Interleukin-10 Inhibits Erythropoietin-Independent Growth of Erythroid Bursts in Patients With Polycythemia Vera

By Klaus Geissler, Leopold Öhler, Manuela Födinger, Eva Kabrna, Marietta Kollars, Sonja Skoupý, and Klaus Lechner

In polycythemia vera (PV) erythroid colonies that grow in vitro in the absence of exogenous erythropoietin (EPO) arise from the abnormal clone that is responsible for overproduction of red blood cells. Although the mechanism of autonomous formation of burst-forming units-erythroid (BFU-E) is not fully understood, a spontaneous release of growth regulatory molecules by PV cells and/or by accessory cells is likely to be involved. Because of its cytokine synthesis inhibiting action, interleukin-10 (IL-10) could be a potentially useful molecule to modulate abnormal erythropoiesis in PV. We studied the effect of recombinant human IL-10 on the BFU-E-independent growth of erythroid bursts derived from peripheral blood mononuclear cells (PBMCs) of patients with PV. IL-10 showed a profound, dose-dependent, and specific inhibitory effect on autonomous BFU-E formation. Ten nanograms per milliliter of IL-10 significantly suppressed spontaneous growth of erythroid colonies in methylcellulose in five of five PV patients tested with a mean inhibition by 81% (range, 72-94). To elucidate the possible mechanism of the inhibitory action of IL-10 we further studied the effect of anticytokine antibodies on autonomous BFU-E growth and the ability of exogenous cytokines to restore IL-10-induced suppression of erythroid colony growth. Among a panel of growth regulatory factors tested (granulocyte-macrophage colony-stimulating factor [GM-CSF], IL-3, granulocyte colony-stimulating factor, stem cell factor, and insulin-like growth factor-1) GM-CSF was the only molecule for which both an inhibition of spontaneous BFU-E formation by its respective antibody as well as a significant restimulation of erythroid colonies in IL-10-treated cultures by exogenous addition was found. Moreover, inhibition of GM-CSF production by IL-10 was shown in PV PBMCs at the mRNA level. Our data indicate that autonomous BFU-E growth in PV can be profoundly inhibited by IL-10 and that this inhibitory effect seems to be at least in part secondary to suppression of endogenous GM-CSF production.

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MATERIALS AND METHODS

Patients. Five patients clinically diagnosed as having PV with the help of the PV Study Group guidelines were used in this study. Clinical and laboratory data of these patients are shown in Table 1. All patients were being managed by phlebotomy at the time of study and none of them had been previously treated with cytostatic drugs or radioactive phosphorus.

Preparation of cells. After informed consent, PB was collected into sterile tubes containing EDTA. MNCs were isolated from PB of patients

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by Ficoll-Hypaque density gradient centrifugation (density 1.077 g/mL, 400g for 40 minutes). The low-density cells were collected from the interface between density solution and plasma, washed twice, and resuspended in Iscove’s modified Dulbecco’s medium (GIBCO, Paisley, Scotland).

Reagents. Recombinant human IL-10 (rhIL-10; specific activity 1.2 × 10^6 U/mg) was kindly provided by Schering-Plough Corp (Kenilworth, NJ) and rhGM-CSF and rhIL-3 by Sandoz (Basel, Switzerland). rhG-CSF was purchased from British Biotechnology (Oxford, UK) and rhEPO from Boehringer Mannheim (Vienna, Austria). Recombinant human stem cell factor (rhSCF) and recombinant human insulin-like growth factor-1 (rhIGF-1) were obtained from Pharmacia Biotechnologie Hannover (Hannover, Germany). Antibodies directed against GM-CSF, IL-3, G-CSF, and SCF were purchased from Genzyme (Cambridge, MA), anti-IGF-1 from Serotec Ltd (Oxford, UK), and anti-IL-10 from R&D Systems Europe Ltd (Abingdon, UK).

 Colony assay. PBMNCs were cultured in 0.9% methylcellulose, 30% fetal calf serum (FCS; INLIFE, Wiener Neudorf, Austria), 10% bovine serum albumin (Behring, Marburg, Germany), α-thioglycerol (10⁻⁴ mol/L) and Iscove’s modified Dulbecco’s medium with or without the addition of cytokines or anticytokine antibodies. Cultures were plated in triplicate at 75 to 120 × 10⁶ MNC/ml. In some experiments a neutralizing antibody against IL-10 was preincubated with IL-10 for 2 hours at room temperature. Neutralizing antibodies against GM-CSF, G-CSF, IL-3, SCF, or IGF-1 were used as recommended by the manufacturer. Plates were incubated at 37°C, 5% CO₂, and full humidity. After a culture period of 14 days, cultures were examined under an inverted microscope. Aggregates with at least 50 hemoglobinized cells, easily recognizable by their red color, were counted as BFU-E-derived erythroid bursts. For cultivation of BFU-E from normal individuals, 2 U/mL EPO was added to culture dishes.

Semi quantitative reverse transcriptase-polymerase chain reaction (RT-PCR) analysis of GM-CSF transcripts. A total of 2 × 10⁵ PV PBMNCs were cultured in suspension both with and without IL-10 (10 ng/mL) for 48 hours. After incubation, cells were washed twice in diethylpyrocarbonate-treated water and 10⁶ cells aliquots were lysed by addition of 1.6 mL RNAzol B (Biotecx, Houston, TX). Total RNA was extracted as described. The integrity of RNA was controlled by densitometry and the system’s volume integration program (Bio Rad 670 Imaging Densitometer and the system’s volume integration program (Bio Rad Gel DOC 1000 system. Molecular Analyst/PC software; BioRad, Richmond, CA). Potential differences of total cellular RNA/cDNA in PCR analyses were corrected by dividing GM-CSF values by the ABL value (mean value of six PCR analyses). The relative level of GM-CSF transcripts was measured in patient samples in three PCR analyses in duplicate using freshly synthesized cDNA.

Statistical analysis. The t-test was used to determine the significance of differences. A P value of <.05 was considered statistically significant.

RESULTS

Inhibitory effect of IL-10 on autonomous BFU-E growth in patients with PV. As it has been originally shown by Prchal and Axelrad,1 erythroid colonies can grow in methylcellulose cultures containing PBMNCs from patients with PV in the absence of exogenous EPO. Treatment of PV cell cultures with IL-10 resulted in a profound and dose-dependent inhibition of autonomous BFU-E growth (Fig 1). The inhibitory effect of IL-10 became apparent at a concentration of 1 ng/mL and was even more pronounced at higher concentrations. This effect was specific, because a neutralizing anti-IL-10 antibody was able to prevent IL-10-induced suppression of BFU-E growth (Fig 1). The effect of 10 ng/mL IL-10 on autonomous BFU-E growth was investigated in PBMNCs from 5 patients with PV (Table 2). As shown in Table 2, autonomous formation of erythroid colonies greatly varied among different patients but was significantly inhibited by IL-10 in all of them. On average, 10 ng/mL of IL-10-inhibited EPO independent BFU-E growth from PV PBMNCs by 81% (range, 71% to 94%).

Effect of anticytokine antibodies on autonomous BFU-E growth from PV cells. To elucidate the possible mechanism of the inhibitory action of IL-10, we first tried to identify the factor responsible for autonomous BFU-E growth in PV by adding neutralizing antibodies against GM-CSF, IL-3, G-CSF, SCF, and IGF-1 to cell cultures from three patients with marked colony growth (Fig 2). Not unexpectedly the IGF-1 antibody significantly inhibited autonomous BFU-E growth in our FCS containing culture system (patient SA), because IGF-1 has been

### Table 1. Clinical and Laboratory Data in Patients With Polycythemia Vera at Time of Study

<table>
<thead>
<tr>
<th>Patient</th>
<th>Age/Sex</th>
<th>Duration (mo)</th>
<th>RBC (×10¹²/L)</th>
<th>Hct</th>
<th>WBC (×10⁹/L)</th>
<th>Platelets (×10¹²/L)</th>
<th>Spleen</th>
<th>Therapy</th>
</tr>
</thead>
<tbody>
<tr>
<td>HR</td>
<td>82/M</td>
<td>5</td>
<td>6.88</td>
<td>42.3</td>
<td>17.4</td>
<td>594</td>
<td>Enlarged</td>
<td>Phlebotomy</td>
</tr>
<tr>
<td>KJ</td>
<td>75/M</td>
<td>216</td>
<td>6.15</td>
<td>42.1</td>
<td>16.1</td>
<td>340</td>
<td>Enlarged</td>
<td>Phlebotomy</td>
</tr>
<tr>
<td>KA</td>
<td>51/M</td>
<td>1</td>
<td>5.63</td>
<td>43.5</td>
<td>9.9</td>
<td>496</td>
<td>Normal</td>
<td>Phlebotomy</td>
</tr>
<tr>
<td>JH</td>
<td>70/F</td>
<td>1</td>
<td>6.59</td>
<td>46.1</td>
<td>6.1</td>
<td>365</td>
<td>Normal</td>
<td>Phlebotomy</td>
</tr>
<tr>
<td>SA</td>
<td>54/M</td>
<td>9</td>
<td>7.65</td>
<td>45.9</td>
<td>39.4</td>
<td>380</td>
<td>Enlarged</td>
<td>Phlebotomy</td>
</tr>
</tbody>
</table>

Abbreviations: RBC, red blood cells; Hct, hematocrit; WBC, white blood cells.
reported as the major EPO-like activity in FCS. Among the other anticytokine antibodies tested only the anti-GM-CSF antibody significantly decreased spontaneous growth of erythroid colonies in all three patients tested, with a mean inhibition by 52% (range, 42% to 63%). This clearly suggested that GM-CSF and IGF-1 were involved in the spontaneous formation of erythroid colonies from PV PBMNCs in this culture system.

**Effect of exogenous growth factors on IL-10-induced suppression of autonomous BFU-E growth from PV cells.** The fact that IL-10 has been shown to inhibit cytokine synthesis in different cell types of the mononuclear cell fraction led us to hypothesize that inhibition of autonomous BFU-E growth by IL-10 was secondary to IL-10-induced suppression of endogenous release of growth regulatory molecules. If this was the case one would expect that exogenous addition of particular cytokines could at least in part reverse growth inhibition by IL-10. In contrast, restoration of colony growth by exogenous growth factors would not be observed if IL-10 had a direct cytotoxic effect on PV cells. In fact, exogenous GM-CSF and IL-3 significantly restimulated erythroid colony formation in IL-10-treated cultures (Fig 3). All other cytokines were not effective in correcting IL-10-induced growth inhibition. Also IGF-1 alone at concentrations up to 100 ng/mL was not able to restore autonomous BFU-E growth in IL-10 suppressed cultures, nor did it significantly potentiate the effects of GM-CSF when added in combination (Table 3). This observation and the suppressive effect of the anti-IGF-1 antibody in the culture system suggest that IGF-1 is important for EPO-independent erythroid colony formation in PV patients but is not involved in growth inhibition by IL-10.

**Inhibitory effect of IL-10 on GM-CSF expression in PV PBMNCs.** The demonstration of both the antiproliferative action of the anti-GM-CSF antibody on the autonomous BFU-E growth and the restoration of IL-10-induced growth suppression

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**Fig 1.** Dose-dependent inhibitory effect of IL-10 on autonomous BFU-E growth from PV cells and its abrogation by an anti-IL-10 antibody in patient HR. A total of $120 \times 10^3$ PBMNC/mL were cultured in methylcellulose with medium alone, with increasing concentrations of IL-10, or with IL-10 plus a neutralizing antibody against IL-10. Colony growth was assessed after 14 days. Results represent the mean values ± standard deviation (SD) from triplicates. * Significant change from control with $P$ value at least <.05.

**Table 2. Inhibitory Effect of IL-10 on Autonomous BFU-E Growth in PV Patients**

<table>
<thead>
<tr>
<th>Patient</th>
<th>MNC/Dish $\times 10^3$</th>
<th>BFU-E $\pm$ SD/Dish</th>
<th>Percent of Inhibition</th>
<th>$P$</th>
</tr>
</thead>
<tbody>
<tr>
<td>HR</td>
<td>100</td>
<td>32.0 ± 3.5</td>
<td>9.3 ± 1.2</td>
<td>71</td>
</tr>
<tr>
<td>Kj</td>
<td>100</td>
<td>43.0 ± 1.7</td>
<td>10.3 ± 1.5</td>
<td>76</td>
</tr>
<tr>
<td>KA</td>
<td>100</td>
<td>5.0 ± 1.0</td>
<td>0.3 ± 0.6</td>
<td>94</td>
</tr>
<tr>
<td>JP</td>
<td>116</td>
<td>15.8 ± 4.2</td>
<td>3.0 ± 0.0</td>
<td>81</td>
</tr>
<tr>
<td>SA</td>
<td>75</td>
<td>107.3 ± 7.6</td>
<td>19.3 ± 2.5</td>
<td>82</td>
</tr>
</tbody>
</table>

PBMNCs from PV patients were cultured as described in the Materials and Methods with or without 10 ng/mL IL-10. Colony growth was assessed after 14 days. Results represent mean values ± SD from triplicates.

**Fig 2.** Effect of anticytokine antibodies on autonomous BFU-E growth from PV cells in three patients. PBMNCs were cultured in methylcellulose with medium alone or with antibodies against GM-CSF, IL-3, G-CSF, SCF, or IGF-1, respectively. Colony growth was assessed after 14 days. Results represent mean values ± SD from triplicates. * Significant change from control with $P$ value at least <.05.

**Fig 3.** Effect of exogenous growth factors on IL-10-induced suppression of autonomous BFU-E growth from PV cells. PBMNCs were cultured in methylcellulose with medium alone or with antibodies against GM-CSF, IL-3, G-CSF, SCF, or IGF-1, respectively. Colony growth was assessed after 14 days. Results represent mean values ± SD from triplicates. * Significant change from IL-10 treated cultures with $P$ value at least <.05.
Results represent mean values indicated in the Table. Colony growth was assessed after 14 days.

Methylcellulose in the presence or absence of exogenous cytokines as isoenzyme type as that expressed by peripheral blood elements. Colonies from PV patients that formed in the absence of exogenous EPO contained the same G6PD as indicated in the Table. Colony growth was assessed after 14 days. Results represent mean values ± SD from triplicates.

by exogenous GM-CSF strongly suggested that the growth inhibitory effect of IL-10 in PV was at least in part caused by suppression of spontaneous GM-CSF production. Therefore, the effect of IL-10 on GM-CSF expression in unseparated PBMNCs from a patient with PV (HR) was studied by a semiquantitative PCR technique. We have previously shown that the PCR technique used by us allows at least semiquantitative measurements of GM-CSF transcript levels by establishing linear ranges of amplifications of GM-CSF as a function of cycle number and cDNA concentrations. After culturing PV cells in the presence of IL-10 (10 ng/mL) for 48 hours, GM-CSF transcript levels were substantially lower than that of cells kept in suspension without IL-10 (Fig 4A). In comparison, ABL transcripts, which served as a control, remained unchanged during the time of culture (Fig 4A). Figure 4C shows corrected GM-CSF mRNA levels in PV PBMNCs at 48 hours. Comparison of corrected GM-CSF transcript levels between PBMNCs cultured with and PBMNCs cultured without IL-10 showed a mean decrease by 47%.

Effect of IL-10 on EPO-dependent BFU-E growth in normal individuals. To investigate the effect of IL-10 on the EPO-dependent in vitro growth of normal erythroid progenitors, PBMNCs from five healthy volunteers were cultured in the presence of EPO with or without 10 ng/mL IL-10 (Table 4). IL-10-induced suppression of EPO-dependent erythroid colony growth was seen in all five individuals, but this inhibition was less pronounced than in PV patients. The mean inhibition of normal EPO-dependent BFU-E growth by 10 ng/mL IL-10 was 45% ± 12% (standard deviation), which was significantly less than the inhibition of EPO-independent erythroid colony growth in PV patients (81% ± 12%, P < .001).

DISCUSSION

Elegant studies by Adamson et al analyzing the isoenzyme pattern in different cell types of PV patients heterozygous for G6PD have shown that circulating red blood cells, granulocytes, and platelets expressed the same isoenzyme, suggesting that these cells were unicellular in origin and derived from an abnormal multipotential progenitor cell. Later, Prchal et al could show that erythroid colonies from PV patients that formed in the absence of exogenous EPO contained the same G6PD isoenzyme type as that expressed by peripheral blood elements.

Thus, the so-called endogenous colonies arose from the abnormal clone that was responsible for the overproduction of red blood cells in PV. When exogenous EPO was added, increasing numbers of erythroid colonies were formed containing cells that expressed the other G6PD isoenzymes, indicating the existence.

Table 3. Effect of IGF-1 Alone or in Combination With GM-CSF on IL-10-Induced Suppression of Autonomous BFU-E Growth From PV Cells

<table>
<thead>
<tr>
<th>Culture Conditions</th>
<th>BFU-E ± SD/Dish</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control (no cytokines)</td>
<td>96.0 ± 8.2</td>
</tr>
<tr>
<td>+ 10 ng/mL IL-10</td>
<td>12.7 ± 3.1</td>
</tr>
<tr>
<td>+ 10 ng/mL IL-10 + 1 ng/mL IGF-1</td>
<td>10.3 ± 0.6</td>
</tr>
<tr>
<td>+ 10 ng/mL IL-10 + 10 ng/mL IGF-1</td>
<td>14.0 ± 2.7</td>
</tr>
<tr>
<td>+ 10 ng/mL IL-10 + 100 ng/mL IGF-1</td>
<td>13.7 ± 1.5</td>
</tr>
<tr>
<td>+ 10 ng/mL IL-10 + 100 U/mL GM-CSF</td>
<td>83.7 ± 4.2</td>
</tr>
<tr>
<td>+ 10 ng/mL IL-10 + 100 U/mL GM-CSF + 10 ng/mL IGF-1</td>
<td>92.7 ± 24.6</td>
</tr>
</tbody>
</table>

A total of 10^5 PBMNC/dish from patient SA were cultured in methylcellulose in the presence or absence of exogenous cytokines as indicated in the Table. Colony growth was assessed after 14 days. Results represent mean values ± SD from triplicates.

Table 4. Effect of IL-10 on EPO-Dependent BFU-E Growth in Normal Individuals

<table>
<thead>
<tr>
<th>Patient No.</th>
<th>BFU-E ± SD/Dish</th>
<th>Percent of Inhibition</th>
</tr>
</thead>
<tbody>
<tr>
<td>Control</td>
<td>+ IL-10</td>
<td></td>
</tr>
<tr>
<td>1</td>
<td>13.0 ± 5.0</td>
<td>8.3 ± 2.1</td>
</tr>
<tr>
<td>2</td>
<td>33.0 ± 2.7</td>
<td>13.7 ± 10.1</td>
</tr>
<tr>
<td>3</td>
<td>36.0 ± 4.4</td>
<td>23.7 ± 5.0</td>
</tr>
<tr>
<td>4</td>
<td>30.3 ± 2.1</td>
<td>19.0 ± 3.0</td>
</tr>
<tr>
<td>5</td>
<td>53.7 ± 3.5</td>
<td>22.7 ± 2.5</td>
</tr>
</tbody>
</table>

A total of 10^5 PBMNC/dish from 5 healthy volunteers were cultured in the presence of 2 U/mL EPO as described in the Materials and Methods with or without 10 ng/mL IL-10. Colony growth was assessed after 14 days. Results represent mean values ± SD from triplicates.
of both malignant and nonmalignant populations of hematopoietic progenitor cells in PV marrow. Here we show a profound inhibitory effect of IL-10 on endogenous erythroid colony formation in vitro in patients with PV. A significant inhibition of spontaneous erythroid colony formation by IL-10 was observed in all five PV patients investigated. The effect of IL-10 was dose-dependent and specific, because a neutralizing anti-IL-10 antibody was able to prevent IL-10-induced suppression of BFU-E growth.

The mechanism of spontaneous erythroid colony formation in vitro and the factors that lead to in vivo expansion of clonal cells in patients with PV remain to be fully understood. Spontaneous erythroid colony formation in the presence of anti-EPO antibodies (3) and in serum-free culture conditions (2) exclude minute amounts of EPO in the culture system as a potential stimulus for endogenous colony formation. The marked reduction of spontaneous erythroid colony formation by cells from PV patients by depletion of monocytes and its restoration by readdition of adherent cells or adherent cell supernatant\textsuperscript{26,27} strongly suggests monocyte-derived molecules as stimulatory factors for erythroid colony formation in PV. Several cytokines including GM-CSF,\textsuperscript{28} IL-3,\textsuperscript{28} SCF,\textsuperscript{29} and IGF-1\textsuperscript{30} have been shown to enhance EPO-independent BFU-E growth if added to semisolid cultures containing PV MNCs. Moreover a hypersensitivity of PV BFU-E to each of these growth factors compared with normal erythroid progenitors has been shown,\textsuperscript{26,30} which may be a principal factor in the profound marrow hyperplasia and increased blood counts of patients with PV. Recently, a defect in phosphatase activity in PV cells has been proposed as the molecular basis for the enhanced response of PV BFU-E to growth regulatory molecules. Thus, increased basal and induced tyrosine phosphorylation of the IGF-1 receptor $\beta$ subunit in circulating MNCs in such patients has been reported.\textsuperscript{31} Moreover, a diminished enhancement of erythroid colony formation in the presence of orthovanadate,\textsuperscript{32} an inhibitor of protein tyrosine phosphatases, in PV cells has been shown.

There is substantial evidence that the inhibitory effect of IL-10 on autonomous BFU-E growth in PV is indirect. Spontaneous formation of erythroid colonies from PV cells has been shown to be decreased after removal of adherent cells.\textsuperscript{26,27} In unseparated PV PBMCs, we found, in agreement with observation by others,\textsuperscript{33} that autonomous BFU-E growth was markedly reduced by addition of an anti-GM-CSF antibody, suggesting spontaneous release of GM-CSF as an important mechanism of EPO-independent erythroid colony formation. Not unexpectedly, an anti-IGF-1 antibody also inhibited autonomous BFU-E growth in our culture system because IGF-1 has been shown to be the major EPO-like activity in FCS.\textsuperscript{23} The antiapoptorative action of anti-GM-CSF antibody on EPO-independent BFU-E formation in PV and the fact that IL-10 has been shown to inhibit cytokine synthesis, including GM-CSF in human monocytes,\textsuperscript{13} led us to hypothesize that inhibition of BFU-E growth by IL-10 was mainly caused by IL-10-induced suppression of endogenous GM-CSF release. Our observation that exogenous GM-CSF was able to, at least in part, reverse growth inhibition by IL-10 strongly supports this concept. In fact, analysis of the effect of IL-10 on GM-CSF expression in MNCs from a patient with PV clearly showed that the IL-10-induced inhibition of erythroid colony growth correlated with a substantial decrease in GM-CSF transcripts in PV PBMCs. Exogenous IL-3 also restimulated autonomous BFU-E formation in IL-10-treated PV cell cultures, but an anti-IL-3 antibody had no antiproliferative effect on spontaneous growth of erythroid progenitors. These data suggest that in addition to GM-CSF-responsive PV BFU-E, there seems to exist an IL-3-responsive population, which however, cannot be sufficiently stimulated by the endogenously released growth factors in unstimulated cultures.

The inhibitory effect of IL-10 on hematopoietic colony formation is not specific for PV BFU-E. However the growth suppression of IL-10 on EPO-independent erythroid colony growth in PV seems to be more pronounced than that on EPO-dependent normal BFU-E. In contrast to a mean growth inhibition by 81% on autonomous BFU-E formation from PV PBMCs, IL-10 decreased the number of EPO-stimulated BFU-E by approximately 45% in PBMC cultures from normal individuals. Moreover, the inhibitory action of IL-10 is not restricted to the erythroid lineage, because we have also shown IL-10-induced inhibition of spontaneous formation of myeloid colonies in normal individuals\textsuperscript{15} and patients with CMML\textsuperscript{16} through suppression of endogenous GM-CSF release. Thus, the inhibition of GM-CSF synthesis by IL-10 makes it useful in indirectly modulating hematopoiesis in a variety of conditions. Despite its well-defined effects in vitro, the in vivo role of GM-CSF in normal and abnormal hematopoiesis is much less clear. Mice deficient in GM-CSF (GM$^{-/-}$) through homologous recombination in embryonal stem cells show no alterations in hematopoiesis\textsuperscript{34} suggesting substantial overlap of function with other colony-stimulating factors. On the other hand, animals transplanted with cells containing the GM-CSF cDNA developed a fatal syndrome resembling myeloproliferation, which however, was non-neoplastic in nature.\textsuperscript{35} In PV patients it is not known if GM-CSF, as in the in vitro system, plays a significant role in the expansion of clonal cells. If this is the case, therapy with IL-10, a cytokine that has been well tolerated in first clinical trials,\textsuperscript{36,37} could be considered as a strategy to suppress the excessive production of red blood cells in PV.
potent growth and differentiation factors for activated human B lymphocytes. Proc Natl Acad Sci USA 89:1890, 1992
Interleukin-10 Inhibits Erythropoietin-Independent Growth of Erythroid Bursts in Patients With Polycythemia Vera

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