Characterization of Cryptic Rearrangements and Variant Translocations in Acute Promyelocytic Leukemia

By David Grimwade, Patricia Gorman, Estelle Duprez, Kathy Howe, Stephen Langabeer, Fiona Oliver, Helen Walker, Dominic Culligan, Jonathan Waters, Mark Pomfret, Anthony Goldstone, Alan Burnett, Paul Freemont, Denise Sheer, and Ellen Solomon

Acute promyelocytic leukemia (APL) is typified by the reciprocal translocation, t(15;17)(q22;q21), leading to the formation of PML-RARα and RARα-PML fusion genes. We have characterized 7 cases of morphologic APL found to lack the t(15;17) on conventional cytogenetic assessment. In 6 of 7 cases, cryptic PML-RARα rearrangements were identified by reverse transcriptase-polymerase chain reaction and fluorescent in situ hybridization (FISH); whereas, in the remaining patient, APL was associated with the variant translocation, t(11;17)(q23;q12-21), leading to the formation of PLZF-RARα and RARα-PLZF fusion genes. In each of the cases with cryptic PML-RARα rearrangements, PML-RARα transcripts were detected in the absence of RARα-PML, consistent with the concept that PML-RARα is the critical oncogenic fusion protein. In 4 of these cases with evaluable metaphase spreads, the occurrence of a nonreciprocal translocation was confirmed by FISH with nonbearing formation of the PML-RARα fusion gene; in 3 cases with morphologically normal chromosomes 15 and 17, RARα was inserted into PML on 15q, whereas in the remaining patient the PML-RARα fusion arose due to insertion of 15q-derived material including PML into RARα on 17q. Immunofluorescence studies were performed using antibodies raised against PML and PIC 1, a ubiquitin-homology domain protein previously identified as an interaction partner of PML. In acute myeloid leukemia (AML) of subtypes other than M3, PIC 1 was localized to the nuclear membrane and colociliated with PML within discrete nuclear bodies. In APL cases with cryptic PML-RARα rearrangements, the characteristic microparticulate pattern of PML staining was detected with partial colocalization with PIC 1, indicative of disruption of the nuclear bodies; whereas in t(11;17)-associated APL, PML and PIC 1 remained colociliated within discrete nuclear bodies, as observed in non-APL cases. Although deregulation of the putative growth suppressor PML and delocalization of other nuclear body constituents have been advocated to play a key role in the development of t(15;17)-associated APL, the present study shows that disruption of PML nuclear bodies per se is not a prerequisite for the pathogenesis of APL.

© 1997 by The American Society of Hematology.
identified as targets for autoantibodies in patients with primary biliary cirrhosis, and PIC 1, a ubiquitin-homology domain protein, which has been found to interact directly with PML. A variety of experimental approaches have implicated PML in immunologic responses (Grumwade and Solomon and references therein) and there is some evidence to suggest that PML itself or components of the nuclear bodies are cell-cycle regulated and can mediate growth-suppressor activity. Cells from a wide range of tissues and blasts from leukemic subtypes other than APL typically demonstrate 10 to 30 discrete nuclear bodies when stained with PML antisera. Whereas, in APL cases associated with the t(15;17), a microparticulate pattern of PML staining is characteristic, reflecting disruption of nuclear bodies due to an interaction between PML and PML-RARα. This process may promote leukemogenesis by delocalizing the putative growth-suppressor PML and other nuclear body components; this, coupled with an abnormal pattern of retinoid responses also mediated by the fusion protein, compounded by RXR sequestration could account for the block in myeloid differentiation that characterizes the disease. Treatment of such cases with ATRA leads to complete remission by terminal differentiation of the leukemic clone associated with release of inhibitory effects of PML-RARα at retinoid response elements and degradation of the fusion protein accompanied by normalization of nuclear body architecture. Rare cases of PML-RARα-mediated APL have been identified that fail to respond to retinoids associated with persistence of the microparticulate PML nuclear staining pattern. On the basis of these findings it has been suggested that disruption of PML nuclear bodies is critical to the pathogenesis of APL; furthermore, it has been advocated that reconstitution of normal nuclear architecture is essential to permit differentiation in the presence of retinoids. Although ATRA therapy is unable to sustain long-term remission in APL, recent studies have shown that a combined treatment approach using ATRA and chemotherapy confers significant improvements in disease-free survival compared with chemotherapy alone. Because a favorable response to ATRA appears to be restricted to patients with the t(15;17), establishing the presence of this cytogenetic change, or in its absence identification of a PML-RARα rearrangement, is fundamental for optimal treatment of patients and meaningful analysis of APL trials. Although early studies in specialized centers reported the presence of the t(15;17) in all cases of APL; it is now clear that some cases with morphologic acute myeloid leukemia (AML) M3 reflect cryptic PML-RARα rearrangements, whereas in others RARα is fused to a partner other than PML. Thus far, three such alternative fusion partners have been identified, including the novel Krüppel-like zinc finger protein PLZF involved in the t(11;17)(q23;q21); nucleophosmin, an RNA processing protein disrupted by the t(5;17)(q32;q12); and most recently NuMA, which is involved in control of mitosis and is rearranged by the t(11;17)(q13;q21). Although such variant translocations are extremely rare, accounting for less than 1% of morphologic APL, elucidation of the mechanisms underlying leukemogenesis in these cases is likely to provide considerable insight into the processes involved in the development of PML-RARα-mediated disease and in particular as to whether disruption of PML nuclear bodies is fundamental to the pathogenesis of APL. Cases with variant fusion translocations also afford the opportunity to dissect out mechanisms leading to leukemic transformation from those mediating the block in myeloid differentiation and its reversal by retinoids. In the present study, we have characterized 7 cases of morphologic APL found to lack the t(15;17) on conventional cytogenetic assessment and considered their implications for the pathogenesis of APL as a whole.

MATERIALS AND METHODS

Patients and cytogenetics. This study considers 7 patients with morphologic APL who were found to lack the t(15;17) on conventional cytogenetic assessment. Five patients were drawn from the MRC ATRA trial; details of the treatment protocol have been described elsewhere. Cytogenetic assessment was undertaken at local centers or by the central UK MRC AML trials cytogenetics service at University College Hospital, London, according to standard methods. In each case in which APL was associated with a normal karyotype, preparations were subject to at least 24 hours of culture and a minimum of 20 metaphases were examined.

Reverse transcriptase-polymerase chain reaction (RT-PCR) and sequence analysis. Details of bone marrow and peripheral blood sample preparation, RNA extraction, and RT-PCR protocols to detect PML-RARα and RARα-PLZF transcripts in conjunction with RARα and PML as controls for RNA integrity have been fully described elsewhere. Using cDNA generated by the same method, PFLZ-RARα and RARα-PLZF transcripts were amplified using nested PCR in material derived from a patient with t(11;17)(q23;q12-21) identified by cytogenetic primers. Used for PFLZ-RARα PCR were as follows: PFLZ external, 5'-TCCAGAGGAGGACTTCCAGC-3'; RARα external, 5'-TCTTCTGGATGTCCTGCAGG-3'; PFLZ internal, 5'-TCAGACCTCTCAGATAAGCAG-3'; and RARα internal, 5'-GGCCGTCAGCCATAGTGGT-3'. Primers for RARα 1-PLZF nested PCR comprised the following: RARα external, 5'-GGCCGACAGACAGCAGCTCCT-3'; PFLZ external, 5'-ATGTCAGTGCCAGTATGGGT-3'; and RARα internal, 5'-GTCGTCCTCCACCGTCTTCT-3'; and PFLZ internal, 5'-CAGTACAGACAAAGGC-3'.

PCR was performed in a 50 μL reaction, with 1 μL of the first round PCR products used as template for the second round of PCR, as previously described. First and second round PCR reactions comprised 35 cycles, each consisting of 1 minute of denaturation at 95°C, 1 minute of annealing at 57°C, and 1 minute of extension at 72°C, followed by 10 minutes of extension at 72°C (Oligene apparatus; Hybaid, Teddington, Middlesex, UK). PCR products were size-separated on ethidium bromide-stained 1.5% agarose gels, as previously described. Bcr 1 (intron 6) and bcr 2 (exon 6) PML breakpoints were distinguished by sequence analysis of PML-RARα PCR products using previously described methods and primers. PLZF-RARα and RARα-PLZF PCR products were similarly sequenced, using appropriate internal primers, with an automated sequencer (377; AB1, Perkin-Elmer, CA).

Fluorescent in situ hybridization (FISH). FISH using ICRF PML and RARα cosmid probes (15.5 and 121, respectively) was performed to detect PML-RARα fusion genes in patients lacking the t(15;17), using previously described methods. PML cosmid 15.5 encompasses the 5′ region of the gene, including exons 1 and 2, whereas RARα cosmid 121 spans the APL breakpoint region on chromosome 17. In APL cases associated with the t(15;17), a dual signal is detected with these probes on the derivative chromosome 15, identifying the site of the PML-RARα fusion, in addition to single hybrid-
ization signals corresponding to the normal PML and RARα loci on 15q and 17q, respectively. FISH was also performed, using PML and RARα probes (Oncor, Gaithersburg, MD), which detect the formation of the reciprocal PML-RARα fusion gene on del(17q) in APL cases associated with the t(15;17), in addition to single signals relating to the normal PML and RARα loci. To further characterize cases with cryptic PML-RARα fusion genes, whole chromosome paints (WCPs; Vysis, Richmond, Surrey, UK) and biotinylated centromere probes (Oncor) for chromosomes 15 and 17 were used, in accordance with the manufacturer’s instructions. Images were captured with a Zeiss Axioskop epifluorescence microscope and cooled CCD camera (Photometrics, AZ), controlled by an Apple Macintosh computer (Apple Computer Inc, Cupertino, CA). Image capture and processing software were obtained from Vysis, UK.

**Immunofluorescence.** The PML immunofluorescence technique as applied to crude bone marrow or peripheral blood smears using polyclonal PML antiserum has been described in detail previously. Dual immunofluorescence studies were also undertaken using a polyclonal antiserum raised against the PML nuclear body constituent, PIC 1, and a monoclonal antibody (PG-M3; kindly provided by P.G. Pelicci, European Institute of Oncology, Milan, Italy) directed against the amino-terminal of PML, which recognizes both wild-type PML and the PML-RARα fusion protein. Bone marrow/peripheral blood smears were fixed in methanol at −20°C for 15 minutes, allowed to air dry, and preblocked with 10% fetal calf serum. Slides were then incubated simultaneously with PG-M3 and PIC 1 antibodies at a dilution of 1/50 and 1/200, respectively, in phosphate-buffered saline (PBS)-Tween 0.5% for 1 hour at room temperature. Slides were subsequently incubated with Texas-Red–coupled antimouse (Dako Ltd, High Wycombe, UK) and fluorescein-coupled antirabbit (Amersham Intl, Amersham, UK) secondary antibodies, each at 1/200 dilution, for 30 minutes at room temperature. All incubations were followed by three washes in PBS followed by a final wash comprising 0.05% Tween in PBS for 10 minutes. Preparations were examined by confocal laser scanning as previously described.

**RESULTS**

**Cytogenetic and molecular characterization of APL cases lacking the t(15;17).** Cytogenetic and molecular findings in 7 patients with morphologic features of APL, but lacking the t(15;17), are presented in Table 1. In 6 of 7 patients, RT-PCR confirmed the presence of a PML-RARα rearrangement, and in each of these cases PML-RARα transcripts were detected in the absence of RARα-PML. In the remaining patient (case no. 7), APL was associated with t(11;17) (q23;q12-21); RT-PCR confirmed expression of both PLZF-RARα and reciprocal RARα-PLZF transcripts (Fig 1). Sequencing of PLZF-RARα and RARα-PLZF PCR products was consistent with breakpoints within the second intron of RARα and within the intron separating the exons coding for the second and third zinc fingers of PLZF, as identified in 5 of 6 previously described cases.

**FISH analysis of APL cases with cryptic PML-RARα rearrangements.** FISH using ICRF probes, PML cos 15.5 and RARα cos 121, confirmed formation of a PML-RARα fusion in each of the 6 APL cases in which PML-RARα rearrangements were identified by RT-PCR (Table 1). In 4 patients with available diagnostic metaphase spreads, the mechanism underlying the PML-RARα rearrangement was further characterized, using ICRF cosmid probes that specifically detect the PML-RARα fusion gene, in conjunction with centromere probes and WCPs for chromosomes 15 and 17. In 3 patients, in each of whom morphologically normal chromosomes 15 and 17 were identified by conventional cytogenetics (cases no. 1 through 3, Table 1), the PML-RARα fusion gene was localized to chromosome 15q (Fig 2a and b). In each case, 15 and 17 specific paints hybridized solely to their respective chromosomes (Fig 2c and d), consistent with an interstitial insertion of RARα into PML on 15q. Furthermore, in each of these cases, using commercially available probes (Oncor), RARα was found to hybridize to two normal appearing chromosome 17s in addition to forming a fusion signal on 15q, again consistent with formation of a PML-RARα fusion on 15q and absence of the reciprocal RARα-PML fusion gene as suggested by RT-PCR analyses. In the remaining patient with evaluable metaphases, PML 15.5 and RARα 121 probes localized the PML-RARα fusion to 17q (case no. 4; Fig 3a and b). In addition to the PML-RARα fusion signal, a more centromeric RARα hybridization signal was observed. WCPs demonstrated insertion of chromosome 15 material into 17q, such that chromosome 17 appeared abnormally large on conventional cytogenetic assessment (Fig 3c and d). These results indicate that the PML-RARα fusion in this patient reflected insertion of PML with more centromeric chromosome 15-derived material into the genomic region spanned by RARα cos 121; again, this was consistent with detection of PML-RARαs in the absence of RARα-PML fusion transcripts by RT-PCR.

**Nuclear architecture in APL cases lacking the t(15;17).** Immunofluorescence studies were performed using polyclonal PML antiseras. In 3 patients, cryptic PML-RARα rearrangements detected by FISH and RT-PCR were confirmed by the presence of the characteristic microparticulate nuclear staining pattern in leukemic cells (Table 1 and Fig 4a), identical to that observed in the NB4 cell-line and APL cases associated with the t(15;17); whereas, in the APL case associated with the t(11;17) leading to a PLZF-RARα rearrangement, a wild-type pattern of PML nuclear staining was observed within the leukemic blasts (Fig 4c), identical to that seen in 2 non-APL AML patients (Fig 4b) and to that previously described for HL60 and U937 cell-lines.

Dual immunofluorescence studies were subsequently undertaken using a PML monoclonal antibody and polyclonal antiserum directed against PIC 1, a newly described constituent of PML nuclear bodies. In 3 non-APL AML cases, PML and PIC 1 were colocalized within discrete nuclear bodies as shown in Fig 5b; as distinct from the pattern detected in a patient with a cryptic PML-RARα rearrangement (case no. 5) in which microparticulate PML staining was observed, with only partial colocalization with PIC 1 (Fig 5a). In the patient with t(11;17)-associated APL, PML and PIC 1 were colocalized within discrete nuclear bodies (Fig 5c), as observed in non-APL cases. In addition to discrete punctate nuclear staining, PIC 1 was also localized to the nuclear membrane, as clearly shown in Fig 5b and c.

**DISCUSSION**

Early studies in specialist cytogenetic centers reported that the t(15;17) could be detected in all cases of APL. In the light of such claims, clinicians encountering AML cases with morphologic features of APL could lose faith in the initial
Table 1. Molecular and Cytogenetic Characteristics of Patients With Morphologic APL Lacking the t(15;17)

<table>
<thead>
<tr>
<th>Case No.</th>
<th>Morphologic Features</th>
<th>Cytogenetics</th>
<th>Fusion Transcripts Detected by RT-PCR</th>
<th>PML Breakpoint</th>
<th>Rearrangement Detected by FISH</th>
<th>PML Immunofluorescence</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Hypergranular M3</td>
<td>47,XX,del(5)(q27q34),-i(8)(q19) [3]</td>
<td>PML-RARa</td>
<td>bcr 3</td>
<td>Interstitial insertion of RARa into PML on 15q</td>
<td>ND</td>
</tr>
<tr>
<td>2</td>
<td>Hypergranular M3</td>
<td>46,XY</td>
<td>PML-RARa</td>
<td>bcr 1</td>
<td>Interstitial insertion of RARa into PML on 15q</td>
<td>Microparticulate</td>
</tr>
<tr>
<td>3</td>
<td>Hypogranular M3 variant</td>
<td>46,XY</td>
<td>PML-RARa</td>
<td>bcr 3</td>
<td>Interstitial insertion of RARa into PML on 15q</td>
<td>ND</td>
</tr>
<tr>
<td>4</td>
<td>Hypergranular M3</td>
<td>46,XY,t(4;16)(p14;q22),t(9;12)(q22;q24),dir insulation(17;15)(q21;q15q22)</td>
<td>PML-RARa</td>
<td>bcr 1</td>
<td>Interstitial insertion of RARa into PML on 15q</td>
<td>ND</td>
</tr>
<tr>
<td>5</td>
<td>Hypergranular M3</td>
<td>46,XX,del(7)(q22q36) [5]</td>
<td>PML-RARa</td>
<td>bcr 3</td>
<td>PML-RARa fusion detected by interphase FISH</td>
<td>Microparticulate partial PML and PIC 1 colocalization</td>
</tr>
<tr>
<td>6</td>
<td>Hypergranular M3</td>
<td>46,XX</td>
<td>PML-RARa</td>
<td>bcr 3</td>
<td>PML-RARa fusion detected by interphase FISH</td>
<td>Microparticulate</td>
</tr>
<tr>
<td>7</td>
<td>Hypergranular M3. Marrow replaced by abnormal promyelocytes with basophilic granules.</td>
<td>46,XY,t(11;17)(q23;q12-21) [12]</td>
<td>PLZF-RARa, RARa-PLZF</td>
<td>NA</td>
<td></td>
<td>Wild-type PML and PIC 1 colocalized</td>
</tr>
</tbody>
</table>

Abbreviations: ND, not determined; NA, not applicable.

Fig 1. Detection of PLZF-RARα and RARα-PLZF transcripts by nested RT-PCR in t(11;17)(q23;q12-21)-associated APL. PLZF-RARα and RARα-PLZF cDNA sequences are shown on the right; the positions of RARα and PLZF fusion junctions are delineated by vertical arrows.
Fig 2. Cryptic PML-RARα fusion resulting from interstitial insertion of RARα into PML on 15q (case no. 2; Table 1). (a) FISH analysis using ICRF PML 15.5 (green) and RARα 121 (red) cosmid probes. The RARα probe hybridized to two chromosome 17s of normal appearance, whereas 1 normal PML locus was observed on chromosome 15. The PML-RARα fusion was detected on 15q (yellow arrow). Localization of the fusion gene was confirmed by subsequent hybridization with a chromosome 15 centromere probe, shown in red in (b). (c) Chromosome 15 paint (red); chromosome 17 centromere probe (green). (d) Chromosome 17 paint (green); chromosome 15 centromere probe (red).

Fig 3. PML-RARα fusion resulting from interstitial insertion of PML with associated chromosome 15-derived material into RARα on 17q (case no. 4; Table 1). (a) FISH analysis using ICRF PML (green) and RARα (red) cosmid probes. PML-RARα fusion gene was detected on 17q (yellow arrow), adjacent to RARα hybridization signal (red arrow). Splitting of the RARα-derived signal on der(17q) was indicative of insertion of PML and adjacent sequence into the genomic region covered by RARα cosmid 121. Localization of the PML-RARα fusion gene was confirmed by subsequent hybridization with a chromosome 17 centromere probe shown in green in (b). (c) Chromosome 15 paint (red) and chromosome 17 centromere probe (green), confirming insertion of chromosome 15-derived material into 17q. (d) Chromosome 17 paint (green), chromosome 15 centromere probe (red). Chromosome 17 paint remained localized to 17; der(17q) showed a region of absent signal corresponding to the inserted region of 15q shown in (c).
Cryptic Rearrangements and Variant Translocations in APL

**Fig 4.** PML immunofluorescence in AML using polyclonal antisera. Phase contrast is shown in left-hand panels and corresponding PML immunofluorescence on the right. In AML cases with cryptic PML-RARα rearrangements, a microparticulate pattern of PML nuclear staining was observed as shown in (a). In non-APL cases, a wild-type pattern of PML nuclear staining was detected, as shown in AML M2 blasts in (b); similar nuclear staining was observed in t(11;17)-associated APL (case no. 7; Table 1), shown in (c).

Clinical diagnosis if subsequent cytogenetic assessment failed to provide appropriate confirmatory evidence. However, since the characterization of the PML-RARα rearrangement that underlies the t(15;17) and identification of the rare variant translocations whereby RARα is fused to partners other than PML, it is clear that absence of the t(15;17) does not preclude a morphologic diagnosis of APL, although it remains the diagnostic hallmark of the disease. Large multicenter studies such as the UK MRC ATRA trial afford the opportunity to determine the frequency of cryptic rearrangements and variant translocations among patients with suspected APL. In this regard, in only 87% of APL patients with molecular evidence of a PML-RARα rearrangement was the t(15;17) detectable by conventional cytogenetics. In most cases, absence of the t(15;17) was a reflection of failed cytogenetics; however, 2% of cases were due to cryptic PML-RARα rearrangements. In the present study, we have characterized a series of 7 patients with morphologic APL found to lack the t(15;17) on successful conventional cytogenetic assessment. In 6 patients, of whom 5 had morphologically normal chromosomes 15 and 17, the diagnosis was confirmed by the presence of a PML-RARα rearrangement; in each case, PML-RARα transcripts were detected by RT-PCR in the absence of the reciprocal derived RARα-PML species. FISH analyses in 4 such patients with evaluable metaphase spreads confirmed the presence of a nonreciprocal translocation and in each case were consistent with formation of PML-RARα as the sole fusion gene as a result of an interstitial insertion event, most commonly due to insertion of RARα into PML on 15q. This phenomenon was observed in 3 cases and has been the subject of two previous case reports in which chromosomes 15 and 17 also appeared normal by conventional cytogenetics. In the remaining patient with evaluable metaphases in the present study, the PML-RARα fusion was found to result from insertion of PML into RARα on 17q, which has not been previously described. Future characterization of the genomic breakpoints of these cases may provide insights not only into mechanisms mediating interstitial insertions but also into mechanisms underlying the more typical classical reciprocal translocation.
Demonstration of PML-RARα as the sole fusion gene formed in each of the APL cases with cryptic PML-RARα rearrangements in this study is consistent with the proposed role of its gene product as a critical mediator of leukemogenesis.24,31 This has recently been confirmed in a transgenic model38 whereby expression of PML-RARα was associated with impairment of normal myeloid differentiation accompanied by accumulation of primitive precursors and predisposition to the development of an APL-like syndrome responsive to ATRA. Although the presence of a latent period before developing the leukemia argues in favor of a requirement for additional mutational events, as has been suggested in theoretical models of tumorigenesis,54 it could also imply that high-level expression of the transgene did not occur within equivalent progenitors to those forming the targets of leukemic transformation in human APL. Although PML-RARα is clearly established as playing a key role in leukemogenesis and determining the differentiation response to ATRA, any role for RARα-PML in the pathogenesis of the disease remains to be determined. Although a single case of APL in which a RARα-PML fusion gene was apparently formed in isolation has been reported,3 the present study demonstrating a series of cases in which PML-RARα was the sole fusion gene formed argues against a significant role for RARα-PML in the pathogenesis of APL. Furthermore, this study, when considered in conjunction with previous series establishing that RARα-PML transcripts are not detectable in approximately 20% APL patients,3 shows that, at least in a proportion of cases, absence of RARα-PML transcripts reflects the occurrence of PML-RARα rearrangements due to interstitial insertion events rather than the classic reciprocal translocation.

In 1 patient in the present study, APL was associated with t(11;17)(q23;q12-21), leading to a PLZF-RARα rearrangement. In common with the index case,3 both PLZF-RARα and RARα-PLZF transcripts could be detected by RT-PCR, consistent with the concept that, in contrast to t(15;17)-mediated APL, both fusion partners are implicated in leukemogenesis.31,55 Whereas PLZF bears no structural similarities to PML, being characterized by an amino-terminal POZ motif, and 9 carboxy-terminal zinc finger domains implicated in its role as a transcription factor,41,33,55-58, the two proteins and their respective fusion products share a number of common features. In particular, PLZF is localized to discrete nuclear bodies56,58 whose formation is dependent on the integrity of the POZ domain57 and exhibits growth-suppressor activity in transformation assays.58,59 Furthermore, in common with PML-RARα, PLZF-RARα may sequester RXR and binds retinoid response elements inhibiting transactivation.56,57,60 These processes involving disrupted growth-suppressor function coupled with deregulation of retinoid signaling pathways appear to provide a final common pathway for the pathogenesis of APL. However, in marked contrast to the disease associated with PML-RARα rearrangements, in vitro differentiation assays have shown that blasts derived from t(11;17)(q23;q21) APL cases are resistant to retinoids.41 This may be accounted for by a number of factors, including differing repertoire and character of response between the respective fusion proteins in the presence of ligand,57,60,61 possibly compounded by the absence of ATRA-induced degradation of PLZF-RARα34 and by upregulation of RARα-PLZF leading to persistent deregulation of the cell cycle.41,55,59

Recent studies suggest that there is at least partial colocalization of PML and PLZF within the nucleus,61,62 raising the possibilities that their growth-suppressor activities might be interrelated and that fusion proteins associated with variant APL translocations could promote leukemogenesis by disruption of PML nuclear bodies. Therefore, in the context of such a model, one might expect ATRA-resistant t(11;17) cases to maintain a disrupted pattern of PML nuclear bodies, reminiscent of that reported in retinoid-resistant PML-RARα-mediated cases.59,35 However, the present study refutes such a hypothesis; disruption of PML nuclear bodies was only observed in APL cases associated with cryptic PML-RARα rearrangements in which PML nuclear staining patterns were identical to t(15;17)-positive cases, whereas a wild-type pattern of PML nuclear staining was detected in
blasts derived from (t(11;17)(q23;q21)-associated APL, reminiscent of that observed in other subtypes of AML. Furthermore, using dual immunofluorescence techniques, we were able to show that PIC 1 (GMP 1, SUMO 1) interacts with PML in a yeast-two hybrid screen, and interacts with PML in vivo in AML blasts as well as in nonhemopoietic cell lines, as previously described. In t(11;17)-associated APL and in non-APL cases, in addition to perfect colocalization with PML within discrete nuclear bodies, a perinuclear pattern of PIC 1 staining was observed consistent with its reported interaction with RanGap1, targeting it to the nuclear pore complex. In contrast, in APL cases associated with cryptic PML-RARα rearrangements, only partial colocalization of PML and PIC 1 was observed, confirming results previously obtained with the APL cell-line NB4, reflecting disruption of PML nuclear bodies in the presence of the PML-RARα fusion protein. Recent work has established that the second heptad repeat of the coiled-coil domain of PML-RARα is critical for nuclear body disruption, which does not appear to be necessary either for the block in differentiation or its reversal by retinoids; both of these effects are dependent on the integrity of the first heptad repeat of the coiled-coil region. Whether this perturbation of nuclear architecture is involved in the process of leukemic transformation is still undetermined. It remains a possibility that disruption of the nuclear bodies is merely a secondary phenomenon reflecting an interaction between the PML-RARα fusion protein and wild-type PML and is of no importance to leukemogenesis. Nevertheless, the present study underlines the fact that disruption of PML nuclear bodies provides a valuable marker for the PML-RARα fusion protein in cases lacking the t(15;17) and confirms PML immunofluorescence as a suitable technique for rapid identification of the subgroup of APL patients likely to benefit from retinoids. In conclusion, although deregulation of the putative growth-suppressor PML and delocalization of other nuclear body constituents have been advocated to play a key role in the development of t(15;17)-associated APL, the present study shows that disruption of PML nuclear bodies per se is not a prerequisite for the pathogenesis of APL.

ACKNOWLEDGMENT

The authors are grateful to all the clinicians who entered patients into the MRC ATRA trial and forwarded material for molecular and cytogenetic analyses, particularly Dr Steve Kelsey. We thank Steve Chatters and Joanne Rogers in the Cytoanalytics Laboratory at University College Hospital, London, and the cytogeneticists involved in karyotyping these patients who provided material for FISH analyses, particularly Debra Lillington, Michael Neat, and David Stevenson. We are grateful to Peter Jordan for assistance with immunofluorescence studies, to the photographic department at ICRF, Lincoln’s Inn Fields, and to Iain Goldsmith and the oligonucleotide synthesis service at ICRF, Clare Hall. We also thank Hans Nicolai, Dr Melissa Brown, Dr Aurelie Catteau, and Dr Chun-Fang Xu for helpful discussions and assistance with sequence analyses.

NOTE ADDED IN PROOF

A wild-type PML nuclear localization pattern has also recently been reported in an APL case with the variant translocation (t(11;17)(q13;q21) leading to a NUMA/RARα rearrangement. PIC 1 has been designated UBL 1 by the Gene Nomenclature Committee.

REFERENCES

1. Rowley JD, Golomb HM, Dougherty C: 15/17 translocation, a consistent chromosomal change in acute promyelocytic leukaemia. Lancet 1:549, 1977
12. Tsai S, Collins SJ: A dominant negative retinoic acid receptor blocks neutrophil differentiation at the promyeloctye stage. Proc Natl Acad Sci USA 90:7153, 1993
17. Daniel MT, Koken M, Romagni O, Babey S, Bazarbachi A,
33. AID Blood 0004 / 5h42$$$$61 11-12-97 14:42:27 blda WBS: Blood
t(5;17) variant of acute promyelocytic leukemia expresses a nucleo-
47. Wells RA, Cazavet-Coles A, Kamel-Reid S: Fusion of retinoic acid
receptor α to NUMA, the nuclear mitotic apparatus protein, by
a variant translocation in acute promyelocytic leukaemia. Nat Genet
17:109, 1997
48. Pandolfo PP: PML, PLZF and NPM genes in the molecular
pathogenesis of acute promyelocytic leukaemia. Haematologica
81:472, 1996
49. Webber LM, Garson OM: Fluorodeoxyuridine synchroniza-
tion of bone marrow cultures. Cancer Genet Cytoenet 8:123, 1982
A, Solomon E: Minimal residual disease detection in acute promy-
elocytic leukaemia by reverse-transcriptase PCR: Evaluation of PML-
RARα and RARα-PML fusion transcripts. Br J Haematol 82:529, 1992
51. Flenghi L, Fagioli M, Tomassoni L, Pileri S, Gambacorta M,
Pacini R, Grignani F, Casini T, Ferrucci PF, Martelli MF, Pelicci
PG, Falini B: Characterization of a new monoclonal antibody (PG-
M3) directed against the aminoterminal portion of the PML gene
product: Immunocytochemical evidence for high expression of PML
proteins on activated macrophages, endothelial cells, and epithelia.
Blood 85:1871, 1995
zinc finger gene and the retinoic acid receptor-α locus due to a
variant t(11;17) translocation associated with acute promyelocytic
leukaemia. EMBO J 12:1161, 1993
53. Vickers M: Estimation of the number of mutations necessary
to cause chronic myeloid and acute promyelocytic leukaemias from
epidemiologic data. Br J Haematol 93:60, 1996 (abstr, suppl 1)
54. Sitterlin D, Tiollais P, Transy C: The RARα-PLZF chimera
associated with acute promyelocytic leukemia has retained a se-
55. Licht JD, Shakhnovich R, English MA, Melnick A, Li J-Y,
Reddy JC, Dong S, Chen S-J, Zelent A, Waxman S: Reduced and
altered DNA-binding and transcriptional properties of the PLZF-
retinoic acid receptor-α chimera generated in t(11;17)-associated
S-J: Amino-terminal-protein-protein interaction motif (POZ-domain)
is responsible for activities of the promyelocytic leukemia zinc fi-
ger-retinoic acid receptor-α fusion protein. Proc Natl Acad Sci USA
93:3624, 1996
is expressed with a speckled nuclear pattern in early hematoo-
58. Reid A, Gould A, Brand N, Cook M, Strutt P, Li J, Licht J,
Waxman S, Krumlauf R, Zelent A: Cyclin A is a candidate target gene for the promyelocytic leuke-
ia zinc finger protein. Blood 88:291a, 1996 (abstr, suppl 1)
fusion proteins generated from the variant t(11;17)(q23;q21) translo-
cation in acute promyelocytic leukaemia inhibit ligand-dependent
transactivation of wild-type retinoic acid receptors. Proc Natl Acad
Sci USA 91:1178, 1994
60. Ruthardt M, Testa U, Nervi C, Riganelli D, Ferrucci PF,
Grignani F, Alcalay M, Puccetti E, Grignani F, Peschle C, Hoelzer
PG, Falini B: Characterization of a new monoclonal antibody (PG-
M3) directed against the aminoterminal portion of the PML gene
product: Immunocytochemical evidence for high expression of PML
proteins on activated macrophages, endothelial cells, and epithelia.
Blood 85:1871, 1995
modification modulates the partitioning of the Ran-GTPase-activat-
ing protein RanGAP1 between the cytosol and the nuclear pore
62. Mahajan R, Delphin C, Guan T, Gerace L, Melchior F: A
small ubiquitin-related polypeptide involved in targeting RanGAP1
to the nuclear pore complex protein RanBP2. Cell 88:97, 1997
63. Dyck J, Warrell RP, Evans RM, Miller WH: Rapid diagnosis
of acute promyelocytic leukemia by immunohistochemical localiza-
Characterization of Cryptic Rearrangements and Variant Translocations in Acute Promyelocytic Leukemia

David Grimwade, Patricia Gorman, Estelle Duprez, Kathy Howe, Stephen Langabeer, Fiona Oliver, Helen Walker, Dominic Culligan, Jonathan Waters, Mark Pomfret, Anthony Goldstone, Alan Burnett, Paul Freemont, Denise Sheer and Ellen Solomon

Updated information and services can be found at:
http://www.bloodjournal.org/content/90/12/4876.full.html

Articles on similar topics can be found in the following Blood collections
Neoplasia (4182 articles)

Information about reproducing this article in parts or in its entirety may be found online at:
http://www.bloodjournal.org/site/misc/rights.xhtml#repub_requests

Information about ordering reprints may be found online at:
http://www.bloodjournal.org/site/misc/rights.xhtml#reprints

Information about subscriptions and ASH membership may be found online at:
http://www.bloodjournal.org/site/subscriptions/index.xhtml