The human basophil: a new appreciation of its role in immune responses

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Basophilic granulocytes (usually referred to as basophils) are a small population of peripheral blood leukocytes containing cytoplasmic granules that stain with basophilic dyes. They were first described in 1879 by Paul Ehrlich, who 1 year earlier had found a morphologically similar cell type present in tissues that he termed Mastzellen. Based on their similarity to mast cells, basophils have often been considered (and neglected) as minor and possibly redundant “circulating mast cells.” Like mast cells, basophils possess high-affinity immunoglobulin (Ig) E receptors (FcεRI) that are cross-linked upon engagement of receptor-bound IgE with corresponding antigens (“allergens”), resulting in the release of a number of mediators that are in part common for both cell types. Until recently, it has been very difficult for most laboratories to obtain basophils without major contaminating cell populations, because the percentage of basophils in peripheral blood is low (<1%) and they share physicochemical properties with other blood cells. This lack of satisfactory purification protocols has considerably hampered basophil research and negatively affected the interest in this cell type. Nevertheless, recent findings have provided new insights into the possible role of basophils in allergic disease and immunity to pathogens. Most notably, the discovery that basophils rapidly produce large amounts of the regulatory cytokines interleukin (IL)-4, IL-13, and IL-3, together with the constitutive expression of CD40L and CCR3 on their surface, has fueled speculations that extend beyond their recognized role as effector cells in IgE-mediated reactions. As discussed in this review, the increasing knowledge about the chemokine network has provided a robust framework for understanding the mechanisms underlying basophil recruitment to the tissues in allergic diseases. This development has been instrumental in identifying very promising new targets for the treatment of asthma and other allergic diseases. We also review recent aspects regarding basophil growth and development and their role in allergic late-phase reactions (LPRs). Furthermore, we have included a section on basophil signal transduction, which expands our understanding of the mechanisms operating downstream of the high-affinity IgE receptor and are also expected to yield new and better targets for pharmacologic intervention. Finally, we discuss the possible roles of basophils in innate immunity and finish with a section on the currently available tools for basophil research.

Growth and development of basophils

Although the phenotype of mature basophils has been extensively studied, the early stages of basophil maturation and their relationship to other cell lineages are not well understood. Basophils, as well as mast cells, monocytes, eosinophils, and neutrophils, are thought to arise from CD34 + progenitors found in cord blood, peripheral blood, and the bone marrow. Basophils, in particular, have been suggested to evolve from CD34+/IL-3Rα+/IL-5+ eosinophil/basophil progenitors, as supported by the occurrence of granulocytes with a hybrid eosinophil/basophil phenotype in patients with chronic or acute myelogenous leukemia or in cell culture.

Although mast cell progenitors have been documented as a separate lineage, eg, characterized as CD34+/CD38+ or CD34+/c-kit+/CD13+, most recently a new monoclonal antibody (97A6) has been described as specific for mature mast cells and basophils and their progenitors. Monoclonal antibody 97A6 did not react with any other hematopoietic or nonhematopoietic cell type. The epitope recognized by 97A6 may therefore be associated with a commitment of the CD34+ precursor to a mast cell or basophil lineage distinct from other lineages. The possibility of a common mast cell or basophil lineage also arises from the surprising observation that basophils with phenotypic features characteristic of mast cells (presence of tryptase, chymase, c-kit, carboxypeptidase A) can be found in patients with asthma, allergy, or allergic-drug reactions. Therefore, the currently predominant concept that mast cells and basophils originate from separate lineages may have to be revised.

Various protocols using different combinations of cytokines and growth factors have been described for the in vitro cultivation of basophils from blood or bone marrow progenitors, with the presence of exogenous IL-3 being a common prerequisite for the successful development and maturation of basophils. Other growth factors for basophils are IL-5, granulocyte-macrophage colony-stimulating factor (GM-CSF), transforming growth factor-β (TGF-β), and nerve growth factor (NGF). There is a general consensus, however, that the main growth and differentiation factor for basophils is IL-3, whereas the growth and development of mast cells requires the presence of stem cell factor (SCF). In conformity with their relative cytokine requirements, mature basophils can be distinguished phenotypically from mast cells by their differential surface expression of IL-3 α chain (CD123, found on basophils and other cells but not mast cells) and SCF receptor (c-kit/CD117, strongly expressed on mast cells but not, or only weakly, on mature basophils).

Role of basophils in allergic late-phase reactions

It is well established that mast cells, which are found in the tissues in the proximity of small blood vessels and postcapillary venules,

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play a key role in the early phase of IgE-mediated allergic reactions.53 Unlike mast cells, which differentiate and mature in the tissues, mature basophils are found in the circulatory system, where they constitute less than 1% of circulating leukocytes. Basophils, eosinophils, and Th2 lymphocytes are recruited to the site of inflammation during LPRs. Because until recently specific markers for immunohistochemical detection of basophils were not available (see “Tools for basophil research”), the participation of basophils during LPRs has been documented mainly by indirect means,32-34 ie, by determining the pattern of mast cell–or basophil-specific mediators like histamine (derived from both), prostaglandin D2 (from mast cells), or leukotriene C4 (LTC4 from basophils). It is now well established that basophils are rapidly recruited to the skin,35 lung,36 or nose37 after allergen challenge.

More recently, a better understanding of the processes leading to inflammatory cell recruitment in LPRs has sprung up from the study of chemokines and their receptor counterparts, identifying potentially important new targets for the treatment of asthma and other allergic diseases.38-40 These studies complement previous knowledge regarding the molecules involved in basophil adhesion to the endothelium,41-43 a critical step in the process of extravasation. The aim of the following section is to highlight the potential roles of basophils in LPRs. We use the new nomenclature for chemokines proposed by the Keystone Chemokine Conference (Keystone, CO, January 18-23, 1999).40

Several reports have pinpointed eotaxin (CCL11) and the eotaxin receptor (CCR3) as key molecules in the selective recruitment of eosinophils to the lung during allergic airway inflammation,44 both in animal models45-49 and asthmatic patients.50-53 Sensitized mice with a disrupted eotaxin gene showed a 70% reduction in the number of eosinophils recruited by antigen in comparison with the identically treated wild-type mice.54 Enhanced eotaxin expression in allergic inflammation may also account for the influx of basophils, which strongly express CCR3 on their surface and respond to eotaxin in vitro.10,55 In addition, human basophils also respond to other chemokines such as the CC chemokines eotaxin-2 (CCL24), RANTES, MCP-3 (CCL7), MCP-1, and MIP-1α,56,57 which also play a role in antigen-induced tissue recruitment of eosinophils.58,59 Moreover, basophils may be responsive to eotaxin-3 (CCL26),60,61 the most recent addition to a growing list of CCR3 agonists. Although it remains to be proven, it is likely that the presence of CCR3 and other CC chemokine receptors shared between eosinophils and basophils account for the previously demonstrated strong temporal association of their influx into tissues such as the nose57,62 or the lung56 during LPRs.

A further insight regarding tissue influx of basophils and their subsequent activation is also given by the actions of hematopoietic cytokines such as IL-3, IL-5, and GM-CSF. An increased expression of these Th2 lymphocyte–derived cytokines in allergen-induced cutaneous LPRs in atopics was clearly demonstrated by Kay and colleagues,64 and these cytokines have also been shown to facilitate basophil migration.65-67 In addition, IL-3, IL-5, and GM-CSF also enhance mediator release following either IgE-dependent stimulation or activation due to platelet-activating factor, C5a, and C3a. Similar properties have additionally been described for the neurotrophic cytokine NGF.68 Moreover, increased levels of NGF have recently been reported in allergic rhinitis69 and asthma.70

IL-3, IL-5, GM-CSF, and NGF have similar efficacies with respect to potentiating IgE-mediated basophil histamine and LTC4 releases, and they bind to receptors of the same subfamily but, of these, IL-3 is the most potent71 and its effects on basophils have been appreciated for some time.72,73 In vitro, IL-3 alone causes little mediator release by itself, with the exception of IL-13 production in some donors. However, a short preincubation or coculture of basophils with IL-3 causes significant enhancement of histamine and LTC4 release to a number of immunologic and nonimmunologic stimuli such as anti-IgE, C5a, C3a, the eosinophil product major basic protein, and platelet-activating factor.74-77 Moreover, studies by Ochensberger78 demonstrate that LTC4 synthesis in basophils primed by IL-3 before stimulation with C5a is continuously maintained over many hours. This clearly highlights the ability of basophils to maintain a state of activity over longer periods than previously anticipated. Additionally, these authors showed an elevated production of IL-4 and IL-13 from basophils under the same conditions,78 which may exacerbate LPRs because of the up-regulatory effects of these cytokines on vascular cell adhesion molecule-1 (VCAM-1), thus promoting further influx of inflammatory cells (discussed in detail below).

The ability of basophils to continuously produce LTC4 and cytokines may also be particularly important in maintaining chronic allergic inflammatory diseases, such as asthma. Furthermore, because IL-3 can, in conjunction with C5a, cause basophil mediator release independently of IgE-receptor cross-linking, this suggests a role for basophils in nonallergic inflammatory diseases caused by either bacterial infections or IgG immune complexes. In addition to the combined effects of C5a and IL-3, C5a by itself is a highly potent and efficacious inducer of histamine release from basophils79 following binding to the C5a receptor, CD88.80 In contrast, most human mast cell subtypes do not express CD88 and are unresponsive to C5a except skin mast cells.80 Moreover, unlike basophils, mature human mast cells, including those of the skin, also do not express receptors for IL-3 and GM-CSF.81 This clearly suggests that, compared with mast cells, basophils are more sensitive to the effects of immunomodulatory cytokines and proinflammatory factors, which are likely to be in abundance in both LPRs and other (nonallergic) inflammatory conditions.

Unfortunately, the interplay of these various basophil activators, in terms of the net clinical outcome of their combined effects, in allergic disease remains to be elucidated. However, the importance of IL-3 in vivo as both a modulator of basophil growth and activation as well as its ability to cause clinical symptoms of allergic inflammation was shown by van Gils et al.82 Using rhesus monkeys treated with the IL-3, the authors observed a clear increase in both basophil numbers together with an up-regulation of IL-3 receptors without a concomitant rise of IL-3R+ monocytes or mast cells.82 Moreover, the same authors also demonstrated that IL-3 treatment caused severe hypersensitivity reactions including urticaria, vomiting, diarrhea, edema, arthritis, and stimulation of hemopoiesis.83 Given the above evidence, IL-3 and related hematopoietic growth factors, as well as NGF, potentially play a pivotal role in exacerbating LPRs in addition to CC chemokines described earlier.

Another indirect line of evidence that reveals basophils as potential major regulatory cells in allergic inflammation comes from the study of their cytokine profile. As described earlier, human basophils are an important source of the proallergic cytokines IL-4 and IL-13. Kasai et al84 have shown that allergen-induced IL-4 from primary cultures of unfractionated peripheral blood leukocytes of atopic patients is mainly basophil-derived. This finding has been confirmed and extended to IL-13 by Devouassoux and coworkers,85 who found that basophils are the main source of both cytokines early after allergen stimulation of peripheral blood cells. IL-4 is of central importance for the recruitment of inflammatory conditions.
cells to the tissues, because it induces the expression of the adhesion molecule VCAM-1 on endothelial cells in vitro and it recruits eosinophils in vivo, eg, when injected in rodents. Transgenic mice expressing IL-4 in the lung under the control of a lung-specific promoter showed eosinophil infiltrations in the airways. Recently, Hickey et al have used intravital microscopy to demonstrate that the VLA (very late antigen)-4/VCAM-1 interaction is necessary and sufficient for IL-4-induced eosinophil recruitment in mice. On the other hand, it has been shown that IL-4, especially in combination with tumor necrosis factor (TNF)-α, induces the production and release of eotaxin by human dermal fibroblasts; moreover, eotaxin-3, but not eotaxin, is up-regulated in vascular endothelial cells by IL-4. The induction of eotaxins by IL-4 could represent a strong positive feedback loop operating in eosinophilic skin diseases (eg, atopic dermatitis), with the induced eotaxins recruiting more eosinophils, basophils, and Th2 lymphocytes and the subsequent release of more IL-4 upon activation (Figure 1). Interestingly, eotaxin can potentiate the antigen-dependent IL-4 production by basophils without having a direct effect on basophils by itself, thus generating a further potential amplification loop in allergic inflammation. The aforementioned proallergic properties of IL-4 therefore make the identification of its cellular source(s) an important prerequisite for the successful therapy of asthma and other allergic diseases.

Recent evidence has implicated the pleiotropic cytokine IL-13, which is also produced in large amounts by activated basophils, as a key mediator of allergic asthma. In humans, IL-13 is secreted following local allergen challenge in subjects with mild atopic asthma. Work performed by 2 independent laboratories, using an IL-13R–Fc fusion protein that specifically neutralizes IL-13, has indicated that this cytokine is necessary and sufficient for establishment of an asthmatic phenotype in a murine model. Administration of soluble recombinant IL-13 was effective in inducing airway hyperresponsiveness and in generating a significant recruitment of eosinophils to the lung, as shown by their appearance in bronchoalveolar lavages. In vitro, IL-13 induces recruitment and prolongs the survival of eosinophils, up-regulates VCAM-1 expression on vascular endothelium, and is a very potent inducer of eotaxin in airway epithelial cells.

The importance of IL-13 in the pathogenesis of asthma is also corroborated by a recent study using transgenic mice that express IL-13 selectively in the lung. Administration of soluble recombinant IL-13 is effective in inducing airway hyperresponsiveness and in generating a significant recruitment of eosinophils to the lung, as shown by their appearance in bronchoalveolar lavages. In vitro, IL-13 induces recruitment and prolongs the survival of eosinophils, up-regulates VCAM-1 expression on vascular endothelium, and is a very potent inducer of eotaxin in airway epithelial cells. The importance of IL-13 in the pathogenesis of asthma is also corroborated by a recent study using transgenic mice that express IL-13 selectively in the lung. The authors of this study found that transgenic mice, in the absence of a specific sensitization, developed a phenotype very similar to the asthmatic phenotype, including eosinophil infiltration, up-regulation of eotaxin, mucus hypersecretion by goblet cells, subepithelial airway fibrosis, and airway hyperresponsiveness to methacholine.

Finally, basophils have also been proposed to play a key role in allergy by directly inducing the switch to the IgE isotype in B cells independently of T cells (Figure 1). Gauchat et al have shown that the basophil cell line KU812, as well as purified peripheral blood basophils, the mast cell line HMC-1, and human lung mast cells, can provide the necessary signals for IgE production in vitro, ie, IL-4, IL-13, and CD40L. Basophils generated from human umbilical cord blood mononuclear cells, after cultivation in the presence of appropriate cytokines, expressed detectable levels of CD40L and induced IgG4 and IgE synthesis in B cells when stimulated with allergen. The induction of IgE synthesis was completely abrogated by neutralizing IL-4 and IL-13 with monoclonal antibodies or CD40L with soluble CD40L.

Taken together, the data about the involvement of IL-4, IL-13, and eotaxin as key players in the pathogenesis of asthma, together with the proven infiltration and activation of basophils in the tissues during LPRs, makes it reasonable to assume that basophils, as a major source of IL-4 and IL-13 and by constitutively expressing CCRI and CD40L on their surface, may play an important but underestimated role in the pathology of asthma and other allergic conditions. The knowledge about the relative roles and contribution of basophils, eosinophils, mast cells, and T cells to these processes as well as about their interplay is still preliminary and needs further refinement.

**Figure 1. LPR scenario emphasizing the putative regulatory roles of basophils in allergic inflammation.** Eotaxin (CCL11) (A) derived from epithelial cells and eosinophils can be detected in the lungs of asthmatic patients and recruits cells expressing the eotaxin receptor (CCR3), ie, eosinophils, basophils, TH2 cells, and mast cells. Recruited basophils are a major source of the regulatory cytokines IL-4 and IL-13 and the up-regulation of eotaxin in airway epithelial cells. Eotaxin has also been shown to potentiate the IL-4 production by activated basophils. The increased IgE production is thought to up-regulate the expression of FcεRI on basophils and mast cells.
characterized for human basophils compared with their counterparts from other species or cell lines. Several examples given below show that a mere extrapolation of observations derived from basophil lines or basophils from other species does not clearly reflect the situation in primary human basophils. Although it may be obvious, this point must be stressed because future successful pharmacologic approaches to reducing basophil mediator production hinge crucially on understanding basophil biology taken from studies on primary basophils.

**Early IgE-dependent signaling events in basophils**

High-affinity IgE-receptor (FcεRI) cross-linking culminates in 3 main results. The first, and most rapid, is fusion of basophil granules to the plasma membrane resulting in histamine release; the second is an activation of lipid metabolism and subsequent LTC4 production; and the third is synthesis of immunomodulatory cytokines. The initial signaling events following FcεRI cross-linking are currently modeled on studies largely carried out on rodent mast cell lines (such as RBL-2H3 cells), where phosphorylation of tyrosine, serine, and threonine residues contained in the β and γ chains of FcεRI receptor have been observed (Figure 2). In human basophils, as in RBL-2H3 cells, this has been shown to involve Lyn and subsequent recruitment of SH-2 domain-containing proteins such as Syk and other ZAP-70–related protein tyrosine kinases. The possibility that Syk may be involved in IgE-dependent basophil signaling was recently also demonstrated by Kepley and coworkers, who showed that nonreleaser human basophils, occurring in about 20% of the population, have a deficiency of this protein. In mast cells, most of the known major regulatory proteins known to be involved in FcεRI activation are downstream of Syk and thus crucially hinged to Syk activity. These include PLCγ1 and PLCγ2, Shc, Vav, mitogen-activated protein (MAP) kinases, phosphatidylinositol (PI)-3 kinase, and JNK, among others, although this has yet to be conclusively demonstrated for human basophils.

Although several of the early IgE-dependent signaling events in basophils are known, there are currently no pharmacologic approaches to the treatment of allergy that specifically modulate very early signaling events following cross-linking with allergen. However, the molecules known to be involved in early post–cross-linking are also used by other immunoreceptor systems affecting many other immune cell functions, thus making selective pharmacologic intervention difficult. Therefore, an alternative strategy is to inhibit either FcεRI expression itself or the binding of IgE molecules to the receptor. It has long been observed that there is a positive correlation between circulating IgE antibodies and FcεRI expression in basophils, and recent evidence clearly demonstrates that IgE does directly induce high-affinity IgE-receptor expression by acting through FcεRI itself. Thus, basophil (and mast cell) function may significantly be down-regulated by disrupting the binding of IgE to its receptor at the sensitization stage. This has been successfully achieved in atopic patients treated with a monoclonal anti-IgE antibody (rhuMAb-E25), which prevents IgE binding to the FcεRI α chain without cross-linking high-affinity receptor-bound IgE, and is a promising new therapeutic advancement in the treatment of allergic disorders.

**PI 3-kinase is a universal regulator of basophil mediator release**

PI 3-kinase is composed of a p85 regulatory subunit and a 110-kd catalytic domain that catalyses the reaction of membrane PtdIns^4,5P_2 principally to PtdIns^3,4,5P_3. Indirect observations using specific PI 3-kinase antagonists suggest that PI 3-kinase plays a crucial role in the release of all major basophil mediator types, suggesting that this enzyme is closely associated with initial events in IgE-receptor signaling. We have recently shown that the PI 3-kinase inhibitors wortmannin and LY-294003 almost completely abrogate the release of histamine, LTC4, IL-4, and IL-13 in human basophils stimulated with anti-IgE. Moreover, inhibition of both IL-4 and IL-13 production did not result in increased cellular contents of the cytokines, showing that PI 3-kinase blockade is not restricted to impairing secretory events. With respect to mast cell and basophil degranulation, there is a universal consensus that PI 3-kinase inhibitors abolish granule-associated release. However, in regard to LTC4 production, Marquardt et al reported that the PI 3-kinase inhibitor wortmannin had no effect on the production of the eicosanoid in murine mast cells. The same authors also failed to detect any effects of this compound on cytokine release, whereas other reports have described inhibition of either leukotriene synthesis or TNF-α production. These differences show that a marked heterogeneity exists between basophils and various mast cell lineages to PI 3-kinase signaling.
Extracellular signal–related kinases control basophil LTC₄ production but not histamine or cytokine release

The extracellular signal–related kinases (ERKs) consist of 2 forms, ERK-1 and ERK-2 (sometimes referred to as p44 and p42 MAP kinases), which are considerably homologous to another in both structure and function. ERKs are activated after FceRI cross-linking via Vav, Ras-Raf1, and MEK1 and activate phospholipase A₂ (PLA₂) as well as several transcription factors (Figure 2). The role of ERKs in basophils, however, has only recently been partially investigated. Studies from our own laboratory and from Miura et al.¹¹⁶ have shown that ERK phosphorylation occurs within 3 to 10 minutes of basophil stimulation, after which there is a decrease to basal levels. The MEK inhibitor PD-098059 abolishes both anti-IgE–induced ERK activation and LTC₄ production in basophils but has little effect on histamine release or cytokine synthesis (IL-4 and IL-13).¹⁰⁷,¹¹⁶ The above studies highlight that ERK activation does not seem to affect cytokine transcription in basophils although it is an important regulator of cJun, cFos, STAT, and other transcription factors in diverse cell types. Therefore, in basophils ERKs appear to be largely involved only in LTC₄ production because of the activation of PLA₂. Nevertheless, we have also demonstrated that ERK activation is not by itself sufficient for leukotriene synthesis because IL-3, which strongly activates ERK in the absence of IgE-dependent stimulation,¹¹⁶ is almost ineffective at causing LTC₄ release.²⁴,¹⁰⁷,¹⁰⁷,¹¹⁷,¹¹⁸ Despite this, in combination with anti-IgE, IL-3 leads to both a striking potentiation and prolongation of ERK phosphorylation compared with the effects of IL-3 or anti-IgE alone and leads to twice as much LTC₄ production than anti-IgE alone. Although activation of PLA₂ is dependent on ERK, translocation of soluble PLA₂ from the cytosol to the membrane is dependent on calcium. However, studies by Krieger et al.¹¹⁵ and MacGlashan and Hubbard¹¹⁸ have shown that the priming caused by briefly (<15 minutes) preincubating basophils with IL-3 neither requires extracellular calcium nor induces calcium elevations. Taken together, these findings strongly suggest that, despite there being an important regulatory input from both ERK phosphorylation and intracellular calcium mobilization regarding PLA₂ activity, one or more additional signals, possibly from protein kinase C (PKC)–dependent routes,¹²⁰ are vital for LTC₄ synthesis in basophils.

Comparing human basophils with other rodent histamine-producing cells, there are important differences with respect to regulation of cytokine synthesis controlled through ERKs. For example, IgE-dependent TNF-α production in RBL-2H3 cells is inhibited by the MEK inhibitor PD-098059, suggesting ERK involvement. In contrast, the release of IgE-dependent TNF-α from MC/9 murine mast cells is not affected by the antagonist. Although basophils have not been shown to produce TNF-α, the above examples highlight further the variable abilities of ERKs to control the transcription of cytokine factors between human basophils and their counterparts in rodents. Further evidence that the MEK-ERK route does not predominantly influence basophil IL-4 production is provided by Miura et al.¹¹⁶ who report that the bacterial peptide fMLF induces basophil ERK activation without IL-4 release.

Calcium dependence of basophil mediator secretion

It has long been appreciated that basophil degranulation is strikingly reduced in the absence of extracellular calcium.¹²¹ Similarly, production of both IL-4 and IL-13 (B.F.G. et al, unpublished data, 1998) is just as dependent on the ion for optimum response to IgE-receptor activation. Calcium responses are in 2 phases.¹²⁴ The first takes place by the release of intracellular calcium stores from the endoplasmic reticulum because of the effects of inositol 1,4,5-trisphosphate (IP₃). The second phase is caused by the opening of calcium channels allowing an influx of the ion from the extracellular milieu and is modulated by cyclic adenosine monophosphate (cAMP)–elevating drugs.¹²⁵ A rise in intracellular free calcium ions can be mimicked by using calcium ionophores such as A23187, which directly transport extracellular calcium into the cells, release internal stores of the ion, and evoke degranulation, eicosanoid generation, and cytokine production.

In terms of IgE-dependent cytokine secretion, both IL-4 and IL-13 are controlled by calcium-calcinurin pathways, which promote the translocation of the transcription factor NFAT to the nucleus.¹²⁶ This pathway is blocked by macrolides such as FK506 and cyclosporin A, which reduce both IL-4 and IL-13 synthesis.¹²⁷ Additionally, the release of these cytokines is also effectively blocked by cAMP-elevating drugs such as theophylline or β-2 agonists (salmeterol), further highlighting the importance of calcium influx in basophil activation.¹²⁸

IL-4 and IL-13 are differentially controlled by IL-3 and PKC

In terms of IgE-dependent stimulation of IL-4 and IL-13, there is no clear evidence of differential control regarding the production of these cytokines. However, the kinetics of their release from basophils is strikingly different, suggesting that factors other than the signaling routes already described above are involved. IL-4 is released rapidly, peaking within 4 hours of stimulation, and is accompanied in some donors by the release of preformed stores of this cytokine.²⁶,¹²⁹,¹³⁰ In contrast, IL-13 is detectable in basophil supernatants beginning 2 hours after FcεRI cross-linking, and the amounts of this factor continue to rise for at least 16 hours.⁶ At present, the reasons for these differing kinetics are unknown, but factors such as autocrine effects are now being investigated.

In terms of the levels of IL-4 and IL-13 produced, there is growing evidence that signals employed by priming factors such as IL-3 favor the production of IL-13 rather than IL-4. IL-3 potentiates the IgE-dependent release of both factors but, in addition to this, Schroeder et al, as well as Redrup et al, have reported that IL-3 stimulation alone is sufficient itself to give rise to IL-13 generation without a marked increase in IL-4 production.¹²⁶,¹²⁷ These authors further showed that, in stark contrast to IgE signaling, this IL-3-dependent IL-13 release is insensitive to FK506, suggesting the involvement of an activatory pathway independent of calcineurin and NFAT.

A further tier of differential control of IL-4 and IL-13 release is seen with respect to PKC. Following PMA-induced PKC activation in basophils, Schroeder and coworkers have reported that although messenger RNA expressions for both IL-4 and IL-13 are increased only IL-13 protein is released.¹²⁶,¹²⁷ It is not yet clear, however, which isozymes of PKC are involved. The same authors have shown that the selective PKC inhibitor bisindolylmaleimide I1 not only reverses the effects of PMA activation but potentiates the anti-IgE induced IL-4 release but not that of IL-13. Interestingly, bisindolylmaleimide I1, as well as FK506, does not affect IL-3–induced secretion of IL-13, thus showing that IL-3–associated signaling in basophils diverges considerably from both calcium and PKC-sensitive responses compared with IgE-dependent pathways.

Future aspects

The brief summary above regarding the control of IgE signaling in basophils highlights 2 major problems for future consideration.
First, functional differences exist between primary basophils and mast cells or basophil cell lines and, second, IgE-dependent signaling is greatly affected by the actions of priming factors such as IL-3. Thus, the net outcome of IgE-triggered basophil activation in vivo is likely to be modulated by varying degrees of the local concentrations of hematopoietic cytokines (such as IL-3, IL-5, and GM-CSF) as well as neurotrophic cytokines (NGF), complement factors, and histamine-releasing factors. Because basophils are geared to the rapid release of Th2-type cytokines, it would be of considerable interest to establish the concentrations of the above factors in the in vivo environment, particularly during an allergic reaction where acute (IgE-mediated) basophil activation occurs.

What is the physiologic role of basophils in the immune system?

Although their existence has been known for more than a century, the physiologic role of basophils in immunity remains a mystery. It is conceivable that basophils, like eosinophils, neutrophils, macrophages, and mast cells, fulfill important roles in innate immunity against pathogenic organisms. Important clues could be derived from the study of natural or experimentally induced gene deficiencies, as happened with W/Wv c-kit–deficient mice, which have proven very useful in dissecting the role of mast cells in immunity to intestinal nematodes, or IL-3−/− mice. Unfortunately, in the absence of a precise knowledge about the unique developmental pathway of basophils, none of the currently available gene-deficient mice are likely to elucidate the exclusive role of basophils in immunity.

 Mast cells and basophils have been proposed to play an important role in the innate immune response to a variety of pathogens. For mast cells such a role is corroborated by an increasing body of evidence, whereas the current evidence for a comparable involvement of basophils is, at best, circumstantial. Such a role would require the ability of basophils to be activated in a non–antigen-specific manner. Indeed, several such non–antigen-specific stimuli, derived from various organisms, have been shown to induce mediator or cytokine release from basophils. For example, protein Fv, an endogenous Ig-binding protein released in the intestine of patients affected by viral hepatitis, as well as the HIV-1 glycoprotein gp 120 have recently been shown to induce mediator or cytokine release from basophils. It is therefore intriguing to note that several parasitic helminths, which are well known to induce a strong Th2 response in their host, have been recently shown to produce or secrete lectins. For instance, infective larvae of the nematode parasite *Toxocara canis* secrete a novel C-type lectin (TEN-32) that surprisingly binds both mannose- and galactose-type monosaccharides, which are both present on the carbohydrate side chains of IgE. The hematophagous parasite *N americanus* produces a major allergen that is a putative homolog of calretulin, a lectin with a carbohydrate affinity that also matches the glycosylation pattern of human IgE. Several pathogenic organisms may induce the activation of basophils by the release of superantigens, a term introduced by Patella et al, which in this context would define molecules with the ability to cross-link FceRI either directly or via receptor-bound IgE in a non–antigen-specific manner.
Taken together, the observation that different classes of molecules derived from pathogens or endogenous sources can directly activate basophils independently of the presence of specific IgE on their surface fulfills the theoretical requirement for a role of this cell type in innate immunity or in bridging innate and specific immunity by skewing the differentiation of naïve T cells to the Th2 phenotype. Thus, more research is needed to address the fundamental question concerning the true physiologic role of basophils in the immune system.

**Tools for basophil research**

Investigation into the role of basophils in health and disease has previously been curtailed by the lack of suitable experimental tools. The following section, therefore, summarizes the currently available tools for basophil research.

**Monoclonal antibodies**

Because basophils are quickly recruited to the tissues in a variety of allergic diseases, monoclonal antibodies allowing specific detection of basophils by immunohistochemical staining would be a highly desirable tool, eg, for studying the relative contribution of this cell type to pathogenesis. Earlier protocols relied on the presence of FcεRI and the absence of tryptase for immunohistochemical detection. It has become clear, however, that the expression of FcεRI is not restricted to mast cells and basophils, because it can be up-regulated on other cells such as Langerhans cells and other dendritic cells, eosinophils, and monocytes. Furthermore, the expression of tryptase cannot be used for discrimination of mast cells from basophils, because the gene for tryptase α is transcribed in the latter cell type and is present in substantial amounts in peripheral blood basophils of patients with asthma or other allergic diseases. Therefore, several laboratories have generated antibodies for specific detection of basophils in flow cytometry or immunohistochemistry.

There are currently 3 basophil-specific monoclonal antibodies: BSP-1, 2D7, and BB-1. BSP-1 is an IgM class monoclonal antibody that recognizes an epitope expressed on the surface of human basophils and was originally raised against the human erythroblastic leukemia cell line (HEL). It reacts with a 45-kd surface antigen on HEL cells in Western blots. Because of its low sensitivity, BSP-1 is not suitable for immunohistochemistry. Monoclonal antibody 2D7 is an IgG1κ monoclonal antibody that reacts with a ligand localized to the secretory granules of human basophils. Western blots with basophil extracts exhibit 2 major bands with apparent molecular weights of 72 and 76 kd and a minor band of 124 kd. Immunologic activation of basophils leads to the loss or attenuation of the staining intensity, indicating release (or decay) of the 2D7 antigen. Monoclonal antibody 2D7 has successfully been used for immunohistochemical staining of basophils in human skin during the LPR after cutaneous allergic challenge.

The most recent addition to the pharmacopoeia is BB-1, an IgG2a monoclonal antibody that recognizes a 124- (± 11) kd antigen (estimated by the method of Hedrick and Smith) localized mainly to the secretory granules with a comparatively small expression on the surface. The BB-1 antigen was released upon activation with anti-IgE or the calcium ionophore A23187. Because the BB-1 antigen was not detected in any other cell types, the authors suggest its use as a discriminating marker of basophil activation, particularly in the tissues, where it has been successfully used for immunohistochemical detection of basophils in lung sections fixed in Carnoy’s fluid.

**Cell lines**

Several studies have used the cell line KU812, originally established from a patient with chronic myelogenous leukemia, and has served as a model for basophil differentiation. Several cytokines can induce its differentiation into basophil-like cells, including TNF-α and IL-6, IL-3, IL-4, and IL-6. As shown by Hara et al, incubation of KU812 cells with IL-4 induced several changes, including a 10-fold increase of total histamine content, appearance of metachromatic staining with toluidine blue, and up-regulation of functionally active FcεRI on the surface after 7 days, with increased transcription of FcεRI α, β, and γ chains’ messenger RNA. Another cell line, termed LAMA-84, was established in 1987 by Seigneurin et al and has also been described as basophil-like. Kepley and coworkers have recently described the in vitro generation of fully functional basophils from cord blood. The protocol consists in pulsing normal cord blood leukocytes with IL-3 for 3 to 4 hours and subsequently incubating them with fetal calf serum. The resulting cells are mostly 2D7+ FcεRI+, express a series of integrins also found on normal peripheral blood basophils (CD11b, CD18, CD29, and CD49d), and display basophil-like morphology by light or electron microscopy. Activation through the high-affinity IgE receptor results in a time- and dose-dependent release of preformed mediators.

Figure 3. Purified human peripheral blood basophils stained with May-Grünwald-Giemsa obtained with a recently published basophil purification protocol. (A) Original magnification × 160. (B) Original magnification × 1000. Courtesy of A. Gronow and K. Haisch, Forschungszentrum Borstel, Germany.
release of histamine. The use of basophil-like cell lines, however, must be cautiously examined given the fundamental differences in morphology and function compared with primary human basophils.

**Purification protocols**

A high degree of basophil purity is often required for functional studies with basophils, especially when mediator secretion from contaminating cells masks what is released by basophils. Pure basophil populations are also mandatory when investigating either intracellular signaling events or to exclude the possibility of priming cytokines (eg, IL-3, GM-CSF), derived from cellular contaminants, to influence basophil function. Therefore, several protocols for basophil purification have been published over the last decade (see references in Haisch et al177). When purifying basophils, one is faced with several firsts. First, the average percentage of basophils in the peripheral blood of healthy individuals is low (<1%), which restricts the possible yield. Furthermore, a large donor variation in the proportion of basophils makes the final recovery of basophils unpredictable, especially when dealing with blood from unknown donors (ie, from buffy coats).178

Most of the currently published purification techniques rely on initial physical separation methods (density gradients or elutriation) that, by themselves, do not reliably result in very high purity but are successful in enriching basophils sufficiently enough for further purification by immunoselection. Some protocols have relied on the expression of the high-affinity receptor FceRI on the surface of human basophils.179,180 This positive selection technique, although resulting in very high basophil purity, is not desirable for many purposes because of the likelihood of activation during purification. The use of negative selection with magnetic beads178,181,182 consistently resulted in viable, functional (nonpreactivated) basophils but, in our experience, high yields were usually obtained only with hyperbasophilic donors.178 A recently published protocol, however, has overcome this limitation (Figure 3) and uses a commercially available kit,177 therefore making the purification protocol widely accessible to any laboratory with an interest in granulocyte function. A similar protocol combines density gradient centrifugation with negative selection with magnetic beads.183

Taken together, the availability of effective protocols for the purification of nearly homogeneous basophils or for the generation of this cell type from peripheral blood precursors as well as the availability of monoclonal antibodies for immunohistochemical detection and of basophil-like cell lines should encourage further study of human basophil function. The development of similar tools for the study of murine basophils would be extremely useful because it would enable the research to be extended to experimental models of disease.

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**References**


137. Patella V, Florio G, Petraroia A, Marone G. HIV-1 gp120 induces IL-4 and IL-13 release from hu-
man Fc epsilon RI+ cells through interaction with the VHR region of IgE. J Immunol. 2000;164:589-
595.
138. Falcone FH, Dahinden CA, Gibbs BF, et al. Hu-
man basophil release interleukin-4 after stimula-
139. Pruzansky JS, Grammer LC, Petterson R, Rob-
erts M. Dissociation of IgE from receptors on hu-
1949-1953.
140. Gregory WF, Blaxter ML, Maizels RM. Differen-
tially expressed, abundant trans-spliced cDNAs from larval Brugia malayi. Mol Biochem Parasitol. 
1997;87:95-95.
141. MacDonald SM, Rahn T, Langdon J, Lichten-
stein LM, MacDonald SM. Human recombinant histamine-
releasing factor. Int Arch Allergy Immunol. 1997; 
142. Xia HZ, Keppler CL, Sakai K, Chelliah J, Irani AM, 
Schwarz LB. Quantitation of thryptase, chymase, Fc epsilon RI alpha, and Fc epsilon RI gamma-chains and can use this 
affinity IgE receptor on eosinophils is involved in defence against parasites. Nature. 1994;367:136-
138.
144. Maurer D, Fiebiger E, Reininger E, et al. Express-
ion of functional high affinity immunoglobulin E 
receptors (Fc epsilon RI) on monocytes of atopic 
145. Kepley CL, Craig SS, Schwarz LB. Identification 
and partial characterization of a unique marker for 
6555.
146. Ishizaka T, Yamada K, Tachibana H. Basophilic dif-
culties in the human basophilic leukaemia cell line. Bioch 
terleukin-4: its role in the switch towards a Th2 
148. Coffman RL, von der Weid T. Multiple pathways 
149. Siraganian RP, Siragania PA. Mechanism of ac-
tion of covalent interaction of an IgE-de-
150. Schroeder JT, Lichtenstein LM, MacDonald SM. 
Recombinant histamine-releasing factor en-
hances IgE-dependent IL-4 and IL-13 secretion by human basophils. J Immunol. 1997;159:447-
452.
151. MacDonald SM. Human recombinant histamine-
releasing factor. Int Arch Allergy Immunol. 1997; 
152. Prazak JS, Grammer LC, Petterson R, Rob-
erts M. Dissociation of IgE from receptors on hu-
1949-1953.
153. Wantke F, Lichtenstein LM, MacDonald SM. 
The human recombinant histamine 
releasing factor: functional evidence that it does not bind to the IgE molecule. J Allergy Clin 
terleukin-4: its role in the switch towards a Th2 
155. Winkfe F, MacLaughlin DW, Langdon JM, Mac-
Donald SM. The human recombinant histamine 
releasing factor: functional evidence that it does not bind to the IgE molecule. J Allergy Clin 
lecitins can induce in vitro release of IL-4 and 
1999;29:918-927.
157. Loukas A, Maizels RM. Helinth C-type lectins 
and host-parasite interactions. Parasitol Today. 
158. Shibasaki M, Sumazaki R, Isomaya S, Takita H. 
Interaction of lectins with human IgE. IgE-binding property and histamine-releasing activity of 
lecitins can induce in vitro release of IL-4 and 
1999;29:918-927.
lecitins can induce in vitro release of IL-4 and 
1999;29:918-927.
157. Loukas A, Maizels RM. Helinth C-type lectins 
and host-parasite interactions. Parasitol Today. 
158. Shibasaki M, Sumazaki R, Isomaya S, Takita H. 
Interaction of lectins with human IgE. IgE-binding property and histamine-releasing activity of 
lecitins can induce in vitro release of IL-4 and 
1999;29:918-927.
lecitins can induce in vitro release of IL-4 and 
1999;29:918-927.
157. Loukas A, Maizels RM. Helinth C-type lectins 
and host-parasite interactions. Parasitol Today. 
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