Anaplastic large cell lymphomas (ALCL) are frequently associated with the t(2;5)(p23;q35). This translocation fuses the nucleophosmin (NPM) gene at q35, which encodes a nuclear protein involved in shuttling ribonucleoproteins from the cytoplasm to the nucleus, to the anaplastic lymphoma kinase (ALK) gene at 2p23, encoding a tyrosine kinase receptor.

In this report, we describe a typical case of ALCL whose malignant cells exhibited a novel (1;2)(q25;p23) translocation. These cells expressed ALK protein, but, in contrast to t(2;5)-positive ALCL (which show cytoplasmic, nuclear, and nucleolar staining), labeling was restricted to the malignant cell cytoplasm. Using a polymerase chain reaction (PCR)-based technique to walk on chromosome 2 from the known ALK gene across the breakpoint, we showed that the gene involved at 1q25 is TPM3, encoding a nonmuscular tropomyosin.

We subsequently identified, using reverse transcription-PCR analysis of cases showing similar ALK cytoplasm-restricted staining, fusion of the ALK and TPM3 genes in 2 other cases of ALCL. The TPM3 gene has been previously found in papillary thyroid carcinomas as a fusion partner with the TRK kinase gene. We showed that TPM3 is constitutively expressed in lymphoid cell lines, suggesting that, in these t(1;2)-bearing ALCL cases, the TPM3 gene contributes an active promoter for ALK expression. Activation of the ALK catalytic domain probably results from homodimerization of the hybrid protein TPM3-ALK, through the TPM3 protein-protein interaction domain. The present cases of ALCL associated with a novel t(1;2)(q25;p23) demonstrate that at least one fusion partner other than NPM can activate the intracytoplasmic domain of the ALK kinase.

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Immunostaining

Polyclonal anti-ALK (antibody p80)\textsuperscript{21} and monoclonal anti-ALK (antibody ALK1)\textsuperscript{5,20} were used, as previously described,\textsuperscript{5,20} diluted 1/100 and as undiluted culture supernatant, respectively. All cases were also immunostained with a panel of monoclonal antibodies detecting B and T antigens and for CD30 (Dako-Ber-H2, Copenhagen, Denmark), for EMA (Dako-EMA/E20), and with antibody BNH9,\textsuperscript{30} which is known to react with ALCL.

Cytogenetics

Karyotyping was performed on cells from the diagnostic lymph node from patient GEL after 24 hours of culture. Reverse-Heating-Giemsa (RHG) banding was performed using hot phosphate buffer, and chromosomal abnormalities were described according to the ISCN 95 nomenclature.\textsuperscript{30a}

DNA Extraction and Digestion

DNA from case GEL was extracted from 10 deparaffinized ModA-MeX-processed sections (10-µm thick) by proteinase K digestion, phenol-chloroform extraction, and ethanol precipitation.\textsuperscript{31} A small quantity (70 µg) of genomic DNA could be extracted from this material, but this DNA was of poor quality, with a maximum size of 6.5 kb when analyzed by agarose gel electrophoresis, precluding the construction and the screening of any genomic library for cloning the breakpoint.

DNA from the CNE2 tumor and the JAR cell line was extracted after solubilization of cell pellets in 5 mol/L guanidium isothiocyanate (GITC), 50 mmol/L Tris, pH 8, and 1% mercaptoethanol by gentle agitation at room temperature. The lysate was then dialysed three times using 100 vol of 10 mmol/L Tris, pH 8, 1 mmol/L EDTA, and 10 mmol/L NaCl (buffer A) before being treated at 37°C successively with 100 µg/mL RNase (Sigma-Aldrich Chimie, L’Isle d’Abeau Chesnes, France) for 2 hours and with 100 µg/mL proteinase K (Merck, Chelles, France) for 2 to 4 hours. The solution was then extracted several times with equal volumes of phenol and chloroform-isooamyl alcohol (24:1) and again dialysed against buffer A.

Aliquots of DNA (5 µg) from GEL, CNE 2, and JAR were digested with Dra I, EcoRV, Pvu II, Sca I, and Ssp I enzymes (New England Biolabs, Montigny-Le-Bretonneux, France), following the manufacturer’s recommendations.

PCR-Based Technique for Walking on Genomic DNA

This technique, described by Siebert et al.,\textsuperscript{22} involves a PCR between a gene-specific primer and a primer annealing to an adaptor sequence previously ligated to the ends of the genomic DNA fragments. The adaptor is designed in such a way that its specific primer cannot anneal to the native adaptor, because the target sequence is only created by the elongation of the gene-specific primer after the first PCR cycle.\textsuperscript{22} Non-specific amplifications between adaptors are thus avoided and specific amplification occurs despite the use of only a single gene-specific primer (Fig 1). To increase sensitivity and specificity, the first amplification is followed by a second nested amplification.

Adaptor ligation. The blunt ends of DNA fragments generated by digestion with Dra I, EcoRV, Pvu II, Sca I, and Ssp I enzymes were ligated to an excess of adaptors, using T4 DNA ligase (New England Biolabs), following the manufacturer’s recommendations. The adaptor was made of two complementary oligonucleotides, one long and one short (Fig 1). Once the two complementary oligonucleotides annealed, the adaptor presented with one blunt end that was able to ligate to any blunt end fragment of genomic DNA. The 3’ extremity of the short oligonucleotide was blocked by an amine group, preventing its elongation by DNA polymerase (Fig 1).

DNA amplification. Long-range PCR was performed with the Expand Long Template PCR system (Boehringer Mannheim, France).
SA, Meylan, France) using a unique enzyme mix containing thermostable Taq and Pwo DNA polymerases. This was performed using GEL DNA fragments for the three first steps and JAR DNA fragments for the fourth (because of the limited amount of DNA from case GEL; Fig 2). Primary PCR reactions were conducted in 50 µL volumes containing 0.5 µg of ligated DNA, 0.3 µmol/L adaptor primer AP122 and ALK gene-specific primer, 1.75 mmol/L MgCl₂, 0.35 mmol/L dNTP, and 6.125 U of DNA polymerase. An initial denaturation step at 94°C for 2 minutes was followed by 30 cycles of denaturation at 94°C for 30 seconds, annealing-extension at 68°C for 5 minutes, and a final extension time of 7 minutes at 65°C. The second (nested) PCR reaction was performed under the same conditions on 1 µL of the primary PCR, using adaptor primer AP222 and a nested ALK gene-specific primer. The adaptor primers AP1 and AP2 and the different ALK gene-specific primers are listed in Table 1.

Identification of the ALK Gene Breakpoint

To determine whether a given PCR fragment contained the breakpoint of interest, the fragment was purified, labeled, and used as a probe in Southern blot experiments on an unrelated DNA (CNE2) digested with the same panel of restriction enzymes: Dra I, EcoRV, Pvu II, Sca I, and Ssp I. We considered that a PCR fragment generated from the GEL DNA (following digestion with a single specific enzyme) that annealed with two different fragments of the unrelated CNE2 DNA digested with the same enzyme probably contained the ALK gene breakpoint.

After digestion of the CNE2 DNA by the same restriction enzymes (Dra I, EcoRV, Pvu II, Sca I, and Ssp I), 5 µg of each digested DNA was loaded on a 0.7% agarose gel, electrophoresed for 16 hours at 1.5 V/cm in TBE, and transferred onto nylon membrane (Genscreen Plus; Dupont, Boston, MA) using 0.4 mol/L NaOH as transfer solution. The membrane was then rinsed, dried, and prehybridized in 10% dextran sulfate, 1 mol/L NaCl, and 1% sodium dodecyl sulfate (SDS) for 30 minutes at 65°C. Long-range PCR fragments were purified after agarose gel electrophoresis using the GenClean III Kit (Bio 101, Ozyme, Saint Quentin Yvelines, France) and used as probes for the Southern blot experiments. Fragments (~25 ng) were labeled with 25 µCi of α-32P-dCTP using the NEBlot kit (New England Biolabs). After the addition of the denatured labeled probe, filters were hybridized overnight at 65°C, washed according to the manufacturer’s recommendations, and exposed to Kodak X-Omat AR films (Eastman Kodak, Rochester, NY) using intensifying screens at ~80°C.

Direct Cycle Sequencing of PCR Products

Long-range PCR fragments and RT-PCR fragments were sequenced using an original protocol with DeepVent(exo²) DNA polymerase (New England Biolabs) and α-35S-dATP. For each PCR fragment, 5′-3′.

**Fig 2.** Long-range amplification on digested and nondigested DNA. (A) The first three steps were performed using a PCR-based technique after adaptor ligation to digested DNA, with an adaptor primer and an ALK gene-specific primer, to determine the chromosome 1 gene involved in the (1;2) translocation. By walking on the chromosome from the ALK gene across the breakpoint to this unknown gene, we demonstrated that this chromosomal rearrangement fused the ALK gene on chromosome 2p23 to the TPM3 gene on chromosome 1q25. The 1.6-kb Ssp I fragment was only partially sequenced to yield the first 170 bp of this fragment at the 5′ end, allowing preparation of primers for the next step. This 170-bp sequence showed a complete identity to the sequence of intron 7 of the TPM3 gene. (B) The next three PCR steps were performed on undigested DNA and confirmed that all PCR fragments previously generated were colinear. The longer 2.2-kb Ssp I fragment was entirely sequenced and showed 99.9% identity with a 1,146-bp sequence in part of intron 7 of the TPM3 gene deposited in the GenBank database (accession no. X79910).
sequences (located close to the adaptor) were first determined to prepare
primers for the next PCR step.

Total RNA Extraction
Total RNA was extracted from frozen sections of normal thyroid and
lymph nodes involved by ALCL (cases no. 1, 2, and 3) and from the
Jurkat, CEM, HSB-2, and SU-DHL-1 cryopreserved cell pellets (10 × 10⁶)
using the RNeasy Midi Kit (Qiagen, Courtaboeuf, France).

Detection of Tropomyosin (TPM3) Transcripts by RT-PCR
Two TPM primers pairs, C-TPM1/C-TPM2 (positions 149-169/451-
431) and C-TPM3/C-TPM4 (positions 497-515/809-788; see Table 1),
were chosen according to the cDNA sequence of TPM3 published by
MacLeod et al., yielding RT-PCR products of 303 and 313 bp,
respectively. Synthesis of the first cDNA strand and PCR amplification
were performed in a single tube with the RT-PCR Access Kit (Promega
France, Charbonnières, France), following the manufacturer’s recom-
mendations. The first step consisted of a gene-specific reverse transcrip-
tion at 48°C for 45 minutes, followed by 30 cycles comprising a
denaturation step at 94°C for 45 seconds, an annealing step at 58°C for
45 seconds, and an elongation step at 72°C for 35 seconds.

Detection of Hybrid TPM3-ALK Transcripts by RT-PCR
This study was performed in 3 ALCL cases that were negative for
NPM-ALK transcripts and that showed ALK staining restricted to the
cytoplasm, resembling that observed in the case GEL. The first round of
RT-PCR was performed as described above, using C-TPM1 (position
149-169) and ALK-1 (position 4211-4187 on the cDNA ALK sequence)
primers. The second round was performed on a 1-µL aliquot from the
first amplification, using nested primers C-TPM3 (position 497-515)
and ALK-2 (position 4171-4148), yielding a RT-PCR product of 307 bp
(see Table 1 for primers sequence).

RESULTS

Immunomorphologic Features
The tumor from patient GEL and the 3 other cases of ALCL
investigated subsequently all contained large neoplastic cells
with horseshoe or kidney-shaped eccentric nuclei (Fig 3). Their
phenotype was also typical of systemic (2;5)-positive ALCL in

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**Table 1. Sequences of the Adaptors and the Different Primers Used**

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<thead>
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<th>Adaptors</th>
<th>Sequences</th>
</tr>
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<td>AD-L-1:</td>
<td>5’ CTA ATA CGA CTC ACT ATA GGG CTC GAG CGG CCG CCG GAG GT 3’</td>
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<tr>
<td>AD-S-1-NH2:</td>
<td>5’ ACC TGC CC-NH2 3’</td>
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<th>Adaptor primers</th>
<th>Sequences</th>
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</thead>
<tbody>
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<td>5’ CTA ATA CGA CTC ACT ATA GGG CTC GAG CGG CCG CCG CCG GAG GT 3’</td>
</tr>
<tr>
<td>AP-2:</td>
<td>5’ ATA GGG CTC GAG CGG CCG CCC GGG CAG GT 3’</td>
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<th>ALK and TPM3 gene-specific primers used on genomic DNA</th>
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<tr>
<td>ALK-2:</td>
<td>5’ GTC GAG GTG CCG AGC TTG CTC AGC 3’</td>
</tr>
<tr>
<td>ALK-5:</td>
<td>5’ GAA ATG GGG AGG GAG CCA GGG AAG G 3’</td>
</tr>
<tr>
<td>ALK-6:</td>
<td>5’ GGC TGG GTG AAC CAG CAG ACT GTG TT 3’</td>
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<tr>
<td>ALK-10:</td>
<td>5’ GTA GAT TCT GTG TGT AAA GCC CAG CCC C’</td>
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<tr>
<td>ALK-11:</td>
<td>5’ TTT AAA TGT GAA ATT GCC GAG C 3’</td>
</tr>
<tr>
<td>ALK-12:</td>
<td>5’ GCC AAC TTC TGC CCA GGA TCT GAA TA 3’</td>
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</tbody>
</table>

| CHRO-1-3: | 5’ CCA CTA GAA AGC ATC CA A CC CCC GGA AC 3’ |
| CHRO-1-4: | 5’ TCA ACC GTC TTC TTT CCC TCC AAT CG 3’ |
| CHRO-1-4R: | 5’ CGA TTG GAG GGA AAG AAG AC G GTT GA 3’ |

| TPM 2: | 5’ GCT TCT CGA GGT CAA GGG ATT CTC CTG T 3’ |

<table>
<thead>
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<th>Tropomyosin-specific primers used for RT-PCR to detect 7PM3 and TPM3-ALK transcripts</th>
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</thead>
<tbody>
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<td>CTPM-1:</td>
<td>5’ CGA GAA GTT GAG GGA AAG AGG 3’</td>
</tr>
<tr>
<td>CTPM-2:</td>
<td>5’ CAA CTT AGC AGC AAC CTC TCC TCC 3’</td>
</tr>
<tr>
<td>CTPM-3:</td>
<td>5’ CTG GCA GAG TCC GTG TGC G 3’</td>
</tr>
<tr>
<td>CTPM-4:</td>
<td>5’ ACT GGG CGT TCT ACA TCT CAT T 3’</td>
</tr>
</tbody>
</table>

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![Fig 3. Immunomorphologic features of the ALCL (GEL) carrying the (1;2) translocation. The morphology in a PAS-stained section is typical of the common-type of ALCL, and a hallmark cell, with an eccentric horseshoe-shaped nuclei, is arrowed. Staining for CD30 shows typical cytoplasmic and membrane-associated labeling of malignant cells. Labeling for ALK protein (antibody ALK1) is confined to the cell cytoplasm.](image-url)
that they were of null (cases no. 1 and 2) or T phenotype (case no. 3) and coexpressed CD30 (Fig 3) and epithelial membrane antigen (EMA). Malignant cells were also positive with the antibody BNH.9. All cells stained for ALK protein as detected by p80 and ALK1 antibodies. However, in contrast to most ALCL, which show labeling of both the cytoplasm and the nucleus, 14,20 ALK staining in these cases was limited to the cytoplasm (Fig 3).

Cytogenetics

Cytogenetic analysis of the diagnostic biopsy from patient GEL showed only abnormal metaphases, with the karyotype 46,XY,t(1;2)(q25;p23),der(22)t(1;22)(q12;p11)t(1;2)(q25;p23)[20].

Cloning of the t(1;2) Breakpoint

Long-range PCR was performed on DNA from the biopsy of patient GEL and from the JAR cell line, after digestion with different restriction enzymes and ligation to adaptors, as described in Materials and Methods. Each PCR step was realized with a couple of nested gene-specific primers and the nested adaptor-specific primers, AP1 and AP2. A PCR walk on chromosome 2 was performed using ALK-1 and ALK-2 gene-specific primers, located in the exon adjacent to the intron where the classical t(2;5) breakpoint occurs (Fig 2).

This first step yielded two fragments of approximately 600 bp on the Sca I-digested GEL DNA. After sequencing, these fragments appeared to be closely related to and probably products of the two chromosomes 2. Nested ALK-5 and ALK-6 primers were chosen in a sequence common to both chromosomes at the 5’ extremity of these fragments and were used for a second PCR step with primers AP1 and AP2 (Fig 2). A new 800-bp fragment was obtained from Dra I-digested GEL DNA. After purification, this fragment was used in a Southern blot experiment to probe DNA from CNE2 digested with Dra I, EcoRV, Pvu II, Sca I, and Ssp I. The GEL Dra I fragment showed two bands of differing intensity in the Dra I-digested CNE2 DNA, suggesting that it was composed of two different sequences that were not colinear in CNE2 DNA and, thus, likely contained the t(1;2) breakpoint. Comparison of its sequence with the sequence of the ALK intron (own unpublished results) confirmed that the 5’ last 80 bp of this fragment were not of ALK origin and putatively of chromosome 1 origin. However, this 80-bp sequence was not found to be homologous to any known sequence present in the GenBank database.

We performed a third PCR step using ALK-10 and ALK-11 gene-specific primers, chosen from the ALK intronic region of the 800-bp Dra I fragment bearing the breakpoint (Fig 2). This amplification yielded a 750-bp fragment from Sca I-digested GEL DNA, but its sequence also showed no homology with sequences in the GenBank database.

Because no more digested and adaptor-ligated DNA from case GEL was available, we performed the next PCR step on DNA from the JAR cell line prepared under the same conditions using gene-specific primers (CHRO-1-3 and CHRO-1-4) chosen from the 5’ end of the previous fragment (Fig 2). This amplification yielded a large 1.6-kb fragment from the Ssp I-digested JAR DNA. The 170-bp sequence at the 5’ end of this fragment showed complete homology in the GenBank database with the intron 7 of the TPM3 (tropomyosin 3) gene. 25

Because our PCR protocol was based on an adaptor-ligated DNA strategy, which can artfactually ligate unrelated fragments, we performed control PCRs (summarized in Fig 2) on undigested DNA from patient GEL using primers defined during our walking through this region. The first control PCR, between ALK-11 and CHRO1-4R, yielded a 650-bp fragment. Because CHRO1-4R has the same sequence (but in an opposite orientation) as CHRO1-4, which allowed the amplification of the last 1.6-kb fragment from JAR DNA, this was considered as the first definitive evidence that ALK-11 and the TPM3 gene were colinear in the DNA from case GEL.

The second control comprised another amplification across the breakpoint, between the ALK-5 and χ1-2R primers, with the latter primer being chosen from the chromosome 1-specific 80 bp of the GEL Dra I fragment. This amplification yielded the expected 700-bp fragment.

Finally, we performed a longer amplification on undigested JAR DNA between primers χ1-2 and TPM-2 chosen from the 5’ end of the 1.6-kb Ssp I fragment. A 2.2-kb fragment was obtained and a 1,146-bp sequence from this fragment was 99.9% identical to part of intron 7 from the TPM3 gene 25 deposited in the GenBank database (Fig 2).

Because ALK-5 and χ1-2, on the one hand, and χ1-2 and TPM-2, on the other, were colinear in DNA from the case GEL, we concluded that ALK-5 and TPM-2 were colinear and that the (1;2)(q25;p23) translocation had juxtaposed the TPM3 gene on chromosome 1 to the ALK gene on chromosome 2 (the accession number of a 3,394-bp fragment of the TPM3-ALK genomic sequence submitted to GenBank is AF 112866). Five Alu sequences have been found in the TPM3 intron, but none in the partial sequence of the ALK intron, and the breakpoint is located at the beginning of an Alu sequence in the TPM3 intron (data not shown).

Detection of Hybrid TPM3-ALK Transcripts and Direct Cycle Sequencing of RT-PCR Products

The 3 cases of ALCL that showed a cytoplasm-restricted staining pattern for ALK (from which no cytogenetic data were available) were analyzed for the presence of TPM3-ALK transcripts. A 307-bp band was found in cases no. 1 and 3 (Fig 4A), and sequencing of nested RT-PCR products confirmed the presence of hybrid TPM3-ALK transcripts (Fig 4B).

Detection of Tropomyosin Transcripts

The tyrosine kinase domain of ALK is constitutively expressed in ALCL carrying the classical t(2;5), under control of the ubiquitously expressed NPM gene promoter. We therefore wished to know whether the TPM3 gene is also active in lymphoid cells. RT-PCR assays of RNA extracted from the Jurkat, CEM, HSB-2, and SU-DHL-1 T-cell lines (and also from thyroid tissue) demonstrated that the TPM3 gene was constitutionally active, 24 because bands of the expected size (303 and 313 bp) were detected with C-TPM1/C-TPM2 and C-TPM3/C-TPM4 primer pairs, respectively (data not shown).
from the SU-DHL-1 cell line was used as negative control (T
2 showing ALK-staining restricted to the cytoplasm. RNA obtained
show a band of 307 bp, indicating the presence of hybridTPM3-ALK
bromide staining. Two of these three cases of ALCL (cases no. 1 and 3)
electrophoresed on 2% agarose gel and visualized with ethidium
amplification using C-TPM3 and ALK-2 primers. PCR products were
involved C-TPM1 and ALK-1 primers, and the second was a nested
RT-PCR was performed using two rounds of PCR. The first round
involved C-TPM3-ALK FUSION BY THE t(1;2) IN ALCL 3093
ALK protein expression in the case GEL was restricted
to part of the
NPM
Fig 4. (A) RT-PCR analysis of TPM3-ALK RNA in 3 cases of ALCL
showing ALK-staining restricted to the cytoplasm. RNA obtained
from the SU-DHL-1 cell line was used as negative control (T-).
RT-PCR was performed using two rounds of PCR. The first round
involved C-TPM1 and ALK-1 primers, and the second was a nested
amplification using C-TPM3 and ALK-2 primers. PCR products were
electrophoresed on 2% agarose gel and visualized with ethidium
bromide staining. Two of these three cases of ALCL (cases no. 1 and 3)
show a band of 307 bp, indicating the presence of hybrid TPM3-ALK
RNA. M, size marker. (B) Nucleotide sequence of hybrid TPM3-ALK
transcripts detected by nested RT-PCR using C-TPM3 and ALK2
primers. Nucleotides are numbered from the deoxyadenosine residue
of the TPM3 initiator ATG codon.

DISCUSSION

Recurrent chromosomal translocations play an important role
in many human lymphoid tumors and are often responsible for
the deregulation of genes involved in the control of cell
proliferation. A number of recent studies have investigated the
(2;5)(p23;q35) translocation, an anomaly associated with ALK
positive ALCL.7 The t(2;5) fuses the nucleophosmin gene
(NPM) with the ALK receptor tyrosine kinase gene (anaplastic
lymphoma kinase) to produce a chimeric protein in which the
116 aminoterminal residues results in oncogenic activation of
NPM-ALK. 12-kb

The breakpoint in the
TPM3-NTRK1 oncogene occurs in the
12-kb
TPM3 intron located between exons 7 and 8, and this was
also the site of the TPM3 breakpoint in the (1;2) translocation
described in this report. Furthermore, the ALK intron
involved in this case is also the intron involved in the classical
(2;5) translocation.

It has been suggested that translocations can occur in
lymphoid tumors through recombinogenic elements such as χ
sequences45 or alternating purine-pyrimidine tracts, which form
negative supercoiled regions (Z-DNA) making DNA more
accessible to the enzymatic machinery.46 We have not found any
of these known recombinogenic elements in the regions se-
quenced around the breakpoint. Alu sequences can also be
implicated in recombinations and translocations, such as the
t(9;22), where both the Abl and bcr introns contain Alu
sequences in an opposite orientation.47 In the case under study
(GEL), sequence analysis demonstrated the presence of numer-
ous Alu sequences in the TPM3 intron, the breakpoint itself
being located at the beginning of an Alu sequence, but none was
found in the disrupted ALK gene intron. Thus, the lack of
recombinogenic elements, including Alu sequences near the
breakpoint in the ALK intron, suggests that novel mechanisms
are involved in the (1;2) translocation.
Whatever the mechanism of the (1;2) translocation, the TPM3 gene promoter appears to drive the expression of the ALK tyrosine kinase domain, because we have demonstrated that this promoter is active in lymphoid T-cell lines. In addition, tropomyosins are known to form dimeric α-coil-coil structures\textsuperscript{14,15} that could induce homodimerization of the chimeric TPM3-ALK protein, a mechanism also involved in the activation of NPM-ALK in the classical (2;5) translocation.\textsuperscript{13} However, further experiments are needed to assess this hypothesis.

The present cases of ALCL, associated with a novel TPM3-ALK hybrid gene, indicate that partners other than NPM can promote oncogenic expression of the ALK intracytoplasmic domain. Moreover, this study demonstrates the usefulness of anti-ALK antibodies for detecting cases containing possible new translocations in ALCL, because when staining is restricted to the cell cytoplasm and not present in the nucleus, a partner other than NPM is likely to be present.\textsuperscript{16,17,19}

**ACKNOWLEDGMENT**

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**REFERENCES**

Lennert K: The expression of the Hodgkin’s disease associated antigen Ki-1 in reactive and neoplastic lymphoid tissue: Evidence that Reed-Sternberg cells and histiocytic malignancies are derived from activated lymphoid cells. Blood 66:848, 1985


A New Fusion Gene TPM3-ALK in Anaplastic Large Cell Lymphoma Created by a (1;2)(q25;p23) Translocation

Laurence Lamant, Nicole Dastugue, Karen Pulford, Georges Delsol and Bernard Mariamé

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