von Willebrand Factor Propeptide in Vascular Disorders: A Tool to Distinguish Between Acute and Chronic Endothelial Cell Perturbation

By J an A. van Mourik, Ria Boertjes, Inge A. Huisveld, Karin Fijnvandraat, Dasja Pajkrt, Perry J. J. van Genderen, and Rob Fijnheer

Before de novo synthesized von Willebrand factor (vWF) leaves the endothelial cell, it undergoes endoproteolytic cleavage of its propeptide (vW antigen II). The processed vWF and propeptide are either released constitutively or, following activation of the endothelium, released through the regulated pathway. In a recent study (Borchiellini et al, Blood 88:2951, 1996), we showed that the half-life of mature vWF and of its propeptide differ fourfold to fivefold. We postulated that the molar ratio of the propeptide to mature vWF could serve as a tool to assess the extent of endothelial cell activation under physiologic and clinical conditions. To test this hypothesis, we measured mature vWF and propeptide in patients with documented acute and chronic vascular disease, including patients with thrombotic thrombocytopenic purpura (TTP), acute sepsis, and diabetes mellitus. These data were compared with experimental conditions in healthy subjects in which perturbation of the endothelium was simulated by physical exercise or by administration of 1-deamino-8-D-arginine vasopressin (DDAVP) or endotoxin.

In all individuals of the latter study group, both vWF and propeptide levels were elevated during the acute phase of the experimentally induced vascular perturbation; at later time points after stimulation, only vWF levels remained elevated. In patients with sepsis and TTP, both vWF and propeptide were elevated several-fold. Thus, this pattern can readily be explained in terms of acute perturbation of the endothelium. In contrast, in patients with diabetes mellitus, propeptide levels were only slightly elevated, whereas vWF levels were elevated twofold to threefold.

These observations support our hypothesis that measurement of both propeptide and vWF levels allows to discriminate between chronic and acute phases of endothelial cell activation in vivo. Measurement of only vWF is less indicative in this respect.

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VON WILLEBRAND FACTOR (vWF) is a large adhesive glycoprotein that mediates the adhesion of platelets at sites of vascular damage and also functions as a stabilizing carrier protein of coagulation factor VIII. It is one of the circulating blood proteins that is produced and released by vascular endothelial cells and is frequently used as an indicator of endothelial cell dysfunction in vascular disorders. Before de novo synthesized vWF leaves the endothelial cell, it undergoes endoproteolytic cleavage of its propeptide (also known as vW antigen II) and, together with the propeptide, is released through both the constitutive pathway and by stimulus-induced exocytosis of specialized secretory vesicles (Weibel-Palade bodies). Both in cultured, resting endothelial cells and in stimulated endothelial cells, the stoichiometry of the released propeptide to the released mature vWF is essentially equimolar.

However, in normal plasma the molar concentration of the propeptide is about one tenth of the concentration of mature vWF. Because the propeptide disappears four to five times faster from the circulation than mature vWF, it seems reasonable to assume that the observed differences in steady-state concentration are due to differences in half-life of the respective polypeptides. Upon perturbation of the endothelium, for instance elicited by experimental disseminated intravascular coagulation (DIC) or administration of 1-deamino-8-D-arginine vasopressin (desmopressin, DDAVP), both mature vWF and propeptide concentrations rapidly increase. Because of its rapid turnover, the propeptide concentration returns to its baseline value much faster after termination of the vascular challenge than the vWF levels. On the basis of these observations, it was postulated that measurement of both propeptide and mature vWF levels could provide a means to assess the extent and time course of endothelial cell activation under clinical conditions. For instance, if both vWF and propeptide levels are elevated, this would be indicative of acute vascular perturbation, whereas conditions in which only vWF is elevated, this would rather reflect chronic endothelial cell activation. In the present cross-sectional study, we have tested this hypothesis. We measured mature vWF and propeptide in patients with thrombotic thrombocytopenic purpura (TTP), sepsis, and diabetes mellitus. Healthy individuals, in whom an acute increase of vWF and propeptide levels were provoked by a standardized exercise test or injection of a low dose of endotoxin or DDAVP, served as a control in this study.

MATERIALS AND METHODS

Patient Selection

TTP. Thirteen patients with TTP were studied (mean age, 36 years; range, 23 to 45). Four patients had TTP after allogeneic or autologous bone marrow transplantation (BMT). All patients had microangiopathic hemolytic anemia, thrombocytopenia (mean platelet count, 28 × 10^9/L; range, 12 to 48), and impaired renal function. Mean hemoglobin level
was 5.1 mmol/L (range, 2.9 to 6.4); mean lactate dehydrogenase (LDH) level was 2,311 U/L (range, 658 to 3,841; normal value, < 640 U/L). The mean values of these parameters were not different in patients with classic TTP and post-BMT TTP. Samples were obtained before treatment was started.

**Sepsis.** Fourteen consecutive patients were recruited into the study from a primary medical-surgical intensive care unit. The entry criteria for patients with suspected sepsis syndrome were (American College of Chest Physicians/Society of Critical Care Medicine consensus conference\(^1\)) the presence of a temperature (>38.3°C or <35°C) and low blood pressure. The sepsis syndrome was verified by positive blood cultures. Seven patients had a pulmonary and seven an abdominal focus for the sepsis. All patients had signs of DIC: all had a prolonged activated partial thromboplastin time (aPTT; 66 ± 8 seconds; range, 53 to 74 seconds; normal value, < 36 seconds) and prolonged prothrombin time (PT; 24.6 ± 6.1 seconds; range, 18 to 38 seconds; normal value, 15.4 ± 4.5 seconds). Fibrin degradation products (FnDPs) were elevated in 11 patients (4.9 to 74 seconds; normal value, 6 to 35 seconds) and decreased in 13 patients (50.6% to 11 patients (4.9 to 74 seconds; normal value, 6 to 35 seconds) and decreased in 13 patients (50.6% to 74 seconds; normal value, 6 to 35 seconds). Antithrombin levels were decreased in 13 patients (50.6% to 6 to 35 seconds) and decreased in 13 patients (50.6% to 74 seconds; normal value, 6 to 35 seconds).

**Diabetes mellitus.** Twenty-two patients with type 1 (n = 7) and type 2 (n = 15) diabetes mellitus were studied. The mean age was 60 years (range, 19 to 83). Ten patients suffered from retinopathy, 10 from nephropathy, and eight from neuropathy. The mean HbA\(_1c\) was 7.3% ± 1.8%. There was no difference in HbA\(_1c\) levels between type 1 and type 2 diabetes. Six patients were insulin-dependent; all other patients used oral antidiabetic drugs. Samples were taken when patients were relatively stable and no acute metabolic complications or infections occurred.

**Propeptide and vWF Levels and Platelet Count**

To examine a possible relationship between vWF and propeptide levels and platelet count, patients with either low or elevated platelet count were also included in this study.

**Bone marrow aplasia.** Five patients with thrombocytopenia due to chemotherapy for acute leukemia, two patients with primary aplastic anemia, and two patients with congenital thrombocytopenia were studied. Their mean age was 40 years (range, 19 to 59). Mean platelet count was 23 \(\times\) 10\(^9\)/L (range, 6 to 45). Patients with signs of infection or thrombocytopenic complications were excluded.

**Essential thrombocytosis.** Seven patients with high platelet count due to essential thrombocytocemia (ET) were studied (mean age, 53 years; range, 28 to 76). Mean platelet count was 910 \(\times\) 10\(^9\)/L (range, 494 to 1,800). All other causes for thrombocytosis were excluded by clinical features and laboratory investigations. Bone marrow examination by cytogenetic analysis was performed to exclude chronic myeloid leukemia. All ET patients were studied during treatment with aspirin.

**Controls.** Controls were subjects referred to the hospital, but who upon serial clinical and laboratory investigations were shown to have no vascular disease, infections, malignancies, diabetes, or other diseases that could affect vWF and propeptide levels. Eighteen individuals were recruited as a control for this study (mean age, 48 years; range, 19 to 77).

**Healthy Subjects**

**Experimental endotoxemia.** This study was designed as described previously.\(^9\) In the present, more extended study, eight healthy male volunteers were treated with endotoxin, administered as a 4-ng/kg injection intravenously in 1 minute. At different time points after the injection of endotoxin, blood samples were collected from the antecubital vein.

**DDAVP.** This study group consisted of nine healthy volunteers (five males and four females; mean age, 31 years) previously studied to assess the half-life of vWF after administration of DDAVP.\(^{12}\) They received 0.4 µg DDAVP/kg body weight.

**Exercise.** Five healthy males (mean age, 40 years; range, 35 to 45) were subjected to a standardized exercise test (cycle ergometer) as described previously.\(^{13}\) Blood samples were collected immediately before the exercise test and at 15 minutes after maximal performance.

**Collection of Blood and Assays**

Blood, after collection in vacutainer tubes containing 3.2% buffered citrate solution (1:9 vol/vol), was immediately placed on ice. After centrifugation at 3,000 g for 20 minutes at 4°C, plasma was aliquoted and stored at −70°C until batchwise assessment. Propeptide and mature vWF concentrations were measured by enzyme-linked immunosorbent assay (ELISA) as described previously.\(^9\) Normal plasma from a pool of 30 donors served as standard. This plasma pool contains 6.3 nmol/L propeptide, as assessed by calibration against purified recombinant propeptide,\(^9\) and 50 nmol/L of vWF (in half homodimers, estimated concentration\(^{14}\)). Soluble P-selectin (sP-selectin), a specific marker of platelet activation,\(^{15-17}\) was measured by ELISA as described previously.\(^16\) Platelets were counted with a Coulter counter. All samples were obtained after informed consent.

**Statistical Analysis**

All data are presented as the mean ± SEM. The means in vWF and propeptide levels were compared by Student’s t-test with vWF and propeptide levels found in the respective control samples. The Pearson correlation coefficient was used as a measure of linear association between two variables.

**RESULTS**

**Effect of Experimental Endotoxemia, Administration of DDAVP, and Exercise on vWF and Propeptide Levels in Healthy Subjects**

As previously shown,\(^9\) combined elevations of vWF and propeptide levels is a typical feature of experimental endotoxemia and DDAVP-induced vascular perturbation. To document this picture in more detail, eight subjects received endotoxin and nine received DDAVP. Subsequently, vWF and propeptide levels were measured at different time points after injection. In all subjects studied, administration of low-dose endotoxin led to a distinct increase of both vWF and propeptide levels after a lag phase of 1 to 2 hours (Fig 1A). The rise of vWF and propeptide concentration was followed by a decline of propeptide levels, whereas the vWF concentration remained elevated for at least 20 hours. The half-life of vWF and propeptide after administration of endotoxin, calculated from the disappearance curves, was approximately 12 and 3 hours, respectively. This observation clearly documents that under conditions in which acute perturbation of the endothelium is induced, the clearance of circulating propeptide is much faster than that of vWF. Similarly, administration of DDAVP resulted in a prompt increase of both propeptide and vWF levels (Fig 1B). The propeptide level returned close to baseline values after about 6 hours, whereas at this time point the vWF level was still twice as high as the vWF concentration before injection of DDAVP. The estimated half-lives of propeptide and vWF differed about threefold to fourfold. To facilitate comparison with clinical data (see later), data of peak levels of vWF and propeptide and vWF and propeptide concentration measured at later time points after endothelial stimulation are summarized (Table 1). We also
tested the effect of physical exercise on the plasma concentrations of propeptide and vWF in healthy volunteers. Similar to DDAVP and endotoxin, exercise enhances vWF release twofold to threefold and the propeptide level fivefold to eightfold (Table 1).

**Propeptide and vWF Levels in Patients With Chronic and Acute Vascular Disease**

A total of 49 patients was studied with different signs of vascular pathology. Patients with diabetes mellitus suffered from chronic vascular dysfunction, whereas patients with TTP and sepsis were admitted to the hospital with acute symptoms of vascular pathophysiology. Eighteen patients, referred to the hospital for underlying disorders other than vascular disease, served as a control group in this study. In the latter study group, the mean vWF and propeptide level was 54.2 ± 6.0 and 7.1 ± 0.7 nmol/L, respectively. The mean level of mature vWF was significantly increased in all patient groups studied (P < .001, Table 2). These values were about twice as high as the mean plasma levels of patients without vascular pathology. In patients with diabetes mellitus, the mean propeptide level was normal (8.0 ± 0.4 nmol/L). However, there was a significant correlation between propeptide and vWF levels (r = .61, P < .01). There was no difference in glycosylated hemoglobin levels and vWF or propeptide levels between type 1 and type 2 diabetes.

![Graph A](image1.png)  
**Fig 1.** Effect of administration of endotoxin (A) or DDAVP (B) on vWF (∙) and propeptide levels (∙) in healthy individuals. Endotoxin (4 ng/kg body weight) or DDAVP (0.4 μg/kg body weight) were administered at time 0. The endotoxin study group consisted of 8 subjects, the DDAVP study group of 9 subjects. Data points represent the mean and bars the SEM.

![Graph B](image2.png)

**Table 1.** vWF and Propeptide Levels in Healthy Individuals After Stimulation of the Endothelium by Endotoxin, DDAVP, and Exercise

<table>
<thead>
<tr>
<th>Perturbant</th>
<th>vWF (mature) (nmol/L ± SEM)</th>
<th>Propeptide (nmol/L ± SEM)</th>
<th>No. of Subjects</th>
</tr>
</thead>
<tbody>
<tr>
<td>LPS</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0 h</td>
<td>29.4 ± 4.7</td>
<td>4.2 ± 0.9</td>
<td>8</td>
</tr>
<tr>
<td>3 h</td>
<td>144.3 ± 11.7§</td>
<td>55.2 ± 4.6§</td>
<td></td>
</tr>
<tr>
<td>24 h</td>
<td>88.7 ± 10.0§</td>
<td>6.0 ± 0.5 (NS)</td>
<td></td>
</tr>
<tr>
<td>DDAVP</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0 h</td>
<td>40.5 ± 7.0</td>
<td>6.1 ± 0.3</td>
<td>9</td>
</tr>
<tr>
<td>1 h</td>
<td>104.8 ± 13.8§</td>
<td>46.2 ± 4.7§</td>
<td></td>
</tr>
<tr>
<td>6 h</td>
<td>81.0 ± 16.0†</td>
<td>14.8 ± 3.1†</td>
<td></td>
</tr>
<tr>
<td>Exercise</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0 h</td>
<td>40.4 ± 3.9</td>
<td>7.1 ± 0.8</td>
<td>5</td>
</tr>
<tr>
<td>5 min*</td>
<td>112.6 ± 17.9t</td>
<td>40.7 ± 11.7t</td>
<td></td>
</tr>
</tbody>
</table>

vWF and propeptide levels of the individuals of the respective study groups were compared with the baseline values before stimulation. Abbreviations: NS, not significant.

*Propeptide and vWF levels were not determined at later time points.

†P < .05.
§P < .01.
¶P < .001.

**Table 2.** vWF and Propeptide Levels in Patients With Vascular Disease

<table>
<thead>
<tr>
<th>Patient</th>
<th>vWF (mature) (nmol/L ± SEM)</th>
<th>Propeptide (nmol/L ± SEM)</th>
<th>No. of Patients</th>
</tr>
</thead>
<tbody>
<tr>
<td>Chronic diabetes mellitus</td>
<td>94.5 ± 6.4§</td>
<td>8.0 ± 0.4 (NS)</td>
<td>22</td>
</tr>
<tr>
<td>Acute TTP</td>
<td>112.7 ± 16.6†</td>
<td>17.3 ± 3.3†</td>
<td>13</td>
</tr>
<tr>
<td>Sepsis</td>
<td>127.8 ± 14.7†</td>
<td>17.8 ± 2.3†</td>
<td>14</td>
</tr>
<tr>
<td>Controls*</td>
<td>54.2 ± 6.0</td>
<td>7.1 ± 0.7</td>
<td>18</td>
</tr>
</tbody>
</table>

Abbreviation: NS, not significant.

*Patients without vascular disease admitted to the hospital. vWF and propeptide levels of patients with vascular pathology were compared with this control group.

†P < .01.
‡P < .001.
The data, on an individual basis, are shown in Fig 2. In the majority of these patients, both vWF and propeptide were elevated. In patients with TTP, a significant correlation between the LDH and propeptide levels was observed \((r = .82, P < .001)\). The correlation between LDH and vWF levels was not significant \((r = .51, P = .09)\). In patients with sepsis, a significant correlation between FnDPs and propeptide was found \((r = .58, P < .03)\). The correlation between vWF levels and FnDPs was not significant \((r = .47, P = .07)\).

**Propeptide and vWF Levels in Patients With Thrombocytopenia and Thrombocythemia**

As both vWF and its propeptide may not only originate from endothelial cells but also from platelets, vWF and propeptide levels were measured in plasma from healthy individuals and patients with low or high platelet counts. Figure 3 shows the relationship between propeptide concentration and platelet count in patients with bone-marrow aplasia, essential thrombocythemia and healthy controls. These parameters did not correlate \((r = .2, \text{difference not significant [NS]})\). Similarly, vWF levels did not correlate with platelet number (not shown). In contrast, there was a significant relationship \((r = .7, P < .001)\) between platelet number and the concentration of plasma sP-selectin (Fig 3), a specific marker of platelet activation.\(^{16,17}\) Also in patients with TTP, sepsis, or diabetes, there was no correlation between platelet count and propeptide or vWF.
concentration (not shown). These observations suggest that in these individuals both propeptide and mature vWF originate from the endothelium, rather than from platelets.

**DISCUSSION**

The primary purpose of this study was to determine the potential value of measurement of plasma concentrations of both vWF and its propeptide as a means to discriminate between acute and chronic vascular disease. This concept is illustrated by a number of control experiments in healthy subjects in which perturbation of the endothelium was provoked by administration of endotoxin or DDAVP, both agents known for their ability to elicit increases of plasma vWF and propeptide levels. DDAVP induces immediate release of vWF and propeptide, whereas lipopolysaccharide (LPS)-induced secretion is preceded by the release of one or more second messengers, which most likely mediate vWF and propeptide secretion through the regulated pathway.\(^8,18,19\) In all subjects studied, the concomitant elevation of vWF and propeptide is a consequence of endothelial cell activation and not due to a release from platelets.

Despite this monophasic response, we found that with most of the control agents there was still a delay in the elevation of propeptide. However, in some cases the vWF and propeptide levels not only would reflect increased secretion, but also plasma clearance. On the other hand, in some TTP patients propeptide and vWF levels reached peak levels (\(\approx 50\) and \(150\) nmol/L, respectively) that were also observed briefly after exposure of healthy individuals to DDAVP or endotoxin. This suggests that the provoking event in these patients had occurred just before admission.

It should be noted that in this cross-sectional study patients were studied at single time points after admission to the hospital because of acute symptoms or an exacerbation of the disease. Prospective, serial studies should reveal whether indeed measurements of both propeptide and vWF levels have a predictive value in terms of disease activity and are useful in monitoring the degree of vascular involvement as well as the response to therapy. In TTP, there was a strong correlation between propeptide concentrations and LDH \((P < .001)\), a marker of hemolysis and severity of disease, whereas the correlation between vWF and LDH was not significant \((P = .09)\). Similarly, in patients with sepsis, propeptide levels and FnDPs, a marker of DIC, were correlated \((P < .03)\), whereas vWF levels and FnDPs were not \((P = .07)\). These data suggest that propeptide is more reliable than vWF as a marker of vascular disease activity, at least in acute vascular disorders. It is possible that the difference in correlation as revealed by ELISA is related to the rather complex quaternary structure of vWF compared with the structure of its propeptide. These differences in biochemical nature could cause differences in the precision of vWF measurements. It should also be noted that in patients with concomitant organ failure, such as nephropathy or liver disease, these dysfunctions could have affected the metabolism of vWF and its propeptide. This would also complicate the interpretation of data on the plasma levels of these proteins. Similarly, altered patterns of posttranslational modifications (in TTP or diabetic patients) or edema formation (in sepsis) could affect the steady-state level of propeptide and vWF.

It could not be excluded that, as suggested previously,\(^23\) in the patients studied platelet activation contributed to the elevated propeptide (and vWF) levels. Indeed, some septic patients and all patients with TTP suffered from severe thrombocytopenia, obviously due to platelet activation. However, there was no relationship between platelet number and propeptide levels (not shown). Also in patients with either high or low platelet counts due to underlying disorders other than TTP or sepsis propeptide levels did not correlate with platelet number (Fig 3). On the other hand, plasma levels of sP-selectin, a specific marker of platelet activation,\(^16,17\) clearly correlated with platelet number in these patients (Fig 3). Further, the amount of propeptide stored in the \(\alpha\)-granules of platelets is probably not sufficient to account for the increased propeptide concentrations.\(^9,10,24\) It seems more likely therefore that vascular endothelial cells are the major source of circulating propeptide and vWF in patients.
and healthy subjects examined in this study. Therefore, another interesting feature of our study is that the vWF propeptide is a genuine marker of acute endothelial cell activation. The question as to the mechanism that induces vWF and propeptide release and pathways through which these proteins are released by the endothelium under pathologic conditions is more difficult to answer. VWF and propeptide can be released through both the regulated and constitutive pathway. However, the storage capacity of endothelial cells is limited, and it seems likely that, in particular cases where the endothelium is exposed to repeated challenges, the vWF/propeptide pool is exhausted. In addition, replenishment of these stores requires about 1 to 2 days after a single exposure to endothelial cell agonists, at least in cultured endothelial cells. Further, only a fraction of newly synthesized pro-vWF is directed to the Weibel-Palade bodies. On the basis of these data, it is to be expected that only in cases of isolated episodes of endothelial cell challenge, such as may occur in TTP or an event preceding overt septicemia, released vWF and propeptide primarily originate from the Weibel-Palade bodies. Pertinent to this point is the observation that in some patients with sepsis or TTP, propeptide and vWF levels approximate the peak levels of propeptide and vWF observed under experimental endothelial cell perturbation (≈50 and 150 nmol/L, respectively; Fig 1). Subsequent and persistent activation of the endothelium and concomitant rises of plasma vWF and propeptide most likely reflect enhanced constitutive release. However, the latter condition would require upregulation of de novo pro-vWF synthesis. Indeed, endothelial cell–specific vWF synthesis can be subject to transcriptional activity. It seems reasonable to assume, therefore, that elevated propeptide and vWF levels in patients with persistent endothelial cell injury not only reflect regulated secretion but also increased transcriptional activity and subsequent enhanced constitutive release. As unprocessed pro-vWF is primarily released through the constitutive pathway, measurement of pro-vWF could provide a means to discriminate between constitutive and regulated release.

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