The combination of chemotherapy and systemic immunotherapy with soluble B7–immunoglobulin G leads to cure of murine leukemia and lymphoma and demonstration of tumor-specific memory responses

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Major mechanisms underlying poor immune responses to autologous tumor-associated antigens are overwhelming tumor kinetics and the absence of effective T-cell costimulation by antigen-presenting cells. To address these issues, leukemia and lymphoma mice were treated with the combination of chemotherapy and systemic immunotherapy with recombinant soluble murine B7–immunoglobulin G (IgG) molecules. In this report, 3 murine models were used, a radiation-induced SJL acute myeloid leukemia, a transplantable spontaneous SJL lymphoma, and the C57BL/6 EL-4 thymic lymphoma. Various treatment modalities were evaluated: single treatments with either B7-IgG or chemotherapy as well as combination therapies. The results demonstrate the following: (1) in all tumor models, the combination of chemotherapy and soluble B7-IgGs is more potent than either therapy alone, leading to cure of tumor-bearing animals; (2) the therapeutic responses are T-cell–dependent, because combined therapy is not efficacious in severe combined immunodeficient mice; (3) the rejection of tumor cells leads to the development of tumor-specific immunity, because cured mice are immune to the rejected tumor but not to a different syngeneic tumor; and (4) 51Cr release assays show that rejection of tumor cells leads to the development of very potent tumor-specific cytotoxic T-lymphocyte activity. On the basis of these results, it is proposed that chemotherapy-mediated tumor reduction, together with consequent augmented tumor-antigen presentation to activated T cells, are primary mechanisms leading to curative responses. The safety profile of the B7-IgG fusion proteins and their synergy with chemotherapy strongly suggest that the combination regimen is a promising strategy in cancer treatment. (Blood. 2001;97:2420-2426)
from these studies was the lack of efficacy against bulk disease; in this context, B7.1-based tumor vaccines were efficacious only when given in early stages of the disease. It has been proposed that this is simply due to drastically increased replication rates of tumor cells that may preclude the opportunity for developing an adequate immune response.25,26

In an attempt to overcome the practical problems currently related to genetic modification of patients' tumor cells, we have developed soluble B7–immunoglobulin G (IgG) (B7.1-IgG and B7.2-IgG) fusion proteins and have recently shown their efficacy as single therapy in several murine solid-tumor models.27 In this report, we tested the efficacy of soluble B7-IgG in murine systemic leukemia and lymphoma models. Our results show that in poorly immunogenic and aggressive systemic tumor models, single therapy with either B7-IgG fusion proteins or cytoreductive chemotherapy has variable efficacy, but neither therapy alone is able to cure tumor-bearing animals. However, when both systemic immunotherapy and chemotherapy are combined, leukemia and lymphoma tumors are rejected and treated mice are cured.

Materials and methods

Mice

Female SJL/J mice and female C57BL/6 severe combined immunodeficient (SCID) mice, 6 to 8 weeks old, were purchased from Jackson Laboratories (Bar Harbor, ME). Female C57BL/6 mice, 6 to 8 weeks old, were purchased from Harlan Sprague-Dawley (Indianapolis, IN). The animals were kept at the animal facility of Genetics Institute (Cambridge, MA) according to the Institute’s guidelines.

Tumor models

We used 3 murine hematological tumor models in this study: a radiation-induced SJL/J acute myeloid leukemia (AML) model,24 a transplantable spontaneous SJL lymphoma (Ly) that has developed at Genetics Institute’s animal facility and has been passed in vivo several times, and the C57BL/6 EL-4 thymic lymphoma. In all experiments, frozen spleen (AML) or lymph node (Ly) mononuclear cells (isolated from moribund leukemia or lymphoma mice) were used. The EL-4 cell line was purchased from American Type Culture Collection (Rockville, MD) and was maintained in vitro at 37°C in RPMI 1640 medium containing 10% fetal calf serum, 2% glutamine, and 1% penicillin-streptomycin. For the establishment of tumors, 105 AML or Ly cells were injected intravenously (IV) in the tail vein of SJL mice, and 106 EL-4 cells were injected subcutaneously (SC) in the flank of C57BL/6 mice. Tumor-bearing animals either died within 2 to 35 days after tumor inoculation (AML/Ly) or were killed when tumors reached a size of approximately 400 to 600 mm2 (EL-4).

Murine recombinant B7-IgG fusion proteins

The development and purification of soluble B7.1- and B7.2-IgG fusion proteins have been previously described.27 Briefly, complementary DNA encoding the signal and the extracellular domains of murine B7.1 or B7.2 were joined to the genomic DNA encoding the hinge–CH2-CH3 domains derived from a murine IgG2a antibody. B7-IgG molecules are dimeric and can bind to CD28 and/or CTLA-4 on T cells and to Fc receptors (FcerS) on macrophages and APCs. In some experiments, either mutated B7.2-IgG fusion protein (the IgG2a region is mutated to ablate binding to FcR) or control isotype murine IgG2a (Adriamycin RDF, Pharmacia & Upjohn, Kalamazoo, MI), 2 consecutive intraperitoneal (IP) injections of 200 mg/kg, 6 hours apart, on days 7 and 21; doxorubicin hydrochloride (Sigma, St Louis, MO), 1 IP injection of 100 mg/kg on days 7 and 21; and complement C1b) or control isotype murine IgG2a was used.

Chemotherapy protocols

Leukemia protocol was as follows: cytarabine (Cytosar-U, Pharmacia & Upjohn, Kalamazoo, MI), 2 consecutive intraperitoneal (IP) injections of 200 mg/kg, 6 hours apart, on days 7, 14, and 21; doxorubicin hydrochloride (Adriamycin RDF, Pharmacia & Upjohn), 1 IP injection of 6 mg/kg on days 7 and 14. Lymphoma protocol was as follows: cyclophosphamide monohydrate (Sigma, St Louis, MO), 1 IP injection of 100 mg/kg on days 7 and 21; doxorubicin hydrochloride, 1 IP injection of 6 mg/kg on days 7 and 21. The choice of the compounds was based on their use in clinical leukemia and lymphoma chemotherapy protocols.

Combination therapy

Mice were injected with live tumor cells on day 0 and subsequently treated with chemotherapy and one injection of 100 µg B7.2-IgG (or 50 µg of each B7.2- and B7.1-IgG) on days 7, 14, 21, and 28. B7-IgG was diluted in 200 µL of sterile saline and administered by subcutaneous injections at the described doses and schedules. Tumor size in EL-4 and survival of mice in AML and Ly were monitored twice weekly. Except for survival experiments, mice were killed when tumors reached a size of approximately 400 to 600 mm2.

Proliferation assays

Spleens were harvested from nontreated leukemic (AML) mice and from mice that had received 1 (day 7) or 2 (days 7 and 14) courses of chemotherapy (AML plus chemotherapy). Single-cell suspensions were prepared as previously described. Cells were cultured at 2 × 106 cells per well in flat-bottomed 96-well plates coated with a suboptimal dose (100 ng/mL) of anti-CD3 monoclonal antibody (mAb) 145-2C11 (Pharmingen, San Diego, CA) and increasing amounts of plate-bound B7.2-IgG. Response to costimulation with anti-CD3 plus anti-CD28 (1 µg/mL) was used as positive control. Proliferation of responder cells was measured after 72 hours by the incorporation of [3H] thymidine (1 µCi per well) for the last 18 hours of incubation. In tumor cell proliferation assays, 105 tumor cells per milliliter (2 × 105 cells per well in U-bottomed 96-well plates) were cultured with 10 µg/mL of soluble B7.1- or B7.2-IgG or control IgG, and their proliferation was measured 72 hours after culture initiation.

Murine IL-2 enzyme-linked immunosorbent assay

Levels of murine IL-2 in culture supernatants were determined by a sandwich enzyme-linked immunosorbent assay that used specific antiumarin mAbs for capture and detection (Endogen, Woburn, MA). The sensitivity of the assay is 5 pg/mL.

Immunostaining and flow cytometry analysis

Splenocytes were isolated from AML and AML-plus-chemotherapy mice and were stained as previously described.24 The following mAbs (Pharminigen) were used for flow cytometry studies: Gr-1 (RB6-8C5), CD3e (145-2C11), CD4 (L3T4), CD8a (53-67).

51Cr release cytoxic T-lymphocyte assays

Spleens were collected from mice 11 weeks after EL-4 tumor inoculation/rejection, and single-cell suspensions were prepared. Splenocytes (5 × 106) were cocultured with irradiated (7335 cGy) EL-4 cells (1 × 106) in 2 mL complete RPMI per well of a 24-well tissue-culture plate (Costar, Cambridge, MA). At 6 days later, splenocytes were harvested and used as effector cells in cytoxic T-lymphocyte (CTL) assays. EL-4 cells or control AML cells (2 × 106) were labeled with 200 µCi 51Cr (New England Nuclear, Boston, MA) for 90 minutes, washed twice, and used as targets (5000 per well) in the CTL assays. The standard 4-hour CTL assays were set up with various effector-to-target (E/T) ratios as previously described.

Statistical analysis

Most individual experiments consisted of 10 mice per treatment group. The data analyzed represent the results of 2 or 3 individual experiments. The statistical survival analysis was performed by means of the standard Mantel-Cox log-rank test. Cytokine values and proliferation results are the mean ± SD. The statistical significance between various groups was analyzed by Student t test.
Results

Effects of systemic chemotherapy

In an attempt to mimic clinical situations as closely as possible, we used chemotherapy regimens consisting of drugs that are major components of leukemia and lymphoma clinical protocols. Because most cytoreductive drugs target dividing cells, it is conceivable that any proliferating cells of the immune system would also be affected. Therefore, we first determined what the in vivo effects of chemotherapy were on both tumor cells and T cells. We treated leukemic mice on days 7 and 14 with Ara-C plus doxorubicin (AML plus chemotherapy). On day 14 or 21 (1 week after the first or second dose of chemotherapy), AML-plus-chemotherapy mice and control AML mice were killed, and their splenocytes were used for in vitro studies. Immunostaining and flow cytometric analysis on day 14 showed that after one course of chemotherapy, AML-plus-chemotherapy spleen had decreased numbers of Gr-1<sup>+</sup> cells (marker for the leukemic cells)<sup>23</sup> and increased numbers of T cells as compared with AML spleen (data not shown). The effect of chemotherapy was more prominent in day-21 flow cytometric analysis. As shown in Figure 1A, there was a dramatic decrease in the percentage of Gr-1<sup>+</sup> cells in AML-plus-chemotherapy spleen as compared with AML (28% vs 87.2%), whereas the percentage of CD3, CD4, and CD8 T cells was significantly increased in AML-plus-chemotherapy spleen (29.1% vs 10.3%, 16.3% vs 2.4%, and 7.2% vs 0.94%, respectively).

We next determined the proliferative and cytokine response of AML-plus-chemotherapy splenocytes to in vitro B7.2-IgG costimulation. On day 14, no differences were observed between AML-plus-chemotherapy and AML splenocytes (Figure 1B), and both appeared to proliferate more vigorously with B7-IgG costimulation than with costimulation by anti-CD28 mAb. Furthermore, as shown in Figure 1C, IL-2 secretion in response to B7-IgG costimulation was comparable to stimulation through CD28. (The IL-2 levels in response to 5 μg/mL AML-plus-chemotherapy culture presented in Figure 1C were not observed in any of our other, similar in vitro costimulation experiments. These data are included because they are part of the series of other parameter measurements in the same experimental system.) Overall, no major differences were observed between AML-plus-chemotherapy and AML splenocytes, indicating that this chemotherapy regimen did not cause detectable immunosuppression of the leukemic mice with regard to thymidine uptake and cytokine secretion. The day-21 proliferation and IL-2 level assays were performed only on AML-plus-chemotherapy splenocytes (as shown in Figure 1A, day-21 AML spleens were heavily infiltrated with Gr-1<sup>+</sup> cells), and the results were comparable to those from day-14 assays (data not shown). Collectively, these results demonstrate that treatment of murine AML with doxorubicin and Ara-C significantly reduces the leukemic burden, without having a detrimental effect on in vitro T-cell proliferation and IL-2 secretion in response to B7.2-IgG costimulation.

In vitro effects of B7.2-IgG on tumor cells

Soluble B7.2-IgG fusion protein can potentially bind on CD28/CTLA-4 and FcγRs on cells. Because EL-4 cells express CD28 and all 3 tumor cell types express FcγRs, we investigated whether the in vitro direct effects of soluble B7.2-IgG were on tumor cells. Tumor cells were cultured as described in “Materials and methods,” and their immunophenotype (fluorescence-activated cell sorting analysis), viability (trypan-blue exclusion), and proliferative response (³H thymidine uptake) were examined. In all types of experiments, no significant differences were observed among the media, IgG, or B7-IgG cultures (data not shown), indicating that there is no direct effect of soluble B7.1- or B7.2-IgG on tumor cells with regard to their immunophenotype, viability, and proliferative profile.

Combination therapy leads to cure of leukemia and lymphoma mice

Single and combined treatment modalities (with B7.2-IgG, B7.1/B7.2-IgG, mutated form of B7.2-IgG, or control IgG) were evaluated in each tumor model, and no statistically significant differences were observed with the use of different sets of circumstances. Each experiment was repeated 2 to 3 times with similar results, and data from one representative experiment per
model will be presented in this report. In the experiment shown in Figure 2A, single B7.2-IgG or control IgG treatment in the AML model had no effect. In most experiments, AML mice treated with single chemotherapy had 1 to 2 weeks’ prolonged survival, as compared with untreated animals (P < .05), but eventually all mice developed lethal leukemia. However, when both treatments were combined, 50% of the mice were cured (Figure 2A).

In the EL-4 model, single therapy with either B7.2-IgG or chemotherapy alone had minimal or no effect. Treatment with B7.2-IgG or the mutated form of B7.2-IgG (which does not bind high-affinity FcγRs and complement C1q)27 plus chemotherapy led to cure of 70% and 60%, respectively (Figure 2B). In this model, a bimodal pattern of tumor growth was observed in 10% to 20% of treated mice; ie, the mice developed palpable tumor masses that then regressed during the following 2 weeks, indicating a therapeutic response to the combination regimen. In the AML and EL-4 models, treatment with both B7.1- and B7.2-IgG fusion proteins and chemotherapy showed comparable efficacy to treatment with B7.2-IgG plus chemotherapy (data not shown).

In the SJL Ly model, single therapy with fusion proteins showed no therapeutic effect (Figure 2C). Chemotherapy alone significantly prolonged the survival of treated mice (P < .01), but no cures were observed. In the same experiment, B7.2-IgG plus chemotherapy led to 90% cure, and the combination of B7-IgGs plus chemotherapy to 70% cure.

Collectively, these results demonstrate that, in all models tested in this study, the combination of B7-IgG fusion proteins with chemotherapy leads to cure of leukemia- and lymphoma-bearing mice.

**Therapeutic responses to combination therapy are T-cell–dependent**

It has been previously shown in tumor models that B7-IgG-mediated curative responses are CD8+ T-cell–dependent.27 To examine the role of T cells in the therapeutic responses to combination therapy, we evaluated single therapies (B7-IgG or chemotherapy) and combination therapy in T- and B-cell–deficient (SCID) mice. C57BL/6 SCID mice bearing EL-4 tumors were treated with B7.2-IgG, chemotherapy alone, or the combination of both. The tumors grew more rapidly in SCID mice than in normal C57BL/6 mice, and by day 28 after tumor inoculation, all mice had succumbed to lethal tumors. As shown in Figure 3, combination therapy had no therapeutic effect in SCID mice, clearly indicating the indispensable role of T cells for the efficacy of the combination regimen. Combination experiments in C57BL/6 SCID mice were also pursued in another systemic leukemia model (C1498 myeloid leukemia model), and similar results were observed (K.D.-J., unpublished data, March 2000).

**Combination therapy leads to long-lasting tumor-specific immunity**

One of the main goals of combining cytoreductive chemotherapy with immunotherapy is to activate the adaptive immune system and thus trigger the development of effector and memory cytotoxic T cells. Whereas effector T cells in this setting may have a beneficial role in eliminating residual tumor cells that have escaped chemotherapy, long-term disease-free survival can be achieved only when potential tumor relapses are under the critical control of memory
cytotoxic T cells. Therefore, we sought to determine if combination therapy was able to support the development of antitumor memory T cells. AML mice that had been cured with combination therapy were challenged 4 months later with live AML cells. As shown in Figure 4A, all mice were immune to challenge and rejected the leukemic cells. At 2 months after the AML challenge and rejection, the same mice were inoculated with syngeneic lymphoma (Ly) cells. As shown in Figure 4B, the challenged mice developed lethal lymphoma at the same time as naive mice that had been inoculated with Ly cells. These findings demonstrate that the mechanisms mediating curative responses in combination therapy also lead to the development of long-lived, tumor-specific memory cells.

Combination therapy leads to long-lasting tumor-specific CTL activity

To further characterize the antitumor memory responses of cured mice, we performed in vitro CTL assays. Spleens were harvested from mice that had been cured of EL-4 lymphoma (with B7.2-IgG plus chemotherapy or the combination of B7-IgG plus chemotherapy), and splenocytes were assayed for in vitro CTL activity. As shown in Figure 5, both groups of cured mice generated very potent cytolytic responses upon stimulation with EL-4 cells. The response was EL-4–specific (H-2b) because the same cells did not lyse alloantigen-presenting AML (H-2a) cells.

Discussion

In this report, we demonstrate that the addition of soluble B7.2-IgG to conventional leukemia and lymphoma chemotherapy regimens has remarkable synergistic effects in murine leukemia and lymphoma models, leading to curative T-cell–dependent antitumor responses. The establishment of long-lived tumor-specific memory T cells was confirmed with in vivo challenge experiments and in vitro CTL assays.

Various mechanisms may contribute to the synergy observed in combination therapy, as opposed to the limited efficacy of single therapies in our studies. Chemotherapy alone, as shown in the AML model, significantly reduces the leukemic burden, but cannot eradicate minimal residual disease. Apparently, this is the reason chemotherapy-treated animals finally succumb to lethal leukemia, a situation resembling the relatively short duration of remission observed in AML patients with partial remission to induction chemotherapy. In addition to tumor-burden reduction, cytoreductive chemotherapy generates a plethora of TAAs, which, as is the case with all non–self-antigens, are eventually expressed in the context of MHC molecules on APCs and can potentially initiate antigen-specific T-cell activation. However, chemotherapy overall fails to generate clinically overt antitumor memory responses, primarily for 2 reasons: first, it temporarily reduces the T-cell pool by targeting the relatively low numbers of proliferating T cells, and second, the sudden availability of abundant tumor antigens is probably not accompanied by signals required for the maturation process of immature dendritic cells (DCs), the most potent APCs and initiators of immunity. In the presence of a maturation signal, DCs express higher levels of costimulatory and MHC molecules and can then activate resting T cells. In our studies, neither tumor reduction and increased TAA presentation (chemotherapy) nor T-cell activation (B7-IgG) alone can cure leukemia and lymphoma mice. The efficacy of the combination regimen suggests that B7-IgG soluble molecules are probably strengthening APC–T-cell interactions during antigen presentation, which in combination with chemotherapy-mediated tumor reduction can lead to curative immune responses.

How does soluble B7-IgG fusion protein work? Because of the nature of the fusion molecule, there is little doubt that in vivo it will bind...
to both CD28/CTLA-4 and FcγR-expressing cells. We have shown that B7.2-IgG indeed binds in vitro to CD3ε, Mac-1, and B220+ murine splenocytes.33 The binding of B7.2-IgG on T cells can potentially involve both ligands, CD28 and CTLA-4. Although at this point we have no experimental evidence what role, if any, CTLA-4 plays in this system, we have reproducible results showing that murine naive T cells proliferate and secrete cytokines (IL-2 and interferon [IFN]–γ) in response to in vitro costimulation with soluble B7-IgG.33 One possible explanation why CD28 binding, rather than CTLA-4–mediated suppression of T cells, might be the primary pathway leading to activation is that CD28 is constitutively expressed on most T cells, whereas no detectable levels of CTLA-4 molecules are expressed on nonactivated T cells.34,35 All together, the in vitro and in vivo results strongly suggest that B7.2-IgG binding on T cells mediates an activation signal, and that most probably CD28 is the dominant responding molecule. Future studies with the use of CD28 knockout (KO) mice will provide more information on the mechanistic function of B7.2-IgG in this matter.

The indispensable role of T cells in combination therapy was confirmed in studies with SCID mice, in which all experimental groups, with or without treatment, rapidly developed lethal tumors. The relatively complex nature of combination therapy has made us hesitant to use additional in vivo compounds for CD4+ and CD8+ T-cell depletion and determination of each subset’s role in combination therapy. We plan, however, on specifically addressing the role of CD4+ and CD8+ cells in future work by (1) in vivo selective depletion of CD4/CD8 cells at the time of rechallenge, (2) adaptive transfer experiments, and (3) T-cell depletion of long-term survivors and determination of whether there is tumor regrowth at that time.

In addition to CD28/CTLA-4, B7.2-IgG also binds in vitro (and potentially in vivo) to FcγR-expressing cells. Experiments in vitro have shown that soluble B7.2-IgG binding in SCID macrophages induces up-regulation of several activation-related molecules, including B7.1, CD40, MHC class I, and adhesion molecules.33 It is conceivable that similar in vivo effects of B7.2-IgG binding on APCs (which are loaded with an excess of tumor antigens) would potentially have significant contribution to the curative responses of the combination regimen. In our studies, the in vivo use of the mutated form of B7-IgG had comparable efficacy to the nonmutated form, indicating that abrogation of high-affinity (CD64) FcγRI binding is dispensable in the combination therapy. It has been shown that murine low-affinity FcγRII (CD16)–mediated endocytosis and antigen presentation are dependent on the immunoreceptor tyrosine-based activatory motif (ITAM) of the associated gamma-chain.36–38 Since murine low-affinity FcγRII (CD32) contains only immunoreceptor tyrosine-based inhibitory motif (ITIMs) signaling motifs and therefore mediates inhibitory signals,39 we believe that any FcγR–binding–related activatory responses occur through FcγRII (CD16)–mediated signals. We aim to specifically define the role of FcγR-B7.2-IgG in vivo interactions with the use of various FcγR KO mice.

A critical issue in the combination regimen is the appropriate timing for the immunoadjuvant. It is well known that chemotherapy induces peripheral leukopenia whose severity and duration depend on the type and dose of chemotherapeutics.40 Whereas high-dose, myeloablative regimens are accompanied by prolonged peripheral leukopenia, conventional chemotherapy is related primarily to neutropenia and, to a lesser extent, to lymphopenia. Several studies in humans have shown that among lymphocyte subsets, B cells are the cells most affected by chemotherapy, followed by CD4+ T cells, with CD8+ T cells remaining relatively well preserved.41-44 Interestingly, the remaining clonogenic T lymphocytes derived from acute leukemia patients with therapy-induced leukopenia have shown a broad cytokine response to in vitro activation.45 We have previously shown in the SJL AML model that absolute peripheral lymphocyte numbers return to normal within a week after chemotherapy with cytarabine and doxorubicin.26 On the basis of these findings, we reasoned that the interval between injections of B7-IgG should be 1 week or longer. Because 2 of the B7-IgG injections in the lymphoma models and 3 in the AML model were given on the same day with chemotherapy, it is conceivable that a percentage of T cells responding to B7/CD28 interactions by proliferation would be affected by chemotherapy. However, the favorable clinical outcome of the combination regimen in our studies suggests that a significant number of B7-IgG–activated T cells can nevertheless resist chemotherapy and eventually become long-lived antitumor memory cells.

A major potential advantage for the use of B7.2-IgG as an adjuvant to chemotherapy is its expected high safety profile. The protein has not shown any in vivo toxicity, even when injections of 500 μg were given to mice (unpublished results, May 1999). Studies in the MethA tumor model (which has reproducibly shown therapeutic responses to single B7.2-IgG treatment) have shown that the efficacy of the fusion protein is not ablated in IFN-γ−/− or IL-12−/− KO mice (unpublished results, October 1999). These observations dissociate the immune mechanisms mediated by IFN-γ and IL-12 and the efficacy of B7-IgG in therapeutic tumor models, and suggest that cytokines with less toxicity than IL-12 and IFN-γ may characterize the microenvironment, where tumor presentation occurs, or, alternatively, that B7-IgG induces cytokines only in specialized microenvironments.

The combination of immunotherapy with chemotherapy is an emerging form of cancer treatment. With the addition of an immune-boosting agent that, in principle, forces “provoked” immunity, conventional cancer therapy could conceivable be made more effective without increasing its toxicity.46 This may be manifested in greater durability of response rather than absolute clinical response rate. It is anticipated that individual immunomodulatory compounds will not synergize with all cytotoxic drugs, owing to differential immunosuppressive effects, and at present it is a great challenge to identify successful combinations in preclinical tumor models. The chemotherapy regimens used in this report apply beyond leukemia and lymphoma, since the combination of anthracyclines and cyclophosphamide is broadly used in the clinic and is one of the most widely used regimens in breast cancer treatment.47 The safety profile of combining B7.2-IgG with chemotherapy in preclinical tumor models, together with the potent therapeutic effect, has directed our efforts toward the development of strategies for clinical evaluation.

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References


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