Risk Factors for Adult T-Cell Leukemia Among Carriers of Human T-Lymphotropic Virus Type I

By Michie Hisada, Akihiko Okayama, Shigemasa Shioiri, Donna L. Spiegelman, Sherri O. Stuver, and Nancy E. Mueller

ADULT T-CELL LEUKEMIA (ATL) is caused by human T-lymphotropic virus type I (HTLV-I) infection, generally many decades after initial infection. It is believed that carriers who have acquired infection early in life are at greatest risk of ATL. One of the features of ATL is the unique morphology of leukemic cells with an indented or convoluted nucleus (“flower cell”). The level of flower cells is a marker to distinguish clinical subtypes of ATL and to monitor progression from smoldering to overt ATL. Asymptomatic HTLV-I carriers often have a very low, but detectable, level of circulating abnormal lymphocytes (Ably) that closely resemble the flower cells.

In the population-based Miyazaki Cohort Study, we have found that the presence and levels of Ably is highly correlated with HTLV-I proviral load. The presence of Ably also is associated with a greater proportion of CD4+/CD25+ cells, a predominant phenotype of ATL cells. Moreover, persistent proliferation of HTLV-I–infected clones has been shown primarily in CD4+ cells in asymptomatic carriers with a high proviral load. These data suggest that HTLV-I–infected T-cell clones are likely expanding among Ably-positive carriers and that the presence of Ably may correlate with the risk of ATL. High levels of Ably in the presence of the symptoms of immune suppression has been shown to predict the subsequent development of ATL. However, it is uncertain if low levels of Ably among asymptomatic HTLV-I carriers predict the onset of ATL.

Although the precise mechanism of HTLV-I oncogenesis is unknown, the enhanced expression of Tax regulatory protein, a viral gene product that transactivates transcription of viral mRNA and other host genes that control cell proliferation, is believed to play a central role in HTLV-I oncogenesis. However, the level of Tax mRNA has been shown to be low in ATL cells, which has led to the hypothesis that a continuous expression of Tax gene product may not be required for the maintenance of the malignant phenotype. This theory is supported by our observation of a lower prevalence of antibody to the Tax regulatory protein (anti-Tax) among ATL cases compared with asymptomatic HTLV-I carriers. A lower prevalence of anti-Tax also has been found in the subset of HTLV-I carriers with a detectable level of Ably (>0.6%) compared with those without Ably, suggesting that anti-Tax reactivity may be low among those at risk of ATL before their clinical diagnosis.

In this prospective study, we describe changes in prediagnostic viral markers of ATL cases and examine risk factors for ATL in a matched case-control study nested within the Miyazaki Cohort Study.

MATERIALS AND METHODS

Study subjects. The Miyazaki Cohort Study was established in 1984 in two HTLV-I endemic villages in Miyazaki Prefecture, Japan. Nearly 27% of 1,960 cohort members enrolled as of August 1996 are HTLV-I seropositive at baseline. The cohort has been followed in the context of free health examinations offered annually by the government for those aged 40 or older. Those younger than 40 years who come to these examinations also are enrolled in the study. The annual screens consist of a physical examination, other routine health examinations, and blood tests. A detailed baseline questionnaire collects demographic data, as well as information on alcohol and smoking status. A shorter instrument completed at each follow-up screening updates information on interval health history and symptoms and current alcohol and smoking behavior. The study protocol was approved by the Institutional Review Boards of the Miyazaki Medical School and the Harvard School of Public Health. Informed consent was obtained from all study subjects.

The subjects of this analysis are the five HTLV-I carriers who developed ATL during the follow-up through December, 1995, and 38 HTLV-I–positive controls. ATL cases were identified through annual census or reports from next-of-kin. The diagnosis was confirmed by medical records for three of the five cases; two others were confirmed by reports from the local government nurses. Using a nested case-control sampling, the controls were selected from within the cohort among HTLV-I carriers who were alive and free of ATL diagnosis when...
Peripheral blood smears were obtained from either the ear lobe or a finger prick, by smearing one drop onto a glass slide. The slides were fixed by methanol and stained with Giemsa. All blood smears were read blinded to ATL diagnosis. The identification of Ably followed the criteria by Kondo et al. The number of Ably among 500 leukocytes was selected for case-control comparison in the present study were retested during the follow-up. Because the standard method of viral marker measurement has changed over time, all sera from the screens were available for subjects who attended more than one screening during the follow-up. Because the standard method of viral marker measurement has changed over time, all sera from the screens selected for case-control comparison in the present study were retested for anti-HTLV-I titer and anti-Tax reactivity in 1996 to attain internal consistency.

Laboratory methods. For each serum sample, positivity of anti-HTLV-I was tested by passive particle agglutination assay (Serodia-HTLV, Fujirebio, Tokyo, Japan) at a dilution of 1:100, 1:500, 1:2500, or higher (+ + + +, + + +, +, or ±), where a lower dilution corresponds to a higher anti-Tax reactivity. Multiple measurements of viral markers were available for subjects who attended more than one screening during the follow-up. Because the standard method of viral marker measurement has changed over time, all sera from the screens selected for case-control comparison in the present study were retested for anti-HTLV-I titer and anti-Tax reactivity in 1996 to attain internal consistency.

Statistical analysis. The mean age, leukocyte count, Ably level, and geometric mean of anti-HTLV-I titer were compared between cases and controls using Wilcoxon signed-rank test. The prevalence of smoking and the presence of anti-Tax reactivity were compared using McNemar’s test. The odds ratio (OR) and 95% confidence interval (CI) for the association of ATL diagnosis with viral markers and Ably level were estimated from exact conditional logistic regression (LOGXACT, 1996, Cytel, Cambridge, MA). For cases with multiple observations, the first measurement of HTLV-I viral markers, Ably, and smoking status were used for calculation of the ORs. In the logistic regression models, anti-HTLV-I titer by serial twofold dilution and anti-Tax reactivity were treated as ordinal categorical variables with 10 and five levels, respectively. Leukocyte count, Ably level, and age were treated as continuous variables. Smoking status was dichotomized, with current- and ex-smokers combined as ever-smokers. Statistical significance was based on two-sided tests.

RESULTS

Five ATL cases (3 men, 2 women) developed during the study period. One additional patient was diagnosed with ATL before his enrollment in the study. The characteristics of these ATL cases are summarized in Table 1. The mean age at death of these six cases was 73 years. Samples had been taken from these cases up to 10 years preceding their death. Four of the five incident cases had multiple measurements of prediagnostic viral markers, which showed virtually no change in level over time. Anti-Tax reactivity was relatively low or undetectable for all cases. Only one case (case 3) had detectable level of Ably (>0.6%) before diagnosis. All male cases were long-term smokers, whereas both female cases were never-smokers.

Because ongoing therapy may have modified the levels of Ably and of viral markers, the prevalent ATL case (case 1) was excluded from further analysis. The characteristics of the five ATL cases as compared with the 38 controls are summarized in Table 2. After accounting for the matching factors, neither the geometric mean of anti-HTLV-I titer (1,782 v 467, P = .31) nor prevalence of anti-Tax reactivity (20% v 32%, P = .57) was different between cases and controls. The prevalence of ever-smokers and the mean leukocyte count were also similar for cases and controls. The mean Ably level was slightly, but not significantly, higher among cases compared with controls (0.36% v 0.24%, P = .13). Table 3 shows the distribution of anti-Tax reactivity among cases and controls by their anti-HTLV-I titer. Among controls, the proportion of anti-Tax positivity increased with anti-HTLV-I titer level. The positive correlation between the levels of anti-HTLV-I titer and anti-Tax reactivity were comparable to those in the entire study cohort of HTLV-I carriers (Table 3). In contrast, there was no apparent correlation between these two viral markers among ATL cases.

Anti-HTLV-I titer was associated with ATL diagnosis (OR = 1.6 per twofold dilution; 95% CI 0.95, 3.8), albeit not to a statistically significant degree (P = .09) (Table 4). Anti-Tax was not predictive (OR = 0.78; 95% CI 0.26, 1.7). With mutual adjustment for anti-HTLV-I titer and anti-Tax reactivity, the OR for anti-HTLV-I titer and anti-Tax reactivity was 1.6 (95% CI 0.95, 3.5) and 0.74 (95% CI 0.21, 1.8), respectively. Smoking and leukocyte counts were not significant predictors of ATL. We were unable to obtain a stable estimate for the association of Ably level and the risk of ATL in the logistic regression analysis due to sparse data.

DISCUSSION

The rare occurrence and long latency period of ATL has posed substantial difficulties in community-based, prospective...
RISK FACTORS FOR ATL AMONG HTLV-I CARRIERS

The mean age of ATL mortality, 73 years, was much higher than the national average of 57 years. Studies of this malignancy, the present study examined the characteristics of ATL cases in a well-defined adult population endemic for HTLV-I infection. The mean age of ATL mortality, 73 years, was much higher than the national average of 57 years among the general Japanese population, reflecting the high average age (60 years) of the HTLV-I-positive cohort members in the Miyazaki Cohort.

Our analysis found a 1.6-fold increase in the risk of ATL per twofold increase in anti-HTLV-I titer, indicating that those carriers with the highest titer (≥1:8,192) will have an approximately 70-fold risk of developing ATL relative to those with the lowest titer level (1:16). Given a strong positive correlation between anti-HTLV-I and proviral load among asymptomatic carriers in our cohort, the observed association of anti-HTLV-I with ATL diagnosis may imply a similar association of proviral load with this malignancy.

As would be predicted from our previous finding of a lower prevalence of anti-Tax reactivity among ATL cases, the analysis of prediagnostic sera of the ATL cases in the present study showed that anti-Tax reactivity was low or undetectable for all cases for up to 10 years preceding their diagnosis. Given the observed higher anti-HTLV-I titer among the ATL cases compared with asymptomatic carriers, as well as our previous report of a strong correlation between anti-HTLV-I titer and anti-Tax positivity among asymptomatic carriers, it seems possible that a loss of anti-Tax occurs in the process of ATL development. Those carriers at risk of ATL also may be inherently more likely to have low anti-Tax reactivity.

Table 2. Comparison of Demographic Characteristics and HTLV-I Viral Markers Between Five ATL Cases and 38 Age- and Sex-Matched Controls in the Miyazaki Cohort Study, Adjusted for the Matching Factors

<table>
<thead>
<tr>
<th>Case (n = 5)</th>
<th>Controls (n = 38)</th>
<th>P Value</th>
</tr>
</thead>
<tbody>
<tr>
<td>Age (mean, years)*</td>
<td>79</td>
<td>76</td>
</tr>
<tr>
<td>Smoker†</td>
<td>3 (60%)</td>
<td>18 (47%)</td>
</tr>
<tr>
<td>Ably level (mean, %)§</td>
<td>0.36</td>
<td>0.24</td>
</tr>
<tr>
<td>Leukocyte count (mean, /mL)‡</td>
<td>4,200</td>
<td>5,300</td>
</tr>
<tr>
<td>Geometric mean anti-HTLV-I titer$</td>
<td>1,782</td>
<td>467</td>
</tr>
<tr>
<td>Positivity for anti-Tax reactivity§</td>
<td>1 (20%)</td>
<td>12 (32%)</td>
</tr>
</tbody>
</table>

*Age of subjects was determined at the death of the index case.
†Viral markers and smoking status were determined at the first screen at which data were available for the index case.
§The mean value was based on data from the screens attended by the index case.

Table 3. Distribution of Anti-Tax Reactivity by the Level of Anti-HTLV-I Titer Among Cases, Controls, and All HTLV-I Carriers in the Study Cohort

<table>
<thead>
<tr>
<th>Anti-HTLV-I Titer</th>
<th>No. of Anti-Tax Positive Subjects (%)</th>
</tr>
</thead>
<tbody>
<tr>
<td>Case (N = 5)</td>
<td>Control (N = 38)</td>
</tr>
<tr>
<td>4,096 ≤ titer</td>
<td>0/2 (0%)</td>
</tr>
<tr>
<td>1,024 ≤ titer ≤ 2,048</td>
<td>1/1 (100%)</td>
</tr>
<tr>
<td>128 ≤ titer ≤ 512</td>
<td>2/2 (100%)</td>
</tr>
<tr>
<td>16 ≤ titer ≤ 64</td>
<td>0/0 (UD)</td>
</tr>
</tbody>
</table>

For cases and controls, cross-classification was by the observation of viral markers at the first screen at which data were available for the index case.

Table 4. Univariate Association of ATL With Viral Markers, Smoking, and Leukocyte Count Using Exact Conditional Logistic Regression

<table>
<thead>
<tr>
<th>Variables*</th>
<th>OR</th>
<th>95% CI</th>
</tr>
</thead>
<tbody>
<tr>
<td>Anti-HTLV-I titer</td>
<td>1.6</td>
<td>0.94-3.8</td>
</tr>
<tr>
<td>Anti-Tax reactivity</td>
<td>0.78</td>
<td>0.26-1.7</td>
</tr>
<tr>
<td>Leukocyte count</td>
<td>0.73</td>
<td>0.40-1.2</td>
</tr>
<tr>
<td>Ever-smoker</td>
<td>1.7</td>
<td>0.04-198.4</td>
</tr>
</tbody>
</table>

Abbreviations: OR, odds ratio; CI, confidence interval.

*The ORs for anti-HTLV-I titer and anti-Tax reactivity are for 1 unit increase in level; smoking status is dichotomous. Referent categories are: anti-HTLV-I titer 1:16, anti-Tax reactivity (−), and never-smokers, respectively. Leukocyte count is a continuous variable.
was clearly limited by the relatively small number of observations.

In sum, the present analysis suggests that HTLV-I carriers with a higher anti-HTLV-I titer are at greatest risk of ATL and that the level of anti-HTLV-I and anti-Tax reactivity is discordant before diagnosis. Additional analysis of Tax mRNA expression, proviral load, HTLV-I clonality, as well as direct measurement of CTL response is needed to provide further insights into the oncogenic process of this virus infection. Investigation of correlates of the host immune response in this population in relation to HTLV-I viral markers may be useful to shed light on the association between host factors and the risk of ATL.

REFERENCES

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