Immune thrombocytopenic purpura (ITP) is an autoimmune disease in which platelet antibodies cause increased platelet consumption, sometimes leading to hemorrhage. The destruction of platelets is mediated by the reticuloendothelial system (RES), particularly by splenic and hepatic macrophages, and management of the disease is associated with concept of decreasing the RES activity. In fact, it includes splenectomy, corticosteroids; intravenous high-dose gamma globulin (IVIg); anti-D immunoglobulins, danazol, and immunosuppression. In recent years the importance of the Fcγ receptors in the uptake of platelets in ITP has been confirmed. Moreover, the available data do not support its use.

In this report we analyze the role of hepatic and splenic macrophages in a mouse model of ITP induced by antplatelet antibodies (Ab). The depletion of these cells was carried out by the intravenous injection of lip-clod. This macrophage “suicide” method has been used in different experimental models and has important characteristics such as: (1) lip-clod inhibits the clearance of IgG-sensitized autologous red cells in mice; (2) the intravenous injection of lip-clod ensures that platelet count is restored to initial levels in these animals. The bleeding times in lip-clod–treated animals was not different from those in control animals. The results presented in this study demonstrate that lip-clod treatment can be effective in the management of experimental ITP.

Materials and methods

Mice

BALB/c mice were bred in the animal facility of the Academia Nacional de Medicina, Buenos Aires. Male and female mice aged 14 to 16 weeks and weighing 23 to 26 g were used throughout the experiments. They were...
maintained under a 12-hour light-dark cycle at a temperature of 22°C ± 2°C and fed with standard diet and water ad libitum. The experiments were conducted according to principles set forth in the Guide for the Care and Use of Laboratory Animals.28

**Liposome-encapsulated clodronate**

Clodronate (dichloromethylene bisphosphonate) was provided by Roche Diagnostics, Mannheim, Germany. Lip-clod was prepared using 86 mg of phosphatidylcholine (Lipoid EPC; LIPOID, Ludwigshafen, Germany), 8 mg of cholesterol (Sigma Chemical Co, St Louis, MO), and clodronate (0.7 mol/L), in a final volume of 4 mL as previously described.25 This lip-clod will be referred to in the text also as original preparation or lip-clod 1:1. Empty liposomes were prepared under the same conditions in phosphate-buffered saline (lip-PBS). The intravenous injection of 0.1 mL/kg body weight of this lip-clod suspension induces the complete depletion of splenic and hepatic macrophages within 24 hours.24

**Preparation of platelets and platelet counts**

Platelet-rich plasma (PRP) was prepared as previously outlined.29 Briefly, blood from mice was collected in plastic tubes containing sodium citrate (3.8% w/v), pH 7.4. After centrifugation at 200 g for 10 minutes, PRP was removed and pooled. Platelet-poor plasma (PPP) was obtained by centrifugation of platelets at 1800g for 10 minutes. When necessary, the concentration of platelets was adjusted by addition of PPP. Platelets were counted in a hemocytometer using a buffer containing ammonium oxalate 1% for erythrocyte lysis. In agreement with others,30,31 the range for platelet count in our BALB/c mice was from 350 to 500 × 10^9/µL (n = 75).

**Rabbit-antimouse platelet antiserum**

Polyclonal antibody against mouse platelets was prepared in rabbits. For this purpose, a pool of PRP of BALB/c mice was centrifuged at 200 g for 10 minutes to eliminate erythrocytes. The supernatant was centrifuged at 1800g for 20 minutes, the platelets resuspended, and injected intravenously into rabbits (10^8 platelets/dose) at days 1, 15, 30, and 40. Ten days later the animals were bled. The sera were adsorbed with BALB/c mice red blood cells to remove a weak antimouse erythrocyte activity. The antiplatelets antibodies belong to the IgG class (purified by Sepharose G; Sigma). To avoid a partial blockade of the RES by IgG aggregates we used, like others,32,33 dilutions of whole serum instead of purified IgG.

**Labelling of platelets with 111In-oxine**

Platelet labeling was performed carefully to avoid cell damage. A pool of blood from 10 BALB/c mice was drawn in acid-citrate-dextrose buffer (ACD-A) (9 vol blood to 1 vol ACD-A). The platelet pellet obtained by centrifugation of PRP at 1800g for 10 minutes (approximately 0.5 × 10^10 platelets) was washed with ACD-A-saline, prior to adjusting to pH 6.8 with NaOH 1 mol/L. The platelets were resuspended in the same buffer and incubated with 200 µCi (7.4 MBq) of 111In-oxine at 24°C for 20 minutes. The reaction was terminated by addition of 4 mL of PPP. After 7 minutes the sample was centrifuged 5 minutes at 1800g to remove free 111In-oxine and the platelets were suspended in PPP. Labeling efficiency measured by counting radioactivity of the pellet and supernatant was 82%.

**Clearance studies**

Mice were injected intravenously with 6 × 10^8 111In-oxine labeled platelets (111In-Plat).34 After 90 minutes (time necessary for 111In-Plat to equilibrate with the splenic pool) the animals were injected with Ab or saline. The distribution of 111In-Plat was determined by serial bleedings. The amount of counts per minute (cpm) in peripheral blood at 1 minute after Ab or saline injection was considered as 100% of cpm in circulation (t = 0 minute). Blood samples were diluted in saline, centrifuged, and free and intracellular 111In-oxine was determined. The total number of counts in the bone marrow was estimated assuming that one sixth of the marrow space was within the femur.35 The 111In-oxine in the blood at 24 hours was obtained considering the total volume as 0.09 mL/g body weight. This value plus the radioactivity present in the organs was considered the total radioactivity. Taking into account the short half-life of 111In (67.4 hours), the radioactivity of different samples from kinetics experiments was evaluated simultaneously at the end of experiments.

**Platelet survival study**

Mice were injected with 200 µL of 111In-Plat as described above. Blood samples were taken at 1, 3, 5, 24, 48, 72, and 120 hours after injection. Free and intracellular radioactivity was distinguished by counting packed platelets and supernatants after washing the cells. Mean platelet survival was determined as previously described.36

**Ex vivo platelet aggregation**

Platelet aggregation studies were performed by the standard turbidimetric technique using a Chrono Log Corporation aggregometer. Aggregation was initiated by addition of 5 to 20 µL of aggregating agents: (1) adenosine diphosphate (ADP) 50 µmol/L, (2) collagen 20 µg/mL, or (3) arachidonic acid 1 mmol/L. Aggregation was quantified as percentage of maximal amplitude 5 minutes after addition of aggregating agents.

**Bleeding time test**

The essay was done by making a standardized incision of a depth of 3 mm in parallel to the tail veins of mice, at a site where no visible vessel was seen. The blood was carefully removed at exactly 30-second intervals with a filter paper, until bleeding stopped completely. The time taken for the blood flow to stop was recorded. The normal bleeding time of mice was less than 2 minutes (n = 15).

**Statistical analysis**

Intergroup contrasts of dimensional variables were compared with one-way analysis of variance (ANOVA) followed by Bonferroni t test. All statistical tests were interpreted in a 2-tailed fashion to estimate P values.

**Results**

**Effect of lip-PBS and lip-clod on platelet, red blood cell, and leukocyte counts**

Mice were injected intravenously with 200 µL of either saline, lip-PBS, or lip-clod, and the platelet count was evaluated at 24, 48, 72, and 96 hours after injection. Lip-clod induced a slight but significant thrombocytopenia in the animals during the period from 48 to 72 hours after treatment, returning to the normal platelet count at 96 hours after treatment. The results, expressed as mean ± SEM of platelets × 10^9/µL were the following: 48 hours, saline, 410 ± 10; lip-PBS, 420 ± 1; lip-clod, 376 ± 3 (*P < .01 versus saline and lip-PBS treated groups, n = 6); 72 hours, saline, 411 ± 4; lip-PBS, 440 ± 6; lip-clod, 320 ± 10 (*P < .001 versus saline and lip-PBS treated groups, n = 6). However, this effect on the platelet count was not observed when lip-clod diluted 1:8 to 1:32 was administered (not shown). The number of peripheral red blood cells and leukocytes was not significantly different from controls at any time after treatment.

**Effect of rabbit antimouse platelet antiserum on the platelet count**

Mice were injected intraperitoneally with a unique dose of 25 µL (100 µL diluted 1:4) of Ab and the platelet count was evaluated at the times indicated in Figure 1A. A severe thrombocytopenia was induced at 4 hours after injection. The effect was platelet specific because the Ab did not modify either the leukocyte count or hematocrit at any time after treatment. However, 24 hours later this
value was slightly increased, and the recovery of control levels was achieved 48 hours after Ab injection. Taking into account this result, another group of mice was injected with a daily dose of Ab to maintain the platelet count at low levels. This schedule was used to obtain a sustained thrombocytopenia throughout the experiments. In addition, the treatment with normal rabbit serum did not modify either the platelet and leukocyte count or hematocrit (not shown). For the ITP model it is important to know whether rabbit Ab at a concentration equivalent to that obtained in vivo did not induce platelet aggregation. Thus, taking into account that the amount of whole blood in normal mice is approximately 7% of the body weight (range, 1.6-1.8 mL),37 25 μL of Ab (amount that induces thrombocytopenia) was incubated with 0.5, 1, and 1.5 mL of whole blood during 1 hour at 37°C. After this period the microscopic aggregation of platelets was not observed. In addition, the platelet counts before and after Ab treatment were similar, confirming that aggregates were not formed. Complement-dependent lysis of platelets was not observed.

Lip-clod inhibits the Ab-induced thrombocytopenia in a dose-dependent manner

Clearance of immune complexes is one of the most important functions carried out by the RES, and it is mostly dependent on the FcγR present on the surface of hepatic and splenic macrophages.38 We have previously shown that clearance of immune complexes can be strongly inhibited by lip-clod in a mice model of sensitized autologous erythrocytes.23 Then, we further analyzed whether lip-clod inhibits the Ab-induced thrombocytopenia. For this purpose, groups of mice were injected intravenously with a unique dose of 200 μL of different amounts of lip-clod. Twenty-four hours later the animals were injected intraperitoneally with Ab. The administration of Ab was repeated at 24-hour intervals during the course of the experiment, and the mice were bled daily for platelet count (before Ab administration). As shown in Figure 2, whereas lip-clod 1:1 inhibits the Ab-induced thrombocytopenia up to 120 hours, a decreasing and dose-dependent effect was observed with lip-clod 1:4, 1:8 and 1:32, respectively. In the lip-clod 1:1-treated group, the inhibition of thrombocytopenia was partial at 172 hours (Ab plus lip-clod: 145 × 10^3 ± 17 × 10^3 versus Ab: 16 × 10^3 ± 4 × 10^3 platelets/μL, P < .001, n = 6), and at 200 hours the level of platelets was similar in both groups (Ab plus lip-clod: 22 × 10^3 ± 5 × 10^3 versus Ab: 11 × 10^3 ± 2 × 10^3 platelets/μL). On the other hand, lip-PBS did not alter the Ab-induced thrombocytopenia at all (not shown). Then, taking into account the long-lasting effect of lip-clod 1:1 (original preparation, see "Materials and methods"), we used this dose throughout the experiments.

Reversal of antibody-induced thrombocytopenia by lip-clod

Taking into account that ITP patients are usually admitted to the hospital with thrombocytopenia, we further investigated if thrombocytopenic animals were capable of recovering the platelet count to a hemostatically safe level after lip-clod treatment. For this purpose, animals were injected intraperitoneally with Ab, and 4 hours later (time = 0 hour, Figure 3) the mice had a platelet count less than 5 × 10^3/μL. After this, the mice were injected intravenously with 200 μL of lip-PBS or lip-clod. The animals were bled daily for platelet count and reinjected with Ab to obtain a sustained thrombocytopenia. As shown in Figure 3, lip-clod treatment rapidly restored (24 hours) the platelet count to approximately 50% of normal values. Moreover, despite additional Ab treatment, mice were able to maintain this level of platelets (48 hours). Although to a lesser extent, the recovery of platelet count was also observed with lip-clod diluted up to 1:64 (24 hours: 11.2% ± 0.3% and 48 hours: 20% ± 1.0% of control, n = 6).

Effect of lip-clod in the organ uptake of 111In-oxine labeled platelets

Because the injection of Ab induced a profound thrombocytopenia, the organ localization of platelets in experimental ITP was analyzed. For this purpose, control and lip-clod–treated mice were injected intravenously with 6 × 10^8 111In-Plat in a volume of 200 μL. After 90 minutes (time necessary for 111In-Plat to equilibrate with the splenic pool) the animals were injected with Ab, and 24 hours later the distribution of radioactivity was analyzed in peripheral blood, spleen, liver, lungs, kidneys, and bone marrow. As shown in Figure 4, a highly significant decrease of labeled...
platelets in circulation was detected in Ab-treated mice. On the other hand, in the mice treated with lip-clod, most of the platelets remained in circulation. The liver was largely the principal organ responsible for platelet sequestration in Ab-treated mice, whereas the radioactivity in bone marrow, lung, and kidneys was negligible and not different from control animals (not shown). However, the treatment with lip-clod significantly blocked the liver uptake. The splenic uptake of platelets was similar in all experimental groups, but significantly different from control. This result indicates that the splenic uptake of platelets induced by lip-clod, which might also explain the transient thrombocytopenic state after treatment.

It is known that in patients with ITP the platelet destruction occurs in the liver and spleen but in most of them the predominant site of destruction is the spleen. However, we did not observe a shift from the liver to the spleen uptake by using different concentrations of Ab (1:15, 1:45, and 1:135) as described by Veerhuis and coworkers. Similar results were obtained in 2 additional experiments using different lots of lip-clod (not shown).

**Platelets sequestered by the liver did not return to circulation**

We analyzed whether platelets entrapped in the liver were able to return to the circulation. For this purpose, mice were injected with $^{111}$In-Plat, and 90 minutes later with saline or Ab. As shown in Figure 5, at 3 and 24 hours after treatment, both the radioactivity in circulation and platelet counts were very low in the Ab-treated group. However, at 48 hours, although the level of radioactivity was similar to that found at 24 hours, the platelet count reached normal values. These results indicate that trapped labeled platelets do not return to circulation and that new cells might be derived from the platelet storage pool or new thrombocytogenesis. These data were confirmed by analysis of excised organs at 48 hours, where most of the radioactivity was still found in the liver (not shown). Moreover, at this time the $59.8\% \pm 2.6\%$, $n = 4$, of the residual radioactivity in circulation in Ab-treated animals was found free in the serum, indicating that platelets had been destroyed. The platelet survival of this group was $0.39 \pm 0.1$ days, $n = 4$. On the other hand, in the control group $94.2\% \pm 0.7\%$, $n = 4$, of the radioactivity was associated with platelets. The progressive decrease in the levels of radioactivity observed in the liver was largely the principal organ responsible for platelet sequestration in Ab-treated mice, whereas the radioactivity in bone marrow, lung, and kidneys was negligible and not different from control animals (not shown). However, the treatment with lip-clod significantly blocked the liver uptake. The splenic uptake of platelets was similar in all experimental groups, but significantly different from control. This result indicates that the splenic uptake of platelets in both lip-clod and lip-clod plus Ab-treated groups cannot be attributable to antibodies. The cause of this effect appears to be the nonspecific splenic uptake of platelets induced by lip-clod, which might also explain the transient thrombocytopenic state after treatment.

It is known that in patients with ITP the platelet destruction occurs in the liver and spleen but in most of them the predominant site of destruction is the spleen. However, we did not observe a shift from the liver to the spleen uptake by using different concentrations of Ab (1:15, 1:45, and 1:135) as described by Veerhuis and coworkers. Similar results were obtained in 2 additional experiments using different lots of lip-clod (not shown).
control group (without Ab) can be attributed to the senescence of platelets, because the platelet survival of BALB/c in our colony was 3.5 ± 0.3 days, n = 4.

**Bleeding time and platelet aggregation in mice treated with lip-PBS and lip-clod**

The bleeding time test is a good in vivo functional assay used for evaluating the arrest of hemorrhage by platelet plug formation. Then, we analyzed the bleeding time in control, lip-PBS, lip-clod, lip-clod plus Ab, and Ab-treated animals. The bleeding times in lip-clod treated animals, either with or without Ab, were not different from controls and lip-PBS–treated mice (≤ 2 minutes, n = 7), demonstrating that the hemostasis was well controlled in these animals. However, animals injected with Ab show a longer bleeding time (range, 4-6.5 minutes; n = 6).

Maximal platelet aggregation induced by ADP (50 μmol/L), collagen (20 μg/mL), or arachidonic acid (1 nmol/L) was reduced in lip-PBS to 60% of control, in lip-clod to 50% of control, and in lip-clod plus Ab-treated mice to 90% of control. Saline- and lip-PBS–treated mice in presence of Ab had less than 20,000 platelets per milliliter; therefore it was not possible to evaluate platelet aggregation in these groups. We also found a similar reduced platelet aggregation in the presence of lip-PBS or lip-clod added during the course of the test (not shown). These results indicated that liposomes can interfere with the normal platelet aggregation.

**Discussion**

Taking into account that the different second-line treatments for ITP are not devoid of several adverse or multiple side effects and the lack of consensus on how to approach the patients with refractory ITP, we investigated a new strategy for ITP treatment using a mouse model.

The significant, although moderate, decrease in platelet count observed in mice after lip-clod administration (48-72 hours) contrasts with the severe displacement of platelets from peripheral blood to the splenic cords observed by others. Although the mechanism of this effect is unknown, factors such as the clodronate concentration, the amount of liposomes injected, and the age and weight of the mice might be potential causes of these differences. A most accurate standardization will be necessary, because, for instance, the significant reduction of about 20% in the weight of spleen described in lip-clod treated mice, was found not significant by the same group in other studies. On the other hand, in agreement with others, the injection of low amounts of lip-clod (dilution 1:8 and 1:32 from the original preparation) did not induce changes in the platelet count, whereas these concentrations of lip-clod exerted a clear blockade of platelet uptake (Figure 2).

The hematocrit and leukocyte count were not affected by lip-clod treatment, and as previously shown, signs of illness or weakness in the treated mice were not observed. Furthermore, the administration of lip-PBS did not modify either hematocrit or platelet and leukocyte counts. The lack of additional side effects in mice treated with lip-clod is in agreement with the nontoxic effect of lip-clod for other cells than macrophages. Moreover, it has been demonstrated that clodronate does not escape from liposomes, and the drug released in the circulation from dead macrophages does not cross cell membranes in the opposite direction.

Lip-clod not only inhibited the uptake of sensitized platelets by the liver, but also induced the recovery of peripheral platelets (from 50,000 to 200,000/μL) in Ab-induced thrombocytopenic animals as soon as 24 hours after lip-clod injection (Figure 3). These values were maintained at least for 48 hours, despite additional Ab injection. This is a crucial point for the experimental model because one of the goals of treatment should be a rapid increase in the platelet count to a hemostatically safe value. In a recent randomized trial carried out in children with acute ITP with platelet counts of less than 20,000/μL, the conventional treatments with IVIg, anti-D, or prednisone, took about 72 hours to reach a platelet count of 50,000/μL. An additional and important issue was that recovery of platelets in circulation was observed at 24 hours after lip-clod injection, time at which the partial thrombocytopenia induced by lip-clod was not yet observed. Thus, the rapid reconstitution of the platelet count counterbalanced the slight but significant thrombocytopenia induced by lip-clod. Moreover, the recovery of platelet count after Ab injection can be achieved, although to a lesser extent, with concentrations of lip-clod (1:8 and 1:32) that did not modify the platelet count.

Although in patients with ITP the platelet destruction occurs in the liver and spleen, the predominant site of destruction is the spleen. In our experimental conditions, however, the liver was almost the exclusive organ of platelet sequestration. This is in agreement with previous reports in which the hepatic uptake of platelets was greater than the splenic uptake in dog  and mouse models of ITP, suggesting that the site of accumulation of sensitized platelets may vary among species. Although it has been reported that the shift of uptake from liver to spleen depends on the number of IgG and C3 molecules on the cell surface, we had no success using different Ab concentration (dilutions 1:15, 1:45, and 1:135 from the original preparation), and therefore the inhibition of splenic uptake of IgG-sensitized red cells by lip-clod in mice could not be observed. On the other hand, as in human ITP, the liver sequestration of platelets in mice is not reversible (Figure 5), discarding any delayed traffic of platelets later returning to the circulation in a complement dependent rebound mechanism. Moreover, this was followed by the destruction of platelets, because about 60% of the radioactivity in circulation was not cell associated, whereas in control animals most of the radioactivity was found in the platelets.

Although the platelet aggregation assay gives a rough estimation of the functionality of platelets, it is frequently used. We found less aggregation in platelets from lip-PBS, lip-clod, or lip-clod plus Ab-treated mice compared to control platelets. The cause of this effect is not known, although the partial platelet aggregation induced by ADP, collagen, or arachidonate in the presence of liposomes (added with the agonists), suggests a physical interference by these particles. In contrast, the bleeding time test, both in animals treated with lip-PBS or lip-clod, was within the normal range, demonstrating that hemostasis was well controlled in the animals studied. Although in most ITPs the bleeding time is prolonged, this parameter does not always correlate with the platelet count. However, this in vivo assay is used to determine the arrest of hemorrhage by platelet plug formation and it is considered a valuable screening test for platelet function.

In this, our first attempt to investigate the management of experimentally induced ITP, we can summarize the potential advantages and disadvantages of lip-clod treatment. Among the potential advantages are (1) lip-clod rapidly restores the platelet count to hemostatic safe values; (2) lip-clod does not induce anemia nor leukopenia; (3) lip-clod is toxic to phagocytes but not to...
other cells, lip-clod does not induce changes in the bleeding time; and (5) lip-clod has a potential low risk of infections compared to plasma-derived products. Another important point is that relatively low concentrations of lip-clod (dilution 1:8 to 1:32; Figure 2) are effective in the blockade of platelet clearance during the first 48 hours after injection. In addition, no effect induced by lip-clod treatments was observed on nonphagocytic spleen cells, dendritic cells, and neutrophils.

On the other hand, among the undesired effects we found the following: (1) Lip-clod exerts a moderate thrombocytopenia after 48 to 72 hours of injection. However, in agreement with others, this effect is null with lip-clod 1:8 and 1:32, though they are still capable of inducing the RES blockade; (2) lip-clod interferes with platelet aggregation; (3) lip-clod induces a depletion of hepatic and splenic macrophages, although in a dose-dependent and reversible manner.

Because lip-PBS are devoid of any visible side effects, confirming the inert nature of this neutral liposome, the possibility of using liposomes with different concentrations of clodronate, or other drugs, or lower amounts of lip-clod, encourages us in the development of this methodology. As previously suggested, the improvement of a controlled manipulation of macrophages, as well as the evaluation of other hematologic parameters related to platelet functions, will be necessary in future experiments.

As far as we know this is the first report demonstrating that liposome-encapsulated drugs can be effective in the treatment of experimental ITP, and we think that because of the possibilities opened by this method further investigation would be worthwhile.

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Treatment with liposome-encapsulated clodronate as a new strategic approach in the management of immune thrombocytopenic purpura in a mouse model

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