6</sup> *Atg7*<sup>±/+</sup> or *Atg7*<sup>−/−</sup> embryonic day 13.5 (E13.5) fetal liver cells were transplanted into lethally-irradiated B6.FVB-Tg(H2K-GFP) mice.<sup>27</sup> The H2K-GFP transgene in the transplant recipients is expressed in all hematopoietic lineages, and allows discrimination between host and donor cells by flow cytometry. In contrast to the *Atg7*<sup>±/+</sup> transplant recipients, which usually survived, approximately half of the *Atg7*<sup>−/−</sup> transplant recipients failed to engraft and died. Surviving *Atg7*<sup>−/−</sup> transplant recipients exhibited lymphopenia, anemia, and reticulocytosis (Table 1). *Atg7*<sup>−/−</sup> erythrocytes showed a modest increase in central pallor, but their morphology was otherwise unremarkable (Figure 1A). Consistent with these findings, the spleens of *Atg7*<sup>−/−</sup> transplant recipients exhibited erythroid-myeloid hyperplasia, and lymphoid hypoplasia (Supplemental Figure S1). By 10 weeks post-transplant, the erythrocytes and
reticulocytes of the surviving Atg7−/− transplant recipients were entirely of donor origin and therefore suitable for further analysis (Supplemental Figure S2).

Mitochondrial clearance is impaired, but not absent, in Atg7−/− reticulocytes

To determine the role of Atg7 in mitochondrial clearance in mice, we performed flow cytometry on blood from engrafted Atg7+/− and Atg7−/− fetal liver transplant recipients, stained with Mitotracker Red (MTR), or doubly stained with MTR and thiazole orange (TO) (Figure 1B). Staining with MTR alone provides optimal separation between MTR-negative and positive populations, whereas staining with both MTR and TO permits the simultaneous assessment of mitochondrial and RNA (i.e., ribosomal) content. In Atg7+/− and Atg7−/− transplant recipients, 0.9 ± 0.8% and 9.9 ± 6.4% of the circulating erythrocytes contained mitochondria, respectively (Figure 1B and Table 1). By contrast, 53.4 ± 10.7% of Nix−/− erythrocytes contain mitochondria, and approximately half of these have cleared their ribosomes (Fig. 1B and ref. 15). These results suggest that if mitochondrial clearance is defective in Atg7−/− transplant recipients, the defect is less severe than in Nix−/− mice.

Since erythrocytes in mice under normal conditions are heterogeneous with respect to age, we considered the possibility that Atg7−/− reticulocytes have a defect in mitochondrial clearance, but are able to clear their mitochondria in vivo, given enough time. To test this possibility, wild type and Nix−/− mice, and Atg7−/− transplant recipients, were phlebotomized to induce reticulocytosis, and their blood was cultured in vitro. As previously shown, wild type reticulocytes readily cleared their mitochondria. 15 By contrast, mitochondrial clearance in Atg7−/− and Nix−/− reticulocytes was significantly impaired (Figure 2A). To exclude the possibility that this was an artifact caused by in vitro culture, we monitored mitochondrial clearance in the circulation of phlebotomized Atg7+/− and Atg7−/− transplant recipients. Mitochondrial clearance was delayed in the Atg7−/− transplant recipients, confirming the in vitro studies (Figure 2B). Collectively,
these studies indicate that mitochondrial clearance is partially impaired in \textit{Atg7\textsuperscript{-/-}} reticulocytes.

\textit{Atg7 is required for ubiquitin-like conjugation activity in erythroid cells}

To confirm that \textit{Atg7} deficiency inactivates the \textit{Atg8} ubiquitin-like conjugation pathways in erythroid cells, we examined the mammalian \textit{Atg8} homologs LC3, GABARAP, and GATE-16\textsuperscript{29-31} for autophagy-dependent modification during erythroid maturation. E14.5 fetal liver cells were expanded in culture for 6 days, then differentiated for 2 days in the presence of erythropoietin. Erythroid maturation was monitored by cellular morphology, and expression of the erythroid marker proteins Ter119 and BCL-X\textsubscript{L}. For all three \textit{Atg8} homologs, generation of the faster-migrating modified form was strictly dependent on the presence of \textit{Atg7} (Figure 3A). Furthermore, conjugation of \textit{Atg12} to \textit{Atg5} required \textit{Atg7}. Thus, both ubiquitin-like conjugation pathways are inactive in \textit{Atg7\textsuperscript{-/-}} erythroid cells.

Given the similarities in the cellular phenotypes caused by deficiency of \textit{Atg7} or NIX, we looked for evidence of cross-talk in the expression of these proteins. However, deficiency of \textit{Atg7} had no effect on the expression of NIX, and likewise, deficiency of NIX had no effect on the expression of \textit{Atg7} (Figure 3B).

\textit{Ultrastructural appearance of \textit{Atg7}-independent mitochondrial clearance}

Our results suggest that mitochondrial clearance proceeds, at a reduced rate, in the absence of \textit{Atg7}. To gain insight into this phenomenon, we examined the ultrastructure of \textit{Atg7\textsuperscript{-/-}} reticulocytes cultured in vitro. \textit{Atg7\textsuperscript{+/+}} and \textit{Atg7\textsuperscript{-/-}} transplant recipients, and \textit{Nix\textsuperscript{-/-}} mice, were phlebotomized to induce reticulocytosis, and their blood was cultured in vitro for 1 day. Mitochondria and degradative vacuoles were assessed by transmission electron microscopy, and quantified (Figure 4). The ratio of degradative vacuoles to mitochondria increased from day 0 to day 1 for reticulocytes of all three genotypes; however, on both days, it was highest for \textit{Atg7\textsuperscript{+/+}}, intermediate for \textit{Atg7\textsuperscript{-/-}}, and lowest for \textit{Nix\textsuperscript{-/-}} reticulocytes. Mitochondria were readily identified inside the vacuoles of both
Atg7+/+ and Atg7−/− reticulocytes, as were vesicles that appeared to be fusing with the
vacuoles (Figure 4, arrows). These results are consistent with the notion that Atg7 has a
quantitative effect on mitochondrial clearance, but is not essential for the incorporation of
mitochondria into vacuoles, vacuolar maturation, or exocytosis.

Role of mitochondrial depolarization in mitochondrial clearance

The role of mitochondrial depolarization is central to our understanding of mitochondrial
clearance. Mitochondrial depolarization can induce mitophagy,9,32 and it has been
suggested that NIX causes mitochondrial depolarization in reticulocytes.16 BAX and
BAK regulate mitochondrial outer membrane permeability, and their activation by BH3-
only proteins causes mitochondrial depolarization. Bax−/−;Bak−/− mice do not exhibit
abnormal erythrocytes with retained mitochondria.15 However, by analogy to Atg7, we
considered the possibility that Bax−/−;Bak−/− reticulocytes have a partial defect of
mitochondrial clearance. To investigate this scenario, we obtained blood from
phlebotomized Bax−/−;Bak−/−, Atg7+/+, Atg7−/−, and Nix−/− mice, and monitored reticulocyte
maturation in vitro over 3 days. Mitochondrial clearance from Bax−/−;Bak−/− reticulocytes
was normal, and indistinguishable from that of Atg7+/+ reticulocytes (Figure 5). By
contrast, mitochondrial clearance from Nix−/− reticulocytes, and to a lesser extent Atg7−/−
reticulocytes, was impaired. Therefore, if NIX causes mitochondrial depolarization, in
reticulocytes, it does not function through BAX or BAK.

ABT-737 is a potent small molecule antagonist of BCL2 and BCL-XL.33 The
ability of ABT-737 to rescue mitochondrial clearance in Nix−/− reticulocytes has been
interpreted as evidence that NIX causes mitochondrial depolarization.16 Indeed,
treatment of Atg7+/+, Atg7−/−, and Nix−/− reticulocytes with ABT-737 for 1 day caused
mitochondrial depolarization (Figure 5). However, treatment of Bax−/−;Bak−/− reticulocytes
with ABT-737 had no effect. Thus, consistent with its role as a BH3 mimetic, ABT-737
functions through a BAX- or BAK-dependent pathway. By contrast, NIX-dependent
mitochondrial clearance in reticulocytes is BAX- and BAK-independent. Therefore, ABT-737 and NIX mediate mitochondrial clearance by distinct mechanisms.

Our results indicate that mitochondria are cleared in reticulocytes independent of BAX, BAK, and the mitochondrial permeability transition pore (MPTP). However, we have not excluded the possibility that a NIX-dependent pathway, which does not employ BAX, BAK, or the MPTP, could cause mitochondrial depolarization and clearance. To address this possibility, we examined the polarization state of mitochondria in $Atg7^{+/+}$ and $Atg7^{-/-}$ reticulocytes cultured for 3 days. Reticulocyte-enriched blood was stained with Mitotracker Green (MTG) and tetramethylrhodamine methylester (TMRM). Polarized mitochondria, excited by a laser at a wavelength of 488 nm, emit red light due to fluorescence resonance transfer from MTG to TMRM. Depolarized mitochondria retain MTG but not TMRM, and emit green light. We previously showed that mitochondria in cultured wild type reticulocytes depolarize within 1 day. In this experiment, we found that essentially all mitochondria in $Atg7^{+/+}$ reticulocytes depolarize within 1 day, whereas most mitochondria in $Atg7^{-/-}$ reticulocytes, remain polarized after 3 days (Figure 6). Since Atg7 regulates autophagosome formation, this result suggests that mitochondrial depolarization in wild type and $Atg7^{+/+}$ reticulocytes is primarily a consequence rather than a cause of autophagosome formation.

Discussion

The replacement of senescent and defective mitochondria is thought to be critical for the maintenance of cellular homeostasis, but the mechanisms through which mitochondria are sequestered and eliminated are not well understood. Reticulocytes, which rapidly eliminate their entire cohort of mitochondria during development, provide a physiological model for the elucidation of these mechanisms. Ultrastructurally, reticulocytes eliminate mitochondria through an autophagy-related process. In the present study, we show
that mitochondrial clearance from reticulocytes is regulated by both autophagy-dependent and independent mechanisms.

When autophagy is induced, dual ubiquitin-like conjugation pathways are activated. These pathways, which covalently link Atg12 to Atg5, and Atg8 to PE, are essential for the growth of autophagosomal membranes. Both pathways require the E1-like enzyme, Atg7. Therefore, to gain insight into the role of autophagy in mitochondrial clearance, we transplanted \textit{Atg7}^{-/-} fetal liver cells into lethally-irradiated recipient mice. \textit{Atg7}^{-/-} transplant recipients exhibited lymphopenia, anemia, and reticulocytosis. The lymphopenia is consistent with peripheral lymphocyte depletion, and impaired T cell survival, in \textit{Atg5}^{-/-} fetal liver transplant recipients.\textsuperscript{35} Interestingly, \textit{Atg5}^{-/-} and \textit{Atg7}^{-/-} T cells have increased mitochondria.\textsuperscript{36,37} The anemia and reticulocytosis we observed are suggestive of an erythroid maturation defect. Indeed, our studies show that \textit{Atg7}^{-/-} reticulocytes have a defect in mitochondrial clearance, and \textit{Atg7}^{-/-} erythrocytes have a shortened life-span in vivo (unpublished results). In this regard, defective mitochondrial clearance has been previously linked to caspase activation and decreased erythrocyte life-span in \textit{Nix}^{-/-} mice.\textsuperscript{16}

Our studies show that mitochondrial clearance is regulated by \textit{Atg7}-dependent, ubiquitin-like conjugation pathways in reticulocytes. Another study of \textit{Atg5}^{-/-} erythrocytes suggested that autophagy may not be necessary for mitochondrial clearance;\textsuperscript{25} however, reticulocyte maturation was not examined, and it was not established that autophagy is impaired in \textit{Atg5}^{-/-} erythroid cells. More recently, it was demonstrated that reticulocytes deficient for the mammalian homolog of Atg1, ULK1, have a defect in mitochondrial clearance.\textsuperscript{38} ULK1 and ULK2 act upstream of Atg7 as initiators of autophagy. The similarity of the erythroid phenotypes caused by ULK1 and Atg7 deficiency reinforces the notion that autophagy has a role in this process.
Several lines of evidence suggest that mitochondrial clearance proceeds, at a diminished rate, in $Atg7^{-/-}$ reticulocytes. First, in contrast to $Nix^{-/-}$ mice, $Atg7^{-/-}$ transplant recipients do not have an abnormal population of circulating erythrocytes with retained mitochondria. This suggests, given sufficient time in vivo, that $Atg7^{-/-}$ reticulocytes are able to eliminate their mitochondria. Consistent with this interpretation, mitochondria are cleared, at a reduced rate, in phlebotomized $Atg7^{-/-}$ transplant recipients. Second, $Atg7^{-/-}$ reticulocytes contain degradative vacuoles, which are ultrastructurally indistinguishable from vacuoles in $Atg7^{+/+}$ reticulocytes, and contain mitochondria. There are fewer vacuoles in $Atg7^{-/-}$ than in $Atg7^{+/+}$ reticulocytes, but more than in $Nix^{-/-}$ reticulocytes. Thus, it appears that mitochondria are cleared through a degradative pathway, in the absence of Atg7. Whether this pathway is authentic autophagy, or a variant, remains to be determined.

NIX is an atypical BH3-only protein that targets mitochondria to autophagosomal membranes in reticulocytes.\(^{15,16}\) Since selective mitochondrial clearance is not well understood, the mechanism whereby NIX accomplishes this feat is of interest.\(^{39}\) One possibility is that NIX targets mitochondria for clearance by triggering depolarization. Accordingly, the BH3-mimetic ABT-737 rescues mitochondrial clearance in $Nix^{-/-}$ reticulocytes.\(^{16}\) However, ABT-737 and NIX differ in their requirement for BAX and BAK, and therefore function through distinct mechanisms. Alternatively, NIX may cause depolarization by activating the MPTP. NIX activates the MPTP in cardiac myocytes, indirectly, by increasing calcium stores in the ER.\(^{40}\) However, compared to cardiac myocytes, reticulocytes have relatively little ER. Furthermore, we have shown that inhibitors of the MPTP fail to prevent mitochondrial clearance.\(^{15}\) Finally, NIX may cause mitochondrial depolarization through a BAX-, BAK-, and MPTP-independent pathway. We addressed this possibility in autophagy-defective $Atg7^{-/-}$ reticulocytes. If NIX causes mitochondrial depolarization, then depolarized mitochondria should accumulate in $Atg7^{-/-}$
reticulocytes in culture. Instead, we observed the persistence of polarized mitochondria. This result suggests that mitochondrial depolarization, which precedes elimination, is a consequence rather than a cause of autophagosome formation. Altogether, these results suggest that NIX does not promote mitochondrial clearance by causing mitochondrial depolarization. On the other hand, we cannot exclude the possibility that NIX causes subtle or transient mitochondrial depolarization, and that this is sufficient to trigger mitochondrial clearance.

Recent studies demonstrating cross-talk between cell death and autophagy pathways suggest another mechanism whereby NIX may regulate mitophagy. These studies have shown that multidomain antiapoptotic proteins, such as BCL2 or BCL-X_L, inhibit autophagy, whereas proapoptotic BH3-only proteins, such as BAD, activate autophagy.\textsuperscript{41,42} The mechanism involves competition between proapoptotic proteins and Beclin-1 for binding to BCL2 or BCL-X_L. Beclin-1 binds to BCL2 and BCL-X_L through its BH3 domain, and can be displaced by another BH3 domain-containing protein, activating autophagy.\textsuperscript{43,44} NIX is upregulated during terminal erythroid differentiation,\textsuperscript{15,45} and could potentially participate in such an exchange. Autophagy is activated in both wild type and \textit{Nix}\textsuperscript{-/-} erythroid cells during differentiation. Thus, NIX is not generally required for autophagy in erythroid cells; however, given its location in the mitochondrial outer membrane, NIX may be required for activating autophagy near mitochondria.

It is interesting that the defect in mitochondrial clearance in autophagy defective strains of mice, such as \textit{Atg5}\textsuperscript{-/-}, \textit{Atg7}\textsuperscript{-/-}, and \textit{Ulk1}\textsuperscript{-/-} is less severe than that in \textit{Nix}\textsuperscript{-/-} mice. Despite inactivation of both ubiquitin-like conjugation pathways in \textit{Atg7}\textsuperscript{-/-} reticulocytes, mitochondrial clearance proceeds, albeit at a reduced rate. There are many possible explanations for this effect, one being that Atg8 homologs are recruited to mitochondria without ubiquitin-like modification. In this regard, it is intriguing that NIX directly interacts with LC3 (Ivan Dikic, personal communication), and GABARAP\textsuperscript{46}. Potentially, NIX can
function as an adaptor to recruit membrane trafficking or autophagy proteins directly to mitochondria.

There appears to be at least two mechanisms of mitochondrial clearance in mammalian cells. First, mitochondrial fission and depolarization trigger clearance,9,32 which is consistent with the observation that apoptotic signals cause mitochondrial clearance, when caspase activation is blocked.10 Although the biological significance of the latter observation is not fully clear, it is reasonable to hypothesize that mitochondrial quality control involves the clearance of senescent, depolarized mitochondria. In this regard, it is interesting that Parkin is recruited to depolarized mitochondria, and is required for their engulfment by autophagosomes.47 Second, studies of NIX and the related protein BNIP3 have shown that in some circumstances, mitochondrial clearance is programmed. This is important in developmental settings, such as reticulocyte maturation, and may also be important in the setting of cellular stress. BNIP3 is regulated by hypoxia and HIF1α,48 and has a role in mitochondrial clearance in hypoxic cells,49 which may be relevant for reports of deregulated BNIP3 expression in cancer (reviewed in ref. 50) Our present studies suggest that programmed mitochondrial clearance can be mediated through both autophagy-dependent and independent mechanisms. Additional studies of Atg7−/− and Nix−/− reticulocytes should help to elucidate these pathways.

Acknowledgments

The authors acknowledge the support of the Animal Resource Center, Flow Cytometry, and Cellular Imaging facilities of SJCRH. The authors thank Masaaki Komatsu and Keiji Tanaka for providing the Atg7flox/flox mice, and Abbott Laboratories for the gift of ABT-737.

This work is Supported by National Institutes of Health grants R21 DK074519 (P.A.N.), RO1 CA076379 (J.L.C.), and KO8 HL084199 (M.K); National Institutes of Health
Cancer Center support grant P30 CA21765; and the American, Lebanese, and Syrian Associated Charities.

Author contributions

J.Z. designed research, performed research, and analyzed data. M.S.R. and M.R.L. performed research. F.C.D., M.K., and J.L.C. designed research and analyzed data. P.A.N. designed research, analyzed data, and wrote the paper. The authors declare no conflict of interest.

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Figure legends

Figure 1. Mitochondrial clearance is preserved in unstressed Atg7<sup>−/−</sup> transplant recipients. (A) Blood smears from Atg7<sup>+/+</sup> and Atg7<sup>−/−</sup> transplant recipients, stained with Wright-Giemsa (original magnification, X 1000). Note the increased central pallor of the Atg7<sup>−/−</sup> erythrocytes. (B) Flow cytometry of blood stained with MTR (upper panels) or doubly-stained with MTR and TO (lower panels) from Atg7<sup>+/+</sup> and Atg7<sup>−/−</sup> transplant recipients, and Nix<sup>−/−</sup> mice.

Figure 2. Mitochondrial clearance is impaired in Atg7<sup>−/−</sup> reticulocytes. (A) Immunofluorescence and flow cytometry of reticulocyte-enriched blood cultured for 3 days in vitro from wild type and Nix<sup>−/−</sup> mice, and Atg7<sup>−/−</sup> transplant recipients. The cells are stained with MTR. The black line in the histograms corresponds to day 0, and the shaded area to day 3. The scale is indicated at the bottom. (B) Flow cytometry of blood from phlebotomized mice stained with MTR (upper panels) or doubly-stained with MTR and TO (lower panels). Mice were phlebotomized for 4 days. Thereafter, a small amount of blood (10 μl) was withdrawn on days 0, 1, and 3 for analysis. Day 0 was the day after the final day of phlebotomy. Genotypes are on the left.

Figure 3. Deficient ubiquitin-like conjugation activity in Atg7<sup>−/−</sup> erythroid cells. (A) Western blots of LC3, GABARAP (GRP), GATE-16 (GT16), Atg12, Ter119, BCL-X<sub>L</sub>, Beclin-1, and β-actin in expanded E14.5 fetal liver cells undergoing differentiation in culture for 2 days. The upper LC3 band corresponds to the unconjugated form of LC3, and the lower band corresponds to the lipid conjugated form of LC3. This same is true for the other Atg8 homologs, GABARAP and GATE-16. Wild type murine embryonic fibroblasts (MEF), provide a positive control for the position of the unconjugated and conjugated proteins. Genotypes and days in culture are indicated at the top. (B) Western blots of Atg7, NIX, Beclin-1, and β-actin in primary E14.5 fetal liver cells. NIX
protein is expressed as two isoforms in erythroid cells, a full-length monomeric form (upper band), and a shorter isoform, which has not been fully characterized (lower band). Beclin-1 is provided as a loading control. Genotypes are indicated at the top.

**Figure 4. Atg7-independent mitochondrial clearance is mediated by degradative vacuoles.** (Top) Ultrastructure of reticulocytes from \( Atg7^{+/+} \) and \( Atg7^{-/-} \) transplant recipients cultured for 1 day. The scale is indicated at the bottom. (Bottom) Quantification of mitochondria and degradative vacuoles in cultured \( Atg7^{+/+}, Atg7^{-/-}, \) and \( Nix^{-/-} \) reticulocytes, expressed as a ratio. Fifty digital images, containing 100-200 mitochondria and vacuoles were quantified per sample. Genotypes and days in culture are indicated at the bottom.

**Figure 5. ABT-737 and NIX differ in their requirement for BAX or BAK.** (Left panels) Flow cytometry of reticulocyte-enriched blood from phlebotomized \( Bax^{-/-};Bak^{-/-}, \) \( Atg7^{+/+}, Atg7^{-/-}, \) and \( Nix^{-/-} \) mice, cultured for 3 days, and doubly-stained with MTR and TO. Genotypes are indicated on the left. (Right panels) Flow cytometry of the same blood samples, cultured for 1 day in the presence (shaded area) or absence (black line) of ABT-737, and stained with MTR.

**Figure 6. Mitochondrial depolarization is a consequence of autophagosome formation in reticulocytes.** Immunofluorescence of reticulocyte-enriched blood from phlebotomized \( Atg7^{+/+} \) and \( Atg7^{-/-} \) transplant recipients, cultured for 3 days, and doubly-stained with MTG and TMRM. The mean immunofluorescence intensity per cell of all cells with mitochondria in a representative field was quantified in the red and green channels, and the ratio of the signals determined. The higher the ratio, the greater the average mitochondrial polarization per cell. The proportion of cells within each range is represented by a chart below each picture (legend at the bottom). Approximately 50-100 cells were quantified per sample (field). Note that mitochondria in the \( Atg7^{-/-} \) reticulocytes remain polarized, whereas mitochondria in \( Atg7^{+/+} \) reticulocytes do not. No
quantification is provided for \( \text{A}l\text{g}7^{+/+} \) reticulocytes on day 3 due to an insufficient number of cells with mitochondria for analysis. The scale is indicated at the bottom.
Table 1. Complete blood counts and erythrocyte indices of *Atg7*⁻/⁻ transplant recipients.

<table>
<thead>
<tr>
<th>Atg7 genotype</th>
<th>RBC, X 10⁶/ml</th>
<th>WBC, X 10⁶/ml</th>
<th>Gran, X 10⁶/ml</th>
<th>Lymph, X 10⁶/ml</th>
<th>Plts, X 10⁶/ml</th>
</tr>
</thead>
<tbody>
<tr>
<td>+/+ , n=7</td>
<td>9.2 ± 0.4</td>
<td>23.0 ± 7.7</td>
<td>4.1 ± 1.4</td>
<td>17.1 ± 6.8</td>
<td>697 ± 184</td>
</tr>
<tr>
<td>−/−, n=11</td>
<td>7.9 ± 0.8†</td>
<td>5.4 ± 2.0†</td>
<td>3.0 ± 1.2</td>
<td>1.8 ± 0.7†</td>
<td>489 ± 189</td>
</tr>
</tbody>
</table>

<table>
<thead>
<tr>
<th></th>
<th>Hct, %</th>
<th>Hb, g/dL</th>
<th>MCV, fl</th>
<th>MCHC, gm/dL</th>
<th>Reticulocytes, %</th>
<th>Mitotracker, %</th>
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<tbody>
<tr>
<td></td>
<td>44 ± 2</td>
<td>15.5 ± 0.5</td>
<td>48 ± 3</td>
<td>35.4 ± 1.5</td>
<td>2.5 ± 1.0</td>
<td>0.9 ± 0.8</td>
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<tr>
<td></td>
<td>37 ± 3†</td>
<td>12.9 ± 1.0†</td>
<td>51 ± 6</td>
<td>34.8 ± 2.3</td>
<td>9.5 ± 5.4†</td>
<td>9.9 ± 6.4†</td>
</tr>
</tbody>
</table>

Values presented as ± SD. †P<0.01
Figure 1

A  

Atg7+/+  Atg7/−

B  

Atg7+/+  Atg7/−  Nix/−

MitoTracker Red (log₁₀ FI)

Thiazole Orange (log₁₀ FI)

For personal use only.on April 16, 2017. For personal use only.
Figure 2

A

Day 0  
Day 3

wild type

Atg7⁻⁻⁻

Nix⁻⁻⁻

Bar=10 μm MTR

B

Day 0  Day 1  Day 3

Atg7⁺⁺⁺

Atg7⁻⁻⁻

MitoTracker Red (log₁₀ FI)

0 31 53

0 22 16

0 48 27

0 22 16

0 45 27

0 22 16

0 51 45

0 22 16

0 49 45

0 22 16

0 72 51

0 22 16

0 66 49

0 22 16

0 75 57

0 22 16

0 19 75

0 22 16

0 66 49

0 22 16

0 19 75

0 22 16

0 66 49

0 22 16

0 19 75

Thiazole orange (log₁₀ FI)

Bar=10 μm
Figure 3

A

<table>
<thead>
<tr>
<th>Atg7 genotype</th>
<th>Fetal liver cell</th>
<th>MEF</th>
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<tbody>
<tr>
<td></td>
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<td>Atg7−/−</td>
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<tr>
<td>Day: 0</td>
<td>1</td>
<td>2</td>
</tr>
<tr>
<td>LC3</td>
<td></td>
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<tr>
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<td>Atg5-Atg12</td>
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<td>Ter119</td>
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<td>BCL-XL</td>
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<td>Beclin1</td>
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<tr>
<td>β-actin</td>
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</table>

B

<table>
<thead>
<tr>
<th>Atg7 genotype</th>
<th>Atg7+/+</th>
<th>Atg7+/−</th>
<th>Atg7+/−</th>
<th>Atg7−/−</th>
<th>Nix−/−</th>
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<tr>
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<td>Atg7</td>
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<td>37 kD</td>
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<td>Beclin1</td>
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<tr>
<td>β-actin</td>
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</table>
Figure 4

*Atg7*+/+  *Atg7*−/−

Bar=0.2 μm

<table>
<thead>
<tr>
<th></th>
<th>Atg7 (+/+)</th>
<th>Atg7 (+/-)</th>
<th>Atg7 (-/-)</th>
<th>Nix (+/-)</th>
<th>Nix (-/-)</th>
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<tr>
<td>day 0</td>
<td>100%</td>
<td>100%</td>
<td>50%</td>
<td>50%</td>
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<tr>
<td>day 1</td>
<td>75%</td>
<td>75%</td>
<td>75%</td>
<td>75%</td>
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</tr>
</tbody>
</table>

- Black: Mitochondria
- Gray: Degradative vacuoles
Figure 6

Atg7^+/+

Day 0  Day 1  Day 2  Day 3

Atg7^-/

Day 0  Day 1  Day 2  Day 3

<table>
<thead>
<tr>
<th>Fluorescence</th>
<th>Red/Green:</th>
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<tr>
<td>0.2-0.3</td>
<td>0.6-0.7</td>
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<tr>
<td>0.4-0.5</td>
<td>&gt;1.0</td>
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<td>0.6-0.7</td>
<td>&gt;1.0</td>
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<tr>
<td>0.8-0.9</td>
<td>&gt;1.0</td>
</tr>
<tr>
<td>&gt;1.0</td>
<td>&gt;1.0</td>
</tr>
</tbody>
</table>

Bar=10 μm
Mitochondrial clearance is regulated by Atg7-dependent and independent mechanisms during reticulocyte maturation

Ji Zhang, Mindy S. Randall, Melanie R. Loyd, Frank C. Dorsey, Mondira Kundu, John L. Cleveland and Paul A. Ney