RAPID COMMUNICATION

High Efficiency Adenovirus-Mediated Gene Transfer to Human Dendritic Cells

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The interest in the use of human dendritic cells in cancer immunotherapy calls for efficient ex vivo methods of dendritic cell education. To extend the range of methods available, we generated phenotypically characteristic dendritic cells from peripheral blood monocytes incubated with granulocyte-macrophage colony-stimulating factor and interleukin-4 and infected them with an adenovirus containing a humanized version of green fluorescent protein as a marker of gene expression. The levels of expressed protein were high, but they were further increased in combination with cationic liposomes. In comparison to transfection efficiency of the homologous expression plasmid, adenovirus-mediated gene transfer was substantially more efficient. With the aid of liposome-mediated infection, gene transfer into CD83+ dendritic cells was highly effective, resulting in more than 90% of the cells expressing the transgene.

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mL conical tubes, washed twice with 50 mL of PBS, and centrifuged at 250g for 10 minutes. The cells were washed with 50 mL cold PBS (PBS with 0.5% bovine serum albumin and 2 mmol/L EDTA), counted, and assayed for viability by trypan blue exclusion.

Peripheral blood mononuclear cells (5 × 10^8 total cells) were used as the source for immunomagnetic isolation of CD14^+ leukocytes according to the manufacturer’s instructions (Miltenyi, Auburn, CA). CD14^+ cells (8 × 10^7) were resuspended at 5 × 10^6 cells/mL in serum-free RPMI-1640 for 1 hour at 37°C. The nonadherent cells were discarded and the attached cells were rinsed with PBS (Fig 1A). Complete medium was supplemented with 1,000 IU/mL of GM-CSF and 1,000 IU/mL IL-4. Within 48 hours, loosely attached grapelike clusters formed with some remaining adherent cells (Fig 1B). Two days after plating with cytokines, the cells were suspended at 2 × 10^6 cells/mL of complete medium with GM-CSF and IL-4 and incubated for an additional 72 hours. Cells were harvested by vigorous pipetting, counted, and used for flow cytometry or gene transfer. At this stage, cells were more than 92% viable as determined by trypan blue exclusion.

**Adenovirus-mediated gene transfer.** For gene transfer, we used the recombinant adenovirus Ad5RSVGFP alone, Ad5RSVGFP in combination with cationic liposomes,^27^ the expression plasmid pRSVGFP containing the adenovirus expression cassette, and pRSVGFP with cationic liposomes. Ad5RSVGFP contained a humanized version of the green fluorescent protein (GFP; GFP-S65T; Clontech, Palo Alto, CA) under the control of the Rous Sarcoma virus (RSV) promoter. Ad5RSVGFP and pRSVGFP were provided by the University of Iowa Gene Transfer Vector Core (Iowa City, IA). The adenovirus contained 1.1 × 10^9 particles/µL and was stored at −70°C until needed.

For infection, AdRSVGFP was prepared by suspending 7.5 × 10^9 particles (5,000 particles per cell) in 50 µL total serum free medium (Opti-MEM; GIBCO BRL). Meanwhile, 1.25 µg cationic liposomes

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**Fig 1.** Morphology of isolated cells in culture. (A) Purified adherent monocytes used for initiation of dendritic cells. (B) After 48 hours in culture with GM-CSF and IL-4, monocytes clustered into aggregates typical of dendritic cell precursors.
Lipofectamine; GIBCO BRL) were suspended in 50 l. serum-free medium. For experiments without liposomes, the adenovirus was suspended in 100 l. serum-free medium. For plasmid transfections, we used the equivalent cell numbers, liposome amounts, and virus expression equivalents (5,000 plasmids per cell) as for Ad5RSVGFP infection. The vector and liposome solutions were mixed and incubated for 15 minutes.

For gene transfer experiments, purified dendritic cells were centrifuged at 250g for 10 minutes and the medium was carefully removed. The cells (1.5 × 10⁶ cells for each trial) were resuspended in 100 l. of serum-free medium and combined with the vector or vector/liposome. The suspension was gently mixed and incubated for 105 minutes at 37°C. (The final adenovirus/cell ratio was 5,000 virus particles per cell, resulting in a multiplicity of infection factor of about 50.) After incubation, the cells were centrifuged, the supernatant was removed, and the cells were resuspended at 0.5 × 10⁶ cells/ml in complete medium containing 400 IU/mL GM-CSF and 200 IU/mL TNF-α. The cells were incubated at 37°C for 72 hours before analysis for GFP expression by flow cytometry and fluorescence microscopy.

Flow cytometric characterization of dendritic cells. The purity of dendritic cells was assessed by flow cytometry with the aid of a FACScan cytometer (Becton Dickinson, Sunnyvale, CA) at the Mayo Flow Cytometry Core Facility. All immunoreagents were obtained commercially and were used with appropriate isotype controls. The reagent for CD56 was obtained from Becton Dickinson (San Jose, CA); for CD14, CD19, CD1a, CD54, CD3, CD4, CD11c, fluorochrome isothiocyanate (FITC) isotype control, phycoerythrin (PE) isotype control, and HLA-DR from BioSource International (Camarillo, CA); for CD83 and CD64 from Coulter (Hialeah, FL); and for CD86 from Ancell (Bayport, MN). For each analysis, 100,000 cells in 100 l. PBS were incubated for 30 minutes on ice with the manufacturer’s recommended amount of PE-labeled or FITC-labeled antibody. After incubation, the cells were washed from the plates and combined with the vector or vector/liposome. Additionally, dendritic cells were visualized with an IM35 Zeiss fluorescent microscope equipped with the fluorescein filter set and photographed with ISO 200 speed Royal Gold or PJM Multispeed film (Kodak, Rochester, NY).

All experiments were performed at least in duplicate with similar results.

RESULTS

Dendritic cells were cultured from monocytes by a technique that required minimum manipulation and resulted in high yields. Positive isolation of monocytes by magnetically labeled CD14-specific antibodies increased dendritic cell purity without reducing cell yield. Starting with 5 × 10⁶ peripheral blood mononuclear cells, we typically ended up with more than 3 × 10⁶ dendritic cell precursors. Figure 2A shows the results of flow cytometric analysis of cells after incubation with GM-CSF and IL-4. The cells contained markers typical of dendritic cell precursors and CD3 (T cells), CD19 (B cells). Significantly, the cells were negative for CD83, a marker of mature dendritic cells. Some dendritic cells in this system expressed CD14 upon development of the ability to adhere, although this has not been consistently documented. In addition to adherent cells, the culture contained some multicell aggregates typical of proliferating dendritic cells (Fig 1B).

Incubation with GM-CSF and TNF-α transformed the immature dendritic cells into veiled cells, strongly adherent cells with dendritic appendages, and some remaining cell clusters. Flow cytometric analysis indicated an increase in expression of CD83, HLA-DR, CD54, and CD86 (Fig 2B). Although the cells were still CD1a⁺ and CD11c⁺, the level of expression of these markers was reduced. CD83 has been characterized as a unique cell surface marker of mature dendritic cells. An analysis of cell size (forward scatter) and cell geometry (side scatter) showed a discrete population of larger and less regular cells (Fig 2A). In this discrete population, we found that more than 50% of the cells were CD83⁺ (Fig 2B). Significantly, more than 90% of cells in this population were also HLA-DR⁺ (Fig 2B). Thus, the large cells in the gated region were a population of mature dendritic cells.

To determine the role of the recombinant adenovirus in GFP expression in dendritic cells, we transfected the cells before maturation with pRSVGFP, the plasmid contained in Ad5RSVGFP (Fig 3B), and with pRSVGFP in the presence of liposomes (Fig 3C). Also, we infected the cells with the equivalent (in comparison to pRSVGFP) amount of Ad5RSVGFP in the absence (Fig 3D) or in the presence of liposomes (Fig 3E).

Whereas cells treated with pRSVGFP alone (Fig 3B) were no more fluorescent than untreated controls (Fig 3A), inclusion of liposomes in the transfection medium resulted in GFP expression in some 5% of cells (Fig 3C). Interestingly, the equivalent amount of Ad5RSVGFP alone resulted in more successful gene transfer; about 20% of dendritic cells expressed GFP (Fig 3D). Adenovirus combined with liposomes was the most effective, resulting in approximately 90% of cells expressing GFP (Fig 3E). In control cells (Fig 3A) and cells transfected with the plasmid (with or without liposomes; Fig 3B and C), the mean fluorescence intensity, a measure of the average level of GFP expression per cell, was low and indistinguishable among the groups. However, cells infected with the adenovirus (Fig 3D) were characterized by the mean fluorescence intensity 2.5 times above control and the cells treated with the adenovirus and liposomes (Fig 3E) were characterized by the mean fluorescence intensity 8.5 times above control. Clearly, both adenovirus infection and liposome treatment increased the levels of transgene expression.

To determine whether cells expressing GFP were mature dendritic cells, we analyzed CD83 and HLA-DR fluorescence, respectively, versus GFP fluorescence. Of CD83⁺ cells, 90% expressed GFP (Fig 4C). Similarly, of HLA-DR⁺ cells, some 90% expressed GFP (Fig 4D). Thus, the GFP⁺ cells harbored the two characteristic markers of dendritic cells. GFP expression was observed in adherent, loosely adherent, and nonadherent cells (Fig 5). The cells were morphologically typical: the unattached cells contained small pseudopodia and the attached cells displayed long bifurcated appendages. Thus,
DISCUSSION

The purpose of this work was to establish a technique for high efficiency gene transfer into human dendritic cells derived from peripheral blood. We used expression of GFP as proof of infection and effective expression of the transgene. To increase the efficiency of transgene expression, we applied the recently described liposome-enhanced adenovirus infection to human dendritic cells. Adenoviruses alone can infect these cells, but they are rather ineffective (i.e., they need high levels of multiplicity of infection). By the use of liposomes, we dramatically increased infection efficiency.

Although the adenovirus combined with liposomes was highly infective, it was unclear whether the virus contributed to infection and transgene expression had no apparent effect on dendritic cell morphology and viability.
gene expression in comparison to the plasmid transfection in the presence of liposomes. To resolve the role of the adenovirus, we studied gene expression in cells transfected with the plasmid containing the adenovirus expression cassette. In cells transfected with the plasmid alone, there was virtually no transgene expression. Liposomes did induce measurable expression, but it was more than one order of magnitude below the level obtained with the adenovirus/liposome combination. Clearly, the role of the adenovirus was critical even when the transgene was transferred across the cell membrane by the liposome-mediated mechanism.\(^{27}\) The practical consequence of this finding is that one can achieve superinfectivity with rather low amounts of adenovirus. Thus, the high levels of gene expression achieved by the use of adenovirus in combination with liposomes make this system well suited for antigen expression and presentation.

GFP expression by cells infected by the adenovirus alone indicates that dendritic cells harbor CD51 (integrin-\(\alpha_v\)), the molecule critical for effective adenovirus infection, or its

Fig 3. GFP expression in human dendritic cells infected with pRSVGFP and Ad5RSVGFP in the absence and the presence of cationic liposomes. Cells in the gated region (Fig 2A) were analyzed for GFP fluorescence. They were untreated (A), treated with pRSVGFP in the absence (B) and in the presence of liposomes (C), or infected with Ad5RSVGFP in the absence (D) and in the presence of liposomes (E).

Fig 4. Expression of CD83 (A and C) and HLA-DR (B and D) versus green fluorescence in uninfected cells (A and B) and cells infected with Ad5RSVGFP in the presence of liposomes (C and D). The vertical and horizontal lines distinguish CD83\(^{+}\) and HLA-DR\(^{+}\) cells (left of and below the line, respectively) from CD83\(^{-}\) and HLA-DR\(^{-}\) cells.
functional equivalent. In a murine dendritic cell line, GM-CSF increased the expression of CD51. Expression of CD51 as part of the functional integrin $\alpha_v\beta_3$ and/or $\alpha_v\beta_5$ raises questions about the role of vitronectin binding integrins in the biology of dendritic cells and particularly in their maturation. In mice, vitronectin receptor was found in thymocytes, splenocytes, and bone marrow cells. In thymocytes, expression of vitronectin receptor depended on the stage of development. Expression of vitronectin receptors in dendritic cells may help colocalize these cells with developing T cells in the course of immune system education. This hypothesis is in line with the observation that blocking adhesion also blocks T-cell proliferation. Work is underway in our laboratory to determine the role of CD51 in human dendritic cells.

A salient feature of this work is that liposomes enhanced the adenovirus-mediated gene expression without any apparent interference with dendritic cell maturation; high levels of CD83 and HLA-DR were measured in infected cells. Because CD83 is highly expressed in adenovirus/liposome-treated cells (this work) and coexpressed with CD1a,b,c, class I, class II, CD80, and CD86, it is likely that the adenovirus/liposome-treated cells contain all the factors required for their function in immunity.

The ability of antigen-presenting dendritic cells to elicit a specific immune response has stimulated interest in the use of dendritic cells as adjuvants in immunotherapy of tumors and of autoimmune and infectious diseases. Currently, clinical trials are conducted to evaluate the usefulness of ex vivo educated dendritic cells in therapy of B-cell lymphoma and prostate cancer, whereas laboratory observations promise that this treatment modality will soon be extended to other malignant
diseases. Thus, the high efficiency gene transfer and transgene expression by dendritic cells can extend the range of potential targets for dendritic cell mediated immunotherapy. To investigate this hypothesis, we are studying antigen-dependent stimulation of the T-cell response by transgene expressing dendritic cells.

The technique that we have described is rapid and requires only a single additional manipulation step to the standard dendritic cell isolation protocol. It results in high levels of dendritic cells under the conditions that we have established. In all cases, we have observed high levels of transgene expression and increased antigenic presentation to T cells. We are also exploring the potential for ex vivo expansion of dendritic cells isolated from human bone marrow to further enhance their therapeutic potential.

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