Murine hematopoietic stem cell characterization and its regulation in BM transplantation

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Using 5-color fluorescence-activated cell sorting, we isolated a subset of murine pluripotent hematopoietic stem cells (PHSC) with the phenotype Lin^{-} Sca^{+} kit^{+} CD38^{-} CD34^{-} that appears to fulfill the criteria for most primitive PHSC. In the presence of whole bone marrow (BM) competitor cells, these cells produced reconstitution in lethally irradiated primary, secondary, and tertiary murine transplant recipients over the long term. However, these cells alone could not produce reconstitution in lethally irradiated recipients. Rapid proliferation of these cells after BM transplantation required the assistance of another BM cell subset, which has the phenotype Lin^{-} Sca^{+} kit^{+} CD38^{-} CD34^{-}. (Blood. 2000;96:3016-3022)

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Introduction

Pluripotent hematopoietic stem cells (PHSC) are a population of cells that reside in the bone marrow (BM) and maintain all cells of lymphoid and myeloid lineage over the long term. In mice, the frequency of long-term reconstituting (LTR) PHSC is about 1 in 100,000 whole BM cells. In accordance with their characteristics after BM transplantation (BMT), cells in murine BM with multilineage repopulating ability can be divided into 2 groups: the LTR cells, which can support hematopoiesis for more than 6 months in irradiated recipients, and the short-term reconstituting (STR) cells, which can repopulate blood elements for several weeks. Using counterflow centrifugal elutriation, Jones et al separated murine BM and found that cells with the STR property provided unsustained early engraftment in BM, whereas cells with the LTR property provided sustained but delayed engraftment in BM, spleen, and thymus. However, the cells that produced sustained repopulation could not protect animals from lethal irradiation. On the basis of these types of observations, it has been hypothesized that LTR cells proliferate slowly and can produce only delayed but sustained engraftment. Thus, to detect the subpopulation, or subset, of PHSC that can produce long-term engraftment, it appeared necessary to cotransplant early- engraftment cells along with the putative LTR cells into lethally irradiated recipients so that the host would survive the initial aplasia. The LTR cells are assumed to be the “true” stem cell.

PHSC can be isolated and characterized according to their immunophenotype by using fluorescence-activated cell sorting (FACS) and antibodies to the cell-surface markers. Spangrude et al defined a BM cell subset with the surface markers Lin^{-} Sca^{-} Thy1^{+} CD34^{-}. These cells were found to be 1000 to 2000 times enriched for the subset of PHSC with LTR ability. Further characterization of these cells indicated that they represent a heterogeneous population that includes both LTR cells and cells able to provide only STR ability. Using 4-color FACS, Osawa et al demonstrated that the most active murine LTR PHSC are CD34 negative (CD34^{+}). A similar proposal was made regarding human stem cells. Three groups have reported data showing that human CD34^{+} cells have LTR activity and may be more primitive than CD34^{-} cells. In addition, Randall et al studied another surface marker, CD38, and reported that murine LTR PHSC are CD38^{-}.

To characterize murine PHSC in more detail and to study their proliferation after BMT, we developed and used 5-color FACS analysis and cell fractionation. The subsets of PHSC that were isolated were studied with long-term competitive repopulation and short-term repopulation assays. Using these approaches, we determined that the most active LTR cells in mouse BM are lineage-negative cells with Sca^{+} kit^{+} CD38^{-} CD34^{-} (hereafter abbreviated 38^{-}34^{-}) surface markers and that another lineage-negative subset, with the phenotype Sca^{+} kit^{+} CD38^{-} CD34^{-} (38^{-}34^{-}), plays an important role in supporting the rapid proliferation of 38^{-}34^{-} cells after BMT.

Materials and methods

Mice

C57BL/6J (Ly5.2) male mice were obtained from B & K Universal Inc (Fremont, CA) and used as the irradiated recipients. Congenic C57BL/6J-Ly5.1-Pep3b (Ly5.1) mice and the first-filial-generation (F1) hybrid of Ly5.2 and Ly5.1 mice were bred at the Jackson Laboratory (Bar Harbor, ME) and the animal facility at the University of Southern California (USC; Los Angeles, CA) and used as sources of the hematopoietic progenitor population. All animals were housed under specific pathogen-free conditions and given acidified drinking water and autoclaved Chow ad libitum. Mice used in the experiments were 8 to 12 weeks of age. The study protocol was approved by the USC Animal Care and Use Committee.
Antibodies

The antibodies used in the lineage cocktail were anti-Mac-1 (M1/70), anti-Gr-1 (RB6-8C5), anti-B220 (RA3-6B2), anti-CD3e (145-2C11), anti-CD4 (RM4-5), anti-CD5 (53-7.3), anti-CD8a (53-6.7), and anti-erythroid (Ter119); all were biotinylated when used. Other antibodies were anti-Ly5.2 (104), anti-Ly5.1 (A20), anti-c-kit (2B8) labeled with allophycocyanin (APC), anti-Sca-1 (E13-161.7) labeled with phycoerythrin (PE), and anti-CD34 (RAM34) labeled with fluorescein isothiocyanate (FITC). All antibodies listed above were from Pharmigen (San Diego, CA). Anti-CD38 (NIM-R5) labeled with red 613 (R613) was conjugated under contract by Southern Biotech (Birmingham, AL). Streptavidin-conjugated peridinin chlorophyll protein (perCP) was from Becton Dickinson (San Jose, CA). The goat antirat or streptavidin-conjugated magnetic beads were from Miltenyi Biotech Inc (Auburn, CA).

Preparation and isolation of hematopoietic progenitors

BM cells were harvested from the femurs and tibias of Ly5.1 or F1 mice. After lysis of red blood cells with ammonium chloride lysis buffer (Ortho-mune Lysing Reagent; Ortho, Raritan NJ), cells were stained with biotinylated antibodies to lineage markers. Lin<sup>−</sup> cells were depleted with streptavidin-conjugated magnetic beads by using a CS column (Miltenyi Biotech). The lineage-depleted cells were collected and incubated with perCP and streptavidin, anti-Sca-1 (PE), anti-c-kit (APC), anti-CD34 (FITC), and anti-CD38 (R613). Stained cells were sorted with a customized Elite machine (Coulter, Miami, FL) equipped with a 15 mW argon laser tuned at 488 nm (for FITC, PE, R613 and perCP excitations) and a 10 mW helium-neon laser tuned at 610 nm for APC excitation. The fourth and fifth photomultiplier tubes (PMTs) were replaced with customized PMTs (4526A photomultiplier; Burle Industries, Lancaster, PA) with increased sensitivity in the higher wavelengths. Forward light scatter was detected with a 488 bp10 and an ND1.0 filter. For FITC, PE, R613 and perCP, 520 to 530, 555 to 595, 605 to 615, and 670 to 680 filters, respectively, were used; for APC, a 670 to 680 filter was used. Because both perCP and APC were detected with the fifth PMT, a time delay of 40 milliseconds was established for the perCP signal. Compensation was adjusted to achieve optimal signals from each fluorochrome when used simultaneously. Restricted sorting variables were chosen with the purity-1 mode, 1-drop-sort envelope, and coincidence-abort system on. Residual erythrocytes, debris, and doublets were excluded by forward- and side-scatter gating. The carryover lineage-positive cells were excluded by gating out the perCP-positive cells. Different subsets were sorted according to the gating variables.

Short- and long-term repopulation assays

The sorted cells were mixed with different numbers of competitor cells and transplanted through a tail vein into lethally irradiated mice that received a single (lethal) dose of 9.5 Gy from dual, opposed sources of cesium 131. For analysis of reconstitution in the mice, either BM or peripheral blood (PB) cells (from the tail) were collected in phosphate-buffered saline (PBS) and assayed for the presence of Ly5.1 (donor) cells of each lineage. Red blood cells were lysed with ammonium chloride lysis buffer and washed with PBS and 1% bovine serum albumin. The remaining nucleated cells were stained for lineage markers and Ly5.1 antibody. Anti-CD3, anti-CD4, and anti-CD8a were used to identify T cells, B220 was used for B cells, anti-Mac-1 for macrophages, and anti-Gr1 for granulocytes.

Assay of colony-forming units–spleen (CFU-S)

CFU-S assays were performed as described previously. Sorted cells from Ly5.1 mice were injected through the tail vein into lethally irradiated animals. Twelve days after the injection, the mice were killed and the spleens were removed and fixed. Macrosopic colonies were counted by inspection.

Results

38<sup>−</sup>34<sup>−</sup> cells are the subset of PHSC with the greatest LTR ability. Previous reports indicated that the LTR activity is in the Lin<sup>−</sup> Sca<sup>−</sup><sup>+</sup><sup>kit</sup> fraction of PHSC. Our data confirmed this observation (data not shown). Lineage-negative BM cells that were positive for Sca<sup>−</sup> and c-kit expression (Figure 1A) were further fractionated by using the expression of both CD38 and CD34 (Figure 1B). In lineage-negative Sca<sup>−</sup><sup>kit</sup> BM cells, there are 4 possible CD38/CD34 expression profiles: Sca<sup>−</sup><sup>kit</sup><sup>−</sup> CD38<sup>−</sup> CD34<sup>−</sup> (38<sup>−</sup>34<sup>−</sup>), Sca<sup>−</sup><sup>kit</sup><sup>−</sup> CD38<sup>+</sup> CD34<sup>−</sup> (38<sup>+</sup>34<sup>−</sup>), Sca<sup>−</sup><sup>kit</sup><sup>+</sup> CD38<sup>−</sup> CD34<sup>+</sup> (38<sup>−</sup>34<sup>+</sup>), and Sca<sup>−</sup><sup>kit</sup><sup>+</sup> CD38<sup>−</sup> CD34<sup>−</sup> (38<sup>−</sup>34<sup>−</sup>) cells. The frequency of each of these subsets in whole BM with our gating variables is shown in Table 1. Post-sorting analysis of the sorted cells indicated that the

Figure 1. Flow cytometric analysis of the surface-marker–expression profile of murine bone marrow (BM) cells. Lineage-positive cells were removed by CS column before flow cytometry. (A) Expression of Sca-1 and c-kit on the cell surface was gated as shown; Lin<sup>−</sup> Sca<sup>−</sup><sup>kit</sup> cells were gated as shown in box A. (B) Expression of CD38 and CD34 on the cell surface of the Lin<sup>−</sup> Sca<sup>−</sup><sup>kit</sup> cells was used to separate the cell population shown in box A in Figure 1A into the following 4 subsets: Sca<sup>−</sup><sup>kit</sup><sup>−</sup> CD38<sup>−</sup> CD34<sup>−</sup> (38<sup>−</sup>34<sup>−</sup>), Sca<sup>−</sup><sup>kit</sup><sup>−</sup> CD38<sup>+</sup> CD34<sup>−</sup> (38<sup>+</sup>34<sup>−</sup>), Sca<sup>−</sup><sup>kit</sup><sup>−</sup> CD38<sup>−</sup> CD34<sup>+</sup> (38<sup>−</sup>34<sup>+</sup>). Two other subpopulations examined (Figure 3) were Sca<sup>−</sup><sup>kit</sup><sup>−</sup> CD38<sup>−</sup> CD34<sup>−</sup> (38<sup>−</sup>34<sup>−</sup>) and Sca<sup>−</sup><sup>kit</sup><sup>−</sup> CD38<sup>−</sup> CD34<sup>−</sup> (38<sup>−</sup>34<sup>−</sup>) (rectangular gates). Cells in each population were sorted and collected for analysis in a competitive repopulation assay.
Individual subsets were isolated by 5-color fluorescence-activated cell sorting (FACS). Then, 10, 45, or 100 Ly5.1 donor cells and the indicated number of Ly5.2 whole bone marrow (WBM) competitor cells were injected into lethally irradiated Ly5.2 mice. Peripheral blood (PB) was collected at various times during the next 12 months and examined for engraftment of Ly5.1 donor cells. Only the data from 12 months after bone marrow transplantation (BMT) are shown. The frequency of formation of the 3 subsets we originally had reconstitution with 100 Ly5.1 cells (PHSC). Bone marrow transplantation (BMT) was performed as follows: 100 Ly5.1 donor cells (from 1 of the 3 subsets, 38°, 38°, or 38°) and 2 x 10^5 Ly5.2 whole BM competitor cells were transplanted into Ly5.2 lethally irradiated mice. Blood samples were collected at the indicated times after BMT and examined for the presence of Ly5.1 multilineage reconstitution (marked on y-axis). The data suggest that all 3 subsets have short-term multilineage reconstituting ability but that only the 3° subset of PHSC has marked long-term reconstituting ability. T indicates T lymphocytes; B, B lymphocytes; M, macrophages; and G, granulocytes. Data are from 2 independent experiments using 3 to 6 mice per group in each experiment; the values presented are means ± SE.
over time (Figure 3). These data indicate that the cells with the CD38+ phenotype were more primitive and had better self-renewal ability than the CD38− cells, although both cell populations had the same level of CD34 expression.

All 3 subsets of PHSC (3834+, 3834−, and 38−34+) start proliferating in BM efficiently after transplantation

For successful long-term engraftment, injected stem cells must reach the proper microenvironment in the BM for proliferation, a process called homing. To compare the long-term engraftment ability of the cell subsets, it is important to determine whether the compared cells can home to and repopulate the BM with equal efficiency. We examined the reconstitution of infused cells in BM early after BMT (8 days) to provide an indicator of whether the cells homed to and repopulated the BM. With 2 × 105 Ly5.2 competitor cells, 200 Ly5.1 cells of each subset were injected separately into Ly5.2 lethally irradiated mice. For all 3 subsets of PHSC, donor-derived cells were detected 8 days after BMT in BM and 14 days after BMT in PB (Table 3). These data indicate that there does not appear to be any preference in regard to homing or proliferation of one cell population over the others in this experimental setting.

Efficient radioprotection requires both 3834− and 3834+ cells

Because we observed both early engraftment (8 days after BMT in BM and 14 days after BMT in PB) and sustained engraftment (12 months after BMT) with 3834− cells, we investigated whether 3834+ cells can function as both radioprotective (STR) cells and LTCs. With a fixed total cell number, either 300 cells (Figure 4A) or 1000 (data not shown), neither 3834− nor 3834+ cells alone could efficiently protect the mice from lethal irradiation. However, when the 2 subsets of PHSC were mixed in a 1:1 ratio (maintaining a total cell number of 300 or 1000 cells), 90% of the mice survived (in both experimental settings; Figure 4A).

Early proliferation of 3834+ cells after transplantation requires the presence of another subset of PHSC

The radioprotection study raised the question of why 3834− cells, which engrafted by 8 days after BMT in the competitive repopulation assay, could not protect lethally irradiated animals. Because 2 × 105 whole BM cells were supplied in the competitive repopulation assay but not in the radioprotection study, we wondered whether some other type of cell in the BM might be necessary to support early proliferation of the 3834− cells. Therefore, we examined the efficiency of engraftment of 3834− and 3834+ cells in the setting of the radioprotection study, i.e., with no competitor cells present. A total of 400 cells from either 1 subset or a mixture of 2 subsets identifiable by different genetic markers were used. Of 19 mice (9 individual experiments) that received 400 3834− cells, 2 had a low level (3%) of donor-cell reconstitution in the BM 8 days after BMT, whereas all the mice that received 3834+ cells or a mixture of cells had donor-cell engraftment (18.0 ± 6.2% and 13.0 ± 3.8%, respectively). We then determined the contribution of each subset in the mixed-cell group. As shown in Figure 5, all the reconstitution in

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Table 2. Reconstitution in secondary recipients from primary 3834+ donor cells

<table>
<thead>
<tr>
<th>Donor cells</th>
<th>Months after BMT</th>
<th>No. of mice</th>
<th>% Ly5.1</th>
<th>T and B lymphocytes</th>
<th>Macrophages (M) and granulocytes (G)</th>
</tr>
</thead>
<tbody>
<tr>
<td>WBM from 1st recipients (1 × 10⁶)</td>
<td>12</td>
<td>7</td>
<td>31 ± 5</td>
<td>T: 33.5 ± 2.9</td>
<td>M: 39 ± 3</td>
</tr>
<tr>
<td>Sorted Ly5.1 cells from 1st recipients (1.4 × 10⁶)</td>
<td>7</td>
<td>7</td>
<td>79 ± 9</td>
<td>T: 70.3 ± 8.2</td>
<td>M: 63 ± 6</td>
</tr>
<tr>
<td>Sorted Ly5.1 cells from 1st recipients (1 × 10⁶)</td>
<td>8</td>
<td>6</td>
<td>40 ± 8</td>
<td>47.5 ± 8.2</td>
<td>49.7 ± 36</td>
</tr>
</tbody>
</table>

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Table 3. Engraftment of subsets of PHSC after BMT

<table>
<thead>
<tr>
<th>PHSC subset</th>
<th>BM 8 days after BMT</th>
<th>BM 14 days after BMT</th>
<th>PB 14 days after BMT</th>
</tr>
</thead>
<tbody>
<tr>
<td>3834−</td>
<td>11.4 ± 3.2 (10)</td>
<td>19.0 ± 5.7 (11)</td>
<td>5.9 ± 1.3 (11)</td>
</tr>
<tr>
<td>3834+</td>
<td>14.5 ± 3.7 (9)</td>
<td>13.1 ± 3.8 (7)</td>
<td>6.6 ± 1.6 (6)</td>
</tr>
<tr>
<td>38−34+</td>
<td>14.5 ± 4.2 (10)</td>
<td>24.2 ± 8.0 (9)</td>
<td>7.7 ± 2.8 (8)</td>
</tr>
</tbody>
</table>

Two hundred Ly5.1 cells sorted from each subset of PHSC and 2 × 10⁵ Ly5.2 competitor cells were transplanted into lethally irradiated Ly5.2 recipient mice. BM and PB were analyzed for donor-cell engraftment (Ly5.1-positive cells) at the indicated times after BMT. The data represent a summary of 4 experiments. Plus-minus values are mean ± SE percentages; values in parentheses are the total number of mice in the experiment.
BM 8 days after BMT was from the $38^{+}34^{+}$ cells. However, reconstitution from $38^{+}34^{+}$ cells in BM was comparable with $38^{-}34^{+}$ cells 14 days after BMT and increased additionally 21 days after BMT. Twenty-eight days after BMT, more than 95% of donor-cell–derived engraftment in PB was from $38^{+}34^{+}$ cells (data not shown). These data suggested the hypothesis that the $38^{+}34^{+}$ cells prepared an environment for rapid proliferation of the $38^{-}34^{+}$ cells when the 2 subsets were cotransplanted. To test this hypothesis, we injected $38^{+}34^{+}$ cells 7 days before the injection of $38^{-}34^{+}$ cells (Figure 6A). Under these conditions, the engraftment of $38^{-}34^{+}$ cells was immediate, ie, it had occurred by 7 days after the second transplantation (Figure 6B).

Discussion

Stem-cell transplantation and stem-cell gene therapy have potential for broad clinical applications. However, there is still insufficient...
understanding of the regulatory mechanisms and kinetics of stem-cell proliferation after BMT. Although BMT is widely used clinically, it is a cumbersome procedure. In addition, efficient gene transfer into LTR human stem cells has been only partly successful, thereby inhibiting development of stem-cell gene therapy. We used a murine model system in studies to increase understanding of hematopoietic stem cells and the proliferation of stem cells after BMT.

The lineage-negative subset of PHSC that is positive for Sca-1, c-kit, and CD38 but negative for CD34 (the 38⁺34⁻ subset) appears to be the pivotal cell in the mouse stem-cell hierarchy. We found that 38⁺34⁻ cells have both long-term and short-term repopulating ability; they can engraft rapidly after BMT (8 days), they can produce reconstitution in animals over the long term (Table 1), they can effect reconstitution in animals receiving secondary and tertiary transplants (Table 2 and data not shown), and they represent 0.0022% of the nucleated cells in the BM (ie, approximately 2 cells/100 000). There is no explanation for this finding than that a low level of LTR activity is an intrinsic property of 38⁺34⁻ cells during cell sorting, we used stringent sorting conditions; thus, contamination seems less likely an explanation for this finding than that a low level of LTR activity is an intrinsic property of 38⁺34⁻, 38⁺34⁺, and 38⁻34⁻ cells.

The combined expression profile of CD38 and CD34 from the FACS analysis (Figure 1B) indicated that most Lin⁻ Sca⁺ kit⁺ CD34⁺ cells are CD38⁺ and that most of the CD34⁺ cells in the Lin⁻ Sca⁺ kit⁻ subset are CD38⁻. Therefore, our data support the conclusions of both Osaka et al⁺¹⁰ and Randall et al⁺¹¹ that the LTR cells in mice are cells with the immunophenotype Lin⁻ Sca⁺ kit⁻ CD34⁻ as well as Lin⁻ Sca⁺ kit⁺ CD38⁺.

Zijlmans et al⁺³ suggested that the early phase of engraftment after murine blood transplantation is mediated by hematopoietic stem cells, which are defined by the phenotype Lin⁻ WGA⁺ Rh⁻. Using modified rhodamine staining, Zijlmans et al subdivided this population into Rh⁻/Rho⁺ (VP⁺), which showed short-term repopulating activity, and Rh⁺/Rho⁻ (VP⁻), which showed long-term repopulating activity. We speculate that the cells with the phenotype Rh⁻/Rho⁺ might be the same cells as the 38⁺34⁺ cells in our study and that the Rh⁺/Rho⁻ cells might be the same as our 38⁺34⁻ cells. However, because the variables assessed in the study by Zijlmans et al were animal survival (> 4 weeks) and blood cell counts, it is difficult to identify the contribution from each subset when evaluating the early-engraftment kinetics. Our experimental design overcame this problem. Using Ly5.1 and F1 mice with different surface markers, we could specifically assess the repopulation from each subset in vivo.

The observation that rapid engraftment of 38⁺34⁻ cells appears to depend on the presence of 38⁺34⁺ cells (or their offspring) is intriguing (Figures 4 and 5). Do the 38⁺34⁻ cells have only a general supportive role or do they have a specific facilitative role? Because 38⁺34⁻ cells cannot begin to proliferate immediately when transplanted at the same time as 38⁺34⁺ cells (Figure 5), we asked whether an immediate proliferation of 38⁺34⁻ cells would occur if 38⁺34⁺ cells were transplanted 7 days before the 38⁺34⁻ cells to provide a proper environment. As shown in Figure 6, 38⁺34⁻ cells did begin to proliferate immediately when 38⁺34⁺ cells were already present. Although a general supportive function of 38⁺34⁻ cells cannot be excluded, a specific facilitative role (with an unknown mechanism) is more likely. Because the 38⁺34⁻ cells were in the mice for 7 days, the facilitating effect might not have come directly from these cells but from cells derived from them. The specific cell type involved in the putative facilitating effect and the mechanism of this effect are currently being studied.

We are not certain what is the immediate progeny cell to the 38⁺34⁻ cell, but our data suggest that the immediate pathway from the 38⁺34⁻ cell is first the 38⁺34⁺ subset and then the 38⁺34⁻ subset. This pathway is based on the percentage of long-term reconstitution in both primary and secondary recipients and the CFU-S data from the 3 subsets of PHSC. We speculate that the next cells in the pathway after the 38⁺34⁻ subset are those with increasing lineage markers. Studies to determine the complete maturation pathway of PHSC are under way.

If the 38⁺34⁻ cells are the most primitive stem cells with the highest LTR ability, should they be called the “true” stem cells? Because 38⁺34⁺ cells (or cells arising from them) appear to be required for 38⁺34⁻ cells to engraft, should the 38⁺34⁻ subset of PHSC be considered support cells? We believe not, because 38⁺34⁺ and 38⁺34⁻ cells do have some LTR ability and it is possible that 38⁺34⁻ cells also have some supporting ability (ie, an ability to support engraftment and proliferation). Therefore, the situation may be more like that proposed by Schofield⁺¹⁷ and Lansdorp⁺¹⁸ and reiterated by Donnelly et al,⁺¹⁹ all of whom speculated that it is perhaps inaccurate to label one cell type a stem cell because sets of cells may have varying degrees of stem-cell potential. We propose that the PHSC subsets 38⁺34⁻, 38⁺34⁺, and 38⁻34 both form a stem-cell compartment.
Recently, Sato et al.\(^2\) (with a covering analysis by Goodell\(^1\)) suggested the intriguing possibility that murine CD34\(^-\) and CD34\(^+\) stem cells may be able to convert into each other under some conditions. This hypothesis was based on data obtained from mice treated with 5-fluorouracil (5-FU). Although we did not examine the effect of 5-FU on individual subsets of PHSC, the concept that CD34\(^+\) cells can convert to CD34\(^-\) cells in vivo is compatible with the model of a stem-cell compartment described here.

What are the implications of our work for the clinical applications of BMT and stem-cell gene therapy? Because the primary LTR cells in mice are in the CD34\(^-\) fraction, this may also be the location of the primary LTR cells in humans, as was suggested previously by Goodell et al.,\(^1\) Bhatia et al.,\(^12\) and Zanjani et al.\(^13\) As pointed out by Donnelly et al.,\(^19\) CD34\(^+\) LTR cells in mice are 100 times more abundant than CD34\(^-\) cells, but the CD34\(^+\) cells are more efficient in long-term reconstitution after BMT than the more abundant CD34\(^-\) cells. Currently, clinical protocols using stem cells are based on purification of human CD34\(^+\) hematopoietic cells. If conditions for expanding CD34\(^-\) PHSC in culture were identified, perhaps far fewer cells would need to be given to patients to effect reconstitution. It is also possible that for human CD34\(^+\) PHSC, a different set of transduction conditions is required for gene transfer and that therefore the current difficulty in carrying out stem-cell gene therapy successfully may be partly due to targeting the wrong (or a less efficient) stem cell.

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References

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