Molecular Characterization of Glucose-6-Phosphate Dehydrogenase (G6PD) Deficiency in Patients of Chinese Descent and Identification of New Base Substitutions in the Human G6PD Gene


The underlying DNA changes associated with glucose-6-phosphate dehydrogenase (G6PD)–deficient Asians have not been extensively investigated. To fill this gap, we sequenced the G6PD gene of 43 G6PD-deficient Chinese whose G6PD was well characterized biochemically. DNA samples were obtained from peripheral blood of these individuals for sequencing using a direct polymerase chain reaction (PCR) sequencing procedure. From these 43 samples, we have identified five different types of nucleotide substitutions in the G6PD gene: at cDNA 1388 from G to A (Arg to His); at cDNA 1376 from G to T (Arg to Leu); at cDNA 1024 from C to T (Leu to Phe); at cDNA 392 from G to T (Gly to Val); at cDNA 95 from A to G (His to Arg). These five nucleotide substitutions account for over 83% of our 43 G6PD-deficient samples and these substitutions have not been reported in non-Asians. The substitutions found at cDNA 392 and cDNA 1024 are new findings. The substitutions at cDNA 1376 and 1388 account for over 50% of the 43 samples examined indicating a high prevalence of these two alleles among G6PD-deficient Chinese. Our findings add support to the notion that diverse point mutations may account largely for much of the phenotypic heterogeneity of G6PD deficiency.

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MATERIALS AND METHODS

Samples and other materials. After obtaining informed consent, blood specimens were collected from normal and G6PD-deficient individuals of Chinese origin residing in the United States, Taiwan, or China. Most of our samples were from Guangdong province. Unless stated otherwise, all biochemicals were purchased from Sigma (St. Louis, MO). Taq polymerase and the polymerase chain reaction (PCR) buffer were obtained from Promega (Madison, WI). The DNA Thermal Cycler used was a Perkin Elmer–Cetus (Norwalk, CT) model.

G6PD activity and characterization. The G6PD activity in fresh red blood cells (RBCs) was quantitatively measured using the kit No. 34-5-B supplied by Sigma. Physicochemical properties of semipurified RBC G6PD were determined in accordance with the recommendations of the WHO Scientific Group.6 G6PD variants were defined as previously described by Du et al.4 DNA analysis. Genomic DNA was extracted from lymphocyte nuclei by the method of Kunkel et al.13 Direct PCR sequencing of the G6PD gene in the genomic DNA was carried out as previously described.4 The final volume of the PCR mixture was 50 μL containing 50 mmol/L Tris, pH 9.0, 20 mmol/L (NH4)2SO4, 1.5 mmol/L MgCl2, 0.2 mmol/L of all four dNTPs, 0.5 to 1.0 μmol/L of all amplification primers, 1 U of Taq polymerase and 1 μg of DNA template. From the genomic DNA, a DNA fragment of 4.5 kb covering exons 3 through 13 was amplified by a first-round PCR using primers, P1L(5'-GCAAGACAGACATGCTTGCGC) and P1R(5'-GCTGGGCTCGGTGAGTAGCGACAGCA). For a higher quality and better yield, a second-round PCR was performed with 2% of the first-round PCR products using internal primers P2L(5'-AGGATGTATGTTAAGTGTC) and P2R(5'-AATGTGCAGCTGAGGTCAAT) to generate a 4.2-kb fragment. After 10 minutes of predenaturation at 94°C, 35 cycles of denaturation (94°C, 1 minute), annealing (55°C, 1 minute) and extension (72°C, 5 minutes) were used for both rounds of PCR. Using unequal amounts of PCR primers (0.25 μmol/L and 25 μmol/L, respectively), asymmetric PCR (APCR) was carried out from a mixture containing 2% of the second-round PCR products to gen-
Table 1. DNA “Variants” of 43 Chinese G6PD-Deficient Samples

<table>
<thead>
<tr>
<th>DNA “Variant” (cDNA no.) (exon no.)</th>
<th>Base Change</th>
<th>Predicted Amino Acid Substitution</th>
<th>n</th>
<th>%</th>
</tr>
</thead>
<tbody>
<tr>
<td>1388 (exon 12)</td>
<td>G to A</td>
<td>Arg to His</td>
<td>12</td>
<td>25.6</td>
</tr>
<tr>
<td>1376 (exon 12)</td>
<td>G to T</td>
<td>Arg to Leu</td>
<td>12</td>
<td>27.9</td>
</tr>
<tr>
<td>1024 (exon 9)</td>
<td>C to T</td>
<td>Leu to Phe</td>
<td>2</td>
<td>4.6</td>
</tr>
<tr>
<td>392 (exon 5)</td>
<td>G to T</td>
<td>Gly to Val</td>
<td>3</td>
<td>7.0</td>
</tr>
<tr>
<td>95 (exon 2)</td>
<td>A to G</td>
<td>His to Arg</td>
<td>8</td>
<td>18.6</td>
</tr>
<tr>
<td>Undefined*</td>
<td>—</td>
<td>—</td>
<td>7</td>
<td>16.2</td>
</tr>
</tbody>
</table>

* The exons of these samples could not be completely sequenced.

RESULTS

The majority of the 43 G6PD-deficient samples examined (36 of the 43) are associated with one of the five different single base changes. The details of the nucleotide substitutions in the G6PD gene in our sample population are listed in Table 1. All these base substitutions have been found only among ethnic Chinese and have not been reported in any other ethnic group. Substitutions at cDNA 1376 and 1388 are by far the most common base changes and account for over 50% of the nucleotide substitutions associated with our G6PD-deficient samples.

Substitutions at cDNA 392 and 1024 are new findings and have not been previously reported except in an abstract form. The DNA sequence abnormality for substitutions at cDNA 392 and 1024 identified by the direct PCR sequencing technique is shown in Figs 1 and 2, respectively. The G to T base substitution at cDNA 392 creates a Dra I restriction endonuclease site, whereas the C to T base substitution at...
cDNA 1024 abolishes a Mnl I restriction endonuclease site (data not shown).

The characteristics of the Chinese G6PD variants studied are summarized in Table 2. Electrophoretic mobility of the variants examined is generally consistent with the predicted amino acid change based on the nucleotide substitution found in the G6PD gene. For instance, the G to T substitution at cDNA 1376, causing a predicted amino acid change of Arg to Leu, is reflected by a faster electrophoretic mobility as compared with normal, whereas the A to G substitution at cDNA 95, causing a predicted change in amino acid from His to Arg, is reflected by a slower electrophoretic mobility as compared with normal. Substitutions involving charged amino acids appear to affect the enzyme activity more severely than those only involving neutral or hydrophobic amino acids. The deamino NADP utilization rate exhibited by the variant with the DNA 1376 substitution is distinctly different from that exhibited by the wild type and that exhibited by the variants with other nucleotide substitutions (Table 2).

### DISCUSSION

In the present study, we have characterized the molecular defects of the G6PD gene and the biochemical abnormalities of the G6PD in a large group of G6PD-deficient patients of Chinese descent. We have identified five unique nucleotide substitutions in the G6PD gene associated with G6PD deficiency in our patient population (Table 1). These five sub-

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**Table 2. Biochemical and Genetic Identification of Chinese G6PD Variants**

<table>
<thead>
<tr>
<th>Genetic Variant (cDNA no.)</th>
<th>Activity*</th>
<th>EPM†</th>
<th>km for G6P</th>
<th>Analogue Utilization Rate</th>
<th>Deamino NADP</th>
<th>Representative Biochemical Variant</th>
</tr>
</thead>
<tbody>
<tr>
<td>1376</td>
<td>4.9</td>
<td>104</td>
<td>28.9</td>
<td>12.3 7.2 113.0</td>
<td></td>
<td>Taiwan-Hakka</td>
</tr>
<tr>
<td>1388</td>
<td>8.0</td>
<td>102</td>
<td>55.3</td>
<td>7.2 3.5 71.8</td>
<td></td>
<td>Kaiping</td>
</tr>
<tr>
<td>95</td>
<td>6.1</td>
<td>93</td>
<td>44.7</td>
<td>6.7 3.7 69.6</td>
<td></td>
<td>Gaohe</td>
</tr>
<tr>
<td>392</td>
<td>20.0</td>
<td>100</td>
<td>56.3</td>
<td>4.7 2.5 48.8</td>
<td></td>
<td>Quing Yuan</td>
</tr>
<tr>
<td>1024</td>
<td>12.0</td>
<td>91</td>
<td>65.0</td>
<td>6.5 3.5 71.5</td>
<td></td>
<td>Mahidol-like</td>
</tr>
<tr>
<td>Wild type</td>
<td>100.0</td>
<td>100</td>
<td>54.0</td>
<td>3.0 2.3 57.3</td>
<td></td>
<td>G6PD B</td>
</tr>
</tbody>
</table>

* Activity is expressed as % of normal.
† Electrophoretic mobility (EPM) is expressed as the percentage of normal.
‡ 2-Deoxy-glucose-6-phosphate (2dG6P).
Substitutions at cDNA 392 and at cDNA 1024 are new findings (Figs 1 and 2). It is interesting to note that both substitutions involve only neutral and hydrophobic amino acids (Table 1). The substitution at cDNA 392 creates a Dra III restriction endonuclease site, whereas the substitution at cDNA 1024 abolishes a Mnl I restriction endonuclease site. These two substitutions account for about 11% of the G6PD-deficient samples examined in our study.

Of the nucleotide substitutions identified in our samples (Table 1), substitutions at cDNA 1376 and 1388 are the two most common nucleotide changes associated with G6PD deficiency, and they have been previously reported by our group,7,17 and others.18 These two nucleotide substitutions account for over 50% of our G6PD-deficient individuals, suggesting that these two alleles are common among Chinese with G6PD deficiency. It is rather interesting that both substitutions are located in exon 12 of the G6PD gene and are only 12 nucleotides apart. Moreover, both substitutions result in a replacement of an arginine residue by other amino acids.

The high frequency of cDNA 1376 and cDNA 1388 substitutions among Chinese with G6PD deficiency (Table 1) may explain the apparent difference in the clinical presentations between G6PD-deficient American blacks and G6PD-deficient Chinese. Beutler4 has recently stated that the more severe form of G6PD deficiency is due to mutations clustering near the putative NADP binding site of G6PD, whereas milder forms of G6PD deficiency are mostly caused by mutations near the amino end of the enzyme. Although both cDNA 1376 and cDNA 1388 substitutions are not in the immediate vicinity of the putative NADP binding domain,8 their close proximity to this region may alter the kinetics of G6PD in such a way that individuals with either of these two variants would be more susceptible to chemical-induced hemolytic crisis. In contrast, the mutations giving rise to the A and the A' variants among African Americans are located far away from the putative NADP binding site toward the amino end. It also should be noted that although the nature of the mutations may be a major determinant of clinical severity in G6PD deficiency, other factors such as nutrition, environment, and additional genetic factors may play a role in the pathophysiology of G6PD deficiency.17

Aside from a previously reported three-base deletion in exon 2,19 the nucleotide substitution at cDNA 95 is the only base change that has been reported in this exon of the G6PD gene.7 This substitution accounts for almost one fifth of our G6PD-deficient Chinese. Nucleic Acids Res 19:6056, 1991

ACKNOWLEDGMENT

The authors are in debt to Dr T.Y. Wang of Chang Gung Medical College, Tao-Yuan, Taiwan, ROC, for his critical review of this manuscript. The authors are also grateful to Jomei Tsai and Peter B.D. Chiu for their excellent preparation of this manuscript.

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Molecular characterization of glucose-6-phosphate dehydrogenase (G6PD) deficiency in patients of Chinese descent and identification of new base substitutions in the human G6PD gene

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