In Vivo Neutrophil and Lymphocyte Function Studies in a Patient With Leukocyte Adhesion Deficiency Type II

By Thomas H. Price, Hans D. Ochs, R. Gershoni-Baruch, John M. Harlan, and Amos Etzioni

We investigated in vivo neutrophil and lymphocyte function in a patient who lacks Sialyl-Lewis-X, a ligand for the selectin family of leukocyte adhesion molecules (leukocyte adhesion deficiency II, LAD II). As assessed by skin chamber and skin window techniques, in vivo chemotaxis of neutrophils was markedly impaired (less than 6% of normal values). A marginal pool was present as determined by an increase in circulating neutrophils after epinephrine injection and calculated recovery of infused radiolabeled autologous neutrophils. Kinetic studies showed a reduced half-life of 3.2 hours (normal 7 hours) and markedly increased turnover rate (cells/kg/d) of approximately eight times the normal value. A normal antibody response to the T-cell-dependent antigen bacteriophage φX174 showed that T/B-cell interaction is not affected in LAD II. These findings provide direct evidence that the selectin family and its ligands play an important role in neutrophil function. © 1994 by The American Society of Hematology.

MATERIALS AND METHODS

Patient. YW is a 5-year-old boy who was noted within the first year of life to have severe mental retardation, short stature, a distinctive facial appearance, neutropenia, recurrent bacterial infections, and the Bombay (hh) blood phenotype. A detailed clinical description of this patient was previously reported. The patient's neutrophil count is typically 20 to 35 × 10⁹/L when he is free of infection and usually exceeds 70 × 10⁹/L during episodes of infection. Severe periodontitis has persisted from early life. In the last 3 years, the frequency of infection decreased and prophylactic antibiotics were discontinued. Informed consent for all studies was given by the patient's parents. All studies were approved by either the University of Washington or the Rambam Medical Center Human Subjects Review Committee.

Hematologic values. Hematocrit, platelet count, white blood cell count, and differential counts were determined using standard hematologic techniques. Lymphocyte subsets were determined by flow cytometry using appropriate monoclonal antibodies (MoAbs).

In vivo chemotaxis. The neutrophil response to cutaneous inflammation was assessed by both skin-chamber and skin-window techniques. Skin-chamber studies were performed as previously described. A 1-cm² area on the left-volar forearm was abraded with a scalpel blade and the abrasion covered with a 10-mL sterile glass chamber that was fastened in place by adhesive compound. The chamber was filled with fluid consisting of 10% autologous serum

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in normal saline to which 100 U/mL streptokinase/streptodornase had been added. The fluid was removed after 24 hours and the cell count and differential count determined. Skin-window leucocyte accumulation was assessed by the technique of Dale et al. A 3- x 3-mm abrasion was created on the right-volar forearm by scalpel blade as above. Sterile 18-mm round cover slips were applied to the abrasion, covered with a sterile cardboard square, and changed at 1, 3, 5, 7, 9, 12, and 24 hours. Differential counts were performed after staining with Wright's stain. The number of cells in the most cellular 1 mm² of the coverslip was quantitated by counting the cells in a photograph of the field. Results were expressed as the geometric mean response, as described by Dale et al.

**Blood neutrophil kinetics.** Neutrophil kinetics were determined as previously described. Seventy-five milliliters of blood was injected subcutaneously and blood samples were obtained for neutrophil and specific activity, the blood was reinfused over a 10-minute period. Three-milliliter blood samples were then obtained at zero-time extrapolation of the disappearance curve. Neutrophil pool sizes were based on an assumed blood volume of 840 mL/kg.

**Epinephrine test.** Epinephrine (0.2 mL; 1 mg/mL; 1:1000) was injected subcutaneously and blood samples were obtained for neutrophil count at 0, 5, 10, and 30 minutes.

**Immunization with bacteriophage φX174.** Bacteriophage φX174 (phage) was produced as previously described. The final concentration was adjusted to 1 x 10¹¹ plaque-forming units (PFU)/mL. After obtaining informed consent, phage was administered intravenously at a dose of 2 x 10⁹ PFU/kg body weight. A second immunization was given 6 weeks later. Normal controls and the patient with LAD I received a tertiary immunization 6 or more weeks after the secondary immunization. Blood samples for antibody titers were collected immediately before immunization and at 1, 2 and 4 weeks after each immunization. Antibody activity was determined by a neutralizing antibody assay and expressed as the first order rate constant (Kv), or phage inactivation, as described. Neutralizing antibody resistant to 2-mercaptoethanol was considered to be IgG. For comparison, the LAD II patient, a patient with absence of the β₂ subunit of the leukocyte integrin (LAD I), and 22 young adults were immunized using the same protocol.

**RESULTS**

**Hematologic values.** The patient's hematocrit was 34% and platelets were 469 x 10⁹/L. His total leukocyte count was 35.4 x 10⁹/L with 23.4 x 10⁹ segmented neutrophils/L, no bands, 9.6 x 10⁹ lymphocytes/L, 2.1 x 10⁹ monocytes/L, and 0.3 x 10⁹ eosinophils/L. Lymphocyte subset populations (x10⁹/L) were 0.8 B cells (CD20); 7.2 T cells (CD3); 6.0 CD4⁺; 1.6 CD8⁺; and 0.1 natural killer (NK) cells (CD16). At the time of the studies, the patient was free of infection.

**Blood neutrophil kinetics.** The results of the kinetic studies are shown in Table 1. Neutrophil emigration was markedly diminished in both tests, the values being ≈1.5% and 6% of normal in the skin-window and skin-chamber tests, respectively. Monocyte migration to the skin-window site was reduced to a similar degree.

**Epinephrine test.** Thirty minutes after the administration of 0.2 mL epinephrine an increase in the neutrophil count from 28,700/mm² to 34,800/mm² was observed, indicating that 82% of the blood neutrophils were in the circulating pool by this measure. Thus, a neutrophil marginal pool does exist in this patient, although apparently reduced when compared with normal.

**Antibody responses to bacteriophage φX174.** In contrast to the patient lacking the β₂ subunit of the leukocyte integrin, the LAD II patient had a normal and high-normal primary and secondary response with normal switch from IgM to IgG (Fig 2). In contrast, the antibody response of the LAD I patient was depressed, the switch from IgM to IgG was incomplete, and the antibody titers decreased quickly to very low values.

**DISCUSSION**

The interaction between neutrophils and endothelial cells, critical for neutrophil localization to and migration across endothelium, is mediated by surface molecules on both neutrophils and endothelial cells. Three families of adhesion molecules involved in several phases of emigration have been described, acting at different steps of the adhesion pro-

**Table 1. In Vivo Chemotaxis**

<table>
<thead>
<tr>
<th></th>
<th>Skin Window</th>
<th>Skin Chamber</th>
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<tbody>
<tr>
<td>Neutrophils</td>
<td>2051 (25-1,864)</td>
<td>1021 (16-644)</td>
</tr>
<tr>
<td>Monocytes</td>
<td>2</td>
<td>1</td>
</tr>
<tr>
<td>PMN</td>
<td>3</td>
<td>3</td>
</tr>
</tbody>
</table>

*No. of polymorphonuclear leukocytes x 10⁶ per cm² of abrasion.
† Geometric mean response (95% confidence range); normal values are from Dale et al., n = 10.
‡ Mean (range), n = 23; 96% ± 2% of these were PMNs.

**Table 2. Neutrophil Kinetics**

<table>
<thead>
<tr>
<th></th>
<th>Patient</th>
<th>Normal*</th>
</tr>
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<tbody>
<tr>
<td>Circulating PMN (x10⁹/kg)</td>
<td>1.76</td>
<td>0.19 ± 0.06</td>
</tr>
<tr>
<td>Recovery (circulating pool/total pool)</td>
<td>0.81</td>
<td>0.42 ± 0.11</td>
</tr>
<tr>
<td>Blood half life (hr)</td>
<td>3.2</td>
<td>7.8 ± 2.4</td>
</tr>
<tr>
<td>PMN turnover (x10⁹/kg/d)</td>
<td>11.4</td>
<td>1.49 ± 0.39</td>
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* Mean ± SD.
† From Dancy et al.
‡ From Price.

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IN VIVO LEUKOCYTE FUNCTION IN LAD TYPE II

Fig 1. Disappearance of neutrophil radioactivity after intravenous injection of \(^{3}H\)-DFP-labeled autologous neutrophils.

cess; selectins, integrins, and the Ig superfamily. The selectins (E, L, and P-selectin) mediate neutrophil "rolling" wherein, under conditions of flow, the neutrophils become loosely adherent to the endothelium.\(^7\)\(^11\) One neutrophil surface ligand for E and P-selectins has recently been determined to be the tetrasaccharide, SLeX.\(^28\)\(^34\) Once the neutrophil, through rolling, is brought into contact with the endothelial cell, the \(\beta_2\) integrin (CD11/CD18) on the surface of the neutrophil and the Ig superfamily molecule (intracellular adhesion molecule 1) on endothelial cells interact to produce firm attachment and to facilitate neutrophil migration between the endothelial cells to tissue.\(^12\)\(^35\) The importance of the integrin receptors for neutrophil emigration has been shown by the identification of patients whose neutrophils lack these molecules. The leukocyte adhesion deficiency (LAD I) syndrome is characterized by a defective CD18 gene, absence or deficiency of the CD11/CD18 complex, inability to deliver neutrophils to sites of infection, and recurrent bacterial infection.\(^96\)

A second leukocyte adhesion molecule defect, LAD II, has recently been described in two unrelated children.\(^17\)\(^18\) Neutrophils from these patients have been shown in vitro to have both motility and adherence defects. The LAD II patients apparently have a general defect in fucose metabolism. As a consequence, they exhibit the Bombay blood phenotype and lack the carbohydrate SLeX on the surface of their neutrophils. SLeX is a ligand for both E-selectin and P-selectin,\(^32\) and in vitro studies have shown that neutrophils from these patients are incapable of binding to the endothelial selectins.\(^18\)\(^30\)a

The present studies were undertaken to determine the effect of this abnormality in the selectin system on in vivo leukocyte function. The skin-window and skin-chamber studies clearly demonstrate a marked abnormality in both neutrophil and monocyte migration to inflammatory sites. The magnitude of the abnormality in emigration seen in this patient is similar to that previously reported in patients with CD18 deficiency\(^15\) in spite of the fact that the latter appear to have more serious problems with bacterial infection. This discrepancy may reflect the limited ability of the skin-chamber and skin-window assays to predict neutrophil traffic to sites of infection, but may also be attributed to the fact that the defect in LAD I also affects lymphocyte function by interference with T/B-cell interaction.\(^36\)\(^37\) It remains to be determined to what extent T/B cell interactions are affected in LAD II.

While a similar defect in in vivo chemotaxis was observed between LAD I and LAD II patients, our findings in regards to neutrophil kinetics in the LAD II patient are different from previously described findings in LAD I patients. Several reports described a normal marginal pool in patients with LAD I.\(^38\)\(^39\) We used both the epinephrine test and the neutrophil labeling technique to show that the LAD II pa-

Fig 2. Antibody responses to bacteriophage \(\phi X174\). Phage was given at the time indicated (i). Responses presented are in normal controls (---, geometric mean; ----, 95% confidence interval), in a patient with LAD II (SLeX deficiency) (○○○) and in a patient with LAD I (\(\beta_2\) subunit [integrin] deficiency) (△△△). Antibody is expressed as phage neutralizing activity (Kv). Percent IgG of this activity is indicated for various time points.
tients also had a marginal pool that appears somewhat reduced. Normally, the marginal and circulating pools are approximately equal in size and are in dynamic equilibrium. The anatomic location of the marginal pool is uncertain. It may represent neutrophils transiently sequestered in the lung or other organs, or perhaps cells rolling along the vessels in the general microcirculation. The degree to which selectin receptors are important for the determination of the marginal pool is not clear. Our results could imply that there are two marginal pools. The first, accounting for approximately 20% of the blood neutrophils, is comprised of neutrophils sequestered in the lungs or other organs, may depend upon leukocyte size and deformability, is selectin independent, and is intact in our patient. The second, accounting for approximately 30% of the blood neutrophils, represents neutrophils rolling in the microvascular, is selectin mediated, and is absent or reduced in our patient. Such a construct would be consistent with observations that the total marginal pool size is normal in LAD I patients—neither component is integrin dependent—but reduced in LAD II patients.

Both LAD I and LAD II patients exhibit persistent neutrophilia. In LAD I, a prolonged intravascular neutrophil survival was observed, which in part may be explained by the inability of the cells to migrate from the blood vessels to the sites of infection. In our LAD II patient, the neutrophil intravascular half-disappearance time, which is normally 6 to 9 hours, was 3.2 hours. This shortened intravascular residence time was unexpected. The neutrophil turnover rate, calculated from the $t_{1/2}$ and the blood neutrophil pool size, was markedly elevated and indicated a marrow neutrophil production of approximately eight times normal. A continuing stimulus to marrow neutrophil production does not seem unreasonable in a patient unable to deliver neutrophils to sites of inflammation, but the fact that the blood neutrophil count did not increase over the period of study suggests that neutrophils were destroyed or used at a high rate somewhere other than in the tissues. It is also possible that the underlying abnormality in fucose metabolism, in addition to causing the absence of SLeX, affects the neutrophil in other ways. An artifically rapid blood neutrophil disappearance rate could result for example, if DFP binding were affected in such a patient.

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REFERENCES


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