Clonal Circulating Cells Are Common in Plasma Cell Proliferative Disorders: A Comparison of Monoclonal Gammopathy of Undetermined Significance, Smoldering Multiple Myeloma, and Active Myeloma

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The blood of most patients with active multiple myeloma (MM) contains cells related to the bone marrow tumor. However, identifying clonal cells in the blood of patients with monoclonal gammopathy of undetermined significance (MGUS) has been difficult. In this study, we analyzed blood mononuclear cells (BMNCs) from 16 patients with MGUS, 2 with amyloidosis, 8 with smoldering MM (SMM), 2 with indolent MM (IMM), and 15 with active MM using three different methods to detect and quantitate clonal cells, i.e., immunofluorescence microscopy (IM) for monoclonal plasma cells, three-color flow cytometry (FC) for CD38+CD45+CD45dim cells, and the allele-specific oligonucleotide polymerase chain reaction (ASO-PCR). Using ASO-PCR, we were able to detect clonal cells in the blood of 13 of 16 patients with MGUS, 2 of 2 with amyloid, 6 of 8 with SMM, 2 of 2 with IMM, and 13 of 15 with MM. In 9 of the 13 patients with MGUS with blood involvement, the number of clonal cells was very small (<0.04% of the BMNCs). The median percentage of clonal cells as determined by ASO-PCR was 0.02 for MGUS, 0.02 for SMM, and 0.24 for MM. Clonal plasma cells or CD38+CD45+CD45dim cells were identified by IM or FC in 8 of 16 MGUS patients, 4 of 8 with SMM, and 11 of 15 with MM. In all cases in which IM or FC detected clonal cells, the ASO-PCR was positive. This study shows that, by using ASO-PCR, clonal cells can be found at very low levels in the blood in most patients with MGUS. However, the number of clonal cells in the blood of MGUS patients is less than those with overt MM (P = .006). In contrast to MGUS, patients with active MM are more likely to have identifiable clonal circulating plasma cells (P = .05).

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MATERIALS AND METHODS

Patient selection. The patients included in this study were seen and evaluated in the Dysproteinemia Clinic at the Mayo Clinic (Rochester, MN) between August 1993 and March 1995. The study was conducted with the approval of the Institutional Review Board of the Mayo Clinic/Mayo Foundation and the patients gave informed consent to provide additional blood and marrow samples when these tests were ordered for clinical purposes. Patients were diagnosed as MGUS by their treating physician using standard clinical and laboratory criteria, i.e., they had a serum monoclonal protein (M-protein) value less than 3 g/dL, ≤10% plasma cells in the marrow, no or a small amount of M-protein in the urine (Bence Jones proteinuria), absence of lytic bone lesions, no anemia due to the plasma cell proliferation, and the production of monoclonal gammopathy of undetermined significance (MGUS), smoldering multiple myeloma (SMM), and multiple myeloma (MM), are characterized by the monoclonal expansion of the plasma cell compartment within the bone marrow (BM) as determined by Ig gene rearrangements (P < .05).

THE PLASMA CELL proliferative disorders, which include monoclonal gammopathy of undetermined significance (MGUS), smoldering multiple myeloma (SMM), and multiple myeloma (MM), are characterized by the monoclonal expansion of the plasma cell compartment within the bone marrow (BM) as determined by Ig gene rearrangements and the production of a monoclonal M-protein in the serum or urine. MGUS is relatively common and is detected in approximately 1% of adults more than 50 years of age and in 3% of those more than 70 years of age.1,2 Long-term follow-up studies of MGUS patients have documented that about 20%3 and 24%4 of MGUS patients develop MM, amyloidosis (AL type), or a lymphoproliferative disease after 10 and 22 years of follow-up, respectively. Because most patients never develop an overt malignancy, treatment should be avoided and the patient should be observed regularly for symptoms or signs of transformation to active disease.

It has been shown by morphologic, immunofluorescence microscopy (IM), flow cytometric (FC), and molecular techniques that malignant cells circulate in the blood of most patients with active MM.5–51 However, it has been difficult to show clonal cells in the blood of patients with MGUS using conventional techniques.5,12–17 The complementarity determining region 3 (CDR3) sequence generated during heavy chain Ig gene rearrangement provides a unique genetic fingerprint for any B-cell clone. The use of the polymerase chain reaction (PCR) along with allele-specific oligonucleotides (ASO) designed to detect the CDR3 sequence of the tumor has proven to be an invaluable tool in monitoring minimal residual disease15–21 and in monitoring purging efficacy for autologous blood stem cell transplants in MM.22–24 We have recently described the use of ASO-PCR to detect malignant cells in the blood of patients with MM.27 This technique is highly specific and is able to detect 1 malignant cell in a background of 100,000 normal cells.

In this study, the ASO-PCR technique was performed on blood mononuclear cells (BMNCs) from patients with MGUS, SMM, indolent MM (IMM), AL-amyloidosis, and active MM. The goals were to learn if the ASO-PCR technique could detect clonal cells in the blood of patients with MGUS and to compare these results with those obtained from patients with SMM and active MM. To determine if monoclonal plasma cells could be observed in the blood of patients with MGUS, we also analyzed the same blood specimens used for ASO-PCR with IM and FC.

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proliferation, hypercalcemia, or renal insufficiency. Because these patients were diagnosed within the last 2 years, it was not possible to fulfill one important criteria of MGUS, ie, stability of the M-protein during long-term follow-up. Patients with SMM had \( \geq 10\% \) monoclonal marrow plasma cells, a monoclonal protein in the serum or urine, no known lytic or sclerotic bone lesions, a hemoglobin level greater than 10 g/dL, normal serum calcium and creatinine, a clinical diagnosis of SMM by the treating hematologist, no prior chemotherapy, and the recommendation that the patient be observed without chemotherapy. IMM patients met the same criteria as SMM patients, except for the presence of anemia felt to be due to the plasma cell proliferation or asymptomatic lytic bone lesions. Patients with active MM had either newly diagnosed or relapsed MM that required chemotherapy. The patients with AL-amyloid had monoclonal marrow plasma cells, a monoclonal protein in the serum or urine, no known lytic or sclerotic bone lesions, and a tissue biopsy positive for amyloid.

Sample collection and distribution. Patient marrow and blood samples were split and analyzed as outlined in Fig 1. Monoclonal plasma cells and the plasma cell labeling index were determined on the marrow cells as previously described. Mononuclear cells were isolated on Ficoll-Hypaque from heparinized blood and split into two samples. No procedures were performed to specifically remove adherent cells. One sample was used for the IM technique and three-color (CD38/CD45/CD138 or CD79a) FC, the other was used for ASO-PCR. The IM, FC, and ASO-PCR tests were performed by separate laboratory personnel blinded to the results of the other tests or the clinical status of the patient. The results from each of the three methods were reported as a percentage of the MNCs for ease of comparison.

FC and IM. The presence and number of circulating monoclonal plasma cells and the blood labeling index (BLI) were obtained by an IM procedure as previously described. Monoclonal plasma cells were identified by their morphology (eccentric nucleus with abundant cytoplasm) and by in situ visual documentation of cytoplasmic Ig (cIg) light chain restriction. Three-color FC was performed as previously described. Briefly, 2 \( \times 10^8 \) MNC were suspended in 100 \( \mu \)L of phosphate-buffered saline (PBS); 10 \( \mu \)L of anti-human CD45-PerCP (Becton Dickinson, Mountain View, CA) and 10 \( \mu \)L of anti-human CD38-phycocerythin (PE; Becton Dickinson) were added to the cells; and the mixture was incubated in the dark for 15 minutes at room temperature and then washed with PBS. Four milliliters of lysosolvensin solution was added to the cell pellet and incubated for 30 minutes in the dark at 4°C. After washing with 3 mL of PBS/3% bovine serum albumin (BSA), the pellet was resuspended in 200 \( \mu \)L PBS/BSA and 100 \( \mu \)L was added to each of two 12- \( \times \) 75-mm tubes. To these tubes were added either fluorescein isothiocyanate (FITC)-conjugated anti-human \( \kappa \) or \( \lambda \) light chain antibody (BioSource International, Camarillo, CA), and the incubation was suspended for 30 minutes in the dark at 4°C. After incubation, the cells were washed with 2 mL PBS, resuspended in 0.5 mL 1% paraformaldehyde, and analyzed on a FACScan FC. Gates were set around the CD38−CD45−CD45s\(^{dim} \) quadrant and the percentage of cells expressing \( \kappa \) and \( \lambda \) were compared to determine monoclonality. CD45s\(^{dim} \) cells were those that occupied the gated region between the cells in the sample that did not stain for CD38 or CD45 and the normal lymphocytes with their characteristic strong CD45 expression. Monoclonal \( \kappa \) was defined as \( \kappa:\lambda \) ratio \( \geq 5 \). Monoclonal \( \lambda \) was defined as a \( \kappa:\lambda \) ratio \( \leq 0.5 \).

DNA and RNA isolation from patient samples. DNA was prepared from the marrow and BMNCs of patients by salt-out extraction of 1 \( \times 10^7 \) cells. Briefly, the cell pellet was resuspended in 1 mL of TE (10 mmol/L Tris, pH 8.0, and 1 mmol/L EDTA) and then 2 mL of TNE (10 mmol/L Tris; pH 8.0, 1 mmol/L EDTA, 50 mmol/L NaCl, and 2% sodium dodecyl sulfate (SDS)) was added to lyse the cells. One milliliter of 5 mmol/L NH\(_4\)Ac was then added, vortexed for 15 seconds, and placed on ice for 30 minutes to facilitate the precipitation of the proteins. Precipitated proteins were pelleted by centrifugation at 3,000 RPM for 15 minutes. The supernatant was saved and the DNA was precipitated by the addition of an equal volume of isopropanol. The DNA precipitate was washed 2× in 70% ethanol, air-dried, and resuspended in 500 mL TE. RNA was prepared from 1 \( \times 10^7 \) bone marrow cells using TRizol reagent (GIBCO-BRL, Gaithersburg, MD) per the manufacturer’s protocol.

Consensus PCR of rearranged light genes. One microgram of marrow DNA was subjected to V\( \beta \)3/\( \text{JH} \) or VH family-specific/JH consensus Ig heavy chain PCR, as previously described. In some cases, use of a V\( \beta \)r1c oligonucleotide was used for consensus amplification with the following modifications: the final Mg\(_2\)Cl\(_2\) concentration was 1.2 mmol/L and the cycling conditions were an initial denaturation of 5 minutes at 94°C followed by 40 cycles of denaturation at 92°C for 15 seconds, annealing at 60°C for 15 seconds, and extension at 74°C for 25 seconds. In some cases, 5 \( \mu \)g of RNA was reverse transcribed using isotype-specific constant region primers and then amplified using the VH family-specific primers or V\( \beta \)r1c along with an internal constant region primer, as previously described. PCR products were separated on 7% polyacrylamide gels, isolated, and sequenced as previously described. Along with each patient sample, a normal marrow is run as a control. In the absence of a clonally expanded population, we do not obtain a single PCR product, but rather a nonspecific smear. To further validate the ASO-PCR results, three samples from normal donors at the Mayo Clinic laboratory were sent blinded to the University of Minnesota laboratory and none yielded a consensus product.

Detection and quantitation of clonal circulating cells by ASO-PCR. Allele-specific oligonucleotides were generated using a PCR-Mate oligo synthesizer (ABI, Foster City, CA) to the most specific portion of the CDR3 region based on BLAST (National Center for Biotechnology Information [NCBI]) sequence search comparisons. ASO-PCR was performed as previously described. Briefly, patient marrow DNA dilution curves were generated by serially diluting the patient DNA into sterile ddH\(_2\)O in 10-fold decrements. An appropriate amount of patient diluted marrow DNA was then mixed with normal marrow DNA to yield a target concentration of 1 \( \mu \)g DNA/50 \( \mu \)L reaction. Along with the diluted marrow samples, 1 \( \mu \)g of patient blood lymphocyte DNA was run in triplicate. The 1\( \times \)PCR buffer consisted of 20 mmol/L Tris, pH 8.4, 50 mmol/L KCl, 1.5 mmol/L MgCl\(_2\), 200 mmol/L of each dNTP, 20 pmol of each oligonucleotide, and 2.5 U of Taq polymerase (Perkin Elmer Cetus, Branchburg, NJ). The ASO in each instance was paired with a JH-specific or JH-consensus oligonucleotide. PCR cycling conditions were as follows: an initial 15 minutes of denaturation at 95°C followed by 15 seconds at 92°C, 15 seconds at the appropriate annealing temperature for the ASO, and 25 seconds at 74°C. Annealing temperatures varied between 58°C and 65°C depending on the calculated Tm of the ASO. To visualize the PCR products, the ASO was end-labeled with \( ^{32} \)P-ATP as described and the PCR product was

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**Fig 1. Schema for the analysis of bone marrow (BM) and peripheral blood (PB) specimens.**

**BM MORPHOLOGY% PC**

- LI
- CONSENSUS PCR/SEQUENCE
- GENERATE ASO
- FLOW CYTOMETRY (CD38+45-IDIM)
- IMMUNOFLOURESCENCE MICROSCOPY

**PB**

- ASO-PCR

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RESULTS

Patient population. Sixty-four patients with monoclonal gammopathies were accrued to the study and patient marrow and BMNC samples were analyzed as diagrammed in Fig 1. The diagnoses of the 64 patients included 24 MGUS, 7 AL-amyloidosis, 9 SMM, 2 IMM, and 22 active MM.

Results were obtained with IM or FC in all 64 patients, whereas only 43 of 64 yielded PCR products after consensus amplification of the marrow tumor-rearranged IgH gene. These 43 patients became the focus of our comparative analysis of the ASO-PCR, IM, and FC techniques. Of the 43 patients, there were 16 MGUS, 2 AL-amyloidosis, 8 SMM, 2 IMM, and 15 active MM (9 were new and untreated and 6 were relapsed).

MGUS. We have previously shown that the blood of patients with active MM contains circulating cells related to the plasma cell tumor as determined by ASO-PCR. However, the exact morphologic identification of these cells was not established because ASO-PCR detects all B cells related to the malignant clone regardless of morphology. In an attempt to detect clonal circulating cells in MGUS and to determine the morphologic characterization of the circulating clone, we used ASO-PCR, IM, and FC on the same group of patients. The results of the detection and quantitation of clonal cells in the blood are summarized in Table 1. For patients with MGUS, the ASO-PCR technique detected clonal cells in 13 of 16; the remaining 3 were below the detection level of the assay. Of the 13 patients that showed blood involvement, only 4 were >0.1% and the remaining 9 ranged from 0.001% to 0.07%, with a median blood involvement of 0.025% (Table 1). DNA samples that were negative by ASO-PCR were also amplified with N-ras gene-specific primers to confirm the integrity of the blood DNA sample (data not shown).

The IM technique detected circulating monoclonal plasma cells in 4 of 14 cases, and FC detected clonal CD38⁺CD45⁻CD45dim cells in 4 of 16 cases (Table 1). The ASO-PCR technique detected clonal cells in each of the 6 patients with MGUS in which either the IM or FC method was positive. In the 4 cases in which FC detected clonal

![Image](https://via.placeholder.com/150)
CD38+CD45+CD45^dim cells, only 2 of these showed detectable circulating clonal plasma cells using IM. This finding suggests that, in some patients with MGUS, clonal cells lacking typical plasma cell morphology can be found in the blood. Figure 2A through C shows the results from a patient with MGUS who was positive by ASO-PCR, and FC showed monoclonal CD38+CD45+CD45^dim cells. However, although plasmacytoid cells were seen (Fig 2B), they did not meet the criteria for monoclonal plasma cells by the IM technique. Figure 2D through E is an example of a patient with MGUS who had a positive analysis with ASO-PCR, circulating monoclonal plasma cells by IM, and clonal CD38+CD45+CD45^dim cells by FC. Because of the limits in sensitivity of the immunologic techniques, the remaining 8 cases were negative by both IM and FC but positive by the more sensitive ASO-PCR.

**SMM.** Of the 8 SMM analyzed by ASO-PCR, 6 were positive and 2 were below the detection level of the assay (Table 1). One patient had blood involvement that was estimated to be 10% by ASO-PCR. The remaining 5 patients had blood involvement between 0.001% and 0.06%, with a median blood involvement of 0.02% for all 8 patients. IM or FC showed clonal cells in 4 of 8 patients, and the ASO-PCR technique detected clonal cells in each of the 4 patients with SMM in which either the IM or FC method were positive. The percentages of blood involvement as determined by each method were generally in good agreement; however, ASO-PCR analysis of SMM found an estimated blood involvement of 10%, which is significantly different from that observed by both IM and FC (0.02% and 0.01%, respectively). The significance of this discrepancy is discussed below. In the remaining cases that were negative by the cellular techniques (SMM4 and SMM5) the percentage of blood involvement as determined by ASO-PCR may be below the detection limits of the other two methods.

**IMM and AL amyloidosis.** All 4 cases of IMM and amyloidosis were positive by ASO-PCR (Table 1). Neither the IM nor FC technique detected clonal cells in the patients with amyloidosis, and the level of involvement detected by ASO-PCR was similar to that found in MGUS. In 1 of the IMM patients, large numbers of clonal cells were detected by both IM and FC.

**Active MM.** ASO-PCR analysis of the 15 MM patients showed 13 patients with clonal cells in the blood; 2 were below the detection limits of the assay (Table 1). IM and FC of the MM samples showed circulating cells in 10 of 14 and 8 of 13 patients, respectively. As found for the patients with MGUS and SMM, the ASO-PCR technique detected clonal cells in each of the 11 patients with MM in which either the IM or FC method were positive.

**MGUS versus SMM versus active MM.** The percentages of patients with MGUS, SMM, and active MM that had clonal plasma cells or CD38+CD45+CD45^dim cells detected by IM or FC were 38%, 50%, and 73%, respectively (P = .05). Patients with active MM were more likely to have clonal plasma cells detected by these methods than patients with MGUS (P = .05). When patients with active disease (IMM or MM) were compared with those with inactive disease (MGUS, SMM, or amyloid), they were also more likely to have clonal plasma cells detected by FC or IM (P = .04).

For the comparison of the percentage of clonal cells detected by ASO-PCR for the different plasma cell proliferative diseases, the patients with amyloidosis were grouped with MGUS and those with IMM were grouped with active MM (Table 2). To increase the numbers of patients for this analysis, we also included the ASO-PCR results from 14 patients with MM from our previous report. In contrast to MGUS and SMM, patients with MM had a wide range of blood involvement as determined by ASO-PCR (Fig 3). The difference between the percentage of clonal cells detected by ASO-PCR in the three groups was statistically significant (P = .01). The patients with active MM had a statistically significant increase in the percentage of clonal cells in the blood when compared with those with MGUS (P = .006) and a borderline significant increase compared with those with SMM (P = .06). There was no difference between the patients with SMM and those with MGUS (P = .62).

**DISCUSSION**

The purpose of this study was to directly compare different techniques that serve to identify and characterize cell populations in the blood that are clonally related to the expanded plasma cell population seen in the marrow. Moreover, this also provided an opportunity to compare several plasma cell disorders to determine if there are differences in blood involvement.

It has been difficult to detect clonal cells in the blood of patients with MGUS using traditional techniques. In this study, we showed that, with ASO-PCR, most patients with MGUS have very small numbers of clonal cells in the blood. Previous analyses of the blood from patients with MGUS using immunohistochemistry or FC with antibodies directed against B-cell and plasma cell-associated antigens were not able to detect circulating monoclonal plasma cells. In addition, four studies using Southern blotting of DNA from patients with MGUS were unable to identify B-cell...
Table 2. Percentage of Clonal Cells in the Blood Detected by ASO-PCR in the Various Plasma Cell Proliferative Groups

<table>
<thead>
<tr>
<th>Diagnosis (n)</th>
<th>Mean</th>
<th>Median</th>
<th>Range</th>
</tr>
</thead>
<tbody>
<tr>
<td>MGUS/amyloid (18)</td>
<td>0.06</td>
<td>0.02</td>
<td>0-0.3</td>
</tr>
<tr>
<td>SMM (8)</td>
<td>1.23</td>
<td>0.02</td>
<td>0-10</td>
</tr>
<tr>
<td>IMM/active MM (31)</td>
<td>1.85</td>
<td>0.21</td>
<td>0-33</td>
</tr>
</tbody>
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gene rearrangements in the blood cells. When cytokines were added to blood cells and incubated for several days, plasma cell outgrowths were obtained from the patients with MM but not from those with MGUS. Büchi et al. studied 23 patients with MGUS and found no alteration in the ratio of blood lymphocytes; however, alterations have been found by others. A B-cell clone from the blood of a patient with MGUS was isolated and demonstrated to have an idiotype determinant similar to the serum M-protein and to contain an identical Ig gene rearrangement. Taken together, these data appear contradictory concerning blood involvement in MGUS. However, in light of our data showing the very low percentage of blood involvement in MGUS compared with MM, it is not surprising that in most instances clonal circulating cells were not detected.

The presence of circulating cells clonally related to the marrow tumor has been shown in previous studies by ASO-PCR and IM for patients with active MM. In fact, the number and labeling index of the circulating cells is a marker of disease activity. The results in this study confirm and extend these results by showing that the percentage of blood involvement is typically higher in MM than in MGUS but varies among patients by as much as 4 logs (Fig 3). Because low levels of clonal circulating cells can be found in MGUS and MM, their presence cannot be used to distinguish these diseases.

In this study, we showed that monoclonal plasma cells and CD38⁺CD45⁻CD45dim cells can be detected in some cases of MGUS by IM and FC. However, in 10 of the patients with MGUS and amyloid that had clonal cells detected by ASO-PCR, we were unable to detect monoclonal plasma cells or CD38⁺CD45⁻CD45dim cells in the blood. In addition, there were several cases (SMM6 and MM3) in which substantially more clonal cells were detected by ASO-PCR than by the IM and FC techniques. In the case of the MGUS and amyloid cases, the discrepancy could be due to the fact that plasma cells were indeed present but below the limits of detection by IM and FC. In the cases in which more cells were detected by ASO-PCR, it is likely that the clonal cells detected by ASO-PCR were less differentiated and that all of them did not have plasma cell morphology or a CD38⁺CD45⁻CD45dim immunophenotype. Further cell sorting studies are needed to examine which B-cell population contains the clonal cells and correlations with overall survival will provide clues to the significance of these different clonal cell populations.

By analyzing the same blood samples with each of the ASO-PCR, IM, and FC techniques, we found that whenever IM and FC detected monoclonal cells in the blood, we always detected clonal cells by ASO-PCR, ie, there were no false-positives with IM or FC. In addition, when ASO-PCR was negative, the IM and FC techniques were also always negative, ie, there were no false-negatives by ASO-PCR. This study indicates that, when molecular techniques detect clonal cells in the blood of patients with monoclonal gammopathies, IM and FC may show cells with plasma cell morphology and immunophenotype. In some instances, FC detected clonal cells at levels that would be expected to have been detected by IM, but the lack of typical plasma cell morphology precluded their detection. Certainly, the FC and the IM methods are not equivalent and provide different measures of clonally related cells. The ASO-PCR technique is much more sensitive and has the advantage of sequence specificity to confirm that the cells are clonally related to...
the plasma cell tumor regardless of their morphology; therefore, the population of cells identified by ASO-PCR would include cells with plasma cell morphology (if they were present) and clonally related cells without plasma cell morphology. To learn the immunophenotype of the clonal cells that do not have the morphology of plasma cells, sorting specific cell populations followed by in situ hybridization using the ASO will be required. These experiments will be important to properly identify the cell population that should be targeted with new therapeutic approaches.

The finding of clonal cells in the blood of patients with MGUS has implications about the way we view the pathogenesis of MM. It is clear that, even at this early, premalignant stage of disease, the clone is present in the blood and has differentiated to the plasma cell stage in the marrow. Because most of these patients will never actually progress to the malignant phase of the disease, there must be additional insults to the plasma cell or precursor cells to cause them to proliferate, secrete cytokines, and produce the signs and symptoms of active myeloma.

Although monoclonal plasma cells were absent or in very low amount in the blood of patients with MGUS, they were easily detected in the marrow of all these patients by IM. Therefore, patients with MGUS and MM are similar in that they both have clonal plasma cells in the marrow, a monoclonal protein in the serum or urine, and clonal cells in the blood. They differ in that patients with MM typically have more M-protein, a greater percentage of plasma cells in the marrow, marrow plasma cells with a higher proliferative rate, and lytic bone lesions. Previous studies have found that, in contrast to active myeloma, marrow plasma cells in patients with MGUS have a low proliferative rate. This reduces the probability that blood involvement in patients with MGUS results from spillover of cells proliferating in the marrow compartment. In addition, a recent report by Sahota et al has shown that cells isolated from the marrow of patients with MGUS show clonally related populations with distinct somatic mutations. These results suggest that cells from patients with MGUS retain clonal diversification and raise the possibility that detection of clonal cells in the blood may reflect cells in transit from distal sites that home to the marrow and differentiate into mature plasma cells. Once in the marrow, the proliferative capacity of MGUS cells is significantly reduced compared with the plasma cells typically found in patients with active myeloma.

We have also shown that the blood from patients with SMM, IMM, and amyloidosis can contain clonal circulating cells in the blood. It is now apparent that clonal cells can circulate in the blood of patients with all of the plasma cell proliferative diseases. We have previously shown that patients with SMM who have an increased number of monoclonal plasma cells in the bone have a shorter time to progression to overt MM than those patients who do not. A recent study by Isaksson et al showed that patients with MGUS who had excess blood B lymphocytes expressing the same light chain isotype as the patient’s M-protein had an increased risk of transformation to a B-cell malignancy. It will be interesting to learn with long-term follow-up studies whether the results of the IM, FC, and ASO-PCR studies on the patients with MGUS in this study can predict transformation to overt malignant disease.

This study shows by both cellular and molecular approaches that clonal circulating cells are a common occurrence in plasma cell disorders. Furthermore, the majority of patients with MM have circulating plasma cells as determined by IM and FC that may reflect their active disease state. This is in contrast to what is observed in MGUS, in which relatively few patients show clonal cells as determined by either cellular method, but are clearly positive by ASO-PCR. It is clear that clonal circulating cells are evident in most patients with MGUS; however, the identification of such cells and their impact on possible progression to overt MM will require longitudinal studies. Based on these results, monoclonal gammopathies should be considered hematopoietic disorders involving many lymphoid compartments rather than purely marrow disorders.

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Clonal circulating cells are common in plasma cell proliferative disorders: a comparison of monoclonal gammopathy of undetermined significance, smoldering multiple myeloma, and active myeloma

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