Immunostimulatory Oligodeoxynucleotides Containing CpG Motifs Enhance the Efficacy of Monoclonal Antibody Therapy of Lymphoma

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Bacterial DNA and synthetic oligodeoxynucleotides containing the CpG motif (Cpg ODN) can activate various immune cell subsets, including natural killer cells and macrophages. We evaluated whether the combination of Cpg ODN and antitumor monoclonal antibody is effective at preventing tumor growth in an immunocompetent murine lymphoma model. Cpg ODN–activated murine splenocytes induced lysis of tumor targets more effectively than unactivated splenocytes. These effector cells were also superior to unactivated splenocytes or cells activated with a control methylated ODN at inducing antibody-mediated lysis of 38C13 murine lymphoma cells. In vivo, Cpg ODN alone had no effect on survival of mice inoculated with 38C13 cells. However, a single injection of Cpg ODN enhanced the antitumor response to antitumor monoclonal antibody therapy. Ninety percent of mice treated with monoclonal antibody alone developed tumor compared with 20% of mice treated with antibody and Cpg ODN. These antitumor effects were less pronounced when treatment consisted of an identical ODN containing methylated CpG dinucleotides. A single dose of Cpg ODN appeared to be as effective as multiple doses of interleukin-2 at inhibiting tumor growth when combined with antitumor monoclonal antibody. We conclude that immunostimulatory Cpg ODN can enhance antibody dependent cellular cytotoxicity and warrant further evaluation as potential immunotherapeutic reagents in cancer.

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MATERIALS AND METHODS

Tumor model. The 38C13 murine B-cell lymphoma model has been used extensively in studies of antitumor MoAb therapy. The 38C13 cells were grown in RPMI 1640 supplemented with 100 U/mL of penicillin and streptomycin, 2-mercaptoethanol, and 10% fetal bovine serum (HyClone Laboratories, Logan, UT) that was heat inactivated at 56°C (complete media). Cells were used in log-phase growth. The idiotype expressed by the surface IgM of 38C13 serves as a highly specific tumor antigen. The IgG2a-anti-idiotype MoAb designated MS11G6 has been previously described and is referred to as antitumor MoAb. It was purified from cell culture supernatant by affinity chromatography using protein A and used in both the in vitro and in vivo assays.
CpG ODN

5’ TCT CCC AGC GTG CGC CAT 3’

Control Methylated ODN

5’ TCT CCC AGC GTG CGC CAT 3’

Oligodeoxynucleotide preparation. Phosphorothioate oligodeoxynucleotides were kindly provided by Genta (San Diego, CA) or purchased from Oligos etc (Wilsonville, OR). An immunostimulatory CpG oligodeoxynucleotide (with the sequence 5’ TCT CCC AGC GTG CGC CAT 3’) was selected for further study when it was found to have a significant effect on murine NK cells with little effect on murine B cells. This CpG oligodeoxynucleotide (henceforth referred to as CpG ODN) at concentrations ranging from 1.25 to 20 µg/mL also had no effect on the in vitro proliferation of 38C13 cells (data not shown). It should be noted that this sequence is also antisense to the human Bcl-2 gene and has partial homology to the antisense to mouse Bcl-2 gene. The control oligodeoxynucleotide (control methylated ODN) had an identical sequence, but contained methylcytosines in the CpG motifs (see Fig 1). Lipopolysaccharide levels were assessed by the Limulus assay and were always less than 12.5 ng/mg of ODN.

In vitro chromium release assay. A standard chromium release assay was used to detect cytotoxicity. Spleens of naive 6- to 8-week-old C3H/HeN mice were homogenized, and erythrocytes were removed by hypo-osmolar lysis with a solution containing 0.15 mol/L NaCl, 1.0 mmol/L KHCO3, 0.1 mmol/L Na2EDTA at a pH of 7.4. B cells were removed by anti-Ig selection using a biotinylated goat antimouse Ig (Sigma, St Louis, MO) and avidin-coated micro-magnetic beads eluted over a magnetic column (Miltenyi Biotech, Auburn, CA). The eluted cells were washed with media and cultured in complete media alone or media supplemented with CpG ODN, control methylated ODN, or 500 µM of recombinant human IL-2 (Chiron, Emeryville, CA) for 24 to 72 hours before use as effector cells. Target cells (38C13) were labeled with 200 µCi of Cr51 (Amersham Life Sciences, Arlington Heights, IL) for 1 hour and washed three times. Cells were divided and incubated at 4°C for 30 minutes in complete media or media supplemented with 5 µg/mL antitumor MoAb before distribution into 96-well V-bottom culture plates (Costar, Cambridge, MA) at the desired cellular concentrations. Effector cells were added at the desired effector to target ratios. Plates were incubated for 4 hours at 37°C in a humidified environment containing 5% CO2 and supernatants evaluated by γ counting. Non-specific release of Cr51 (percent release from target cells incubated for 4 hours without effector cells or MoAb) was less than 20% for all experiments. Samples were run in triplicate, and the percentage of lysis was determined.

In vivo tumor therapy. 38C13 cells grow rapidly and consistently in syngeneic, immunocompetent C3H/HeN mice. Female C3H/HeN mice, obtained from Harlan-Sprague-Dawley (Indianapolis, IN) and housed in the Animal Care Unit at the University of Iowa, were used at 6 to 8 weeks of age. Mice were inoculated with 2,500 38C13 tumor cells intraperitoneally (IP), and therapy with CpG or control methylated ODN, IL-2, and antitumor MoAb was administered IP beginning 2 days later as indicated. CpG ODN and control methylated ODN were used at a dose of 300 µg per mouse because pilot studies demonstrated systemic effects at that dose. Survival was determined, and significance with respect to time to death was assessed using Cox regression analysis.16 We therefore assessed whether treatment with CpG ODN allows for more effective MoAb therapy when treatment with MoAb is delayed beyond the time when...
MoAb is usually effective (Fig 3). Mice were inoculated with 2,500 38C13 cells and divided into groups of 10. Mice received no therapy (group 1), treatment with a single IP injection of 300 μg CpG ODN on day 2 (group 2), therapy with a single injection of 50 μg antitumor MoAb IP on day 3 (group 3), or both CpG ODN and antitumor MoAb (group 4). No toxicity was noted in any group. All mice in the control group (group 1) died within 24 days. Mice that received CpG ODN alone (group 2) had slightly improved survival, but all developed lymphoma and died within 34 days. Mice treated with antitumor MoAb alone had prolonged survival when compared with untreated mice, but only 1 of 10 mice achieved long-term survival. In contrast, 8 of 10 mice treated with CpG ODN followed by antitumor MoAb survived without evidence of tumor. The survival of this group was highly significant when compared with the other groups (\( \nu \) CpG ODN alone, \( P < .001 \); \( \nu \) MoAb alone, \( P = .0109 \)).

**Comparison of antitumor therapy with CpG ODN and antitumor MoAb to therapy with IL-2 and antitumor MoAb.** IL-2 has been shown to enhance the efficacy of anti-idiotypic MoAb therapy in the 38C13 model. IL-2 is most effective when administered repeatedly.\(^{11}\) We therefore compared in vivo therapy of antitumor MoAb with CpG ODN to therapy with MoAb and IL-2 (Fig 4). Mice in the IL-2 groups received 50,000 U IL-2 twice daily for 3 days. Mice received no therapy (group 1), treatment with a single IP injection of 300 μg CpG ODN on day 2 (group 2), therapy with a single injection of 50 μg antitumor MoAb IP on day 3 (group 3), both CpG ODN and antitumor MoAb (group 4), IL-2 alone (group 5), or IL-2 and MoAb (group 6). CpG DNA plus antitumor MoAb therapy resulted in long-term survival of 70%, whereas IL-2 plus antitumor MoAb resulted in long-term survival of only 40%. This difference did not reach statistical significance (\( P = .20 \)).

**Comparison of antitumor therapy with CpG ODN and antitumor MoAb to therapy with control methylated ODN and antitumor MoAb.** In vitro studies suggested the control methylated ODN had little effect on enhancement of ADCC. However, it was possible other mechanisms, such as antisense activity, contributed to the antitumor effect seen in vivo with CpG ODN and MoAb. We therefore evaluated therapy in vivo with MoAb and control methylated ODN in parallel with the IL-2 study outlined above. Control methylated ODN added little to the therapeutic efficacy of MoAb alone (control methylated ODN and MoAb \( P = .12 \)) (Fig 5) in contrast to the studies outlined in Fig 3 where therapy with MoAb and CpG ODN was more effective than therapy with MoAb alone.
Fig 5. Therapy with CpG ODN and MoAb enhances survival more effectively than therapy with control methylated ODN and MoAb. Immunocompetent C3H mice were inoculated with 2,500 tumor cells IP on day 0. Therapy consisted of MoAb alone, control methylated ODN alone, CpG ODN alone, control methylated ODN and MoAb, or CpG ODN and MoAb. Mice were followed for tumor development, toxicity, and survival. Each group contained 10 mice. Curves depict percent survival over 60 days. No mice living at day 60 developed tumor, and all survived more than 5 months. No toxicity was observed in any group. (●) No therapy; (○) MoAb on day 3; (▲) control methylated ODN on day 2; (■) CpG ODN on day 2; (△) control methylated ODN on day 2 and MoAb on day 3; (□) CpG ODN on day 2 and MoAb on day 3.

DISCUSSION

B-cell lymphomas are among the most sensitive tumors to MoAb-based immunotherapy. Agents known to enhance NK activity, such as IL-2, have been shown to enhance the antitumor effects of antilymphoma MoAb. We used a well-established lymphoma model to evaluate whether the antitumor effects of MoAb can be enhanced by CpG ODN that induce NK activation. There was clear synergy between CpG ODN and antitumor MoAb in this model and the most likely explanation for this finding is enhanced ADCC. It is unlikely that the CpG ODN has a direct effect on tumor cells, given tumor proliferation was not inhibited in vitro by CpG ODN and only minimal therapeutic benefit was seen in the group treated with CpG ODN alone. An alternative explanation is that the CpG ODN-induced production of cytokines rendered the tumor cells more sensitive to ADCC. These effects were largely eliminated when an ODN containing identical sequences but having methylated cytosines in the CpG motif was used. Because the 5-methyl cytosine base substitutions actually increase the affinity for hybridization to complementary mRNA, this finding demonstrates that the antitumor effect of this ODN cannot be attributed solely to an antisense mechanism.

Anecdotal reports of tumor regression after systemic bacterial infection have been observed for centuries. Experimental antitumor therapy with heat-inactivated bacteria was reported by Dr William Coley in the 1890s. Dr Coley’s original attempt to use bacteria as an antitumor agent involved the use of live cultures of streptococci. This resulted in tumor regression, but proved to be toxic with the first patient almost dying of erysipelas. Subsequent studies by Coley involved a mixture of heat-killed Streptococci and Serratia (then known as Bacillus prodigiosus). It was this preparation that is now known as Coley’s toxin. Much of the antitumor activity of Coley’s toxin is currently attributed to endotoxin. A number of cytokines induced by endotoxin (such as TNF-α and IFN-γ) have been produced in recombinant form and have been shown to have antitumor activity. However, it is curious to note that Coley’s original success was with an organism that does not produce endotoxin. It is possible bacterial DNA with its unmethylated CpG motifs played a role in the antitumor effects seen in Coley’s original preparation. Whether the responses seen by Dr Coley were related to the immunostimulatory effects of streptococcal DNA, the data presented above indicate that motifs found in bacterial DNA can have antitumor effects, particularly when used with other agents such as MoAb.

A number of important questions remain to be answered. The mechanisms by which bacterial DNA and CpG ODN interact with the cell surface of immune effector cells are internalized and induce activation signals that lead to NK cell and macrophage stimulation need to be clarified. It will also be important to understand more thoroughly how different subsets of immune cells respond to CpG ODN and what aspects of that response are related directly to CpG ODN stimulation or indirectly to cytokines such as IL-6, IL-12, or IFN. Although we observed no toxicity in vivo, the toxicity profile of CpG ODN will require definition. We detected no direct effect of the CpG ODN on 38C13 lymphoma cells; however, it is possible the CpG ODN induced changes in the tumor cells that rendered them more sensitive to MoAb.
therapy. These studies therefore need to be confirmed in another tumor model and using other CpG ODN.

As we learn more about the immune system, it is becoming obvious that an effective antitumor immunotherapy will need to integrate several aspects of the immune system, including establishment of appropriate cytokine profiles, modification of tumor immunogenicity, enhancement of tumor antigen recognition, and appropriate effector cell expansion and activation. The studies outlined above demonstrate immunostimulatory DNA in general, and CpG ODN in particular, may have a role to play in the development of such a therapy.

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REFERENCES
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