CLINICAL OBSERVATIONS, INTERVENTIONS, AND THERAPEUTIC TRIALS

Recombinant Human Thrombopoietin in Combination With Granulocyte Colony-Stimulating Factor Enhances Mobilization of Peripheral Blood Progenitor Cells, Increases Peripheral Blood Platelet Concentration, and Accelerates Hematopoietic Recovery Following High-Dose Chemotherapy

By George Somlo, Irena Sniecinski, Anna ter Veer, Jeffery Longmate, Gaylord Knutson, Stanimir Vuk-Pavlovic, Ravi Bhatia, Warren Chow, Lucille Leong, Robert Morgan, Kim Margolin, J ames Raschko, Stephen Shibata, Merry Tetef, Yun Yen, Stephen Forman, Dennie J ones, Mark Ashby, Gwen Fyfe, Susan Helmann, and James H. Doroshow

Lineage-specific growth factors mobilize peripheral blood progenitor cells (PBPC) and accelerate hematopoietic recovery after high-dose chemotherapy. Recombinant human thrombopoietin (rhTPO) may further increase the progenitor-cell content and regenerating potential of PBPC products. We evaluated the safety and activity of rhTPO as a PBPC mobilizer in combination with granulocyte colony-stimulating factor (G-CSF) in 29 breast cancer patients treated with high-dose chemotherapy followed by rhTPO infusion. Initially, patients received escalating single doses of rhTPO intravenously (IV) at 0.6, 1.2, or 2.4 μg/kg, on day 1. Subsequent patients received rhTPO 0.6 or 0.3 μg/kg on days −3, −1, and 1, or 0.6 μg/kg on days −1 and 1. G-CSF, 5 μg/kg IV or subcutaneously (SC) twice daily, was started on day 3 and continued through aphereses. Twenty comparable, concurrently and identically treated patients (who were eligible and would have been treated on protocol but for the lack of study opening) mobilized with G-CSF alone served as comparisons. CD34+ cell yields were substantially higher with the first apheresis following rhTPO and G-CSF versus G-CSF alone: 4.1 × 10⁶/kg (range, 1.3 to 17.6) versus 0.8 × 10⁶/kg (range, 0.3 to 4.2), P = .0003. The targeted minimum yield of 3 × 10⁶ CD34+ cells/kg was procured following a single apheresis procedure in 61% of the rhTPO and G-CSF-mobilized group versus 10% of G-CSF-mobilized patients (P = .001). In rhTPO and G-CSF mobilized patients, granulocyte (day 8 v 9, P = .0001) and platelet recovery (day 9 v 10, P = .07) were accelerated, and fewer erythrocyte (3 v 4, P = .02) and platelet (4 v 5, P = .02) transfusions were needed compared with G-CSF-mobilized patients. Peripheral blood platelet counts, following rhTPO and G-CSF, were increased by greater than 100% and the platelet content of PBPC products by 60% to 110% on the first and second days of aphereses (P < .0001) with the greatest effect seen with repeated dosing of rhTPO at 0.6 μg/kg, rhTPO is safe and well tolerated as a mobilizing agent before PBPC collection. Mobilization with rhTPO and G-CSF, in comparison to a comparable, nonrandomized G-CSF-mobilized group of patients, decreases the number of apheresis procedures required, may accelerate hematopoietic recovery, and may reduce the number of transfusions required following high-dose chemotherapy for breast cancer.

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rhTPO ENHANCES PROGENITOR-CELL AND PLATELET MOBILIZATION

MATERIALS AND METHODS

Patients. All patients gave their written, voluntary informed consent. This study was approved by the Institutional Review Board of the City of Hope National Medical Center (Duarte, CA).

Between July 1996 and November 1997, 29 patients with responding stage IV or high-risk stage III/III primary breast carcinoma were accrued. Patients with a Karnofsky performance status greater than 80%, adequate organ function, left ventricular cardiac ejection fraction $\geq 55\%$, and a measured creatinine clearance $\geq 80$ mL/min were eligible. The use of aspirin, nonsteroidal anti-inflammatory agents, and anticoagulant drugs was not allowed (except to maintain line patency). Chemotherapy or radiation therapy within 3 weeks before mobilization, or the use of growth factors other than G-CSF, was prohibited.

Study design. Patients had a double-lumen Hickman catheter (Bard Access Systems, Salt Lake City, UT) inserted before mobilization of PBPC. Safety of rhTPO, at any given dose level, had to be documented in three patients before accrual began at the next dose level. Initially, groups of patients received a single intravenous (IV) injection of rhTPO on day 1, escalating the dose through 0.6, 1.2, or 2.4 µg/kg (arm A). Completion of accrual on arm A was required before accrual to the subsequent cohort. Subsequent patients received rhTPO on days $-3$, $-1$, and 1 at doses of 0.6 µg/kg or 0.3 µg/kg (arm B) or rhTPO 0.6 µg/kg on days $-1$ and 1 (arm C). G-CSF (Neupogen; Amgen, Thousand Oaks, CA) 5 µg/kg twice daily, subcutaneously (SC) or IV was started on day 3 and continued through aphereses. rhTPO (Genentech Inc, South San Francisco, CA), a full-length glycosylated molecule, was reconstituted in preservative-free normal saline before injection.

Aphereses were started on the fourth day of G-CSF administration (day 6) using a Spectra (Cobe Laboratories, Lakewood, CO), or Fenwall CS-3000 Plus cell separator (Baxter, Deerfield, IL); approximately 10 L of blood was processed during each collection. The mobilization schema is illustrated in Fig 1. A minimum target of $3 \times 10^6$ CD34$^+$ cells/kg (and $\geq 7 \times 10^5$ mononuclear cells/kg) was procured. A cohort of 20 concurrently treated patients were mobilized with G-CSF 5 µg/kg twice per day, SC or IV, daily. These patients were eligible and otherwise similar to those on-study and would have been treated on protocol but for the lack of study openings at the time of their evaluation. Starting on the fourth day of G-CSF administration, the duration and procedure of apheresis were identical to those of the rhTPO study group; $\geq 7 \times 10^5$ mononuclear cells/kg were collected. PBPC were cryopreserved by simple immersion in a $-130^\circ$C freezer.15

High-dose chemotherapy. Patients received both cisplatin 125 mg/m$^2$ and etoposide 30 mg/kg on days $-12$ and $-5$ followed by cyclophosphamide 100 mg/kg on day $-3$. PBPC were reinfused on a divided schedule: 25% of PBPC were infused on day $-2$ following high-dose chemotherapy and 75% were infused on day 0. All patients received G-CSF 5 µg/kg twice daily IV from day $-2$ through absolute granulocyte recovery to $\geq 1,500$ µL.$^{18}$ Blood products were filtered and irradiated at 2,500 cGy and were transfused to maintain a platelet count of greater than $20 \times 10^9$ /L and a hematocrit of $\geq 25$ or as clinically indicated.19

Patient monitoring. Patients were monitored for adverse reactions with each injection of rhTPO. Total WBC, differential, and platelet counts were measured on the days of rhTPO and G-CSF administration and during aphereses. Serum chemistries, coagulation parameters, and samples for anti-rhTPO antibodies were monitored during the mobilization, and following high-dose chemotherapy and PBPC reinfusion (transplant) phase. No more than two episodes of grade 3 toxicity were allowed at any dose level; any grade 4 toxicity related to the administration of rhTPO defined the maximum-tolerated dose of rhTPO. Development of neutralizing antibodies to rhTPO or a platelet count greater than 600,000/µL persisting beyond 24 hours were also considered dose-limiting toxicities.

Hematologic end points. The number of days needed to reach an absolute granulocyte count $\geq 0.5 \times 10^9$/L and platelet independence (defined as the day after the last platelet transfusion) were calculated from day 0 of the “transplant phase.” The number of erythrocyte and platelet transfusions required were tabulated. Single apheresis and pooled platelet products were each counted as one product.

Flow cytometry studies. PBPC apheresis or peripheral blood samples were stained with either phycoerythrin (PE)-conjugated anti-CD34 monoclonal antibody and dual stained with anti-CD45-fluorescein isothiocyanate (FITC) or, for assessment of the presence of megakaryocytic lineage samples, were stained with anti-CD34-PE/anti-CD41-FITC (Becton Dickinson, San Jose, CA). Samples of 300 µL containing approximately 1 million cells were incubated with antibody for 30 minutes at 4°C. After the addition of 2 mL erythrocyte lysing solution (Ortho-mune; Ortho Diagnostic Systems, Raritan, NJ), samples were incubated for 10 minutes at room temperature. Cells were then washed twice in phosphate-buffered saline (PBS) (Buffer Reagent (Coulter, Miami, FL) and resuspended in 500 µL PBS. Analysis was performed using a FACS Calibur or FACS Sort flow cytometer equipped with argon-ion laser, tuned at 488 nm (Becton Dickinson), and analyzed with CellQuest 3.1 software (Becton Dickinson). 100,000 cells were acquired for each analysis. CD34/CD45 analysis was performed as described earlier.16 For the CD34/CD41 analysis, the primary gate was set up using forward scatter versus side scatter. All CD34$^+$ cells were analyzed for side-scatter properties and expression of CD41; subsequently, CD34$^+$ cells within the low side-scatter population were further analyzed for CD41 expression.

Progenitor-cell assays. Peripheral blood was collected in heparin before each injection of rhTPO and on the days of apheresis. PBPC were diluted 1:5 with Iscove’s modified Dulbecco’s medium (IMDM) in heparin. Samples were shipped at room temperature overnight to the Stem Cell Laboratory (Mayo Clinic Cancer Center, Rochester, MN).

Mononuclear cells were isolated by density gradient centrifugation in Lymphocyte Separation Medium (LSM; Cappel, ICN Biomedicai, Aurora, OH; 1.077 g/mL) and diluted with IMDM to no more than 5 $\times 10^6$ cells/mL. Eight milliliters of cell suspension was centrifuged at 400g for 30 minutes at 22°C. Cells at the interface were collected and washed twice with 3 vol of IMDM.

Colonforming unit—granulocyte, macrophage (CFU-GM), colony-forming unit—granulocyte, erythrocyte, megakaryocyte, macrophage (CFU-GEMM) and burst-forming unit—erythroid (BFU-E) were assayed in a methylcellulose medium (MethoCult FH4434; StemCell Technologies, Vancouver, Canada). Cells were plated at a final cell density of 1 or 2 $\times 10^3$/mL (peripheral blood) and 1 or 3 $\times 10^5$/mL (PBPC). After 14 days at 37°C in 5% CO$_2$, the colonies were identified by morphology and scored. Colony-forming unit—megakaryocyte (CFU-MK) was assayed in an agarose medium (MegaCult; StemCell Technologies); cells were plated at a final density of 4 $\times 10^5$/mL (peripheral blood) or 1 $\times 10^6$/mL (PBPC). After 18 days at 37°C in 5% CO$_2$, the colonies were fixed with methanol-acetone and immunohistochemically...
detected by the anti-GPⅡb/Ⅲa antibody and alkaline phosphatase detection system supplied by the manufacturer.

**Statistical analysis.** The study was primarily designed to identify tolerable doses and schedules of rhTPO. Comparison of rhTPO doses and schedules, and comparison to patients not receiving rhTPO was planned to evaluate hematologic response in the context of a phase I study, and not to achieve any specified power. The G-CSF–only mobilized group consisted of comparable, concurrently treated patients who were eligible to enroll on study but did not for lack of an opening on the trial. Nonevaluable patients were excluded from analysis as described in the Results. Patients receiving rhTPO were combined in a single group for comparison to G-CSF–only patients using rank-sum tests; an F test was applied for overall comparison between these two groups after stratifying for prior exposure to radiation therapy. Comparisons among rhTPO regimens were evaluated by Kruskal-Wallis tests; 95% confidence intervals (CIs) for percent (fold) differences were based on $t$ statistics for log-percent difference. The effects of dose and schedule were evaluated by fitting multiple regression models, using logarithms of responses as described in the Results. Separate regressions were fit to the first and second days of cell collections to evaluate the relationship between platelet content for stem cell products and peripheral blood platelet counts. Correlations and coefficients of determination are provided as rough summaries. An F test was applied to verify the presence of significant correlation.

**RESULTS**

The effects of rhTPO and G-CSF on PBPC mobilization were assessable in 26 of 29 patients (two patients were removed from the study due to bacteremia associated with Hickman catheter infection; one patient who had experienced prolonged leukocytopenia after prior standard chemotherapy mobilized so poorly that she was taken off study). Twenty-three of the 26 patients were assessable both during the mobilization and the “transplant” phase: three patients were not evaluable during the transplant phase due to delay of their high-dose chemotherapy (progression, $n=1$; superior vena cava occlusion [likely due to a previous catheter infection], $n=1$; bacteremia after apheresis, $n=1$).

Characteristics of the 26 assessable rhTPO- and G-CSF-treated patients and 20 concurrently and identically treated G-CSF–only mobilized patients are listed in Table 1. Most characteristics, such as age, stage, prior chemotherapeutic exposure, and number of days between apheresis and PBPC reinfusion were similar except for a higher percentage of G-CSF–only–mobilized group (control group) having been exposed to prior radiation treatment.

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>TPO Study Group (n = 26)</th>
<th>Control Group (n = 20)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (yr)</td>
<td>Median: 44</td>
<td>Range: 33-60</td>
</tr>
<tr>
<td>No. of prior chemotherapy regimens</td>
<td>1</td>
<td>1-3</td>
</tr>
<tr>
<td>No. of prior chemotherapy cycles</td>
<td>6</td>
<td>3-23</td>
</tr>
<tr>
<td>Days from apheresis to reinfusion</td>
<td>30</td>
<td>29-45</td>
</tr>
</tbody>
</table>

Table 1. Patient Characteristics

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>No.</th>
<th>%</th>
<th>No.</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>Stage II/III disease</td>
<td>16</td>
<td>62</td>
<td>12</td>
<td>60</td>
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<tr>
<td>Stage IV</td>
<td>10</td>
<td>38</td>
<td>8</td>
<td>40</td>
</tr>
<tr>
<td>Prior radiation</td>
<td>4</td>
<td>15</td>
<td>6</td>
<td>30</td>
</tr>
</tbody>
</table>

Table 2. PBPC Yield and Transfusion Requirements

<table>
<thead>
<tr>
<th>Variable</th>
<th>Prior Radiation</th>
<th>TPO Group</th>
<th>Control Group</th>
<th>P Value*</th>
</tr>
</thead>
<tbody>
<tr>
<td>No. of aphereses to procure</td>
<td>No</td>
<td>1</td>
<td>1-3</td>
<td>22</td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>1</td>
<td>1-2</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>All patients</td>
<td>1</td>
<td>1-3</td>
<td>26</td>
</tr>
<tr>
<td>CD34+ x 10^6/kg/first apheresis</td>
<td>No</td>
<td>3.6</td>
<td>1.3-18</td>
<td>22</td>
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<td></td>
<td>Yes</td>
<td>5.1</td>
<td>2.4-15</td>
<td>4</td>
</tr>
<tr>
<td></td>
<td>All patients</td>
<td>4.1</td>
<td>1.3-17.6</td>
<td>26</td>
</tr>
<tr>
<td>No. of RBC transfusions†</td>
<td>No</td>
<td>3</td>
<td>2-7</td>
<td>21</td>
</tr>
<tr>
<td></td>
<td>Yes</td>
<td>3.5</td>
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<td></td>
<td>All patients</td>
<td>3</td>
<td>2-7</td>
<td>23</td>
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<td>No. of platelet transfusions†</td>
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<tr>
<td></td>
<td>Yes</td>
<td>3</td>
<td>2-4</td>
<td>2</td>
</tr>
<tr>
<td></td>
<td>All patients</td>
<td>4</td>
<td>0-10</td>
<td>23</td>
</tr>
</tbody>
</table>

*Comparisons between the rhTPO v control groups were performed using the Wilcoxon rank-sum test. F test, adjusting for prior radiation, was used to combine strata in the overall analysis.
†Transfusion requirements were evaluated in 23 assessable patients.
In 16 of 26 patients in the rhTPO-mobilized group, the targeted yield of $3 \times 10^6$ CD34+ cells/kg was reached with the first apheresis procedure versus in only 2 of 20 G-CSF mobilized patients ($\chi^2, P < .001$), apheresis from three additional patients in the G-CSF–only group resulted in $3 \times 10^6$ CD34+ cells/kg beyond day 6 (data not shown). In the G-CSF–mobilized group, the highest numbers of CD34+ cells were procured during the first apheresis (day 4 of G-CSF) in 8 and during the second procedure (day 5 of G-CSF) in 5 patients; CD34+ cell yield from 7 patients peaked evenly between days 6 and 8. The median CD34+ yield was 4.1 $\times$ 10^6/kg (range, 1.3 to 17.6 $\times$ 10^6/kg) in products collected on the first scheduled day of apheresis from patients in the rhTPO-mobilized group versus 0.8 $\times$ 10^6/kg (range, 0.3 to 4.2 $\times$ 10^6/kg) in the concurrently treated and G-CSF–mobilized control group. A 3.5-fold increase in CD34+ cell content per apheresis (95% CI, 2.7-to 6.6-fold increase in geometric mean) following administration of rhTPO was observed: the median number of (range) CD34+ cells per kilogram per apheresis was 3.8 $\times$ 10^6 (1.1 to 12.7) in the rhTPO-mobilized group versus 1.1 $\times$ 10^6 (0.5 to 4.7) in the control group ($P = .014$). Higher yields of mononuclear cells were procured following administration of rhTPO: 3.7 $\times$ 10^8 mononuclear cells/kg/apheresis (range, 0.5 to 6.5) versus 2.8 $\times$ 10^8 mononuclear cells/kg/apheresis (range, 1.6 to 4.5). We observed an approximate 30% increase in the mononuclear-cell yield per apheresis (95% CI, 16% to 59% increase in geometric mean), with a statistically significant difference seen even after stratification for the possible effects of prior radiation treatment ($P = .04$ data not shown). Both RBC (3 $v$ 4, $P = .02$) and platelet (4 $v$ 5, $P = .02$) transfusion requirements were decreased in the rhTPO-mobilized group compared with G-CSF–mobilized controls. Hematopoietic recovery was also faster in the rhTPO group: granulocyte and platelet recoveries were accelerated, as illustrated in Fig 2A and B (8 $v$ 9 days [$P = .0001$] and 9 $v$ 10 days [$P = .07$], respectively). Progenitor-cell yield and hematopoietic recovery were improved following mobilization with the rhTPO/G-CSF combination versus G-CSF alone, in patients with or without prior exposure to radiation therapy.

Figure 3 displays the mean CD34+ yield for cohorts of patients receiving different doses and/or schedules of rhTPO on arms A, B, and C. The Kruskal-Wallis test suggested a dose/schedule dependent effect ($P = .03$); the highest yield (15.1 $\times$ 10^6/kg $v$ 7.5 $\times$ 10^6/kg) was observed when three doses of rhTPO 0.6 $\mu$g/kg were administered every other day ($P = .004$, F test).

Table 3 compares the effects of rhTPO/G-CSF mobilization versus G-CSF alone on peripheral blood platelet counts on the days of aphereses and depicts the platelet content of each apheresis product. We observed an approximately 100% and 130% increase in peripheral blood platelet counts on the first and second days of apheresis and 60% and 110% increase in platelet yields in the apheresis products after mobilization with rhTPO.
apheresis products was directly related to peripheral blood with schedule- and dose-dependent effects. Platelet content of mobilized with three doses of rhTPO at 0.6 µg/kg, consistent schedules of rhTPO. Both peripheral blood platelet counts (P = .01) and total platelet content (P = .02) were highest in patients mobilized with three doses of rhTPO at 0.6 µg/kg, consistent with schedule- and dose-dependent effects. Platelet content of apheresis products was directly related to peripheral blood platelet concentration, as shown in Fig 4.

Table 5 describes the percentage of CD41+ cells within the CD34+ cell population: increased percentages of CD41+ cells were seen following administration of either single or repeated injections at the higher (1.2 to 2.4 µg/kg) rhTPO doses. We observed the highest percentage of CD41+ cells within the CD34+ population following sequential administration of rhTPO at 0.6 µg/kg (P = not significant [NS]).

We analyzed the effect of different rhTPO doses and schedules on CFU-GM, CFU-GEMM, CFU-MK, and BFU-E formation from peripheral blood samples. We found a significant increase over baseline in the number of CFU-GM (5.15-fold mean increase; range, 0.9 to 31; 95% CI, 3.6 to 7.5), CFU-GEMM (2.5-fold mean increase; range, 0.4 to 15; 95% CI, 1.7 to 3.6), and CFU-MK (2.5-fold increase; range, 0.2 to 37; 95% CI, 1.4 to 4.4) and BFU-E (2.5-fold mean increase; range, 0.4 to 15; 95% CI, 0.8 to 3.7) following administration of rhTPO and G-CSF; for all CFUs, the largest number of colonies was observed on the day of apheresis (day 5 and 6 after the last dose of rhTPO). rhTPO dose and schedule had no significant effect on the rate of CFU formation from baseline to postmobilization.

Table 5. Platelet Content of PBPC Products and Peripheral Blood Platelet Concentration on the First and Second Days of Apheresis by Mobilizing Regimen

<table>
<thead>
<tr>
<th>Variable</th>
<th>Median (Range)</th>
<th>Median (Range)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>rhTPO 0.6-2.4 µg/kg and G-CSF 5 µg/kg, Twice Daily</td>
<td>218-995</td>
<td>130-369</td>
<td>&lt;.0001</td>
</tr>
<tr>
<td>G-CSF 5 µg/kg, Twice Daily</td>
<td>142-890</td>
<td>77-236</td>
<td>&lt;.0001</td>
</tr>
</tbody>
</table>

Abbreviations: PB, peripheral blood; d1, day 1; d2, day 2; PLTs, platelets.

There was no statistically significant trend favoring any particular rhTPO dosing schedule when we assessed the amount of CFU-MK, BFU-E, CFU-GEMM, and CFU-GM in the stem-cell products, although the highest yields were seen following sequential administration of rhTPO 0.6 µg/kg, as illustrated in Table 6.

**Toxicities.** We observed no allergic reactions associated with the administration of rhTPO. The superior vena cava occlusion in one patient assigned to receive a single injection of rhTPO 0.6 µg/kg preceded the administration of rhTPO and was most likely due to a previous catheter infection; her platelet counts remained within the normal range (234 × 10^9/L and 151 × 10^9/L) on the days of apheresis. She underwent high-dose therapy 6 months later without any thromboembolic complications.

We observed platelet count elevations greater than 600 × 10^9/L lasting for 24 hours in two of six patients (peaks of 995 × 10^9/L and 964 × 10^9/L) treated with rhTPO 0.6 µg/kg on days –3, –1, and 1 (arm B). Both peaks occurred on the first day of apheresis, and platelet counts decreased by the second day of apheresis. To assess further the possibility of excessive thrombocytosis following repeated administration, an additional rhTPO dose level was added, and six patients received three doses of rhTPO 0.3 µg/kg every other day. Platelet counts in these patients never reached 600 × 10^9/L.

Serially tested serum prior and subsequent to the administration of rhTPO and post–high-dose chemotherapy remained negative for the presence of any neutralizing antibodies in all subjects.

**DISCUSSION**

C-Mpl ligand, by expanding, and possibly by protecting, the progenitor-cell pools of megakaryocytic, myelomonocytic, and erythroid lineage, can prevent or ameliorate hematopoietic toxicity inflicted by cytotoxic therapy.15-17 Thrombopoietin may also accelerate hematopoietic recovery following chemotherapy. When administered after carboplatin and cyclophosphamide a truncated, pegylated form of the C-Mpl ligand, PEG-RhHuMGDF (MGDF) enhanced platelet recovery in a dose-dependent fashion when given together with G-CSF. CD34+ cell subsets, in patients treated at higher doses of MGDF (0.3 to 5 µg/kg), were increased and platelet counts increased to greater than 1,000,000/µL, when administered after carboplatin and paclitaxel, resulting in less pronounced and shorter median platelet nadirs compared with controls.22 Similarly, single injections of rhTPO can attenuate thrombocytopenia when administered before
high-dose carboplatin; rhTPO 1.2 µg/kg given after high-dose carboplatin may prevent the need for platelet transfusions. Both the full-length and pegylated, truncated molecules are capable of expanding the progenitor pool. rhTPO increased the number of multilineage progenitors in the bone marrow while mobilizing CD34^+ cells into the peripheral blood in cancer patients. MGDF in combination with a variety of growth factors has been shown to expand the number of megakaryocyte progenitors ex vivo.

We have found a substantial increase in the CD34^+ progenitor-cell content of apheresis products when rhTPO, in combination with G-CSF, was administered several days before apheresis. While one can argue that prolonged administration of 10 µg/kg G-CSF alone for up to 9 days (rather than 3 days) before initiating apheresis may increase the CD34^+ yield, there are data to the contrary. In a study conducted in normal volunteers, administration of 10 µg/kg G-CSF resulted in peak peripheral blood CD34^+ cell counts on day 4 in two of five healthy subjects, while a broader distribution of multiple peak CD34^+ cell counts (between days 3 and 7) was observed in three other volunteers. Since the peripheral blood CD34^+ cell count predicts for progenitor-cell yield in the apheresis product, we applied such short G-CSF course before apheresis in our 20 control patients. Indeed, the highest CD34^+ yields were procured during the first and second apheresis in 65% of patients; yields in the other 35% of patients were highest between days 6 and 8. No patient in the control group yielded ≥3 × 10^6 CD34^+ cells/kg beyond the sixth day of administering G-CSF, making it unlikely that prolonged administration of G-CSF would lead to a decrease in the number of aphereses needed to reach our target.

Patients in the rhTPO– and G-CSF–mobilized group and the control group were comparable in their exposure to the number of prior chemotherapy regimens and cycles; there were, however, half as many patients in the rhTPO-treated cohort with a history of prior radiation exposure (15% vs 30%). Since we procured the targeted numbers of CD34^+ cells/kg in 61% of patients in the rhTPO-mobilized group versus in 10% of controls with only one apheresis procedure, it is likely that this sixfold difference was the consequence of exposure to rhTPO and not due to any pretreatment imbalance between the study cohort and controls. Indeed, when we analyzed our data after stratifying for exposure for prior radiation therapy, the beneficial effect of rhTPO on procurement and engraftment had been confirmed. Although the number of patients with prior radiation exposure was small, a trend suggesting favorable effects of rhTPO on CD34^+ cell yield and hematopoietic recovery was observed in all groups. Should our findings hold true in larger

### Table 5. Percentage of CD41^+ Cells Within the CD34^+ Low Side-Scatter Population From PBPC Products by rhTPO Dose Level

<table>
<thead>
<tr>
<th>rhTPO (µg/kg)</th>
<th>PBPCd1</th>
<th>PBPCd2</th>
<th>PBPCc1</th>
<th>PBPCc2</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>CD41^+ Cells (%)</td>
<td>CD34^+ 10^6/kg (%)</td>
<td>CD41^+ Cells (%)</td>
<td>CD34^+ 10^6/kg (%)</td>
</tr>
<tr>
<td></td>
<td>Median</td>
<td>Range</td>
<td>Median</td>
<td>Range</td>
</tr>
<tr>
<td>Arm A</td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>0.6 × 1*</td>
<td>9</td>
<td></td>
<td>6.1</td>
<td></td>
</tr>
<tr>
<td>1.2 × 1</td>
<td>48</td>
<td>29-74</td>
<td>3.4</td>
<td>2.8-6.1</td>
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<td>2.4 × 1</td>
<td>58</td>
<td>19-65</td>
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<td>1.7-6.8</td>
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<td>Arm B</td>
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</tr>
<tr>
<td>0.3 × 3</td>
<td>22</td>
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<td>4.9</td>
<td>1.7-6.6</td>
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<td>0.6 × 3</td>
<td>68</td>
<td>23-89</td>
<td>8.1</td>
<td>3.4-18.1</td>
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<tr>
<td>Arm C</td>
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<tr>
<td>0.6 × 2</td>
<td>24</td>
<td>4-31</td>
<td>2.5</td>
<td>2.2-4</td>
</tr>
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</table>

*Available only from 1 patient at this dose level.
The observed decrease in erythrocyte transfusion benefit of rhTPO in patients undergoing high-dose chemotherapy with stem-cell support versus a comparable and identically treated nonrandomized control group. A prospective, randomized study to further clarify the optimal dose, schedule, and benefits resulting from the administration of rhTPO and G-CSF versus G-CSF alone is planned.

NOTE ADDED IN PROOF

The most active dose and schedule of rhTPO and G-CSF that were identified in our study have been tested in a randomized, placebo-controlled, phase II study. Preliminary results suggest that this regimen has significant activity in mobilizing PBPC compared with G-CSF alone.38
ACKNOWLEDGMENT

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