G-CSF treatment of Severe Congenital Neutropenia reverses neutropenia but does not correct the underlying functional deficiency of the neutrophil in defending against microorganisms.*

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Abstract

The treatment of children affected by severe congenital neutropenia (SCN) with G-CSF strongly reduces the risk of sepsis by reversing neutropenia. However, SCN patients who respond to the treatment with the growth factor still have an elevated risk to succumb of sepsis. Because the disease is usually caused by heterozygous mutations of \textit{ELA2}, a gene encoding for neutrophil elastase (NE), we have investigated in G-CSF-responder and non-responder patients affected by SCN the expression of polypeptides which constitute the anti-microbial machinery of these cells.

In peripheral-blood-derived neutrophils of patients with heterozygous mutations of \textit{ELA2} who were treated with G-CSF, NE was nearly absent as detected by immunofluorescence and immunoblotting, suggesting that production of the mutant protein interferes with normal gene expression. This defect was associated with abnormal expression of other granule-associated proteins such as myeloperoxidase, lactoferrin, cathepsin-G and human-neutrophil-peptide. Moreover, in one patient with partial response to G-CSF, we observed an impairment of neutrophil antimicrobial activity against \textit{Candida albicans}, and, at lower extent against \textit{E. coli}. Thereby, we propose that the treatment with G-CSF is not sufficient to correct all of the functional deficiency of neutrophils and this might account for the consistent risk of infections observed in SCN patients.
Introduction

Neutrophils are essential components of innate immune system because constitute the first line of defense against bacterial and fungal pathogens. An efficient response against these microorganisms needs that neutrophils carry a fully operational machinery including proteases, antimicrobial peptides and reactive oxygen \(^1\,^2\). In contrast, a reduction of neutrophil blood counts or a defect in their anti-microbial apparatus exposes the host to threats from many pathogens as observed in chronic granulomatous disease and in other functional defects of phagocytes \(^3\,^6\).

Severe congenital neutropenia (SCN) is an uncommon hematologic disorder characterized by reduction of absolute neutrophil counts (ANC; usually < 0.2 x10\(^9\) cells/L), due to maturation arrest of neutrophil precursors in the bone marrow at the promyelocyte stage. If left untreated, the large majority of children affected by SCN dies in the first years of life from invasive infections \(^7\,^8\). However, the empiric use of the polypeptide granulocyte colony stimulating factor (G-CSF) for the treatment of SCN has drastically changed the clinical outcome of this condition by increasing absolute neutrophil count values and reducing the episodes of infection in the vast majority of patients \(^7\,^10\). Nonetheless, SCN patients who are receiving G-CSF are at high risk of myelodisplastic syndrome (MDS) or acute myeloid leukemia (AML). In particular, a higher risk of MDS/AML is observed in SCN children who display a poor response to the treatment with G-CSF and/or receive large doses of the growth factor (above 8 \(\mu\)g/kg/day). In addition, even if the introduction of G-CSF in the treatment of SCN patients has dramatically improved survival and reduced morbidity from infections, infectious complications in treated patients are still observed at a rate of 0.9% per year \(^10\,^11\).

In the last years the identification of the genetic basis of SCN has underlined the key role of neutrophil elastase (NE) in myelopoiesis and focused the attention of many researchers on the strict link between granule formation and neutrophil differentiation. Indeed, heterozygous mutations of the gene which encodes for neutrophil elastase \((ELA2)\) constitute the molecular basis of this autosomal dominant inherited disease \(^12\,^13\). Molecular screening of \(ELA2\) in patients with
Congenital neutropenia has shown that patients with specific mutations (eg., G185R) are inclined to present a more severe clinical phenotype, requiring higher doses of G-CSF and, possibly, are at higher risk of developing MDS/AML\textsuperscript{14,15}. In this context, a primary question related to the use of this growth factor in this genetic disease is if G-CSF is capable to restore normal neutrophil functions besides correcting neutropenia.

In order to address this question, we have investigated in children affected by SCN receiving G-CSF whether the growth factor is able to correct neutropenia and/or to reconstitute the entire antimicrobial machinery of fully-matured neutrophils. Because granule protein synthesis and packaging of primary granules and thereafter of specific granules go along with promyelocytic differentiation up to mature neutrophils\textsuperscript{16-18}, we analyzed expression of NE, myeloperoxidase (MPO), lactoferrin, cathepsin G, human neutrophil peptide (HNP), lysozyme and NADPH oxidase components in cells isolated from SCN receiving G-CSF. We observed that \textit{ELA2} genotype strongly influenced the ability of G-CSF to correct neutropenia, but the expression and activity deficiency of many granule-associated proteins was evident despite the correction of neutropenia by treatment with the growth factor.
Methods

Approval for these studies was obtained from the Spedali Civili of Brescia institutional review board. Informed consent was provided according to the Declaration of Helsinki for the five with SCN, two with idiopathic neutropenia subjects and for the healthy control subjects.

Patients and clinical details

Five SCN affected children (P1, P2, P3, P4, P5) and two with idiopathic neutropenia (C3 and C4) were recruited from the haematological unit of Ospedale dei Bambini in Brescia. All these subjects were under treatment with G-CSF (5-40 µg/kg) at the time when the study was performed. Diagnosis of all patients were made by experts on the basis of clinical and hematological data, valuating conventional criteria of persistent severe neutropenia, severe bacterial and fungal infection, and bone marrow maturation arrest at the myelocyte-promyelocyte stage. Patients P1 and P2 have been subjected to bone marrow transplantation while the other patients are currently receiving G-CSF treatment. However, patient P2 has recently lost the engraftment of donor’s bone marrow and is currently receiving G-CSF. For each experiment performed, at least a healthy subject and/or an age matched child were included for comparison. The control group was used upon informed consent and consisted of healthy adult donors; two children with idiopathic neutropenia or age matched children who were admitted to the hospital for minor head trauma.

Mutational analysis

DNA was extracted from peripheral blood leukocytes using standard techniques. All five exons of ELA2 and at least 15 bases of flanking regions were amplified by PCR (primers and amplification conditions available upon request). PCR products were purified and bi-directionally sequenced as previously described using Big Dye Terminator Chemistry (Applied Biosystems) on an ABI PRISM 310 Genetic Analyzer (Applied Biosystems). For each new mutation found at least 50 healthy controls were screened using PCR amplification and direct sequencing.
Neutrophils

Peripheral neutrophil cells were purified by Ficoll-Hypaque separation density-gradient centrifugation (Amersham Pharmacia Biotech, Uppsala, Sweden) from 5-10 ml of heparinized blood, followed by red cell lysis (NH4Cl 0.829%, EDTA 0.125mM, NaH2CO3 0.1%) and washing with PBS. Cell recovery varied from 1.5 x 10^6 cells (patients P1 and P2) up 4 x 10^6 cells (P3). Neutrophil purity was assessed on the basis of May-Grünwald Giemsa stain, and of G-CSF receptor/CD15 expression at flow cytometry analysis, (neutrophils > 97%). O₂ release was estimated by cytochrome C reduction. Briefly, neutrophils (2x10⁵) were resuspended in HBSS/Ca²⁺/Mg²⁺ pH 7.4 containing 80 µM ferricytochrome C type III (Sigma, St. Louis, MO) and stimulated with 100 nM fMLP, 20 ng/ml PMA or 200 µg/ml opsonized zymosan from S. cerevisiae (Sigma) for 60 min. When required cells were pre-incubated with 50 ng/ml G-CSF (Lenograstim, Chugai Pharmaceutical, Tokyo, Japan) or 50 ng/ml GM-CSF (Schering-Plough, Kenilworth, NJ) for 30 min before fMLP addition. Cells were placed in clear bottomed 96-well plates and cytochrome C reduction was evaluated at 550 nm.

Lactoferrin production by neutrophilic cells or lactoferrin plasma concentration was detected by sandwich ELISA system, using BIOXYTECH® Lactof-EIA™ assay provided by OXIS International Inc. (CA, USA), with a detection limit of 2 pg/ml. When indicated, neutrophils were stimulated with fMLP (10 nM), CXCL8 (10 ng/ml) or medium alone for 15 minutes. In the collected supernatants or plasma samples lactoferrin concentration was measured by ELISA and expressed as average ± SD of two distinct experiments.

Immunocytochemistry and immunofluorescence

Peripheral blood neutrophils were purified by Ficoll separation medium gradient centrifugation, followed by red cell lysis, and washing with PBS. Cells were counted and utilized for cytopsin
preparations, then slides were air-dried and used for immunocytochemistry and
immunofluorescence staining. Neutrophil elastase expression was analyzed using rabbit polyclonal
antibody to human neutrophil elastase (1:300) from Calbiochem (San Diego, CA), revealed with
FITC-conjugated (1:25, DAKO Cytomation, Glostrup, Denmark); nuclei were counterstained for 1
min in 0.5 mg/ml 4,6-diamidino-2-phenylindole. Myeloperoxidase expression was analyzed using
rabbit polyclonal antibody to human myeloperoxidase (1:3000, Dako C.) followed by streptavidin-
biotin complex immunoperoxidase technique, with diaminobenzidine as chromogen; nuclei were
counterstained with Mayer’s hematoxilin. For antigen retrieval the slides were immersed in citrate
buffer and microwaved for 5 min at 750 W. Cells were photographed using an Olympus BX60
fluorescence microscope and objectives with numeric apertures of 0.40 (10 X), 0.70 (20 X), 0.85
(40 X), and 0.90 (60 X), equipped with a DP-70 Olympus digital camera (Olympus, Melville, NY).
Images were acquired using analySIS Image Processing software (Soft Imaging System, Munster,
Germany). At least five fields of magnification 20X, and 60X for each sample were randomly
selected and analyzed.

**Immunoblotting**

Cells were suspended in HBSS pH 7.4 containing 1 mM Di-isopropylfluorophosphate (DFP, Sigma). After 5 min this solution was discarded, and the cells were lysed with electrophoresis
sample buffer (60 mM Tris/HCl, 20% (v/v) glycerol, 4% (W/v) SDS, 2% (v/v) 2-mercaptoethanol,
pH 6.8) and boiled for 5 min. Total cell lysates were subjected to SDS/PAGE on 12% or 14% gels
and then transferred to nitrocellulose membranes (Amersham). The blots were then rinsed in TBS-T
(50mM Tris, 170mM NaCl, 0.2%(v/v) Tween 20; pH 7.5) and incubated for 60 min in TBS-T
containing 5% BSA pH 7.5 (blocking buffer), before overnight incubation (4°C) with the following
rabbit antibodies: anti-gp91phox, anti-p47phox, anti-p67phox, anti-p40phox, anti-p22phox (kindly
provided by dr. F.B. Wientjes, Department of Medicine, University College, London, UK), anti-
human neutrophil elastase (Calbiochem, San Diego, CA), anti-human neutrophil myeloperoxidase
(Assay Design Inc., Ann Arbor, MI), anti-human lactoferrin (Sigma), anti-cathepsin G (BIODESIGN Int., Saco, ME) or anti-β-actin (Sigma); mouse anti-human lysozyme (BIODESIGN Int.); goat anti-HNP (C-19) (Santa Cruz Biotechnology Inc., Santa Cruz, CA) diluted 1:500 in TBS-T containing 1 mg/ml BSA. The blots were incubated with horseradish peroxidase-conjugated anti-rabbit, anti-mouse IgG (Amersham), or anti-goat IgG (Santa Cruz Biotechnology). Bound antibodies were detected by enhanced chemiluminescence western blotting detection reagents (Amersham).

MPO activity assay

The EnzChek® Myeloperoxidase (MPO) Activity Assay Kit (E33856, Molecular Probes, Leiden, The Netherlands) was used for rapid and sensitive determination of MPO chlorination activity in neutrophil lysates. Fluorescence was measured with a fluorescence microplate reader using fluorescence excitation and emission at 485 and 530 nm, respectively. The background fluorescence measured for each zero-MPO control reaction was subtracted from each fluorescence measurement before plotting.

Phagocytosis assay

Neutrophils from patient 2 (P2) and from an healthy subject (CTR) were incubated for 40 min at 37°C with serum-opsonized zymosan (10 particles/cell) or serum-opsonized *C. albicans* (4 particles/cell), washed twice with PBS, transferred onto glass slides and stained with May-Grünwald Giemsa before examination under light microscopy.

Killing assay

Neutrophils were resuspended in HBSS/Ca²⁺/Mg²⁺ pH 7.4 and prewarmed for 10 min at 37°C in a shaking water bath. Opsonized microorganisms were added at a ratio of three bacteria/neutrophil (*E. coli* ATCC 25922) or two blastospores/neutrophil (*C. albicans* ATCC 24433), with a final
neutrophil concentration of $3 \times 10^5$ cells/100µl. Microorganisms were also incubated in the absence of neutrophils to account for growth during the assay. The tubes were incubated at $37^\circ$C in a shaking water bath. After 90 min, samples were diluted in 1 ml pyrogen-free distilled water brought to pH 11.00 with NaOH just before use; all the samples were then inverted twice. After standing for 5 min at room temperature and vortexing vigorously for 5 s, samples were diluted in PBS solution to give a bacterial or fungal concentration of $2 \times 10^3$/ml. Triplicates of 100 µl aliquots of each dilution was plated on LB or Sabouroud agar Petri dishes. The CFU were counted after an overnight incubation at $37^\circ$C (E. coli) or at $30^\circ$C (C. albicans). The percent killing was calculated as follows: $(1 - (\text{cfu after incubation with cells/cfu control culture})) \times 100$.

**Statistical analysis**

Comparison of values between normal donors and patients were performed where indicated by ANOVA for unpaired data. For single comparison amongst groups, Bonferroni correction was applied. Differences were defined significant for $p$ values lower than 0.05.
Results

We have recently identified 5 SCN patients (ANC < 0.1 x10^9 cells/L) with various heterozygous mutations of *ELA2* (Table 1). In order to identify the biological correlates of the response to G-CSF in SCN patients, we evaluated the expression of neutrophil elastase and of other granule-associated protein in children with mutations of *ELA2* during the treatment with the G-CSF (Table 1). We found that patients P3 (P176fsX182) and P4 (N34-H41del) showed a satisfactory increase of neutrophil counts (3.6 and 1.9 x10^9 cells/L), while the remaining three (P2, G185R; P5 A98D) presented a blunted response to the treatment (0.8 x10^9 cells/L) or were not responsive (P1, G185R), even at high doses of G-CSF (up to 40 µg/kg).

Then we asked which was the effect of G-CSF on the expression of neutrophil elastase and myeloperoxidase, which are major components of primary granules and are synthesized at promyelocytic stage of maturation. To this aim, we separated neutrophils from patients P1, P2 and P3, while they were receiving G-CSF and analyzed the expression of neutrophil elastase and of other granule-associated proteins. Immunofluorescence staining with anti-neutrophil elastase rabbit polyclonal antibody showed that expression of the protein was detectable only in a minority of cells derived from SCN patients, while it was abundantly present in cytoplasm of neutrophils from control subjects (Figure 1A). Neutrophil elastase is a monomeric polypeptide which is synthesized in endoplasmic reticulum as a 32 kDa, and subjected to glycosilation in Golgi apparatus. Subsequent processing with removal of aminoterminal and carboxyterminal peptides takes place during its transport to primary granules where it stored as mature polypeptide of about 27-29 kDa. Analysis of neutrophil elastase expression by immunoblotting in SCN patients showed that the extent of protein expression was severely reduced in neutrophils of G-CSF-treated SCN patients, as compared to neutrophils of control subjects, in spite of G-CSF treatment (Figure 1B). Moreover, the relative amount of the different proteolytic polypeptides, of sizes ranging from the unprocessed 32 kDa form to the 29 kDa matured polypeptide, were dramatically altered, suggesting that normal
neutrophil elastase processing is not restored in SCN patients during G-CSF treatment. Of note, analysis of NE expression in two patients with idiopathic neutropenia (C3 and C4), but normal \textit{ELA2}, showed normal processing of the protein (Figure 1B). An analysis of NE expression in other 5 patients with severe neutropenia due to other causes in comparison to an additional SCN patient (P4) showed that the abnormal protein processing was not related to the treatment with G-CSF, but was strictly associated to \textit{ELA2} mutations (data not shown). Next, we evaluated \textit{ELA2} expression at the mRNA level in hematopoietic cells isolated from bone marrow of a SCN patient (P3). Analysis of \textit{ELA2} mRNA by Real Time PCR showed that the extent of expression in the cells derived from the patient was comparable to the control subject (data not shown). This is in accord to previous observations showing that \textit{ELA2} mutations lead to abnormal processing and intracellular targeting of the protein\textsuperscript{15,20}.

Next we investigated whether MPO, the other major constituent of primary granules is expressed and normally processed as tetrameric glycoprotein, consisting of two heavy (60 kDa) and two light chains (15 kDa), in neutrophils of SCN patients receiving G-CSF\textsuperscript{22}. Immunocytochemical analysis of neutrophils obtained from SCN patients showed that MPO is absent or poorly expressed in cells, even while patients were receiving G-CSF therapy (Figure 2A). In addition, western blot analysis showed that the electrophoretic mobility of the light chain was similar both in normal and in neutropenic patients, while the heavy chain had an abnormal size, suggesting an altered cleavage and/or glycosilation of MPO subunits (Figure 2B). Similarly to the 60 kDa heavy chain, a band of 40 kDa, most likely representing a degradation product of the heavy chain\textsuperscript{23}, showed an altered electrophoretic mobility (Figure 2B). We then investigated whether abnormal MPO expressed by SCN patients was functional. To this purpose we analyzed the MPO-dependent chlorination activity in lysates of neutrophils isolated from patient 2 and we found that this activity was greatly reduced (Figure 2C).

We then evaluated the presence of other antimicrobial proteins in neutrophils and blood serum of G-CSF treated SCN patients. First we looked at lactoferrin, a constituent of specific granules, which
is also detectable in large amounts in blood circulation. Specific granules are rapidly mobilized to cell surface in neutrophils activated by various stimuli, including chemoattractancts such as the formyl-MLP peptide (fMLP) and CXCL8, thereby leading to lactoferrin secretion. Analysis of lactoferrin secretion in neutrophils from SCN patients showed that the antimicrobial protein was undetectable in supernatants obtained after stimulation with fMLP or IL-8, suggesting a reduced cellular content of the protein (Figure 3A). Indeed, blood lactoferrin concentration was severely reduced in all SCN patients (Figure 3B). In order to investigate intracellular expression of lactoferrin in neutrophils of SCN patients under treatment with G-CSF, we performed an immunoblot analysis of the protein. We found that lactoferrin was undetectable in neutrophil lysates from SCN patients bearing the G185R mutation (P1 and P2) while it was expressed at reduced levels in neutrophils of patient 3 (P3), as compared to levels detected in control subjects (Figure 3C).

We then analyzed the expression of other defensive proteins contained in primary and/or secondary granules of neutrophils by western blot analysis. We found that cathepsin G and mature HNP were undetectable, whereas HNP precursor was present in SCN neutrophil lysates (Figure 4). Moreover, lysozyme was normally expressed in neutrophils of the patients (Figure 4). Thereafter, we investigated the expression and function of NADPH oxidase, the multicomponent enzyme which plays a central role in host defense by catalyzing superoxide anion production. For its activity, it is required the assembly of the cytosolic subunits p47phox, p67phox, p40phox and Rac with the membrane-associated heterodimer of gp91phox and p22phox. An immunoblot analysis showed that the NADPH oxidase cytosolic components p47phox, p67phox, and p40phox were expressed at similar levels in control and SCN neutrophils, whereas the membrane-associated subunits gp91phox and p22phox were reduced in neutrophils of patients (Figure 5A). In order to determine whether this alteration of NADPH oxidase subunits expression could affect the enzymatic activity of the complex, we evaluated superoxide anion release in neutrophils from SCN patients under treatment with G-CSF. We found that superoxide anion production was normal after SCN neutrophil
challenge with an optimal concentration of the phorbol myristate acetate (PMA), which activates
NADPH oxidase by direct interaction with protein kinase C, while it was significantly reduced in
SCN neutrophils stimulated with fMLP, a receptor-dependent activator of the enzyme (Figure 5B).
Flow cytometry analysis of CXCR1 and fMLP receptor expression showed that the two G-protein
coupled receptors were detectable in neutrophils of patient P3 as well as in cells of normal subjects
(data not shown), suggesting that the impairment of fMLP-mediated oxidative burst is probably
related to a defective signaling machinery.
Phagocytosis experiments showed that SCN neutrophils normally internalized both opsonized
Candida albicans and zymosan, and zymosan ingestion induced superoxide anion production
(Figure 5C), suggesting that the mechanisms involved in pathogens uptake and in phagocytosis-
dependent oxygen radical production are conserved in these cells.
We finally investigated whether the reduction and/or abnormality of defensive proteins observed in
SCN neutrophils results in a decreased anti-microbial activity. To this aim we analyzed the ability
of neutrophils from one SNC patient to kill C. albicans and E. coli. We found that these cells were
unable to inhibit Candida growth, whereas their capacity bactericidal activity against E. coli was
significantly reduced (p< 0.05. Figure 5D), suggesting a defect of neutrophils from SCN patients to
control infections from these microorganisms.
Discussion

Because of the ability of G-CSF to restore the neutrophil counts in many conditions characterized by neutropenia, this highly effective growth factor of myeloid cells has been empirically experimented in a large series of disorders of myelopoiesis including SCN. Recent studies on the pathogenesis of this inherited disease have focused our attention on the effect of G-CSF on granule formation and expression of anti-microbial polypeptides in children affected by SCN who were treated with the growth factor. Analysis of NE in granulocytes from SCN patients under treatment with G-CSF has shown that the protein was detectable at low amounts as assessed by immunofluorescence, suggesting that the expression of the mutant interferes with the production of the mature protein, even from the normal allele. Moreover, immunoblot analysis has revealed that NE was undetectable in neutrophils of SCN childrens, suggesting that NE processing and possibly intracellular trafficking were abnormal despite the treatment with G-CSF. This finding is accord with the observation that the absence of β3A subunit of the adaptor protein 3 complex, which mediates intracellular trafficking of neutrophil elastase from trans-Golgi network to primary granules 20, results in abnormal delivery of NE to plasmamembrane and in neutropenia, as observed in Hermansky-Pudlak 2 patients and in gray collie dogs with AP-3 deficiency 20,28,29. Moreover, expression of the G185R mutation in retrovirally transduced HL-60 promyelocytes further demonstrated that the NE is mislocalized to plasmamembrane 15. Mislocalization of NE at plasmamembrane will probably account for the abnormal myelopoiesis because the enzyme might inappropriately interact with regulatory proteins of hematopoiesis including CXCR4, CXCL12, G-CSF, G-CSF receptors, and Notch proteins as potential substrates of this serine protease 30-34.

We have shown by immunofluorescence staining that abnormal expression and cellular targeting of NE in neutrophils of SCN patients will affect the generation of primary granules. This is also supported by the contemporary severe reduction of MPO expression which was observed by immunohistochemistry analysis of SCN neutrophils. Immunoblotting analysis of MPO demonstrated that the protein was still expressed, but its post-translational processing was not
completed as suggested by the atypical pattern of the MPO subunits detected in cell lysates of SCN patients. While the light MPO subunit (15 kDa) was still detectable, but at reduced levels, the high molecular weights peptides corresponding to the heavy MPO subunit (60 kDa) as well as its degradation product (40 kDa) were absent or significantly reduced, while an intermediate molecular weight peptide was detectable in neutrophils of SCN patients. This observation suggests that in the absence of normal NE expression and/or trafficking, even MPO cannot completely mature in neutrophils of SCN patients under treatment with G-CSF. Moreover, our results account for previous findings reporting that mRNA expression of NE and MPO is impaired in bone marrow-derived myeloid precursors of severe congenital neutropenia patients \(^{35,36}\). In contrast to these findings, we were unable to detect differences of \(ELA2\) mRNA levels in total hematopoietic cells isolated from bone marrow of P3; but, the effect of \(ELA2\) mutations on mRNA expression might not be detectable in total hematopoietic cells from bone marrow but only in selected myeloid precursors.

Investigation on NE, MPO and lactoferrin expression in neutrophils of the two SCN patients with the G185R mutation has shown that the defect of the abnormal processing of these polypeptides is more striking in these patients suggesting that the corresponding mutant has a profound effect on the early stages of neutrophil differentiation. It is interesting that transfection studies of the G185R mutant into RBL cell line have shown that this protein cannot be revealed by immunofluorescence while it is still detectable by immunoblotting \(^{21}\). Moreover, we found that children with a refractory response to G-CSF and the G185R genotype expressed an extremely low amount of lactoferrin, while the antimicrobial protein was still detected in cell extracts of patients with other \(ELA2\) mutations or with other causes of neutropenia under treatment with G-CSF \(^{36,37}\) suggesting that an early death of promyelocytes during their differentiation prevented the formation of specific granules \(^{15,38}\).

This succession of events is not fully restored by treatment with G-CSF, although we observed a great degree of variability in the severity of the maturation defect among children with different
types of ELA2 mutations. Therefore, the incomplete maturation of neutrophils in SCN may explain why a significant number of cases of patients die from sepsis (8% of cumulative incidence over 10 years) even if they had shown a satisfactory response to the treatment with G-CSF\(^\text{11}\). In fact, while the observed risk to die of sepsis was higher in the patients less responsive to the growth factor (14%) as compared to children with good response (11%), the use of G-CSF was not sufficient to prevent lethal infections in these children. The increased susceptibility to infections might be related to the abnormal expression in neutrophils of SCN patients of antimicrobial polypeptides, such as NE, lactoferrin, MPO, cathepsin G and HNP, in spite of the treatment of the patients with G-CSF. It is conceivable that the decreased expression of these peptides by neutrophils of SCN patients could lead to an impairment of their antimicrobial functions. In fact, we report that these cells were unable to kill \textit{C. albicans} and showed a decreased killing activity against \textit{E. coli}. Therefore, it is likely that the antimicrobial activities which are dependent on MPO, lactoferrin, cathepsin G, HNP and, other unknown abnormal proteins, are not correctly functioning in neutrophils of SCN patients; whereas the defense mechanisms which require polypeptides such as lysozyme which are correctly expressed in cells of SNC patients, are still effective against some microorganisms. Notably, we found that phagocytosis is apparently normal in SCN neutrophils, suggesting that the reduced killing ability of these cells does not rely on defects of microbial internalization. Moreover, although the expression of membrane-associated NADPH oxidase subunits is decreased, oxygen radicals production in response to PMA and zymosan particles is conserved in SCN neutrophils, whereas it is defective upon fMLP stimulation. Therefore, activation mechanisms of oxygen-dependent killing machinery appear to be only partially conserved in SCN neutrophils, in spite of the treatment of patients with G-CSF.

Clearly, further studies are needed to determine whether the defect of granule formation can be generalized to all SCN patients treated with G-CSF, but these new results and previous evidence that G-CSF therapy accelerates leukemogenesis in SCN call into question the overall efficacy of treatment with G-CSF and raise the issue of whether it should even be used for the treatment of this
condition. This is particularly evident in SCN children with unsatisfactory response to the therapy, whose elevate risk to die of sepsis or to develop MDS/AML makes them as the best candidates for bone marrow transplantation.

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REFERENCES


Table 1*

Absolute neutrophil counts in SCN children receiving G-CSF

<table>
<thead>
<tr>
<th>Patient (ID)</th>
<th>Age at diagnosis</th>
<th>ELA2 amino acid</th>
<th>ANC (10^9 cells/L)</th>
<th>G-CSF (µg/kg)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td>At diagnosis</td>
<td>After therapy</td>
</tr>
<tr>
<td>P1</td>
<td>2 months</td>
<td>4924G&gt;A</td>
<td>G185R</td>
<td>40</td>
</tr>
<tr>
<td>P2</td>
<td>10 days</td>
<td>4924G&gt;A</td>
<td>G185R</td>
<td>20</td>
</tr>
<tr>
<td>P3</td>
<td>8 days</td>
<td>4899del</td>
<td>P176fsX182</td>
<td>5</td>
</tr>
<tr>
<td>P4</td>
<td>4 months</td>
<td>1903-1926del</td>
<td>N34-H41del</td>
<td>5</td>
</tr>
<tr>
<td>P5</td>
<td>3 months</td>
<td>4498C&gt;A</td>
<td>A98D</td>
<td>5</td>
</tr>
</tbody>
</table>

*del: deletion; fs: frame shift. Nucleotide position is given according to sequence Genebank Y00477. Aminoacid numbers begin from the first residue after the presignal peptide cleavage.
FIGURE LEGENDS

Figure 1. Abnormal expression of neutrophil elastase in neutrophils of SCN patients during treatment with G-CSF. (A) Immunofluorescence staining of NE in circulating neutrophils from SCN patients. PMN were obtained from peripheral blood, cytocentrifugated and stained with anti-NE antibody; nuclei were counterstained with DAPI. The majority of cells from a control subject (CTR) show a strong cytoplasmatic expression of NE, whereas NE is detectable in a minority of cells obtained respectively from patients 1, 2 and 3. Experiment shown are representative of two performed. (B) Cell lysates of neutrophils purified from three patients (P1, P2, and P3), two healthy control subjects (C1, C2) and two idiopathic neutropenia patients (C3, and C4) were subjected to Western Blot analysis with anti-neutrophil elastase antibody. β-actin was used to normalize protein levels. The 29 and 32 kDa polypeptides are indicated by arrows. The band indicated by a dot could correspond to the previously identified elastase-alpha1 antitrypsin complex 21.

Figure 2. Abnormal expression of myeloperoxidase in neutrophils of SCN patients during treatment with G-CSF. (A) Immunocytochemical staining of MPO in circulating neutrophils from SCN patients. PMN were obtained from peripheral blood, cytocentrifugated and stained with anti-MPO antibody; nuclei were counterstained with Mayer’s hematoxilin. In cells from a control subject (CTR), a strong granular expression of MPO is observed in the cytoplasm. In contrast, MPO is detectable in a minority of cells obtained from patient 3 (P3) and undetectable in cells from patient 1 (P1). (B) The Western Blot shown in Figure 1B has been reprobed with specific anti-myeloperoxidase antibody. The three myeloperoxidase bands are indicated by arrows. (C) MPO chlorination activity in neutrophil cell lysates (three dilutions of each sample) from a normal subject (CTR) and patient 2 (P2) was analyzed by EnzChek® Myeloperoxidase (MPO) Activity Assay Kit (see methods). Results are expressed as relative fluorescence and are shown as mean values ± SD of three experiments. Statistical analysis demonstrates that the chlorination activity in neutrophil cell lysates is lower in SCN patients as compared to healthy subjects (p <0.05).
Figure 3. Lactoferrin expression and release in neutrophils of SCN patients under treatment with G-CSF. (A) Neutrophils isolated from patients 1, 2 and 3 (P1, P2 and P3) were stimulated with fMLP (10 nM), CXCL8 (10 ng/ml) or medium alone for 15 minutes. In the collected supernatants lactoferrin concentration was determined by ELISA and expressed as average + SD of two distinct experiments. Statistical analysis demonstrates that the amount of lactoferrin released in supernatants derived from SCN patients is significantly lower than lactoferrin detected in supernatants from control subjects (p<0.05). (B) Lactoferrin concentration in plasma of SCN patients (P1, P2 and P3) was determined in at least three samples. Average + SD of three distinct determinations are presented on y axis. Statistical analysis demonstrates that the amount of plasma lactoferrin is lower in SCN patients as compared to healthy subjects (p <0.05). (C) The Western Blot shown in Figure 1B and 2B has been reprobed with a specific anti-human lactoferrin antibody as described under methods.

Figure 4. Expression of cathepsin G, lysozyme and HNP in neutrophils of SCN patients during treatment with G-CSF. Cell lysates of neutrophils purified from patients 1, 2 and 3 (P1, P2, and P3), an healthy control subject (C1) and two idiopathic neutropenia patients (C3 and C4) were subjected to Western Blot analysis with antibodies raised against neutrophil cathepsin G, lysozyme, HNP precursor and HNP mature protein. β-actin was used to normalize protein levels.

Figure 5. NADPH oxidase expression and activity in neutrophils from SCN patients. (A) Cell lysates of neutrophils purified from two patients (P1 and P3) and two healthy subjects (C1 and C3) were subjected to Western Blot analysis with anti-gp91phox, p47phox, p67phox, p40phox, or p22phox specific antibodies. The gp91phox appears as a broad smear because it is highly glycosilated. β-actin was used to compare protein levels. (B) Neutrophils (2x10^5) from patient 1 (P1) and 3 (P3) and from an healthy subject (CTR) were stimulated with 100 nM fMLP or 20 ng/ml PMA for 60 min. When required cells were pre-incubated with 50 ng/ml G-CSF or GM-CSF for 30
min before fMLP addition. O$_2$ release was measured by cytochrome C reduction. Results shown are expressed as the mean value ± SD of three independent experiments. (C) Phagocytosis was assayed in neutrophils from patient 2 (P2) and from a healthy subject (CTR) after 40 min incubation with opsonized Candida yeasts or zymosan (see methods). Phagocytic index is shown as mean value ± SD of ingested particles per 100 cells. In parallel, neutrophil (2x10$^5$) were stimulated with 200 µg/ml opsonized zymosan for 60 min and O$_2$ release was measured by cytochrome C reduction. Results are expressed as the mean value ± SD of two experiments. (D) Neutrophils from patient 2 (P2) and a healthy subject (CTR) were incubated with opsonized C. albicans blastospores or E. coli for 90 min. Neutrophils were then lysed, diluted and plated on Sabouroud or LB agar Petri dishes. The CFU were counted after an overnight incubation and the percentage of killing was evaluated as in methods. Results shown are expressed as the mean value ± SD of two experiments. Statistical analysis demonstrates that antimicrobial activities of neutrophils against C. albicans and E. coli are reduced in SCN patient as compared to a healthy subject (p < 0.05).
Figure 1

A

CTR

P1

P2

P3

B

kDa

50

36

P1  C1  C2  C3  P2  P3  C4

32 kDa

29 kDa

β-actin
Figure 2

A

CTR

P1

P3

B

kDa

64 -

50 -

36 -

22 -

16 -

P1  C1  C2  C3  P2  P3  C4

C

Relative fluorescence

MPO (ng/ml)  10  50  200

CTR  P2
Figure 3

A

B

C

kDa

77
Figure 4

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Figure 5

**A**

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**B**

O$_2$ nmol/2x10$^5$PMN/60min

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**C**

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<td>ops-Candida</td>
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O$_2$ nmol/2x10$^5$PMN/60min

| ops-zymosan      | 3.56±0.94  | 4.16±0.82  |

**D**

% killing

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G-CSF treatment of Severe Congenital Neutropenia reverses neutropenia but does not correct the underlying functional deficiency of the neutrophil in defending against microorganisms

Marta Donini, Stefania Fontana, Gianfranco Savoldi, William Vermi, Laura Tassone, Francesca Gentili, Elena Zenaro, Daniela Ferrari, Lucia Dora Notarangelo, Fulvio Porta, Fabio Facchetti, Luigi Daniele Notarangelo, Stefano Dusi and Raffaele Badolato