Mitomycin C–Induced DNA Damage in Fanconi Anemia: Cross-Linking or Redox-Mediated Effects?

To the Editor:

The article by Carreau et al1 reports on the in vivo effects of mitomycin C (MMC) in mice carrying the Fanconi anemia (FA) group C mutation (Fac−/−). Among the mechanistic scenarios underlying FA pathogenesis, the authors refer to a phenotypic feature of FA cells related to oxygen hypersensitivity. Unfortunately, the use of citations on this subject appears to be quite inappropriate. First, the authors attributed a “secondary” role for oxygen sensitivity in FA cells2 which, however, may have been made oxygen-resistant after the immortalization procedure. In fact, the loss of O2 sensitivity in transformed cells has been recognized as a general phenomenon, not confined to FA cell lines.3 A general statement was then made4 about the published results of studies which “have demonstrated overproduction of reactive oxygen species (ROS) and increased susceptibility to oxygen, as well as an increase in ROS-induced DNA lesions, particularly 8-hydroxy-2′-deoxyguanosine (8OHdG).5 Unfortunately, the three references reported6–8 (cited as 37–39 in the report) neither dealt with FA nor with ROS-induced DNA damage. The above statement about excess ROS production and 8OHdG formation in FA was true, but rather should refer to the reports by Takeuchi and Morimoto7 and Degan et al.8 It is worthwhile to consider the subject of oxidative stress in FA based on both in vitro and ex vivo evidence, as reviewed by us recently.9

A role for oxidative stress in FA has been documented for two decades, with reports providing evidence for an improvement of either chromosomal instability or cell growth after exposure of either primary lymphocyte cultures or fibroblasts from FA patients to: (1) catalase or superoxide dismutase, (2) low-molecular-weight antioxidants, or (3) decreased oxygen levels.10–14 Ag2 cell cycle delay, observed in FA cells, was counteracted by culturing cells in 5% O2,15 and a major role was suggested for free iron in inducing G2 arrest in FA cells.16 The report by Takeuchi and Morimoto7 provided evidence for excess oxidative DNA damage (8OHdG) in FAA cells challenged with H2O2 that was related, at least in part, to catalase deficiency. A recent report by Ruppitsch et al17 provided elegant evidence for the loss of both MMC and diepoxybutane (DEB) sensitivity of FAA cells transfected with CDNAs causing overexpression of thioredoxin, a nonenzymatic antioxidant protein.18 Hence, both exogenous and endogenous antioxidants can decrease the phenotypic defect of FA cells, both including O2 and MMC sensitivity. In turn, the action mechanisms of MMC can either be ascribed to DNA cross-linking or to redox cycling, as reported in early studies of MMC19,20. That MMC sensitivity in FA cells may be attributed to redox mechanisms rather than to DNA cross-linking has been shown by independent reports11.12.21.22 focused on as many different endpoints (chromosomal instability, cytotoxicity, apoptosis, and mutagenesis). Together, the results of these studies showed that: (1) MMC-induced toxicity was confined to normoxic conditions which, unlike hypoxia, were associated to enhanced redox-cycling mechanisms, not to DNA cross-linking,21,22 and (2) MMC toxicity was both removed by antioxidants and by low-molecular-weight antioxidants.11,13

The observation of redox abnormalities in FA is not confined to in vitro conditions. A series of ex vivo studies provided evidence for abnormal O2 metabolism in FA patients and in their parents. Freshly drawn white blood cells from both FA homozygotes and heterozygotes produced excess ROS as detected by luminol-dependent chemiluminescence (LDCL),21,24 and displayed excess 8OHdG levels that were significantly correlated with LDCL as well as with chromosomal instability.4 Thus, both ex vivo and in vitro evidence pointed to a direct link between ROS formation, oxidative DNA damage, and chromosomal breakages in FA.

Based on the available evidence, one might suggest that the authors could carry out a new series of experiments by exposing Fac−/− mice to different oxygen levels, with or without MMC administration. As additional endpoints worth being evaluated in Fac−/− mice, one might suggest to include the evaluation of oxidative DNA damage as well as of ROS-detoxifying activities. This study could provide a formidable insight both into the FAC defect and the in vivo action mechanisms of MMC.

In conclusion, the current view attributing the FA-associated defect(s) to the phenotypic sensitivity to MMC and DEB related to cross-linking mechanisms may be viewed as a fading dogma relying on the definition of FA as a DNA repair disorder. While no conclusive evidence has thus far related FA gene products to any function in DNA repair, a thriving body of evidence has associated MMC (and DEB) sensitivity to an impairment of redox balance in FA cells, both in vitro and in vivo. This evidence should no longer be disregarded in the forthcoming studies of FA.

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REFERENCES


7. Takeuchi T, Morimoto K: Increased formation of 8-hydroxydeoxyguanosine, an oxidative DNA damage, in lymphoblasts from Fanconi’s anemia patients due to possible catalase deficiency. Carcinogenesis 14:1115, 1993

First regarding the references, we believe that one reference by Takeuchi et al1 was omitted due to formatting of the paper and was overlooked on our part. The references cited as 37-39 regard the Bcl2 knockout mice and are discussed and referred to later in the paper.

Second, our paper dealt with mitomycin C (MMC) hypersensitivity of the Fancc−/− mouse model we generated. We believe that our discussion is in fact an overview of the possible in vivo effects of MMC, and we did not dismiss oxygen radical formation as a possible effect during the metabolism of MMC. Nonetheless, one still does not know if reactive oxygen species (ROS) formation is responsible for the hypersensitivity of the Fancc−/− mice treated with MMC, although we believe that the effect we observed may result from a defect in DNA repair. In fact, more information is now becoming available regarding a DNA repair defect in FA.

1. FA cells were shown to be specifically sensitive to interstrand crosslinks and not intrastrand crosslinks confirming the specificity of the defect in crosslink repair.2
2. FA cells were shown to lack a repair complex that specifically binds DNA crosslinks.3
3. The increased ROS-induced lesion 8OHdG, in FA patients cells also supports the idea of a lack of a repair mechanism; without repair, the lesions remain in the DNA.
4. FANCA and FANCC have been shown to interact in a complex and translocate to the nucleus; this implies a more direct role of FANCC in repair.4
5. FA cells have been shown to be defective in double-strand break repair.5
Oxidative DNA damage is repaired by the BER pathway, which may share steps with the crosslink repair pathway. Thus, increased sensitiv-
To the Editor:

Methylenetetrahydrofolate reductase (MTHFR) is a key enzyme in the remethylation pathway of homocysteine. The study by Girelli et al. showed that the C677T mutation of the gene encoding for MTHFR is common in Italy, is not associated with coronary atherosclerotic disease, and influences the plasma levels of total homocysteine (tHcy) only in subjects who have low serum concentrations of folic acid. These findings are important confirmations of previous reports. In addition to the above issues, Girelli and his coworkers addressed the problem of the influence of the MTHFR genotype on the increases of plasma tHcy levels after an oral methionine loading. The methionine loading test, which was originally developed to detect heterozygosity for the deficiency of cystathionine-β-synthase, a key enzyme in the trans-sulfuration pathway of homocysteine, is useful not only to detect subjects with defects of the trans-sulfuration pathway, but also to identify subjects at risk for thrombosis. It has been suggested that abnormal post-methionine loading (PML) increases in the plasma levels of tHcy reflect abnormalities of the trans-sulfuration pathway of homocysteine metabolism and are insensitive to the remethylation pathway, in contrast with fasting plasma levels of tHcy, which are sensitive to the remethylation pathway. If this hypothesis were true, the T/T mutation at nucleotide 677 of MTHFR, which is associated with an enzyme activity of about 50% of normal, should not influence the results of the methionine loading test. Girelli et al commented that their results agree with this hypothesis, because they showed that homoyzygosity for the C677T mutation of MTHFR is not associated with abnormally high PML tHcy increments. However, they found higher PML tHcy increments in individuals carrying the mutant T/T genotype (25.5 µmol/L) than in those with normal (C/C) or heterozygous (C/T) genotypes (23.6 µmol/L), although the difference was not statistically significant (Table 1). In conclusion, the available experimental evidence indicates that the fasting levels of tHcy in plasma are very sensitive to an abnormality of the remethylation pathway of homocysteine, such as that associated with the T/T genotype of MTHFR; however, they do not allow the conclusion that the PML increases of tHcy levels in plasma only reflect abnormalities of the trans-sulfuration pathway of homocysteine, because they also tend to be abnormal in individuals with the T/T genotype of MTHFR and in those with low serum levels of folic acid.

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Table 1. Increases of Plasma tHcy After an Oral Methionine Loading as a Function of MTHFR Genotypes

<table>
<thead>
<tr>
<th>Study</th>
<th>No. of Subjects</th>
<th>C/C</th>
<th>C/T</th>
<th>T/T</th>
<th>P*</th>
</tr>
</thead>
<tbody>
<tr>
<td>J. Jacques et al, 1996†</td>
<td>365</td>
<td>17.9</td>
<td>18.5</td>
<td>18.2</td>
<td>NS</td>
</tr>
<tr>
<td>Cattaneo et al, 1997†</td>
<td>231</td>
<td>14.4</td>
<td>14.8</td>
<td>18.6</td>
<td>.006</td>
</tr>
<tr>
<td>Legnani et al, 1997†</td>
<td>63</td>
<td>17.0</td>
<td>19.0</td>
<td>23.1</td>
<td>NS</td>
</tr>
<tr>
<td>Verhoef et al, 1997†</td>
<td>318</td>
<td>23.6</td>
<td>26.4</td>
<td>27.7</td>
<td>&lt;.05</td>
</tr>
<tr>
<td>Girelli et al, 1998†</td>
<td>415</td>
<td>23.6</td>
<td>23.6</td>
<td>25.5</td>
<td>NS</td>
</tr>
</tbody>
</table>

Values of PML tHcy increase are arithmetic means (†) or geometric means (†). The dose of oral methionine was 3.8 g/m² body surface area in the study by Cattaneo et al, and 100 mg/kg body weight in the other studies.

Abbreviation: NS, not significant.

*Genotype T/T vs genotype C/C.
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