DESPITE THE AVAILABILITY of high-dose therapy and peripheral blood stem cell transplantation (PBSCT) for the treatment of multiple myeloma (MM), recurrent disease remains a major problem for patients with MM. Immunotherapy may provide additional control of residual disease. The unique biology of MM allows for the MM idiotypes (Id) and their peptide fragments to serve as tumor-specific antigens and potential targets for adjuvant immunotherapy. One of the most potent means of stimulating immune responses is through the use of dendritic cells (DC). Therefore, we have conducted a feasibility study to evaluate a DC-based Id vaccination strategy for MM patients after PBSCT.

Prior studies by Eisen et al.1-2 showed that myeloma-derived Ig can serve as an immunogen in a murine mineral oil-induced plasmacytoma (MOPC) model. When the myeloma protein was used in a vaccine it stimulated a tumor-specific protective immune response.1,2 The specificity of the immune response was restricted to unique determinants (idiotype) located on the antigen-binding sites of the Ig molecule. Other investigators have confirmed Eisen’s findings in various myeloma model systems including a more physiological model based on the spontaneously arising ST2 murine myeloma.3,4 Studies of the immune mechanisms contributing to Id-specific tumor protection have been discussed in detail and have led to the assertion that Id-specific T cells are important for tumor protection in MM.5 Bogen et al.6-8 have shown that Id-specific CD4+ T cells are important for tumor protection in the MOPC 315 model. Others have investigated Id-specific cytotoxic CD8+ T-cell clones, but their role in tumor protection remains to be determined.9,10 The ability to make these anti-Id immune responses, however, was impeded by the presence of circulating myeloma protein.11

The importance of Id-specific T cells in myeloma is not well studied in humans. A small number of Id-specific T cells has been reported in patients with early stages of MM.12 Osterborg et al.13 reported on the Id immunization of five MM patients that stimulated T-cell immune responses in all five. Id presented by monocyte-derived DC was able to stimulate a T-cell proliferative response to the MM protein.14 The immunogenicity of a human MM Id protein vaccine in a normal person and the subsequent transfer of Id-specific T cells to a patient has been shown in the setting of allogeneic bone marrow transplantation (BMT).15

Id vaccination for patients with B-cell non-Hodgkin’s lymphoma has succeeded in inducing specific immune responses in these patients.16,17 There was a strong correlation between the presence of an anti-Id immune response and freedom from disease progression as well as overall survival. Furthermore, the...
patients who were in a complete remission at the time of their vaccination had a greater probability of making an immune response.\textsuperscript{17}

A prospective randomized study has recently shown that high-dose chemotherapy with autologous BMT is superior to standard chemotherapy for the treatment of MM. High-dose therapy resulted in a higher response rate, a prolonged event-free survival, and an increased overall survival.\textsuperscript{18} Id vaccinations are, thus, likely to be most efficacious in MM patients who have completed a PBSCT and have achieved a status of minimal residual disease (MRD).\textsuperscript{5}

Dendritic cells are capable of antigen uptake, processing, and presentation on major histocompatibility complex (MHC) class I and II molecules. They are efficient in inducing strong and protective immune responses, especially T-cell immune responses.\textsuperscript{19} Immunization with antigen or tumor peptide pulsed autologous DC has been successfully used in various animal models\textsuperscript{20,21} and has more recently been introduced into clinical trials for lymphoma\textsuperscript{22} and melanoma.\textsuperscript{23} Based on these observations we have conducted a study to evaluate the feasibility of DC-based Id vaccination in MM patients after high-dose therapy and PBSCT.

MATERIALS AND METHODS

Patients and PBSCT. Fourteen eligible MM patients treated at our institution with PBSCT were offered adjuvant immunotherapy on this phase I study. Two patients declined to participate. All patients had laboratory, histopathologic, and radiologic findings consistent with a diagnosis of MM. Institutional review board–approved informed consent was obtained from all study patients. Serum and BM samples were collected early in their disease, preferably before initiation of chemotherapy. Patients received high-dose therapy if they had chemotherapy-sensitive disease (shown by a >50% decrease of monoclonal Ig and a decrease of BM plasma cell involvement to <20% with initial chemotherapy), a Karnofsky performance status of greater than 70%, and no renal, hepatic, cardiac, and pulmonary impairment. PBSC were identified by ELISA as above except using a goat anti-human IgG subclass. IgG1, IgG2, and IgG4 were purified with protein A affinity chromatography followed by ion exchange chromatography. Specifically, serum was diluted with PBS pH 7.5 and passed through a Protein A Sepharose CL-4B column (Sigma). The bound Igs were specific for the different human heavy-chain isotypes which were detected with peroxidase-conjugated mouse monoclonal antibodies which were specific for the different human heavy-chain isotypes (Southern Biotechnology Assoc, Birmingham, AL).

Ig proteins were purified according to their heavy-chain isotype and subclass. IgG1, IgG2, and IgG4 were purified with protein A affinity chromatography followed by ion exchange chromatography. Specifically, serum was diluted with PBS pH 7.5 and passed through a Protein A Sepharose CL-4B column (Sigma). The bound Igs were then eluted with 0.1 mol/L citric acid in sequential pH steps at pH 5.5, 5.0, 4.5, 4.0, and 3.5. IgG2 Igs eluted from Protein A at a higher pH and IgG1 and IgG4 at a lower pH.\textsuperscript{24} The MM protein containing fractions were pooled and dialyzed against 0.01 mol/L Tris pH 9.0. The Id was then bound to a strong anion exchange (AEX) resin (Macro-Prep High Q support; Bio-Rad, Hercules, CA) and eluted with a 0 to 0.3 mol/L NaCl salt gradient in 0.01 mol/L Tris pH 9.0. Fractions containing the Id protein were identified by ELISA as above except using a goat anti-human

<table>
<thead>
<tr>
<th>Table 1. MM Id Vaccine Patient Characteristics</th>
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<td><strong>Patient (UPN)</strong></td>
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<td>11 (1504)</td>
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<td>12 (1521)</td>
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Abbreviations: UPN: unique patient number; VAD: vincristine, adriamycin, dexamethasone; VMCP: vincristine, melphalan, cyclophosphamide, prednisone; PSC-VAD: PSC 833, vincristine, adriamycin, dexamethasone; MP: melphalan, prednisone; MEL 140: melphalan at 140 mg/m\textsuperscript{2}; CVP: cyclophosphamide, vincristine, prednisone; BMVP: BCNU, melphalan, vincristine, prednisone; SD: stable disease.
heavy-chain antibody and detected with a peroxidase conjugated goat anti-human kappa or lambda antibody.

Id proteins with an IgG3 isotype were isolated by a combination of Protein G Sepharose, Protein A Sepharose, and AEX chromatography. Serum was diluted with PBS and passed through a Protein G Sepharose (Sigma) column. The bound protein was eluted with 0.1 mol/L citric acid, pH 3.5, and the eluate was then passed over a Protein A column to bind contaminating normal serum IgG1, IgG2, and IgG4 Ig subclones. The Protein A column flow through which contains IgG3 lgs was further purified using a strong AEX chromatography as above.

IgA Id proteins were purified by precipitating twice with ammonium sulfate (45% vol/vol), dialyzed against 0.01 mol/L Tris, pH 9.0, and separated by strong AEX chromatography as described above. IgA-containing fractions were pooled and then depleted of IgG by Protein A affinity chromatography. Purified Id proteins were extensively dialyzed against sterile normal saline and filtered through a 0.45-µm membrane.
The Id proteins were tested for endotoxin using the LAL clotting assay as per the manufacturer’s instructions (Cape Cod Inc, Falmouth, MA) and for sterility.

**DC vaccines.** The MM patients received their DC vaccines 3 to 6 months after completing their PBSCT. Each patient was given two Id-pulsed DC immunizations separated by 4 weeks. DC vaccines were prepared as previously described. Briefly, mononuclear cells were collected from peripheral blood by leukapheresis. The precursor DC were isolated from the mononuclear cells by a series of density gradient centrifugations. The DC preparations were cultured with purified Id for 36 to 48 hours, collected, washed free of Id protein, and administered IV over 60 minutes in a volume of 100 mL. The composition of DC preparations was monitored by light microscopy and flow cytometry.

DC were identified by high MHC class II expression and the absence of T- and B-cell markers. An average of 5.1 × 10^5 ± 2.9 × 10^5 DC were infused with each immunization.

**Idkeyhole limpet hemocyanin (KLH) vaccines.** Five Id/KLH vaccines were administered after the DC vaccines. The Id/KLH vaccines were prepared as previously described. Briefly, purified Id proteins were conjugated with glutaraldehyde to the immunologic carrier KLH at a 1:1 (wt/wt) ratio. The conjugates were dialyzed against normal saline and then mixed with freshly prepared ISAF adjuvant. ISAF consisted of 10% (vol:vol) squalane (Aldrich Chemical, Milwaukee, WI), 5% (vol:vol) pluronic L121 (BASF, Parsippany, NJ), and 0.4% (vol:vol) Tween 80 (Aldrich Chemical, Milwaukee, WI) in PBS.
The Id/KLH vaccinations were started 4 weeks after the second Id/DC vaccine and were administered at 4-week intervals.

**Identification of MM Id encoding genes.** The myeloma-derived heavy- and light-chain Ig variable region genes were isolated from BM aspirate specimens obtained before or in the early part of induction chemotherapy. BM mononuclear cells (BM-MNC) were isolated from the aspirate by Ficoll-Paque density centrifugation (Pharmacia, Piscataway, NJ). RNA was isolated from 5 to 10^7 cells by Ficoll-Paque density centrifugation (Pharmacia, Piscataway, NJ). RNA was isolated from 5 to 10^7 cells by Ficoll-Paque density centrifugation (Pharmacia, Piscataway, NJ). The RNA was isolated from 5 to 10^7 cells by Ficoll-Paque density centrifugation (Pharmacia, Piscataway, NJ). The RNA was isolated from 5 to 10^7 cells by Ficoll-Paque density centrifugation (Pharmacia, Piscataway, NJ). RNA was isolated from 5 to 10^7 cells by Ficoll-Paque density centrifugation (Pharmacia, Piscataway, NJ).

The myeloma Id heavy-chain variable region genes were amplified from the cDNA using polymerase chain reactions (PCR). Six 5’ primers corresponding to the leader sequences of the six human heavy-chain variable region families (VH1 to VH6) were used in individual PCR. The 3’ primers used in the PCR corresponded to the appropriate gamma (5’-CTTGGCAAGG CAAGCCAGGC-3’) or alpha heavy-chain constant region (5’-GAGGTCAGC GGGAAACGAC CT-3’). PCR conditions were as described except for a few reactions where the annealing temperature was reduced from 55°C to 50°C. In all cases, electrophoretic analysis of the PCR reactions showed a predominant band of the expected length in only one of the six VH leader primer reactions. The PCR product was purified by electrophoresis through agarose and extracting the DNA from the agarose using the QIAquick gel extraction kit (Qiagen). The isolated PCR product was then ligated into the vector, pCRIII (Invitrogen, San Diego, CA) and transfected into TOP10’ bacteria. Plasmid DNA was isolated from cultures derived from individual bacterial colonies and sequenced using dye termination PCR (Perkin-Elmer, Norwalk, CT). A sequence for a heavy-chain variable region was determined to be myeloma derived if it was identified in DNA isolated from three bacterial colonies generated from the cloning of two independent PCR products.

Identification and cloning of the MM-derived light-chain variable (VL) region was performed using the same PCR-based approach as for the VH-chain region. The sequence for the kappa leader primers are: VL1, 5’-ATACAGACAG TCTCACCAGT GTTGTCAGCA CCCCCGTGTC-3’; VL2, 5’-ATACAGACAG TCTCACCAGT GGRGCCCGCG CKCAGCCT-3’; VL3, 5’-ATACAGACAG TCTCACCAGT GACATGA GGG TCCCGGCTA G-3’; and VL4, 5’-ATACAGACAG TCTCACCAGT GCACACAGGG CCCCCACTA G-3’. The sequence for the lambda leader primers are: AL1, 5’-ATACAGACAG TCTCACCAGT GCCTGCGCTG TCTGCCTCC-3’; AL2, 5’-ATACAGACAG TCTCACCAGT GCCTGCGCTG CACTCTCC-3’; AL3, 5’-ATACAGACAG TCTCACCAGT ACCTGCGCTG CACTCTCC-3’; AL4, 5’-ATACAGACAG TCTCACCAGT GCCTGCGCTG CACTCTCC-3’; and AL5, 5’-ATACAGACAG TCTCACCAGT ACTGCGCTG CACTCTCC-3’.

The 3’ primers were derived from the joining regions of either kappa or lambda and the sequence for the kappa joining region (KJ) is 5’-TGCACTGCGC TACGCGTGA CACGCTCC-3’ and the lambda joining region (LJ) is 5’-CTGACCGCTG AGGGCTCC-3’.

**Generation of an Id-encoding recombinant adenovirus (IdAd).** A recombinant adenovirus containing a patient’s MM-derived variable region genes was made for selected cases. VH genes used for cloning were obtained by reamplification of Id VH using Phi DNA polymerase (Stratagene, La Jolla, CA) and leader and constant region primers containing unique restriction sites to aid in the cloning. The primers used for identifying and isolating the VL genes contained restriction sites so that the initial PCR product could be used for cloning into an expression vector without reamplification. The patient’s MM VH and VL genes were cloned into the bicistronic Ig expression vector, pTC.

This vector allowed for the cloning of the heavy- and light-chain variable region genes in frame with their respective constant regions. Cytomelagovirus promoters were used for each Ig gene. The expression cassette containing the Id genes was then moved from pTC to the adenovirus transfer vector, pXCL1. The Id-gene–containing pXCL1 plasmid was cotransfected with the unpackageable adenovirus type V (Adv) plasmid pMM17 into 293 cells as previously described.

Intracellular recombination between these two plasmids in this Adv permissive cell line resulted in the generation of replication-deficient Ad. The recombinant virus was plaque purified and then expanded on 293 cells. The titers of the adenovirus stocks were determined by limiting dilution. Integrity of the Ig genes was determined by assaying the supernatant of infected 293 cells for Ig expression by ELISA as described above.

**T-cell assays.** Id-specific T-cell immune responses were monitored by a T-cell proliferation assay as previously described except for the use of the serum-free medium, AIM V (GIBCO-BRL, Grand Island, NY) instead of an Iscove’s modified Dulbecco’s medium–based culture medium. Briefly, heparinized blood samples were obtained before each Id vaccination and 4 and 12 weeks after the last vaccination. Peripheral blood mononuclear cells (PBMC) were isolated from blood using Ficoll-Paque (Pharmacia) density centrifugation. After washing, the PBMC were plated at 5 × 10^6 cells per well in a 96-well “U”-bottom plate in AIM V medium supplemented with titrating amounts of either Id protein, an isotype-matched control Ig, or KLH. The PBMC were cultured for 3 days and then split between two 96-well plates. Fresh medium containing IL-2 was added to the wells such that the final
ELISA for isotype or light chain restriction and sodium dodecyl sulfate-polyacrylamide gel electrophoresis (data not shown).

Id vaccination. The MM patients were immunized with a combination of Id-pulsed DC vaccines and Id/KLH conjugate vaccines. Ten of the 12 patients completed their series of two Id-pulsed DC vaccinations plus 5 Id/KLH booster injections. Patient 7 received two Id-pulsed DC and two Id/KLH vaccines before developing acute leukemia. Patient 10 developed progressive disease during his Id immunization and elected to withdraw from the study after his second Id/KLH vaccine. Despite numerous cycles of chemotherapy and high-dose chemotherapy with PB SCT, a sufficient number of DC precursors could be routinely isolated from the leukapheresis products (Table 2). The immature DC precursors were enriched approximately 100-fold using a series of density gradient centrifugations resulting in a cell population consisting of 3% to 63% DC. The DC yield from a leukapheresis collection ranged from 1 to 11 × 10^6 cells. The purity of the DC preparations was somewhat lower than that seen with lymphoma patients, which was likely due to increased circulating immature cells which copurified with the precursor DC. After isolation, the immature DC were incubated in medium containing the patient’s Id protein, washed, and administered IV. The patients tolerated the DC infusions well with the vast majority of patients experiencing no side effects or complications. Two of the patients (patients 6 and 8) developed transient low-grade fevers and chills while being infused with their first Id-pulsed DC vaccine and one patient (patient 7) developed a postinfusion thrombophlebitis after his second DC infusion.

The patients also tolerated the subcutaneous Id/KLH vaccines well. Most patients experienced only minor side effects consisting of local erythema, induration, and soreness at the injection site. These side effects were transient and were controlled with oral acetaminophen.

T-cell proliferation. Before immunization, none of the 12 MM patients had a measurable T-cell proliferative response to either their Id protein or KLH. However, the Id immunizations stimulated an Id-specific T-cell proliferative response in two patients. Patient 1 developed Id-specific proliferation that was first detectable after receiving two Id-pulsed DC vaccines and five Id/KLH booster immunizations (Fig 1A). The T-cell

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**Table 2. Dendritic Cell Vaccine Characteristics**

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<tr>
<th>Patient (UPN)</th>
<th>Time of 1st Id Vaccine*</th>
<th>No. of DC Infused (&gt;10^9) (1st, 2nd vaccine)</th>
<th>DC Purity (1st, 2nd vaccine) (%)</th>
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<td>3.7, 3.3</td>
<td>10, 10</td>
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*Time from reinfusion of the PBSC to time of first Id-pulsed DC infusion in months.

†Percent of infused cells that were B- and T-cell lineage marker negative and high MHC class II expressors by flow cytometry analysis.
proliferation to an irrelevant isotype matched Ig that was isolated in an identical manner was less than one half that seen with the Id protein. The amount of T-cell proliferation measured depended on the dose of stimulating Id. Patient 1 maintained a measurable T-cell proliferative response for more than 3 months after completing the immunizations. Patient 9 developed a T-cellular immune response after two Id-pulsed DC vaccinating and a single Id/KLH boost (Fig 1B). Similar to patient 1, the T-cell response was specific for the patient’s Id protein and dose-dependent. Similar Id-specific T-cell proliferation could be measured after his second and third Id/KLH immunizations.

Eleven of the 12 patients developed a strong, dose-dependent and sustained T-cell proliferative response to the carrier protein KLH. The patient who did not mount an immune response to KLH (patient 7) developed acute myelogenous leukemia after his second Id/KLH vaccine. The T-cell proliferative immune responses to KLH were detected after a single Id/KLH vaccination and a single Id/KLH boost (Fig 1B). Similar to patient 1, the T-cell response was specific for the patient’s Id protein and dose-dependent. Similar Id-specific T-cell proliferation could be measured after his second and third Id/KLH immunizations.

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Fig 1. Id-specific T-cell proliferation. PBMNC were isolated preimmunization and after each immunization and were cultured for 6 days in medium alone (med.) or medium containing Id at 0.1, 1.0, 10, and 100 µg/mL (0.1, 1.0, 10, and 100 respectively). Overnight ³H thymidine incorporation into DNA was determined on day 6. (A) T-cell proliferation assay of patient 1 before any immunization (prevaccine) and 3 months post-Id/KLH vaccine no. 5. Proliferation in medium containing an irrelevant isotype-matched Ig at 100 µg/mL was statistically significant less (P = .014) with 3,296 ± 445 cpm. Comparable results were obtained in experiments performed 1 and 2 months post-Id/KLH vaccine no. 5. (B) T-cell proliferation assay of patient 9 before any immunization (prevaccine) and after Id/KLH vaccine no. 1. Proliferation in medium with an irrelevant isotype matched Ig at 100 µg/mL was at 519 ± 93 cpm (P = .05). Id-specific proliferation was also seen after Id/KLH vaccine nos. 2 and 3. (C) T-cell proliferation of patient 1 in response to the immunologic carrier KLH at 3 months post Id/KLH vaccine no. 5. (D) T-cell proliferation of patient 9 in response to the immunologic carrier KLH after Id/KLH vaccine no. 1. The assays were run in triplicate or quadruplicate and mean values are depicted. Error bars denote one standard deviation. Using the Mann-Whitney U test, prevaccine and postvaccine Id-specific responses were significantly different at all Id concentrations studied (P = .014 patient 1, P = .05 patient 9).
suggesting that the amplified DNA was derived from myeloma cells. The variable region genes were then cloned into a bicistronic expression/adenovirus transfer vector containing the heavy- and light-chain constant regions. This transfer vector was used with an adenovirus plasmid to generate IdAd. Fibroblast lines were established from skin punch biopsy samples obtained from the patients. Sufficient numbers of fibroblasts necessary for the CTL assay could be readily obtained after 3 to 4 weeks of culture. The fibroblasts were infected with the adenovirus without significant cytotoxicity. Approximately 60% to 80% of the fibroblasts expressed the Id protein after adenovirus infection using an MOI of 100 to 1,000. Expression could be detected within 15 hours of infection using an anti-Id protein antibody. Approximately 60% of the fibroblasts internalized 51Cr well with 5 × 10³ cells routinely incorporating 1 SD. 3H thymidine incorporation is reported as counts per minute (cpm). Background proliferation in medium was less than 1,500 cpm.

As previously reported, patient 1 showed an Id-specific CTL response 4 weeks after the second Id-pulsed DC vaccination. The effector cells (E) for the CTL assay were autologous PBMNC that were cultured in medium containing a high concentration of Id protein. After Id stimulation, these effector cells lysed 51Cr labeled Id expressing autologous fibroblast targets (T) in a dose-dependent manner (Fig 2) and the cytotoxic activity required prior stimulation with Id. PBMC cultured with a control Ig did not stimulate any cytotoxic T-cell activity against Id expressing targets or control targets. The cytotoxic activity was also specific for Id-expressing fibroblasts. When cultured with the Id-stimulated effector cells, autologous fibroblasts infected with an IdAd released significantly more 51Cr compared to fibroblasts infected with control adenovirus (AdLacZ) (P ≤ .05 at an E:T ratio of 100:1, 33:1, and 11:1 as determined by the Mann-Whitney U test). The requirement for Id stimulation and specific lysis of Id targets strongly suggest that the cytolytic activity is I-specific and not due to nonspecific cytolytic activity, such as that seen with natural killer cells. The fibroblasts targets expressed class I but lacked class II expression as determined by fluorescence-activated cell sorting (data not shown) providing indirect evidence for CD8⁺ class I-restricted CTLs as the effector cell population. This Id-specific CTL activity was only measurable 1 month after patient 1 received his second Id-pulsed DC vaccine. Because of the complexity of the assay, only three patients were evaluated for Id-specific CTL activity. The other two patients studied (patients 3 and 9) had no measurable Id-specific CTL activity.

Clinical observations. The date of the last follow-up is May 31, 1998, with a minimum follow-up of 16 months from PBSCT and 3 months from last Id-KLH injection. Of the 12 treated patients, 9 are alive 16 to 30 months after transplantation. Two patients have maintained a CR at 30 months (patient 1) and 17 months (patient 9) after PBSCT. One patient (patient 6) had stable disease after PBSCT and is now 20 months after PBSCT. Without any further chemotherapy treatment, this patient’s M-spike has gone from 2.3 g/dL before her Id vaccination to 1.8 g/dL after completing her vaccinations. The myeloma progressed in 5 patients at 8 to 17 months after PBSCT. Two of these patients (patients 3 and 4) died at 11 and 22 months after PBSCT. One patient (patient 7) died 10 months after PBSCT of a secondary acute myeloid leukemia.

DISCUSSION

Compared to conventional chemotherapy, high-dose therapy with autologous stem cell support offers improvements in CR rates, freedom from disease progression, and overall survival, and is emerging as the therapy of choice to achieve MRD. Unfortunately, the majority of MM patients relapse after...
PBSCT, even those patients who obtain CRs and good PRs. Additional chemotherapy dose intensification is unlikely to significantly improve the survival of MM patients. New treatment approaches are needed for this disease. The rationale for immunizing MM patients after PBSCT has been discussed and is based on several premises. The Id protein is a unique immunologic marker for the malignant MM cells and an immune response directed against the Id protein may stem the regrowth of MM. However, the immune system’s ability to mount a response to the MM Id is likely greatest when at a stage of minimal disease. In mouse MM models, the tumor burden directly correlated to the development of Id-specific T-cell unresponsiveness or anergy. Therefore, Id immunizations may be the most efficacious in the post-PBSCT period.

The work presented in the report shows that DC-based Id vaccination is feasible. We isolated sufficient quantities of individual Id proteins and DC to immunize 12 patients. The vaccinations were well tolerated with no serious toxicity and 10 of 12 patients finished the planned course of immunizations. Id immunizations stimulated Id-specific T-cell proliferative responses in two patients and an Id-specific CTL response was detected in one of these patients. While this patient (patient 1) developed a measurable cytolytic immune response after only two Id/DC IV vaccinations, the Id-specific proliferative response was not detectable until completion of the subsequent 5 Id/KLH booster immunizations. The Id-specific proliferative response of patient 9 developed already after two DC/Id immunizations. The ability to stimulate or at least detect an anti-Id immune response correlated with the clinical status of the patient post-PBSCT. The two patients in CR after PBSCT were the ones who made a detectable immune response. One patient had a decrease in M-protein while receiving Id immunization.

We used a DC-based vaccine approach because of the potent immunostimulatory capacity of DC. We were able to detect Id-specific immune responses in only two of the vaccinated patients. It is possible that our T-cell proliferation assay is not sufficiently sensitive and more patients may have made immune responses. Using a single-cell assay (ELISPOT), Bergenbrant et al. found Id-reactive T cells in three of five MM patients immunized with Id mixed in alum. When they used granulocyte-macrophage (GM)-CSF instead of alum, they were able to detect Id-reactive T cells by ELISPOT in all five of the immunized MM patients. Only 1 of the 5 Id/GM-CSF vaccinated patients made an Id-specific T-cell response that could be detected using a T-cell proliferation assay. Yi et al. immunized patients with early stage MM (stage I and II) because of their earlier finding of Id-reactive T cells in similar MM patients. It is possible that patients with early stage MM are better able to make an immune response to their Id than patients with more advanced disease, especially after high-dose chemotherapy. However, we were able to detect strong cellular immune responses to KLH in nearly all of our immunized patients, showing that these patients have an ability to make an immune response to new antigens. These T-cellular immune responses were clearly detectable in 11 of 12 patients after one to three Id/KLH vaccines and peaked usually within less than 1 year after high-dose therapy with PBSCT (Table 3). This is in line with a recent review on immune reconstitution after autologous hematopoietic stem cell transplantation. Other investigators have immunized MM patients with Id after high-dose therapy and have also been able to detect T-cell immune responses. Using a cytokine release assay, Kwak et al. found that four of five Id vaccinated MM patients made an immune response. Similarly, Wen et al. describe immunizing an MM patient with Id-pulsed DC pre- and post-PBSCT, which stimulated a T-cell response.

When similarly prepared DC-based Id immunizations were administered to low-grade lymphoma patients, a much higher proportion of patients made an anti-Id immune response. One major difference between the two groups of patients is that MM patients often have easily detectable circulating Id protein. This circulating Id may have prevented the development or possibly the detection of a T-cell immune response. This observation is consistent with the observation of Bogen where Id-specific T cells become anergic or even undergo apoptotic death when exposed to soluble Id protein. It is intriguing that the two Id-specific T-cell responses we observed were found in the two patients who entered a CR after PBSCT. However, others have observed T-cell immune responses in immunized patients with substantial levels of serum Id. Differences in MM patient characteristics and the method of immunization may account for some of the differing observations. More immunized MM patients need to be evaluated before we can determine whether residual M-protein inhibits the development of an immune response.

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strategies to induce clinically significant Id-specific cellular immune responses in a larger cohort of MM patients.

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