Immunization of allogeneic bone marrow transplant recipients with tumor cell vaccines enhances graft-versus-tumor activity without exacerbating graft-versus-host disease

Larry D. Anderson, Jr., Cherylyn A. Savary, and Craig A. Mullen

Allogeneic bone marrow transplantation (BMT) induces 2 closely associated immune responses: graft-versus-tumor (GVT) activity and graft-versus-host disease (GVHD). We have previously shown that pretransplant immunization of allogeneic BMT donors with a recipient-derived tumor cell vaccine increases both GVT activity and lethal GVHD because of the priming of donor T cells against putative minor histocompatibility antigens (mHAgs) on the tumor vaccine cells. The work reported here tested the hypothesis that tumor cell vaccination after BMT would produce an increase in GVT activity without exacerbating GVHD. C3H.SW donor bone marrow and splenocytes were transplanted into major histocompatibility complex-matched, mHAG-mismatched C57BL/6 recipients. One month after BMT, recipients were immunized against either a C57BL/6 myeloid leukemia (C1498) or fibrosarcoma (205). Immunized recipients had a significant increase in survival and protection against tumor growth in both tumor models, and significant tumor protection was seen even in recipients with preexisting micrometastatic cancer before immunization. Alloreactivity appeared to contribute to the in vivo anti-tumor cytolytic activity, but in vivo immunity was tumor specific, and no exacerbation of GVHD was observed. Although the immunodominant mHAG B6dom1 was shown to be expressed by all B6 tumors tested and was largely responsible for the alloreactivity resulting from tumor immunization of donors, the in vitro alloreactivity of immune recipients was more restricted and was not mediated by recognition of B6dom1. In conclusion, post-transplant tumor immunization of allogeneic BMT recipients against either a leukemia or a solid tumor can increase GVT activity and survival without exacerbating GVHD.

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Bone marrow transplantation

BMT recipients received 850 cGy total body irradiation using a 60Co source 1 day before BMT. On the day of BMT, 2 to 4 × 10^8 bone marrow cells and 5 to 10 × 10^8 spleen cells were injected intravenously together in a total volume of 0.2 mL HBSS. Bone marrow was isolated from donors by flushing each femur and tibia with RPMI. Spleen cells were isolated by macerating the spleens between 2 frosted glass slides, followed by lysis of erythrocytes. Fewer than 5% of recipients were excluded from these studies before the initiation of immunization because of signs of severe GVHD.

Materials and methods

Animals

Female C57BL/6 mice were purchased from the National Cancer Institute (Frederick, MD), and female C3H.SW-H2bSnl (C3H.SW) mice were purchased from the Jackson Laboratory (Bar Harbor, ME). They were used for experiments at 6 to 12 weeks of age. Mice were housed in conventional rooms with food and water ad libitum. From 2 or 3 days before BMT until day 14, water was acidified (pH 2.5) and supplemented with 2 g/L neomycin sulfate (Sigma, St Louis, MO).

Cell lines

The cell line 205 is a weakly immunogenic methylcholanthrene-induced C57BL/6 fibrosarcoma cell line.10 This tumor is not spontaneously metastatic but reproducibly forms multiple lung nodules when at least 1 × 10^3 cells are injected intravenously into C57BL/6 mice. 205IL-2/TK is a 205 cell line modified to express both the interleukin-2 (IL-2) gene and the herpes simplex virus thymidine kinase (TK) suicide gene using the LXSNI and pBabePuro retroviral vectors, respectively. The pBabePuro vector has a puromycin resistance gene,11 and the LXSNI vector contains a neomycin resistance gene.12 Transduced cells were selected in 2.5 µg/mL puromycin, 1 mg/mL G418, or both. C1498 is a C57BL/6 myelomonocytic leukemia cell line of spontaneous origin (ATCC, Rockville, MD). When injected intravenously, 1 × 10^3 cells is a near-uniformly lethal dose of cells; animals die with gross evidence of leukemia, including hepatomegaly and splenomegaly. Both 205 and C1498 express major histocompatibility complex (MHC) class I but not MHC class II. B16F10 is a spontaneous, weakly immunogenic C57BL/6 melanoma cell line (a gift from Dr I. J. Fidler, M.D. Anderson Cancer Center). EL4 is a C57BL/6 lymphoma cell line (ATCC). P815 is a lymphohkine activated killer (LAK)-sensitive DBA/2 mastocytoma cell line (ATCC). Yac-1 is a natural killer (NK)-sensitive A/Sn lymphoma cell line (ATCC). Cells were grown in tissue culture using RPMI-1640 (RPMI) supplemented with 5% heat-inactivated fetal bovine serum (Biowhittaker, Walkersville, MD), and 2 mmol/L L-glutamine.

BMT recipient immunization

205IL-2/TK + ganciclovir immunization. We have shown that ganciclovir-mediated ablation of live 205IL-2/TK cells induces systemic immunity to unmodified 205 in C57BL/6 mice13 and in C3H.SW donors.4 In the initial experiments recipients were injected subcutaneously in the flank with 3 to 5 × 10^4 live 205IL-2/TK cells in 0.2 mL Hank’s balanced salt solution (HBSS) 1 month after BMT and then received 1 mg ganciclovir intraperitoneally in 0.2 mL phosphate-buffered saline daily for 1 week starting 3 days after the cell injection. Subsequent vaccines were given as above at 2-week intervals.

Irradiated 205IL-2/TK or 205 immunization. Some recipients (and control C3H.SW donors) were injected subcutaneously in the flank with 3 to 5 × 10^4 irradiated 205IL-2/TK or 205 cells in 0.2 mL HBSS 1 month after BMT. Vaccines were given at 1-week intervals.

C1498 tumor immunization. Mice were injected subcutaneously in the flank with 10 × 10^3 irradiated C1498 cells in 0.2 mL HBSS starting 1 month after BMT. Vaccines were given at a 1-week interval unless the vaccine was given at the time of intravenous leukemia challenge, which was 10 days after the previous vaccine.

In vivo tumor inoculation

Micrometastatic lung tumors were established by injecting C57BL/6 mice with 1 × 10^8 205 tumor cells intravenously in 0.2 mL HBSS. C1498 leukemia was established by injection of 1 to 1.5 × 10^6 cells intravenously. The timing of tumor challenge in relation to the vaccine schedule is specified in the text and legends.

Evaluation of GVHD

Recipients were weighed weekly and observed daily for signs of GVHD (weight loss, alopecia, dermatitis, hunched posture, and death). In some experiments histologic analysis of livers for GVHD was performed. Liver sections stained with hematoxylin and eosin were examined for characteristic mononuclear cell infiltrates in portal triads.

Cytotoxicity assays

For use as effector cells, spleen cells were cultured in 6-well plates at 1 × 10^6 cells/mL and 10 mL/well in RPMI supplemented with 10% FBS (Summit Biotech, Fort Collins, CO), 100 U/mL penicillin, 100 µg/mL streptomycin, 2 mmol/L L-glutamine, 100 mmol/L sodium pyruvate, 0.1 mmol/L nonessential amino acids, and 50 mmol/L L-2-mercaptoethanol (complete medium). In assays for allosecreativity, 30 Gy-irradiated C57BL/6 spleen cells or B6dom1 peptide-pulsed C1498 spleen cells were used as stimulators at a concentration of 1 × 10^6 cells/mL or 1.25 × 10^6 cells/mL, respectively. Peptide-pulsed stimulators were incubated for 1 hour at 37°C with 35 µg/mL B6dom1 peptide (AAPDNRRET, synthesized by the Peptide Synthesis Core Laboratory of M.D. Anderson Cancer Center). In assays for combined tumor-reactivity and allosecreativity, 250 Gy-irradiated 205 tumor cells or 100 Gy-irradiated C1498 leukemia cells were used as stimulators at a concentration of 5 × 10^4 cells/mL or 5 × 10^5 cells/mL, respectively. After 4 to 5 days in culture with the appropriate stimulator cells, effector cells were harvested and plated in triplicate with 5 × 10^5 51Cr-labeled target cells per well at effector:target ratios ranging from 200:1 to 12.5:1. Target cells were labeled by combining 5 × 10^5 cells in 0.1 mL complete medium with 20 µL FBS (Summit) and 0.1 mL (approximately 100 µCi) sterile isotonic Na2 51 CrO (Amersham, Arlington Heights, IL) for 60 minutes at 37°C. Concanaevalin A (ConA) lymphoblast (CAB) targets were generated by stimulating C57BL/6 or C3H.SW spleen cells for 2 days with 2 µg/mL ConA at 2 × 10^6 cells/mL in complete medium. They were labeled with 51Cr as above for 45 minutes. Some C3H.SW CAB were loaded with B6dom1 peptide by incubation for 90 minutes with 40 µg peptide (45 minutes before adding 51Cr plus 45 minutes after adding 51Cr). Labeled targets were washed 3 times before plating with effectors in a total volume of 0.2 mL/well in 96-well round-bottom plates. Plated cells were incubated for 4 hours at 37°C, after which 0.1 mL supernatant was counted in a gamma counter (Wallac, San Francisco, CA). Percentage lysis was calculated as 100 × (experimental cpm – spontaneous cpm)/(maximum cpm – spontaneous cpm)]. Spontaneous release was usually less than 20% and always less than 30% of the maximum release.

Statistical analysis

Prism 3.0 software (GraphPad Software for Scientists, Sorrento, CA) was used for statistical evaluation of data. When more than 2 groups of lung nodules were compared, a nonparametric 1-way analysis of variance (Kruskal-Wallis test) was performed. If P < .05 overall, then the groups were compared using a Dunn’s multiple comparison test. When only 2 groups were compared, a Mann-Whitney U test was used. To compare
numbers of mice with no lung nodules, the Fisher exact test was used. To compare Kaplan-Meier survival curves, the log-rank test was used.

Results

Tumor immunization of allogeneic BMT recipients increases GVT activity

Experiments were conducted to test the hypothesis that tumor immunization of allogeneic BMT recipients could protect against the growth of metastatic cancer in BMT recipients. We first assessed the kinetics of immune function recovery and found that recipients in this model could not mount a detectable cytolytic immune response at 2 weeks after BMT but began responding to tumor cell antigens by 1 month after BMT and began responding to influenza nucleoprotein antigen by 5 weeks after BMT, as assessed by in vitro cytotoxicity assays (data not shown). Therefore, in the experiments described in this study, we initiated vaccination at least 1 month after BMT. C3H.SW → C57BL/6 recipients in the initial experiments were immunized with 205IL-2/TK cells plus ganciclovir. As seen in Figure 1, recipients immunized only a single time were not protected against tumor growth, but those immunized again at the time of tumor challenge had complete protection against growth of pulmonary tumor nodules \( (P < .05) \) and significantly enhanced survival \( (P = .0040) \) compared to nonimmunized recipients. All deaths among recipients immunized twice resulted from growth of a nonpulmonary metastasis. Nonpulmonary metastases usually consisted of tumors in the connective tissue of the sacral/pelvic/paraspinal region or ovary. Nonpulmonary tumors also grew in some nonimmunized recipients.

Because of evidence of greater efficacy with multiple vaccine treatments, recipients in subsequent experiments were immunized 4 times. Recipients immunized 4 times with irradiated 205-IL-2/TK cells had markedly extended survival \( (P < .0001) \) and often had complete prevention of lung nodule growth \( (P = .0011) \) (Figure 2B). Furthermore, 92% (11 of 12) survived more than 100 days without signs of tumor or GVHD. Recipients immunized 4 times with irradiated, unmodified 205 cells also had enhanced survival \( (P = .0016) \) and a significant reduction of lung nodules \( (P = .0022) \). Fifty percent (3 of 6) survived more than 100 days, and 83% (5 of 6) were completely protected against lung nodules.

We next tested the influence of post-BMT vaccines in a second tumor model, the C57BL/6 C1498 model of acute myelogenous leukemia. Using the immunization method shown by Boyer et al to induce a T-cell-mediated immune response in syngeneic C57BL/6 mice, C3H.SW → C57BL/6 BMT recipients were immunized 4 times with irradiated C1498 cells. Immunization significantly extended survival \( (P < .0001) \) compared to nonimmunized recipients, and 71% (5 of 7) of recipients achieved long-term survival for more than 100 days without signs of leukemia or GVHD. In all recipients that died, there was gross evidence of leukemia, including hepatomegaly, but not GVHD.

**GVT activity is induced by tumor cell vaccines in BMT recipients with preexisting micrometastatic cancer**

Both 205 and C1498 tumors progress rapidly in vivo, often leading to death within 1 month. Because the induction of primary T-cell responses usually does not occur until 7 to 10 days after the
administration of vaccines, we predicted that vaccine efficacy would be reduced in mice with established, progressively growing tumors. Indeed, in mice inoculated with C1498 leukemia cells 1 week before immunization, survival was not prolonged by tumor vaccination (Figure 5). However, in recipients with C1498 leukemia established 3 days before the initiation of vaccination, survival was significantly increased compared to nonimmunized recipients ($P = .0351$; Figure 5). In the 205 tumor model, recipients that began vaccine treatment 3 days after the establishment of metastases had a significant reduction in pulmonary tumor nodules ($P = .0140$, Figure 6). In a separate experiment in which long-term survival was the endpoint, the observed increase in median survival after 205 tumor challenge (from 31 to 41 days) was not statistically significant (data not shown). Death in these vaccine-treated animals resulted from growth of either nonpulmonary tumors or progressive growth of a small number of pulmonary nodules.

**Tumor immunization of allogeneic BMT recipients does not exacerbate GVHD**

Recipients surviving tumor challenge in the above experiments did not develop signs of acute GVHD (eg, weight loss, fur loss, diarrhea, dermatitis, or histologic evidence of GVHD in the liver). This suggested that tumor cell vaccines administered after BMT did not increase GVHD. To test this hypothesis directly, we also conducted 3 experiments in which recipients were immunized 4 times against 205 but were not challenged with tumor. All immunized recipients in both experiments ($n = 16$) survived more than 100 days without signs of GVHD, including weight loss.

**GVT activity induced by tumor immunization of recipients is tumor-specific in vivo**

We have previously shown that the GVT activity induced by tumor immunization of allogeneic BMT donors is mediated by alloreactive T cells that cross-react broadly with recipient tumors and nonmalignant tissues. Therefore, experiments were performed to determine whether recipient immunization would induce cross-protection...
against another immunologically unrelated recipient tumor. Recipients were either not immunized (nonimmune) or immunized against 205 (205-immune) or C1498 (C1498-immune) and then challenged with either 205 or C1498. As seen in Figure 7, the GVT activity of 205-immune recipients was tumor specific. The 205 vaccine again induced protection against 205 (P = .0006; Figure 7B) but did not protect against C1498 challenge compared to nonimmune controls (Figure 7A). Similarly, vaccination against C1498 increased survival after challenge with C1498 immunization (Figure 7A) but not after challenge with 205 (Figure 7B). Reduction in 205 pulmonary metastases was seen with the use of 205 vaccines but not with the use of C1498 vaccine (Figure 7C).

Tumor-immunized allogeneic BMT recipients exhibit in vitro cytolytic antitumor activity but only limited alloreactivity

To evaluate further the specificity of the antitumor response, in vitro cytotoxicity assays were performed. Splenocytes from immunized BMT recipients or control immunized donors (C3H.SW) were stimulated in vitro with tumor cells and tested for their ability to kill various target cells (Figure 8). Immunization of C3H.SW donors with either 205 or C1498 vaccines generated broadly alloreactive CTL activity, lysing all C57BL/6 targets. In contrast, BMT recipients immunized with 205 or C1498 generated cytotoxic cells with a different target specificity profile. In 3 independent experiments, BMT recipients generated cytotoxic cells that killed the immunizing tumor, but cross-reactivity was only seen against B16F10 in the case of 205-immune recipients and against 205 for C1498-immune recipients. Neither immune recipients nor control immunized C3H.SW donor splenocytes killed Yac-1 or P815, indicating that the cytolytic response was not caused by nonspecific NK or LAK activity.

Unresponsiveness against the immunodominant recipient minor histocompatibility antigen B6dom1 is not reversed by post-transplant tumor immunization of recipients

Perreault et al15-17 have identified an immunodominant peptide mHAg (B6dom1), which is presented by H-2Db on normal C57BL/6 cells and is at least partially responsible for GVHD in the C3H.SW → C57BL/6 BMT model. Therefore, we wanted to determine whether tumor cell vaccines induce a response to B6dom1 in C3H.SW mice, whether C57BL/6 tumor cell lines express B6dom1, and whether BMT recipients fail to respond to B6dom1, which would help explain their limited alloreactivity compared to immunized donors. First, C3H.SW mice were immunized with either C57BL/6 spleen cells (B6-immune) or irradiated 205IL-2/TK tumor cells (205-immune). Control mice were not immunized (naïve SW). After stimulation of immune splenocytes with B6dom1 peptide in

Figure 6. Recipient immunization with a tumor cell vaccine increases GVT activity against preexisting 205 tumor. One month after SW→B6 BMT, micrometastatic fibrosarcoma was established by intravenous injection of 1 × 105 205 tumor cells. Recipients were either not immunized (nonimmune) or immunized 4 times with 50 GY-irradiated 205IL-2/TK cells (205-immune) at a weekly interval starting on day 3 after challenge. Lung nodules were counted 1 month after tumor challenge. The day 3 205-immune group had a significant reduction in lung nodules (P = .0140) compared to the nonimmune recipients.

Figure 7. GVT activity after tumor immunization of BMT recipients is tumor-specific in vivo. One month after BMT, SW→B6 recipients were immunized twice with irradiated C1498 leukemia cells (C1498-immune) or were irradiated 205IL-2/TK cells (205-immune) at a 1-week interval. Control recipients (nonimmune) were not immunized. Ten days after the 2nd vaccine, 1.5 × 104 C1498 cells (A) or 1 × 105 205 cells (B, C) were injected intravenously to simulate relapse after BMT, and immune recipients were immunized twice more at 1-week intervals. Lung nodules were counted in the 205-challenged mice at the time of death or after 100 days. Although tumor immunization against either 205 or C1498 induced protection (P < .05) against the immunizing cell type, it did not induce cross-protection against the other tumor (P > .05 for both survival tests and lung nodules). Results were pooled from 2 independent experiments.
in vitro, we tested their ability to specifically kill C3H.SW ConA lymphoblasts loaded with B6 dom1 peptide (B6 dom1 /SW CAB) versus control unpulsed SW CAB. As seen in Figure 9A, both 205-immune and control B6-immune donors developed a specific and almost identical response to B6 dom1 , demonstrating that 205 tumor cells do express B6 dom1 as an immunodominant antigen. We next tested the ability of B6-immune, B6 dom1 -stimulated C3H.SW cells to kill various C57BL/6 tumor cell lines (Figure 9B). All lines tested (205, C1498, B16F10, and EL4) were killed efficiently and therefore express B6 dom1 , explaining why 205-immune C3H.SW splenocytes were broadly cross-reactive against all of them as seen in Figure 8.

Because tumor-immunized recipients do not have broad cross-reactivity against all C57BL/6-derived targets (unlike tumor-immunized C3H.SW), it is possible that they do not mount a substantial response to B6 dom1 . To test this hypothesis we examined the relative ability of 205-immune BMT recipient splenocytes to kill B6 dom1 -loaded SW CAB and regular SW CAB (Figure 10). Although 205-immune C3H.SW controls again mounted a strong response to B6 dom1 , 205-immune BMT recipients had virtually no response to B6 dom1 .

Unresponsiveness against recipient mHAgs responsible for GVHD is not reversed by secondary adoptive transfer of lymphocytes from tumor-immunized BMT chimeras into new irradiated C57BL/6 hosts

Although use of tumor vaccines after BMT did not increase severe acute GVHD (as measured by death, weight loss, dermatitis, and histology), it is possible that tumor vaccines may have produced subclinical exacerbation of GVHD. To test this possibility, we transferred tumor-immunized BMT recipient splenocytes into newly irradiated C57BL/6 hosts. This is potentially a more sensitive bioassay for the detection of GVHD because in it the secondary hosts may generate the total body irradiation-induced cytokine storm, a critical cofactor in the generation of severe GVHD.18-22 Long-term survivors of C3H.SW → C57BL/6 BMT and irradiated 205IL-2/TK immunization were reimmunized twice before transferring their bone marrow and spleen cells to irradiated C57BL/6 recipients. Secondary transfer of 205-immune BMT recipient (chimera) splenocytes did not induce any signs of GVHD in any of the recipients tested (Table 1).
compared to final weight of recipients of nonimmune C3H.SW. Flow cytometry, which showed that the majority of Thy1.2
the weight of the surviving mice at day 108 or the weight at death for the 2 dead mice.

adoptively transferred to C57BL/6 recipients that were given 850 cGy total body
splenocytes were stimulated for 5 days in vitro with B6dom1 -loaded C3H.SW spleen
205IL-2/TK cells at a 1-week interval. T en days after the second vaccine, their
One month after BMT , alloge
lymphocytes into irradiated recipients does not induce GVHD
Table 1. Secondary adoptive transfer of 205-immune BMT chimera
lymphocytes into irradiated recipients does not induce GVHD

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C3H.SW or long-term surviving C3H.SW — B6 BMT chimeras were either not
immunized (nonimmune) or immunized (205-immune) with 5 × 10^6 irradiated
205IL-2/TK cells twice at a weekly interval. Ten days later, their splenocytes were
adoptively transferred to C57BL/6 recipients that were given 850 cGy total body
irradiation 1 day earlier. Survival was monitored until day 108. Final weights represent
the weight of the surviving mice at day 108 or the weight at death for the 2 dead mice.
Secondary transfer recipients were engrafted with C3H.SW T-cells as assessed by
flow cytometry, which showed that the majority of Thy1.2- cells in the spleen after
sacrifice were CD5.1-.

This work tested the hypothesis that immunization of MHC-
matched allogeneic BMT recipients with a tumor cell vaccine would substantially increase GVT activity and prolong tumor-free
survival without exacerbating GVHD. Tumor vaccination of recipients
against either a fibrosarcoma or a myeloid leukemia produced a
substantial increase in GVT activity, which was capable of
complete protection against tumor growth and of preventing the
growth of preexisting micrometastatic cancer cells. Furthermore,
BMT recipients did not develop signs of acute GVHD after tumor
cell vaccination, demonstrating that GVT activity can occur
independent of GVHD.

This study shows that tumor cells of diverse histology are
capable of both expression of a minor histocompatibility antigen
(mHAg) (B6dom1) and induction of an immune response against this
mHAg, which is an immunodominant GVHD target antigen in this

BMT model.15-17 This finding confirms at the molecular level our
previous findings in the allogeneic donor immunization model1
and elucidates the mechanism whereby tumor-immunized donor lymphocytes can kill various recipient targets and cause GVHD.
Specific restimulation of tumor-immune donor cells with the
H-2Dk-restricted B6dom1 peptide in vitro caused a similar level of
tumor cell lysis compared to tumor cell restimulation, indicating
that much of the alloreactive cytolytic T-cell activity seen after
donor immunization with tumor cells is caused by the recognition
of B6dom1.

Although the same vaccines used in this study have been shown to
increase both GVT activity and lethal GVHD when used to treat
donors before BMT,4 they did not cause GVHD when administered
after BMT. This is consistent with the hypothesis that after
transplantation, naive donor-derived T cells eventually become
unresponsive to the immunodominant mHAg's that are potential
targets for GVHD. Because the antitumor response of recipients
after BMT does not exacerbate GVHD, it is likely that much of
the immune response is directed against other antigens, such as
nonimmunodominant mHAg's, with restricted tissue distribution or
possibly even tumor-specific antigens. This is supported by the
observation that in vivo tumor protection in this study was tumor
specific; the fibrosarcoma and leukemia vaccines induced immu
nity that did not cross-protect against growth of the other tumor
(Figure 7), despite the presence of common mHAg's on both tumors
(Figure 9).4 The hypothesis that allogeneic BMT recipients become
unresponsive to immunodominant mHAg's that are targets of
GVHD is also supported by the finding that although donor
immunization with recipient tumor cells induced a potent response
to B6dom1, immunization of recipients 1 month after BMT failed to
induce a significant response to B6dom1. Such T-cell unrespon
siveness to B6dom1 explains why immune recipient cytolytic activity is
not broadly cross-reactive against all recipient targets. The mecha
nism for this unresponsiveness is unknown. These data are
consistent with either clonal deletion, anergy, or the generation of
suppressor cells. Further studies are being conducted to help
elucidate the mechanism of unresponsiveness to recipient mHAg's,
which will be helpful in learning how to better apply tumor
vaccination strategies to human BMT recipients.

The limited pattern of cytolytic activity seen in vitro was likely
caused by activation and expansion of some T-cell clones that
recognized tumor antigens and other T cells that recognized
mHAg's other than B6dom1. These other putative mHAg's appear to
be expressed on some but not all tumor cell lines and with limited
or no expression on normal lymphoblasts or targets of GVHD. For
example, 205-immune recipient splenocytes recognized both 205
and B16F10 tumors (Figure 8A). (205 and B16F10 do not induce
cross-reactivity in normal, syngeneic C57BL/6 mice and therefore
do not share tumor-specific antigens.)13 That this cross-reactivity
was attributable to the recognition of shared mHAg's is supported
by the observation of lysis of 205 and B16F10 after in vitro
allostimulation of 205-immune recipient splenocytes with normal
C57BL/6 splenocytes that do not express tumor antigens (data not
shown). Nevertheless, the mHAg's recognized by post-transplant
BMT recipients must be different than those recognized by
immunized donor because of the lack of broad cross-
reactivity against all recipient targets, which was seen after donor
immunization (Figure 8). The lack of expression of the target antigens
of alloreactivity by target cells of GVHD is further supported by the
observation that the alloreactive T cells did not induce GVHD even
when transferred to secondary irradiated recipients (Table 1).
Although useful in many respects, the experimental BMT model used in these studies does not exactly replicate the clinical situation in which the BMT recipient is in clinical remission but harbors minimal residual tumor at the time of BMT that may reappear clinically in months or years. As was shown, the experimental murine tumors showed rapid growth producing death in approximately 1 month. In these murine transplants, as in human transplants, the capacity of the host to respond to immunization to antigens in the first month is very poor. Therefore, it was not practical within this model to test the efficacy of the tumor vaccines in the setting of preexisting tumor. The results must be viewed with this in mind. It is possible that the results of post-BMT tumor vaccination would be different in such a setting. One could hypothesize that minimal residual tumor at time of BMT could make the donor-derived lymphocytes tolerant to antigens on host tumor cells. We are developing BMT models to test this hypothesis.

Use of post-BMT tumor vaccines in human trials will necessarily require careful scrutiny of the potential of such vaccines to exacerbate GVHD. Because of the complex and uncharacterized expression of most mHAgS on tumor cells, caution must be exercised about using whole tumor cells as vaccines until methods are developed for identifying mHAgS, their pattern of tissue distribution, and their relative ability to induce GVT activity and GVHD. However, in this study we showed that the induction of GVT activity can occur independent of GVHD if recipients are immunized with cancer cells after the development of unresponsive-ness to immunodominant mHAgS. In some human malignancies, such as melanoma, tumor-specific antigens and antigens with relatively restricted tissue distribution have been identified. Perhaps the identification of such tumor antigens for other malignancies may lead to the ability to immunize BMT donors or recipients without the risk for exacerbating GVHD.

References

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