Cyclosporin A and Cyclosporin SDZ PSC 833 Enhance Anti-CD5 Ricin A-Chain Immunotoxins in Human Leukemic T Cells

By Jean-Pierre Jaffrézou, Branimir I. Sikic, and Guy Laurent

Recent studies have shown that cyclosporin A (CsA) may affect ricin A-chain immunotoxin (RTA-IT) therapy. In this study, we evaluated the ability of CsA and its nonimmunosuppressive analog, SDZ PSC 833, to enhance anti-CD5 T101 RTA-ITs in vitro. Both 4 μmol/L CsA and 4 μmol/L SDZ PSC 833 significantly and specifically enhanced the cytotoxic activity of T101 RTA-IT on the human lymphoblastic T-cell line, CEM III (101-fold and 105-fold, respectively). Furthermore, these CsA also enhanced the cytotoxicity of the more potent T101 F(ab)2 RTA-IT (ninefold and eightfold, respectively). The effect of human plasma, originating from four patients enrolled in a phase I high-dose CsA regimen, was examined on T101 RTA-IT cytotoxicity on CEM III cells. In each case, with plasma CsA levels between 3,060 and 4,860 ng/mL (2.5 to 4 μmol/L), a significant increase in T101 RTA-IT–mediated cytotoxicity was observed ranging from 31% to 60%. Neither CsA nor SDZ PSC 833 affected the rate of RTA-IT binding, internalization, intracellular trafficking, or degradation. Analysis of internalized T101 RTA-IT molecules showed that these were essentially intact, which suggests that these enhancers may act only on a small population of RTA-ITs that escapes present investigational techniques. In conclusion, because the concentrations used are clinically achievable, CsA appear to be promising agents for in vivo enhancement of RTA-ITs.

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IMMUNOTOXINS (ITs), which are composed of a monoclonal antibody (MoAb) coupled to a plant or bacterial toxin, are potentially highly cytotoxic and specific agents for the targeting of cancer cells. Ricin A-chain immunotoxins (RTA-ITs), for example, are among the most potent ITs being evaluated for their potential use as anticancer agents. T101 RTA-IT and T101 F(ab)2 RTA-IT directed against the lymphoid differentiation antigen, CD5, have been found in vitro to be both specific and cytotoxic to lymphoid leukemia cells. However, compared with the native ricin toxin, these RTA-ITs, especially T101 RTA-IT, present limited antitumor efficacy probably because of the low rate of cell intoxication and/or unfavorable intracellular trafficking.

One of the most active protein inhibitors at the acellular level, ricin A-chain is comparable with other toxins of bacterial (diphtheria toxin and Pseudomonas exotoxin) and higher-plant (pokeweed antiviral protein and gelonin) origin. However, the absence of the ricin B-chain, which plays a role in facilitating the translocation of the A-chain from the endosome to the cytosol, greatly decreases the activity of RTA-ITs. Therefore, although these conjugates show stringent specific cytotoxicity, their clinical potential depend on ways of increasing their cytotoxic activity.

Potentiation of RTA-ITs, including T101 RTA-IT, cytotoxicity has been achieved in vitro with the use of agents such as carboxylic ionophores, l-lysosomotropic amines, and calcium antagonists. Some of these enhancers can be used ex vivo such as in bone marrow (BM) purging in humans, but most of them are ill-suited for in vivo use where the major obstacles are either their toxicity or short half-life after intravenous administration.

Recent studies have suggested that cyclosporin A (CsA) increases the efficacy/toxicity of certain RTA-ITs. Therefore, we evaluated the potential RTA-IT enhancement effect of CsA and its nonimmunosuppressive analog, SDZ PSC 833. Both CsA and SDZ PSC 833 were found to be both potent and specific enhancers of T101 and T101 F(ab)2 RTA-IT in vitro. We also investigated the impact of these RTA-IT enhancers on the rate of T101 RTA-IT intoxication, binding, internalization, intracellular degradation, and subcellular localization.

MATERIALS AND METHODS

Cells. The human lymphoblastic T-cell line CEM III is permanently maintained in our laboratory by serial passage. CEM III was subcloned from CEM using a FACs IV cell sorter (Becton Dickinson, Mountain View, CA). These cells express a mean CD5 density of 30,000 molecules per cell. Cells were cultured in RPMI 1640 medium (Mérieux, Lyon, France), supplemented with 10% heat-inactivated fetal calf serum (Flow Laboratories, McLean, VA), 2 mmol/L glutamine, and antibiotics (200 U/mL penicillin, 100 μg/mL streptomycin).

Immunotoxins. T101, T101-F(ab)2, anti-CD19 MoAb, and ricin A-chain were purified and then conjugated by the cross-linking agent N-succinimidyl-3-(2-pyridyldithio)propionate (Pharmacia, Montigny, France) as described previously. Mouse monoclonal IgG2a antibody T101 and its fragment T101-F(ab)2 react with the CD5 surface antigen. An average of two ricin A-chain molecules were linked per antibody or Fab fragment. The amount of RTA-ITs, which contained no detectable ricin B-chain, is expressed as molar concentrations of bound A-chain.

Drugs and reagents. CsA and SDZ PSC 833 were kindly supplied by Sandoz (Basel, Switzerland) and Pharmaceutical Inc (East

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EnhanCeMent of anti-CD5 immunoToxins

Hanover, NJ). These were dissolved in absolute ethanol at 1 mmol/L and stored at −20°C. All other reagents were purchased from Sigma (St Louis, MO).

Human plasma. Plasma samples were obtained from four patients enrolled in a phase I trial of high-dose CsA for modulation of multidrug resistance.24 Samples were collected before and 2 hours after CsA infusion. CsA levels were measured using a non-specific fluorescence-polarization immunoassay that cross-reacts with both CsA and metabolites (TDX; Abbott Laboratories, North Chicago, IL) and found to be between 3,090 and 4,860 ng/mL.

Protein-synthesis inhibition assay. RTA-IT efficacy was evaluated by the inhibition of [3H]-Leucine incorporation as described previously.2 Briefly, 1 X 10^6 cells/mL in complete medium were treated during 18 hours at 37°C with various concentrations of toxin or RTA-IT and, when specified, in the presence of Cs. [3H]-Leucine (specific activity, 150 Ci/mmol; 1 μCi/100 μL well; Amersham, Arlington Heights, IL) was added 6 hours before the end of treatment. Washed cells were harvested on filter paper (Cambridge Technology, Inc, Watertown, MA) and radioactivity was counted. Results of triplicate experiments are expressed as the percentage of control (cells not treated by toxin or RTA-IT). In experiments using human plasma, 2 X 10^6 CEM III cells were pelleted and resuspended in 100% human plasma. After addition of 1 X 10^4 mol/L T101 RTA-IT, the final plasma concentration was 87%.

125I labeling of T101 RTA-IT. T101 RTA-IT (50 μg) was labeled with 1 μCi of 125I (Amersham) in the presence of chloramine T.25 After 1 minute of incubation, the reaction was stopped by the addition of sodium metabisulfite followed by potassium iodide. Free 125I was removed by chromatography on a phosphate-buffered PD-10 Sephadex G-25 M column (Pharmacia, Piscataway, NJ). The collected fractions that contained labeled protein were pooled and the radioactivity counted. The specific radioactivity of labeled T101 RTA-IT was approximately 7.3 μCi/mg protein. Iodination did not modify the activity of T101 RTA-IT (data not shown).

Rate of T101 RTA-IT internalization in CEM III cells. The kinetics of T101 RTA-IT internalization was measured according to the previously described method.26 Briefly, CEM III cells were incubated for 1 hour at 4°C with 125I labeled T101 RTA-IT (at 10-fold the membrane-saturable concentration: 1 X 10^-8 mol/L) in complete medium. A negative control was performed by incubating the cells at 4°C with cold T101 RTA-IT (1 X 10^-8 mol/L) 2 hours before addition of 125I-T101 RTA-IT. After three ice-cold washes with saline solution (removing excess 125I-RTA-IT), cells (1 X 10^6/mL) were incubated at 37°C, with or without 4 μmol/L CsA, for various incubation times. At the end of these times, cells were pelleted by centrifugation and washed twice. The radioactivity of the collected supernatants, which represented non-cell-associated 125I-ligand, was measured by liquid scintillation counting. The negative control, represented less than 10% of the total recovered radioactivity. The time to reach 50% of the total recovered radioactivity was determined using a gamma counter.

Analysis of T101 RTA-IT degradation in CEM III cells. After protease treatment, washed cells were lysed with 50 mmol/L Tris-HCl, 0.5% sodium dodecyl sulfate (SDS), pH 8. Intracellular non-degraded and degraded 125I-ligand were assessed by measuring the 10% trichloroacetic acid (TCA)-precipitable and soluble radioactivities, respectively.

Percol gradient fractionation of organelles. The intracellular trafficking of 125I-T101 RTA-IT in CEM III cells was evaluated according to the previously described methods27 with slight modifications. Cells (1 X 10^6) were incubated with 1 X 10^-4 mol/L 125I-T101 RTA-IT for 4 hours at 37°C with or without 4 μmol/L CsA or 4 μmol/L SDZ PSC 833. Cells were then washed twice in ice-cold phosphate-buffered saline and resuspended in 1 mL of cold buffer (250 mmol/L sucrose, 1 mmol/L EDTA, and 10 mmol/L triethanolamine), and homogenized with 25 strokes of a Dounce homogenizer (Cole-Parmer Instrument Co, Niles, IL). After centrifugation at 250g for 10 minutes at 4°C to sediment nuclei and unbroken cells, the supernatant was recovered and carefully layered onto the surface of a tube containing 9 mL of 20% Percoll in cold buffer resting on a cushion of 0.5 mL 2.5 mol/L sucrose. Percoll gradients were generated by centrifugation at 20,000g for 90 minutes at 4°C. Fractions of 0.5 mL were collected from the top and the radioactivity counted.

Molecular-weight evaluation of internalized molecules. After 4 hours of incubation with 125I-T101 RTA-IT at 37°C, cell-surface radioactivity was eluted from CEM III cells by treatment with Streptomyces griseus protease as described above. Analysis of the molecular weight of internalized 125I-T101 RTA-IT was performed according to the previously described method.2 Cells (5 X 10^5) were solubilized for 30 minutes at 4°C in 3.5 mL of 0.5% Nonidet-P40, 1% Triton X-100, and 0.5% sodium deoxycholate containing protease inhibitors phenylmethylsulfonyl fluoride (100 μg/mL), pepstatin A (1 μg/mL), leupeptin (2 μg/mL), and aprotenin (2 μg/mL). The lysates were obtained after centrifugation at 1,000g and analyzed by SDS-polyacrylamide gel electrophoresis (SDS-PAGE) on a 5% to 15% gradient gel.24 Gels were autoradiographed at −70°C on Kodak X-OMAT AR film with Du Pont Cronex Lightning-Plus intensifying screens (Eastman Kodak, Rochester, NY) and analyzed by densitometry.

RESULTS

Enhancement of RTA-IT activity by CsA. The ability of CsA and SDZ PSC 833 to enhance anti-CD5 RTA-ITs was evaluated by treating the human lymphoblastic T-cell line CEM III with various concentrations of the drugs and RTA-IT. Figure 1 illustrates the relatively weak cytotoxicity of T101 RTA-IT compared with native ricin (>50,000-fold). Nevertheless, specificity and cytotoxicity are superior to ricin A-chain alone (about 10-fold). Coincubation of T101 RTA-IT with either 4 μmol/L CsA or 4 μmol/L SDZ PSC 833 resulted in a significant increase in cytotoxicity (Table 1). Enhancement of T101 RTA-IT cytotoxicity was achieved within a narrow range of CsA concentrations. At 1 μmol/L neither CsA nor SDZ PSC 833 had a significant effect on T101 RTA-IT cytotoxicity. A significant effect of 20- to 30-fold was observed at 2 μmol/L for both CsA, which further increased to greater than 100-fold at 4 μmol/L (a nontoxic concentration). However, CsA toxicity reached about 20% at 6 μmol/L, which prevented their use at higher concentrations (data not shown).

T101 F(ab')2 RTA-IT cytotoxicity on CEM III was about 10-fold more potent than its whole-fragment counterpart. This RTA-IT was also enhanced by CsA and SDZ PSC 833. At 2 μmol/L both CsA and SDZ PSC 833 had a limited...
effect of about twofold but this increased significantly to about 10-fold at 4 μmol/L. The enhancement effect of both CsA and SDZ PSC 833 was specific, because the toxicity of ricin A-chain alone and the control anti-CD19 RTA-IT was unaffected. Furthermore, neither T101 RTA-IT or T101 F(ab')2 RTA-IT showed any change in nonspecific toxicity on the irrelevant human Burkitt's lymphoma cell line, RAJI (data not shown).

Effect of CsA on the rate of protein-synthesis inhibition by anti-CD5 RTA-ITS. The inhibition by T101 RTA-IT (1 × 10^-8 mol/L) of protein synthesis reached approximately 50% after 24 hours, whereas in the presence of 4 μmol/L CsA and 4 μmol/L SDZ PSC 833 it increased to 90% and 94%, respectively (P < .025) (Fig 2A). Compared with its whole-fragment counterpart, T101 F(ab')2 RTA-IT, reached 94% inhibition of protein synthesis within 24 hours, which was further increased to almost 99% with both 4 μmol/L CsA and 4 μmol/L SDZ PSC 833 (P < .05) (Fig 2B).

Enhancement of T101 RTA-IT in human plasma. The cytotoxicity of T101 RTA-IT on CEM III cells was evaluated in a prospective experiment using human plasma (Fig 3). T101 RTA-IT, 1 × 10^-8 mol/L, inhibited protein synthesis in the presence of four pre-CsA control-plasma samples by 43%, 52%, 62%, and 50%, respectively. In the presence of patient plasma obtained 2 hours post-CsA infusion, T101 RTA-IT cytotoxicity increased to 61%, 77%, 85%, and 77%, respectively.

Effects of CsA on internalization and degradation of 125I-T101 RTA-IT. CEM III cells were incubated with a membrane-saturating concentration of 125I-T101 RTA-IT (1 × 10^-8 mol/L) for 1 hour at 4°C. After washing in ice-cold saline solution (removing excess 125I-RTA-IT), CEM III cells were incubated at 37°C; with or without 4 μmol/L CsA. At specified times, cells were pelleted, membrane-bound labeled ligand was removed by a protease treatment, and the radioactivity of the pellet was determined. Figure 4 shows the slow internalization rate of T101 RTA-IT in CEM III cells. Furthermore, treatment by 4 μmol/L CsA did not significantly modify the kinetics of internalization.

After protease treatment, CEM III cells were lysed and intracellular nondegraded and degraded 125I-ligand was

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Table 1. Comparative RTA-IT Enhancement on CEM III Cell Line

<table>
<thead>
<tr>
<th>RTA-IT</th>
<th>Drug</th>
<th>μmol/L</th>
<th>ICso (μmol/L)</th>
<th>Fold Enhancement</th>
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<tr>
<td>T101</td>
<td>No drug</td>
<td>25,000</td>
<td>± 5,000</td>
<td></td>
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<tr>
<td></td>
<td>CsA</td>
<td>1.0</td>
<td>18,000 ± 2,000</td>
<td>1.4</td>
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<td></td>
<td>2.0</td>
<td>1,233 ± 2,516</td>
<td>20.3</td>
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<td></td>
<td>4.0</td>
<td>247 ± 50</td>
<td>101.2</td>
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<tr>
<td></td>
<td>PSC 833</td>
<td>1.0</td>
<td>19,333 ± 1,154</td>
<td>1.3</td>
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<tr>
<td></td>
<td>2.0</td>
<td>866 ± 1,527</td>
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<td>4.0</td>
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<td>2,366 ± 321</td>
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<td>4.0</td>
<td>253 ± 25</td>
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<td>PSC 833</td>
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<td>1,953 ± 117</td>
<td>1.2</td>
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<td></td>
<td>4.0</td>
<td>290 ± 36</td>
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<tr>
<td>Anti-CD19</td>
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<td>&gt;50,000</td>
<td></td>
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</tr>
<tr>
<td></td>
<td>CsA</td>
<td>4.0</td>
<td>&gt;50,000</td>
<td></td>
</tr>
<tr>
<td></td>
<td>PSC 833</td>
<td>4.0</td>
<td>&gt;50,000</td>
<td></td>
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<tr>
<td>Ricin [A-chain]</td>
<td>No drug</td>
<td>273,333 ± 25,166</td>
<td>1.1</td>
<td></td>
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<tr>
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<td>CsA</td>
<td>4.0</td>
<td>240,000 ± 40,000</td>
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<tr>
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<td>PSC 833</td>
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<td>246,686 ± 45,092</td>
<td>1.1</td>
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CEM III cells were incubated with different concentrations of RTA-IT and drugs for 18 hours. Then protein synthesis was measured and ICso values were determined. Values are mean of three independent experiments ± SD.
evaluated by measuring the 10% TCA-precipitable and solu-
ble radioactivities, respectively. Figure 5 shows that the
degradation of internalized T101 RTA-IT increased steadily
until it reached approximately 30%. This implies that
greater than 70% of the internalized T101 RTA-IT re-
ained intact. Treatment with 4 μmol/L CsA had no sig-
nificant impact on internalized ligand. In a control experi-
ment, preincubation of CEM III cells for 1 hour at 37°C with
4 μmol/L CsA before 125I-T101 RTA-IT treatment did not
modify the plasma-membrane binding characteristics of the
ligand (data not shown).

Subcellular distribution of T101 RTA-IT. The intracel-
ular trafficking of 125I-T101 RTA-IT in CEM III cells was
evaluated by density-gradient centrifugation of lysed cells
treated with 1 × 10−8 mol/L T101 RTA-IT for 4 hours at
37°C with or without 4 μmol/L CsA. The distribution of
cell organelles within the gradient reflects the distribution of
T101 RTA-IT within the cell, ie, Golgi and plasma mem-

![Fig 2. Effect of CsA and PSC 833 on the kinetics of protein-synthesis inhibition by anti-CD5 RTA-ITs.](image)

![Fig 3. Cytotoxicity of T101 RTA-IT in human plasma on CEM III cells.](image)

![Fig 4. Internalization rate of 125I-T101 RTA-IT in CEM III cells.](image)
were incubated for 4 hours at 37°C with $1 \times 10^{-8} \text{mol/L}$ $^{125}\text{I}$-T101 RTA-IT with or without $4 \mu\text{mol/L}$ CsA or $4 \mu\text{mol/L}$ SDZ PSC 833. The cells were then treated with protease to remove membrane-bound labeled ligand, solubilized in detergent, and analyzed by SDS-PAGE. Figure 7A and B show that no detectable changes in toxin/MoAb ratio were observed between untreated $^{125}\text{I}$-T101 RTA-IT and $4 \mu\text{mol/L}$ CsA or $4 \mu\text{mol/L}$ SDZ PSC 833. No free ricin A-chain was detected (data not shown).

**DISCUSSION**

Recently, two studies have reported an interaction of CsA in combination with RTA-IT therapy. Przepiorka et al.\(^{11}\) noted that among allogeneic BM-transplantation patients who received an anti-CD5 RTA-IT regimen as graft-versus-host disease prophylaxis, the incidence of RTA-IT toxicity (capillary leak syndrome [CLS]) substantially increased in patients who also received conventional CsA immunophylaxis. More recently, Yefenof et al.\(^{10}\) described that the combination of CsA and an RTA-IT directed against the radiation leukemia virus envelop glycoprotein (gp70) prevented the development of malignancy in mice inoculated with radiation leukemia virus-induced preleukemic cells. This report proposed that CsA increased the effectiveness of the RTA-IT regimen by blocking interleukin-4 (IL-4) secretion, thereby limiting preleukemic cell autostimulation.

In light of these findings, we investigated the potential of CsA to directly increase the cytotoxicity of RTA-ITs. The ability of CsA and its nonimmunosuppressive analog, SDZ PSC 833, to enhance the in vitro activity of anti-CD5 RTA-ITs on the human lymphoblastic T-cell line, CEM III, was evaluated. The cytotoxicity of anti-CD5 T101 RTA-IT was evaluated by over 100-fold by $4 \mu\text{mol/L}$ CsA and $4 \mu\text{mol/L}$ SDZ PSC 833 in vitro. The F(ab')\(_2\) fragment counterpart of the whole T101 Ig RTA-IT, although more potent\(^{4\text{a}}\) was less
ENHANCEMENT OF ANTI-CD5 IMMUNOTOXINS

Fig 7. Analysis of internalized 125I-T101 RTA-IT by SDS-PAGE.

CEM III cells were incubated with 1 $\times$ 10^{-8} mol/L 125I-T101 RTA-IT for 4 hours at 37°C, and then treated with protease to remove cell-surface radioactivity. The cells were then lysed and analyzed by SDS-PAGE on a 5% to 16% gradient gel. Dried gels were autoradiographed at -70°C on Kodak X-OMAT AR film with intensifying screens. Densitometry-analysis plots were superimposed after normalizing backgrounds. (A) The two bands that migrated at 180 kD were control 125I-T101 RTA-IT; lane 2, internalized 125I-T101 RTA-IT in CEM III cells; lane 3, internalized 125I-T101 RTA-IT in 4 pmol/L CsA-treated CEM III cells; lane 4, internalized 125I-T101 RTA-IT in 4 pmol/L PSC 833-treated CEM III cells. (B) (-----), control 125I-T101 RTA-IT; (-----), internalized 125I-T101 RTA-IT in CEM III cells; (-----), internalized 125I-T101 RTA-IT in 4 pmol/L CsA-treated CEM III cells; (-----), internalized 125I-T101 RTA-IT in 4 pmol/L PSC 833-treated CEM III cells.

The RTA-IT enhancement effect of CsA was specific because the toxicities of neither ricin A-chain alone nor the irrelevant anti-CD19 RTA-IT (and an anti-HLA-DR RTA-IT, data not shown) were affected in CEM III cells. Furthermore, the nonspecific toxicity of both anti-CD5 RTA-ITs was also unaffected in the irrelevant human Burkitt's lymphoma cell line RAJI.

We extended our investigation by evaluating the cytotoxicity of T101 RTA-IT in CEM III cells incubated in human plasma. We have previously shown that the activity of potent RTA-IT enhancers, such as monensin and perhexiline, is inhibited by serum components. At the membrane saturating concentration of 1 $\times$ 10^{-8} mol/L, mean T101 RTA-IT protein-synthesis inhibition in four plasma samples was about 50%, which was not significantly different from that observed in regular culture media. However, the cytotoxicity of the RTA-IT was increased to an average of 75% when CEM III were incubated in patient plasma after high-dose CsA infusion. The CsA levels in the plasma were between 3,090 and 4,860 ng/mL (2.5 to 4 µmol/L) as assessed by the immunoassay, TDX.

The fact that effective CsA levels can be achieved in vivo is a crucial observation. We have recently shown that high-dose CsA can be administered with acceptable toxicity. Indeed, short-term infusion led to steady-state CsA levels up to 4 µmol/L, without long-term immunosuppressive consequences. Even though our in vitro experiments using patient's plasma were performed for 18 hours, the kinetics of cytotoxicity experiments, using the CEM III cell line, strongly indicated that shorter incubation times would also lead to significant enhancement of RTA-IT cytotoxicity. Moreover, prolonged exposure to both RTA-IT and CsA would be unnecessary because about 50% of the bound RTA-IT are already internalized in CEM III cells in little over 2 hours.

The mechanism by which drugs enhance RTA-IT cytotoxicity remains unclear. Certain agents such as the calcium antagonists perhexiline and SR33557 appear to act by blocking the intracellular degradation of RTA-ITs and profoundly modifying its routing. However, lysosomotropic amines, such as NH4Cl, and carboxylic ionophores, such as monensin, do not alter either degradation or intracellular trafficking of internalized RTA-ITs. These agents do increase the kinetics of cytotoxicity. T101 RTA-IT has a distinctive slow rate of cell intoxication, and enhancers such as monensin and NH4Cl are able to increase the rate of cell kill dramatically. In our investigation, we found that both 4 µmol/L CsA and 4 µmol/L SDZ PSC 833 similarly increased the kinetics of protein synthesis inhibition of T101 and T101/ F(ab')2 RTA-ITs. Unlike the calcium antagonist, but much like monensin and NH4Cl, CsA had no effect on membrane binding, internalization, intracellular degradation, or intracellular trafficking of T101 RTA-IT. Hence, the mechanism of CsA on CEM III cells leading to RTA-IT enhancement remains unclear. Unlike the calcium antagonists, CsA did not have any inhibitory effect on lysosomal acid phospho-myleasine and electron-microscopy studies did not show the presence of osmiophilic-laminated structures (data not shown) that are indicative of the perturbation of the lipid metabolism observed in perhexiline and SR33557-treated cells. However, CsA has been reported to induce lipid partitioning as well as membrane polarity and lipid order.

Among the limitations of RTA-IT therapy, CLS appears to be the most important side effect. The recent report
that CLS substantially increased in patients who also received conventional CsA immunoprophylaxis may limit their combined use.\textsuperscript{21} However, CLS can be treated by glucocorticoids,\textsuperscript{24} and because CsA significantly enhances the rate of RTA-IT cytotoxicity, shorter RTA-IT treatment times may prove effective. CLS is a major side effect of IL-2 therapy,\textsuperscript{24} and may possibly be linked to the immunosuppressive activity of CsA. This would make the nonimmunosuppressive CsA analog, SDZ PSC 833, a more appropriate RTA-IT enhancer.

In conclusion, agents such as Cs could significantly improve the clinical potential of RTA-ITs by enhancing their cytotoxicity. Although animal studies are required to confirm our observations, CsA and its nonimmunosuppressive analog, SDZ PSC 833, appear to be promising agents for in vivo RTA-IT enhancement.

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Cyclosporin A and cyclosporin SDZ PSC 833 enhance anti-CD5 ricin A-chain immunotoxins in human leukemic T cells

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