Improved Isolation of Normal Human Reticulocytes Via Exploitation of Chloride-Dependent Potassium Transport

By Martin P. Sorette, Kathleen Shiffer, and Margaret R. Clark

Studies on normal human reticulocytes have been limited by a lack of methods for effective reticulocyte enrichment. This study shows a convenient new approach for selective enrichment of reticulocytes from normal blood samples. We have developed a modified arabinogalactan density gradient that contains high potassium levels, approximating the internal cation composition of red blood cells (RBC). The low-density populations from this gradient are enriched in reticulocytes, and the highly selected lowest density fraction shows a much higher reticulocyte enrichment than that obtained with high sodium chloride arabinogalactan density gradients, or other previously reported density gradient methods. We found that this improved isolation is caused by suppression of potassium loss and reticulocyte dehydration via chloride (KCl) cotransport. When the low-density fraction of RBC from a high-potassium gradient was subsequently incubated in high sodium chloride medium and reseparated on a sodium chloride density gradient, the reticulocytes dehydrated and were recovered in high-density fractions. The highest-density fractions from this secondary gradient yield 95% to 99% reticulocytes. We anticipate that this method will benefit investigators who require reticulocyte enriched populations for a wide variety of applications.

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MATERIALS AND METHODS

Chemicals and reagents. Arabinogalactan (Stractan II) was obtained from the St Regis Paper Co (Tacoma, WA). Ethidium Bromide, EGTA (Ethylene glycol-bis-(β-amino-ethyl ether) N,N'-Tetra-Acetic Acid), EDTA (ethylenediaminetetraacetic acid), microcrystalline cellulose (Sigma Chemical Co (St Louis, MO)), REPS (N-2-hydroxyethylpiperazine-N'-2-ethane sulfonic acid), HEPES Na salt, and PIPES (piperazine-N,N'-bis (2 ethanesulfonic acid) were products of Sigma Chemical Co (St Louis, MO). New Methylene Blue N (Breccher formula) was purchased from J.T. Baker Chemical Co (Phillipsburg, NJ). Reagent grade Na chloride, Na nitrate, K chloride, and magnesium chloride were purchased from Fisher Scientific Co (Fair Lawn, NJ).

Collection of blood samples. After obtaining informed consent under a protocol in accordance with the principles of the Helsinki Declaration and approval by the Human and Environmental

From the Department of Laboratory Medicine and the Cancer Research Institute, University of California, San Francisco; and Bio Rad Inc, Hercules, CA.

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Address reprint requests to Margaret R. Clark, PhD, Box 0905, University of California, San Francisco, CA 94143.

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Protection Committee of the University of California, San Francisco, venous blood was collected from healthy normal volunteers in EDTA or acid-citrate-dextrose (ACD) (Vacutainer; Becton Dickinson, Mountain View, CA). Leukocytes and platelets were removed by filtration through a column of cellulose.\(^{13,14}\)

**Arabinogalactan density gradients.** Arabinogalactan (Stractan II) was decolorized using magnesium oxide\(^5\) and activated charcoal and purified according to Corash.\(^7\) For initial experiments at pH 7.4, stock arabinogalactan solutions were prepared that, based on available water, contained HEPES (20 mmol/L), \(MgCl_2\) (1 mmol/L), \(NaH_2PO_4\) (1 mmol/L), glucose (10 mmol/L), EGTA (0.5 mmol/L), adenine (2 mmol/L), inosine, (12 mmol/L), penicillin (100 U/mL), streptomycin (100 \(\mu\)g/mL). \(NaCl\) gradients contained KCl (10 mmol/L), NaCl (14 mmol/L), and KCl gradients contained NaCl (12 mmol/L), KCl (115 mmol/L) added to the stock solution. The pH was adjusted to 7.4, and the osmolality to 290 mOsm using a vapor pressure osmometer (Model 5100 C; Wescor Inc, Logan, UT). For subsequent experiments, \(NaNO_3\) and \(NaCl\) gradients contained PIPES, pH 6.8 (20 mmol/L). glucose (10 mmol/L), adenine (2 mmol/L), inosine, (12 mmol/L), ouabain (0.2 mmol/L), piretanide (0.1 mmol/L), and either \(NaNO_3\) (165 mmol/L) or \(NaCl\) (150 mmol/L). The pH was adjusted to 6.8 and the osmolality to 290 mOsm. Discontinuous gradients were prepared by dilution of stock Stractan with isosmolar buffers of the same composition, using nine layers ranging in equal steps from 1.083 to 1.124 g/mL density. These were layered in 17 mL tubes onto a cushion of at least 1.133 g/mL density. The gradients were centrifuged in a swinging bucket rotor in an ultracentrifuge (Beckman L3-50) for 45 minutes at 74,000g at 20°C. The separated cells were collected from the gradient interfaces using a Pasteur pipet, during which the walls of the tube were washed with buffer to avoid contamination of subsequent layers. The separated cells were washed three times to remove residual Stractan and were resuspended to a defined volume.

**RBCs and reticulocyte density distribution.** RBC distribution in the density gradient was determined by counting the cells in each gradient population with an electronic cell counter (Model Z40; Coulter Electronics, Hialeah, FL). Mean cell hemoglobin concentration (MCHC) was determined from a spectrophotometric measurement of hemoglobin as the cyanomet-hemoglobin complex. Manual reticulocyte counts were made on 1,000 cells/sample stained with New Methylene Blue N.\(^{16}\) Reticulocyte counts by flow cytometry were performed on ethidium bromide stained samples\(^{17}\) using the Becton Dickinson Facscan flow cytometer (Beckton Dickinson, Mountain View, CA). Gating on the light-scattering signal was used to discriminate RBC from particulate debris. Fluorescence was measured with excitation at 488 nm provided by an argon ion laser and detected through narrow-band emission filters at 530 nm (FL1, Green) or 585 nm (FL2, Red). The fluorescence measurements were standardized using fluorescent microspheres (Calibrite; Becton Dickinson).

**Staining for reticulocytes bearing the transferrin receptor.** A mouse monoclonal antihuman transferrin receptor IgG1 raised against MLA 144 Gibbon leukemic cells (Chemicon International Inc, Temecula, CA) was diluted to 10 \(\mu\)g/mL in phosphate-buffered saline (PBS)/1.0% bovine serum albumin. Fifty microliters of washed cells from density gradient fractions were suspended in 200 \(\mu\)L of antibody and incubated for 1 hour at 37°C. Cells were washed three times in PBS and then incubated for 1 hour at 37°C in 200 \(\mu\)L of a goat antimouse IgG-FITC conjugate diluted to 10 \(\mu\)g/mL in PBS. After washing three times, cells were stained in an equal volume of .002% ethidium bromide for 1 hour at room temperature, placed on ice, and immediately analyzed by flow cytometry.

**Quantification of protein 4.1 a and b components.** RBC ghosts were prepared from separated samples by hypotonic lysis in 40 volumes of ice-cold 5 mmol/L Tris-HCl, 1 mmol/L EDTA, 1 mmol/L diisopropylfluoro-phosphate, pH 8.0. Membranes were solubilized and proteins were separated by sodium dodecyl sulfate-polyacrylamide gel electrophoresis (SDS-PAGE) (7% acrylamide) according to the method of Laemmli.\(^{18}\) Proteins were stained with Coomassie Brilliant Blue R-250, and the bands corresponding to 4.1a and 4.1b were cut out and the dye eluted in 25% Pyridine and quantified spectrophotometrically.\(^{19}\)

![Density distribution of RBC from one individual in high NaCl and KCl gradients, pH 7.4. The gradients are identical with respect to density. Note that there is a shift to lower densities in the K gradient compared with the Na gradient. A distinct population of low-density, reticulocyte-enriched RBC (indicated by arrow) is clearly isolated in the high-K gradient. This population is markedly reduced in the high-Na gradient.](image-url)
significantly improved isolation of reticulocytes in the low-density fractions of the KCI gradient when compared with the NaCl gradient. Data indicated. Reticulocytes in each fraction were quantitated by flow cytometric analysis of ethidium bromide stained cells. Note the improved isolation of reticulocytes in the low-density fractions of the KCI gradient when compared with the NaCl gradient. Data presented as percent reticulocytes (SEM).

Hemoglobin A\textsubscript{c} measurements. The relative amount of hemoglobin A\textsubscript{c} in each gradient fraction was measured by ion exchange high-performance liquid chromatography (DIAMAT Fully Automated Hemoglobin Analyzer System; Bio Rad, Hercules, CA).

RESULTS

We hypothesized that the CI-dependent K loss from reticulocytes might alter their density during density gradient centrifugation, resulting in less than optimal reticulocyte separation under conditions most commonly used. In an effort to enhance reticulocyte separation, we tested the effect of various factors that influence the activity of CI-dependent transport. Activation of the transport by reduced pH\textsubscript{c} and suppression of the transport by reducing the transmembrane K\textsubscript{c} gradient and substituting NO\textsubscript{3} for CI were investigated.

Effect of gradient composition on RBC and reticulocyte distribution. Comparison of the density distribution of total RBCs on high-K and high-Na gradients showed that the major difference was an increase in the proportion of cells in the two lowest density layers of the high-K gradient (Fig 1). When the reticulocyte distributions in gradient populations were determined, it was found that these two least dense populations from the high-K gradient were markedly enriched in reticulocytes (Table 1). In contrast, reticulocytes had a much broader distribution on high-Na gradients and did not show such remarkable enrichment in the lowest-density populations.

If the presence of higher-density reticulocytes on high-Na gradients were caused by CI-dependent K loss via the KCI cotransport pathway, it should be accentuated by reducing the pH of the NaCl medium to 6.8, the pH optimum for activation of this pathway in cells containing HbC.\textsuperscript{21} As expected, incubation and separation of the cells in high-Na medium at pH 6.8 did result in even greater increase and broadening of reticulocyte density (data not shown).

Effect of transient exposure to low pH in ACD anticoagulant. Having found that separation in high-Na medium at low pH resulted in reticulocyte dehydration, we asked whether transient exposure to low pH of blood samples drawn into ACD anticoagulant would affect reticulocyte distribution during subsequent density separation on high-K gradients at pH 7.4. Parallel samples were drawn into ACD (final pH 6.4 to 6.7) and EDTA and were then immediately separated on KCI gradients at pH 7.4. A perceptible shift to higher density was observed in the ACD sample, and the percentages of reticulocytes in the lowest-density layers were reduced (data not shown). When the ACD was adjusted to pH 7.4 before adding the blood, the subsequent density and reticulocyte distribution was identical to that of the EDTA sample. This indicated that it was exposure to low pH, not the effect of an ACD component, that influenced the density distribution. All of the data presented in this report were obtained using samples drawn into EDTA.

<table>
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<th>Fraction</th>
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<td>Density (g/mL)</td>
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<td>1.0829</td>
<td>1.0880</td>
<td>1.0919</td>
<td>1.0964</td>
<td>1.1009</td>
<td>1.1055</td>
<td>1.1102</td>
<td>1.1148</td>
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<td>KCI</td>
<td>67.6</td>
<td>25.3</td>
<td>5.2</td>
<td>0.3</td>
<td>0.17</td>
<td>0.23</td>
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<td>4.9</td>
<td>(5.2)</td>
<td>(1.6)</td>
<td>(0.04)</td>
<td>(0.03)</td>
<td>(0.03)</td>
<td>(0.03)</td>
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<tr>
<td>NaCl</td>
<td>31.8</td>
<td>6.2</td>
<td>1.9</td>
<td>1.3</td>
<td>0.56</td>
<td>0.27</td>
<td>0.37</td>
<td>0.2</td>
<td>0.17</td>
<td>0.23</td>
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<tr>
<td>(6.7)</td>
<td>(1.2)</td>
<td>(0.23)</td>
<td>(0.2)</td>
<td>(0.08)</td>
<td>(0.07)</td>
<td>(0.09)</td>
<td>(0.0)</td>
<td>(0.03)</td>
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Cellulose filtered whole blood from three subjects was separated on NaCl and KCI gradients as shown in Fig 1. Fractions correspond to the densities indicated. Reticulocytes in each fraction were quantitated by flow cytometric analysis of ethidium bromide stained cells. Note the significantly improved isolation of reticulocytes in the low-density fractions of the KCI gradient when compared with the NaCl gradient. Data presented as percent reticulocytes (SEM).
Table 2. Reticulocyte Distribution

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<tr>
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<td>A</td>
<td>13.0</td>
<td>4.4</td>
<td>3.7</td>
<td>6.5</td>
<td>28.4</td>
<td>64.8</td>
<td>89.9</td>
<td>88.4</td>
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<tr>
<td>B</td>
<td>7.1</td>
<td>1.05</td>
<td>2.0</td>
<td>4.1</td>
<td>10.4</td>
<td>33.1</td>
<td>71.3</td>
<td>87.7</td>
<td>97.0</td>
</tr>
</tbody>
</table>

Reticulocyte distribution in cells from one subject that were isolated from low-density fractions of a KCl gradient, incubated in high-NaCl medium, pH 6.8, and reseparated on NaCl gradients. Layers A and B denote the lowest and next lowest density fractions from the original KCl gradient separation. Layers 1 to 9 represent populations recovered from the secondary NaCl gradients. Data presented are percent reticulocytes quantitated by manual counts of 1,000 New Methylene Blue stained cells from each fraction.

Chloride dependence of reticulocyte dehydration in high-Na medium. To further verify that the improved separation of reticulocytes on high-K gradients was caused by suppression of Cl-dependent K loss, the effect of Cl substitution by NO3 was tested (Fig 2). When low-density cells obtained from a high-K gradient were incubated and recentrifuged on a NaN03 gradient at pH 6.8, they returned to the same density as those reseparated on a high-K gradient at pH 7.4. In contrast, a parallel sample reseparated on a NaCl gradient, pH 6.8, yielded a markedly broadened density distribution attributable to cellular dehydration. Quantitation of reticulocytes in each of the density fractions showed that it was the reticulocytes that became dehydrated, thus concentrating in the most dense fractions of the NaCl gradient (Table 2).

Reticulocyte maturation and KCl cotransport activity. Next, we asked whether the level of KCl cotransport activity was correlated with two indices of reticulocyte maturation: RNA content and expression of the transferrin receptor. To address this question, a reticulocyte enriched fraction from the high-K gradient was reseparated on a NaCl gradient, pH 6.8. The lowest, highest and an intermediate density fraction were stained with ethidium bromide and monoclonal antitransferrin receptor IgG, and two-color–fluorescence analysis was performed by flow cytometry (Fig 3). The lowest-density fraction was enriched in mature RBCs (22% reticulocytes) with a small percentage (7.2%) of cells staining positive for the transferrin receptor (Fig 3B). The intermediate fraction contained a higher percentage of reticulocytes, and a greater proportion of reticulocytes bearing the transferrin receptor (Fig 3C). The cells having the most active transport activity, isolated in the highest-density fraction of the chloride gradient (Fig 3D) are a nearly pure fraction of reticulocytes (96.0%). A significant percentage (39%) stained positively for transferrin receptor. Moreover, these reticulocytes stained more intensely for RNA and the transferrin receptor, suggesting that this fraction is enriched in the youngest circulating reticulocytes. These results indicate that the least mature reticulocytes are those that become most dehydrated via Cl-dependent K loss.

Age-related markers in fractions from high K+ gradients. In an effort to assess the relative age of the mature cell...
IMPROVED ISOLATION OF RETICULOCYTES

DISCUSSION

Several density-based or rate-sedimentation techniques for separating reticulocytes from erythrocytes, and for separating erythrocytes into populations of differing mean cell age, have been developed.25-29 Our study differs from these in showing that highly enriched populations of reticulocytes can be obtained from normal blood in a one-step density separation on gradients containing a high-K concentration to approximate the internal cation composition of RBC. This method results in isolated populations of up to 80% reticulocytes, a much greater enrichment of normal reticulocytes than previously achieved using high-Na medium or other density-based methods.25 For even greater reticulocyte enrichment, low-density populations isolated on KCl gradients are incubated and recentrifuged in NaCl medium, which promotes selective increase in density of the youngest reticulocytes. The most dense fractions from the secondary separation are almost all reticulocytes. Although affinity based techniques also yield highly enriched reticulocyte populations, they only select fibronectin or transferrin receptor-bearing reticulocytes, and recovery of the cells after isolation in an undamaged state is problematic.

We have shown that the improved isolation of reticulocytes in high-K medium is attributable to the suppression of KCl loss and cell dehydration via the K-Cl cotransporter, which is active in reticulocytes but is suppressed upon erythrocyte maturation.26 It should be noted that even if high-K gradients are used, reticulocyte separation is less effective if the blood is drawn into ACD anticoagulant. This is caused by transient activation of KCl cotransport resulting in irreversible dehydration of some reticulocytes. This mechanism of pH effects on RBC-density-distribution differs from that discussed by Walter et al.27 They found that separation at low pH resulted in an increase in volume and a decrease in the density of mature rat RBC. Although this would cause increased overlap in density between reticulocytes and erythrocytes, the swelling of mature cells would be largely reversible with return to pH 7.4, whereas the KCl cotransport mediated dehydration of reticulocytes would not.

The distribution of putative markers for aging of mature RBCs HbA1c and protein 4.1a was consistent with the reticulocyte distribution, showing low levels of both markers in reticulocyte-enriched cell populations. There was a progressive increase in both markers with increasing cell density, but this tended to plateau in the last few most dense populations. This suggests that although there may be a general increase in average cell age with increasing density of layers from the KCl gradients, the very highest density cells may not represent a highly enriched population of the oldest cells. These studies show a convenient, highly effective approach for selective separation of reticulocytes from mature erythrocytes in normal blood samples via exploitation of their C1-dependent K transport function. We anticipate that its capacity to provide improved recovery of reticulocytes in free suspension will facilitate investigation of a wide variety of fundamental biological questions.

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REFERENCES

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MP Sorette, K Shiffer and MR Clark