Acute promyelocytic leukemia (APL) is a neoplasm with the unique chromosomal translocation t(15;17), which involves the retinoic acid receptor α gene. All-trans retinoic acid (ATRA) has been used for APL patients as a potent therapeutic agent to induce differentiation of leukemia cells. Although polymorphonuclear leukocytes (PMNs) appearing in the blood and bone marrow during ATRA treatment often possess Auer rods, indicating their neoplastic origin, other morphological abnormalities of PMNs have not been elucidated. We studied the morphological changes of APL cells during ATRA treatment at the ultrastructural level. Although most aberrant primary granules, including Auer rods, became morphologically normal in response to ATRA therapy and the nuclei showed chromatin condensation and lobulation, resulting in the emergence of PMNs, the lobulated nuclei often had nuclear filamentous connections and/or nuclear blebs, indicating some pathological process. Furthermore, PMNs, particularly early in ATRA treatment, lacked neutrophil secondary granules as did the PMNs appearing in a culture of APL cells incubated with ATRA, findings consistent with previously reported data that acute myeloid leukemia cell lines do not produce secondary granule proteins even after induction of differentiation towards mature neutrophils. The present data indicate that ATRA is incapable of inducing complete morphological maturation of APL cells and that secondary-granule deficiency may be a hallmark of aberrantly differentiated leukemic cells.

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shaped as cellular maturation proceeds. The fourth maturation stage is characterized by maturation of the nuclei, as shown by heterochromatin condensation and lobulation, involution of protein synthesis-associated cell organelles, and an increase in the number of glycogen particles, but no clearcut distinction between this and the preceding stage can be made in terms of cytoplasmic granules and MPO reactivity.

Previous electron microscopic studies with ultrastructural cytochemistry for MPO have revealed various abnormalities in acute myelogenous leukemia (AML), including the absence of secondary granules, primary granules, or MPO activity in primary granules, not only in immature leukemic cells but also in mature PMNs. However, it has not been elucidated whether these abnormal PMNs are cells of neoplastic origin. To resolve this question, the PMNs that appear during ATRA treatment of APL seem to be a good subject to study because they have been shown to be derived from leukemic clones. Except for the Auer rods that are occasionally observed in the PMNs of APL patients during ATRA treatment, no morphological abnormalities of mature neutrophils have been reported, because, to our knowledge, no studies have been conducted on ultrastructural aspects of differentiating APL cells. To identify morphological changes of APL cells during the differentiation process induced by ATRA and to determine whether ATRA-induced maturation of APL cells follows the normal morphological differentiation pathway, we investigated the ultrastructural characteristics of APL cells in bone marrow and in cultures treated with ATRA.

**MATERIALS AND METHODS**

**Patients and cells.** Three patients, two with a diagnosis of M3 (patients no. 1 and 2, 55- and 71-year-old men, respectively) and one with the M3 variant (patient no. 3, a 43-year-old woman), were studied. Patient no. 3 had an elevated leukocyte count of 111,100/µL with 97% of blasts and promyelocytes, whereas patients no. 1 and 2 had marked leukopenia without obvious leukemic cells in their bone marrow; although 77% and 78%, respectively, of the cells in their bone marrow were blasts and promyelocytes. Chemotherapy, including cytosine arabinoside, was performed from the time of diagnosis in patients no. 1 and 3 for 7 and 5 days, respectively, but failed to induce remission. Then, with a 2-week interval without treatment of either patient with any antileukemic drugs, 60 mg/d ATRA was orally administered continuously to patients no. 1 and 3 for 36 and 50 days, respectively. In patient no. 2, 70 mg/d ATRA was orally administered from the time of diagnosis for 58 days continuously. Hematological abnormalities improved after ATRA treatment, and administration of ATRA was continued until complete remission was obtained in all patients (5 to 8 weeks, depending on the patient). After the patient gave informed consent, peripheral blood and/or bone marrow aspirates were obtained for light and electron microscopic examinations at the time of bone marrow evaluation at the onset and during ATRA treatment (after 1 to 8 weeks of the treatment).

Although the number of leukemia cells in the bone marrow in patients no. 1 and 3 was reduced by the initial chemotherapy, the morphology of the leukemia cells remained unchanged. In addition, as described below, similar morphological changes in APL cells in response to ATRA were observed in all three patients. These findings indicate that the chemotherapy performed before ATRA treatment in patients no. 1 and 3 did not significantly affect our morphological evaluation of APL cells.

**Cell culture methods.** T cells were depleted from mononuclear cell fractions obtained from heparinized peripheral blood of patient no. 3 by using the two-cycle Ficoll-Hypaque procedure as previously described, the second after the formation of E-rosettes to remove T cells. These cells were cultured overnight at 37°C in suspension in α minimum essential medium (αMEM; GIBCO, New York, NY) containing 20% fetal calf serum (FCS; growth medium) to remove monocytes by plastic adherence. The leukemic cell populations thus
obtained were washed in αMEM and cultured at a concentration of
10⁶ cells/mL in 24-mm Linbro tissue culture multwell plates (Flow
Laboratories, McLean, VA) in 1 mL of growth medium containing
10⁻⁶ mol/L ATRA (Sigma, St Louis, MO) or 10 ng/mL granulocyte
colony-stimulating factor (G-CSF; Chugai, Tokyo, Japan). For light
microscopic examination, an aliquot of the cell suspension was taken
from the culture daily, and cytopsin specimens were prepared. For
electron microscopic examination, the cells were obtained on day 7
of culture and resuspended in FCS. They were then centrifuged, and
the cell pellets were fixed by gently replacing the supernatant with
the fixative as described below.

Bone marrow mononuclear cells obtained from two patients with
idiopathic thrombocytopenia and one patient with nonhematologic
disease were used as controls. The cell separation procedure using
Ficoll-Hypaque almost completely eliminated mature neutrophils
from the samples. These cell populations were cultured with 10⁻⁶
mol/L ATRA in the same manner as described above and examined
for morphological changes under both light and electron micro-
scopes.

Light microscopic observation. Bone marrow and peripheral
blood smears and cytopsin specimens of cultured cells were stained
with Wright-Giemsa, and morphological changes of leukemia cells
were observed under a light microscope.

Electron microscopic observation. Electron microscopic exami-
nation was performed as previously described.²⁶ Bone marrow aspirates
and cultured cells resuspended in FCS were centrifuged at
1,000g for 10 minutes and, after removing plasma or FCS, buffy
coats and cell pellets were fixed in 0.1 mol/L cacodylate buffer (pH
7.4) solution containing 1.5% glutaraldehyde for 1 hour at 4°C and
washed in cacodylate buffer for 2 to 3 hours. Some of the samples
were processed for enzyme cytochemistry for MPO, and the re-
mainring samples were processed for ordinary electron microscopic
examination without cytochemistry. For MPO staining, samples
were embedded in agar and sliced into approximately 30-μm-thick
sections using a tissue sectioner (McIlwain Tissue Chopper; The
Mickle Laboratory Engineering, Gomshall, Surrey, England). They
were then preincubated in 3-3’-diaminobenzidine (DAB; Wako Pure
Chemical Industries, Tokyo, Japan) solution without H₂O₂ at room
temperature for 1 hour and then exposed to DAB (0.1%) -H₂O₂
(0.01%) solution for 10 minutes for the peroxidase reaction. The
samples, both with and without peroxidase reaction, were further
fixed in 1% osmium tetroxide for 1 hour, dehydrated in alcohol and
acetone at graded concentrations, and embedded in Epon. Ultrathin
sections were counterstained with uranyl acetate, followed by lead
citrate, and examined under an electron microscope (JEOL 1200EX;
Japan Electric Optical Laboratory, Tokyo, Japan) at an accelerating
voltage of 80 kV.

RESULTS

Light microscopic observation. The leukemic cells in
patients no. 1 and 2 contained abundant cytoplasmic azuro-
philic granules, including Auer rods, and were intensely posi-
tive for peroxidase, a typical finding for the diagnosis of
M3 in the FAB classification. In addition to Auer rods, the
cells of patient no. 1 possessed large cytoplasmic inclusions
with a crystalloid irregular shape (Fig 1a) that were positive
for peroxidase. Compared with patients no. 1 and 2, the
cells of patient no. 3 had a higher nuclear/cytoplasmic ratio, smaller numbers of visible azurophilic granules, and irregularly shaped nuclei with occasional deep nuclear indentation, findings consistent with the diagnosis of the M3 variant form.

The morphology of the leukemic cells changed in a similar manner in response to ATRA treatment in all three patients. After 1 week of treatment, the leukemic cells in bone marrow showed chromatin condensation and nuclear lobulation and the azurophilic granules had lost their characteristic color (Fig 1b). After 2 weeks of treatment, mature neutrophils with somewhat abnormally shaped nuclei appeared (Fig 1c), and some of them contained Auer rods. After 5 to 8 weeks of treatment, when a CR had been achieved, the bone marrow consisted of normal hematopoietic cells of multiple lineages at various stages of differentiation, including neutrophils, eosinophils, monocytes, the erythroid series, and megakaryocytes. Most of the neutrophils appeared normal, but a small number of immature myeloid cells with irregular nuclei were still present.

**Electron microscopic observation.** The leukemic cells of all three patients possessed abundant spherical (type I) primary granules containing electron-dense homogeneous material, with a peripheral halo in some granules and dilated rER (Fig 2a). Although type I primary granules predominated in all patients, small numbers of cytoplasmic granules containing crystalloid structures (type II primary granules) were also present in patient no. 1. Nuclear configurations were irregular, and the chromatin was only slightly condensed along the nuclear membrane. In addition to numerous Auer rods, the cells of patient no. 1 contained large cytoplasmic inclusion bodies also seen under the light microscope. These are the structures surrounded by a unit membrane and filled with parallel tubular material identical to that of Auer rods and belong to the pseudo-Chediak-Higashi (C-H) category of granules (Fig 2b). The cells of patient no. 3 contained smaller cytoplasmic granules than those of the other patients, and Auer rods were less frequently observed, a finding consistent with the M3 variant. Stellate rER complexes and fibrillar structures in the rER, both of which are thought to be structures specific to APL, were detected in this patient.

Common morphological changes of bone marrow leukemia cells were observed during ATRA treatment. After 1 week of ATRA treatment, leukemic cells showed heterochromatin condensation and nuclear lobulation (Figs 3 and 4). Although the lobulated nuclei of some cells resembled those of PMNs, these nuclei were irregular in shape and nucleoli were occasionally present. The lobulated nuclei often had nuclear blebs (Fig 3a) or were connected by thin
nuclear material (Fig 4a), called filamentous connections,\textsuperscript{13} that consisted of heterochromatin delimited by the nuclear membrane. In addition to these changes, the irregularly shaped primary granules became morphologically normal. This was most evident in patient no. 1, in whom the majority of both Auer rods and irregularly shaped inclusions, as shown in Fig 2, dramatically vanished and the remaining primary granules appeared normal, being spherical in shape and relatively homogeneous in size (Fig 3a), although abnormal primary granules and/or Auer rods were infrequently observed in some cells. Rarely, a structure composed of an Auer rod coupled with a primary granule was found, appearing as though the seemingly normal primary granule had emerged from the Auer rod (Fig 3b). Type I primary granules still predominated in all patients, but type II primary granules with a nucleoid structure were rarely observed in patients no. 1 and 2. Some maturing neutrophils with abnormally shaped lobulated nuclei in patient no. 2 possessed unusual, possibly immature, primary granules containing central electron-dense material (Fig 4a and b), indicating asynchronous maturation of nuclei and cytoplasmic granules.

After 2 to 3 weeks of ATRA treatment, maturation of leukemia cells progressed further, and cells resembling normal PMNs appeared (Fig 5). These cells contained multilobulated nuclei with marginally condensed heterochromatin and glycogen particles, characteristic of normal mature neutrophils. rER and free ribosomes became rare as the maturation process proceeded. However, these cells were still rich in primary (type I) granules, which somewhat varied in size and shape, including small and/or elongated forms. A small number of ellipsoid type II primary granules with a nucleoid structure were occasionally observed in patients no. 1 and 2, but not in patient no. 3. Conversely, MPO-positive, small, possibly type III, primary granules were rarely observed intermingled among the larger primary granules in maturing neutrophils in patient no. 3 alone. Despite the ultrastructural features otherwise characteristic of normal mature PMNs, specific granules, which are electron-lucent, elongated or dumbbell-shaped small granules, were not present in the majority of these PMN-like cells, as clearly shown by electron-microscopic cytochemical staining for MPO (Fig 5). The cells contained only MPO-positive primary granules and no MPO-negative secondary granules. A few abnormal primary granules, including Auer rods or C-H type granules, were still present in some cells, and these cells too were consistently devoid of specific granules (Fig 6). Another unusual finding in PMN-like cells at this stage was the presence of large spherical lipid droplets (Fig 5a), which may indicate altered physiological metabolism in these cells,\textsuperscript{29} because lipid droplets are usually not seen in normal PMNs. Although few in number, PMNs having both primary and secondary granules were also observed (Fig 7), and less

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![Fig 4. (a) A PMN in patient no. 2 after 10 days of ATRA treatment. The nuclei are irregularly lobulated, with some of them being connected by nuclear filamentous structures, and heterochromatin is condensed along the nuclear membrane. (Original magnification × 13,100.) (b) Higher magnification of a portion of the cell in (a). The cytoplasmic granules are large and spherical, and some of them contain flocculent material and central cores with higher electron density. (Original magnification × 20,200.)](image-url)
mature intermediate forms with indented single nuclei or irregularly lobulated nuclei and numerous primary granules were still present at this stage of ATRA treatment.

After 5 to 8 weeks of ATRA treatment, when the bone marrow was considered to exhibit a CR at the light microscopic level, it consisted of cells of multiple lineages at various stages of differentiation. Secondary granule-deficient PMNs, rarely containing Auer rods, still persisted in bone marrow but in very small numbers, and the majority of PMNs contained both primary and secondary granules and were morphologically normal even at the ultrastructural level. At this stage, promyelocytes with numerous primary granules and dilated rER but without definite abnormalities diagnostic of leukemic cell origin were observed.

**APL cells in culture.** APL cells in patient no. 3 were cultured in the presence of ATRA or G-CSF for 7 days, and their morphological changes were examined under a light microscope. After 5 days of culture with ATRA the cells displayed a tendency toward maturation, including nuclear lobulation, chromatin condensation, and a decrease or disappearance of azurophil granules, similar to that observed in bone marrow cells. The cells cultured with G-CSF for 7 days, on the other hand, retained a promyelocytic appearance with numerous azurophil granules that even seemed to have increased in size and/or number (data not shown). Electron microscopic examination showed the nuclei of the cells cultured with ATRA to be irregularly lobulated with marginally condensed heterochromatin and the lobulated nuclei to be often connected by nuclear filamentous structures, in a manner similar to the cells seen in the bone marrow (Fig 8). These cells contained numerous MPO-positive primary granules, including irregularly shaped granules, but no MPO-negative specific granules were identified.

In contrast to the APL cell cultures, although only small numbers of PMNs were observed in the control cultures of nonleukemic bone marrow cells with ATRA, possibly because of the absence of added growth stimulants, they possessed both MPO-positive primary granules and MPO-negative secondary granules (Fig 9). The former outnumbered the latter, which was the reverse of what is observed in normal PMNs, and this may be because of decreased cell division of neutrophil precursors in the cultures in the absence of added growth stimulators, with resultant retention of primary granules. In addition, large lipid droplets were also present in some PMNs (Fig 9). These findings suggest that secondary granule deficiency in PMNs is a leukemia-specific phenomenon but not a general effect of ATRA to inhibit secondary granule formation and, conversely, that the accumulation of lipid droplets is caused by nonspecific effects of ATRA.
DISCUSSION

The key findings in the present electron microscopic study of APL cells during the process of ATRA-induced differentiation are summarized as follows. (1) ATRA-induced maturation of nuclei, namely, nuclear lobulation and marginal condensation of heterochromatin, resulting in generation of nuclei of mature neutrophils, although they were irregularly shaped with occasional nuclear blebs and very often connected by nuclear filamentous structures (filamentous connections) that appeared to be of pathologic significance, similar to the nuclear blebs reported in AML. (2) The majority of aberrant primary granules, including Auer rods, lost their abnormal contour in response to ATRA administration, although small numbers of PMNs with Auer rods or C-H type abnormal granules persisted throughout ATRA treatment. (3) The PMNs that emerged in the bone marrow of APL patients during ATRA treatment, particularly at an early stage, and in cultures of APL cells with ATRA were devoid of neutrophil secondary granules.

Normalization of the size and shape of primary granules of APL cells in response to ATRA treatment was shown by the present electron microscopic study. This change was associated with the loss of azurophilic color in primary granules in Wright-Giemsa-stained smear specimens and, therefore, may indicate maturation of the primary granules. Sulfated glycosaminoglycans (GAGs) have been shown to be present in immature primary granules and to be responsible for their azurophilia, and the lack or masking of the sulfate group has been suspected of causing loss of azurophilic staining in mature primary granules. Because it has been shown that GAGs are also present in the primary granules and Auer rods of leukemic cells and that some Auer rods lack GAGs, suggesting maturational sequence of Auer rods, similar chemical modifications in primary granules may have occurred following the cellular maturation process of APL cells induced by ATRA. Although the process of normalization of primary granules was not clearly detected morphologically in this study, the unusual structure of a primary granule and an Auer rod connected to each other encountered in patient no. 1 (Fig 3b) suggests that normally appearing primary granules may have emerged from abnormal primary granules, including Auer rods.

A deficiency of secondary granules was noted in most of the PMNs that emerged in the bone marrow during ATRA treatment. Although there has been no direct evidence that these secondary-granule-deficient PMNs are neoplastic.
cells, they are likely of neoplastic origin. First, the secondary-granule−deficient PMNs decreased in number in the course of ATRA treatment and were replaced by normal PMNs when remission was obtained. This is consistent with the results of molecular studies using premature chromosome condensation and the fluorescence in situ hybridization technique, showing that a substantial proportion of PMNs during ATRA treatment had chromosome 17 translocations, whereas diploid cells without such abnormalities prevailed in the blood and bone marrow in remission. Second, PMNs with Auer rods, clear evidence of neoplastic origin, were consistently devoid of secondary granules. Third, it is known that AML cell lines that can be induced to differentiate towards mature neutrophils do not express any secondary-granule−associated proteins, including lactoferrin (LF), even after induction of differentiation. These findings suggest that secondary-granule−deficient PMNs are abnormally matured leukemic cells. Moreover, our observation that PMNs with secondary granules appeared in the bone marrow during ATRA treatment and in control cultures of nonleukemic bone marrow cells with ATRA indicates that normal hematopoietic precursor cells can give rise to mature neutrophils with secondary granules regardless of the presence or absence of ATRA. Therefore, secondary-granule deficiency in PMNs in APL is probably because of an intrinsic defect in leukemic cells, not because of an adverse effect of ATRA on secondary-granule formation.

Secondary-granule deficiency has been found in PMNs in AML, consistent with this, we and others have shown a deficiency of LF, a constituent of neutrophil secondary granules, in the PMNs of patients with AML. As mentioned above, it has been shown that, to date, no leukemic cell lines that are chemically induced to differentiate towards mature granulocytes, including HL60, PLB-985, and NB4, express any messages for secondary-granule−associated proteins, including LF, even after induction of differentiation. These findings suggest that secondary-granule deficiency is an abnormality closely associated with and perhaps a hallmark of aberrantly differentiated leukemic blasts in AML and may not be restricted to APL alone, but may be common to all subtypes of AML. In conclusion, genetic control of the neutrophil maturation process in APL is probably defective not only at the promyelocyte stage but in later stages, in a similar manner to that in other subtypes of AML. Although ATRA is highly effective in inducing differentiation of APL cells, it is incapable of inducing complete neutrophil maturation. Because it has been reported that G-CSF, when used in combination with ATRA in vitro, induces alkaline phosphatase activity in APL cells and augments the differentiation-inducing effects of ATRA, other
Fig 8. (a) A PMN that emerged in the APL cell culture of patient no. 3 in the presence of ATRA for 1 week. From the lobulated nuclear configurations and condensed heterochromatin, this cell looks like a PMN. However, the cytoplasm contains many MPO-positive primary granules but no secondary granules. (MPO-stained section, original magnification × 14,900.) (b) Higher magnification of a portion of the cell in (a), showing that the cytoplasmic granules are all MPO-positive. (MPO-stained section, original magnification × 30,100.)

Fig 9. Representative appearance of a portion of a PMN that appeared in a culture of nonleukemic bone marrow mononuclear cells incubated in the presence of ATRA for 1 week. Both MPO-positive and -negative (arrow) cytoplasmic granules are present, with the former outnumbering the latter. Large spherical lipid droplets (L) are also present. N, nucleus. (MPO-stained section, original magnification × 17,700.)
bioactive substances such as G-CSF might be required, to-gether with ATRA for APL cells, to undergo the final neutro-
phil maturation steps.

REFERENCES
2. Borrow J, Goddard AD, Sheer D, Solomon E: Molecular analy-
ation t(15;17) in human acute promyelocytic leukemia fuses RARα with a novel putative transcription factor, PML. Cell 66:663, 1991
15. Brederoo P, van-der-Meulen J, Daems WTH: Ultrastructural localization of peroxidase activity in developing neutrophil granulo-
cytes from human bone marrow. Histochemistry 84:445, 1986
16. Pryzwansky KB, Breton-Gorius J: Identification of a subpop-
ulation of primary granules in human neutrophils based upon matura-
tion and distribution: Study by transmission electron microscopy
17. Bainton DF, Abnormal neutrophils in acute myelogenous leu-
kemia: Identification of subpopulations based on analysis of azuro-
19. Dittman WA, Kramer RJ, Bainton DF: Electron microscopic and peroxidase cytochemical analysis of pink pseudo-Chediak-Hig-
20. Miyauchi J, Watanabe Y, Enomoto Y, Takeuchi K: Lactofer-
in-deficient neutrophil polymorphonuclear leukocytes in leukaem-
21. Schofield KP, Stone PCW, Stuart J: Quantitative cytochemis-
23. Grigoriou HM, Rowley JD, Kinter JR, Bloomfield CD, Sundberg RD, Brun-
kemia. Cancer Res 34:2887, 1974
kemia. Cancer Res 34:2887, 1974
31. Parmley RT, Eguichi M, Spicer SS: Ultrastructural cytochem-
32. Parmley RT, Hurt RE, Takagi M, Spicer SS, Austin RL: Glycosaminoglycans in human neutrophils and leukemic myelo-
34. Lübbers M, Herrmann F, Koefler HP: Expression and regula-
Neutrophil Secondary-Granule Deficiency as a Hallmark of All-Trans Retinoic Acid–Induced Differentiation of Acute Promyelocytic Leukemia Cells

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