Presence of Cross-Reactive Antibody Between Human Immunodeficiency Virus (HIV) and Platelet Glycoproteins in HIV-Related Immune Thrombocytopenic Purpura

By Ali Bettaieb, Patricia Fromont, Fawzia Louache, Eric Oksenhendler, William Vainchenker, Najib Duédari, and Philippe Bierling

We previously reported the presence in platelet eluates of autoantibodies directed against epitopes of the platelet glycoprotein (GP)IIb/IIIa complex in acquired immunodeficiency syndrome (AIDS)-free human immunodeficiency virus (HIV)-infected patients with immunologic thrombocytopenic purpura (ITP). We investigated whether HIV antibodies recognized platelet membrane antigens to determine whether the virus might be directly or indirectly responsible for the thrombocytopenia in this context. Direct eluates of platelets from 25 patients with HIV-related ITP contained IgG reacting to epitopes localized on the glycoprotein (GP)IIb/IIIa complex. A syndrome similar to ITP frequently occurs in patients with or without thrombocytopenia and was drug addicts, and hemophiliacs. Immunologically mediated platelet destruction has not yet been established in such patients, but the precise mechanism of the thrombocytopenia has not yet been established. Two main hypotheses have been advanced. The first hypothesis implicates nonspecific deposition of circulating immune complexes on the platelet surface with subsequent clearance of the opsonized platelets. This hypothesis is supported by the presence of anti-HIV antibodies and the absence of detectable HIV antigens on the platelet surface. Certain evidence indicates that anti-F(ab')2 antibodies directed against HIV antibodies (antiidiotypic antibodies) might play a role in the formation of such immune complexes. The second hypothesis, supported by the findings of two independent teams of investigators, is based on the presence of specific platelet antibodies. Thus, von dem Borne et al reported the presence of platelet autoantibodies in most HIV-infected patients, who observed that the cross-reactivity is dependent on the glycosylation or conformational structure of the GP. We conclude that molecular mimicry between HIV-GP160 and GPIIb/IIIa may explain at least some cases of ITP in AIDS-free HIV-infected patients.

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CROSS-REACTIVE Ab BETWEEN HIV-GP AND PLATELET GP

Patient Bou, born in 1960, was a drug addict from 1978 to 1983. HIV seropositivity (CDC stage IVC2) and ITP were diagnosed in 1988. Treatment with zidovudine was started in August 1988 and the platelet count increased to near-normal levels (142 × 10^9/L). Pulmonary tuberculosis, diagnosed in January 1990, resolved with specific treatment.

Platelet autoantibody determination. Platelet-associated IgG (PAIgG) was detected according to the method described by von dem Borne et al. Samples (sera, eluates or F(ab')2 fragments) were tested for antiplatelet membrane activity by means of an indirect immunofluorescence test (PIFT) on paraformaldehyde-treated platelets with fluorescein-conjugated F(ab')2 antihuman IgG or antihuman light chains ( Dakopatts, Copenhagen, Denmark).

In some cases, sera and eluates were also studied in immunoprecipitation experiments. Platelets 1 × 10^10 were surface-labeled with 125Iodine using the lactoperoxidase catalyzed method. The platelets were subsequently solubilized in 1 mL buffer A [containing Tris-HCl 20 mmol/L NaCl 150 mmol/L (Tris-buffered saline-TBS), Triton X-100 1%, EDTA 5 mmol/L, phenylmethylsulfonyl fluoride (PMSF) 2 mmol/L, and 0.5 μg/mL leupeptin] for 30 minutes at 4°C, and the lysate was centrifuged for 30 minutes at 17,600g at 4°C. Fifty microliters of lysate were then incubated overnight at 4°C with 50 μL serum, 100 μL eluate, or 10 μL purified monoclonal antibody (MoAb). Two hundred microliters of a 10% suspension of protein A-Sepharose (Pharmacia, Paris, France) were added to the lysate and incubated for 1 hour at 4°C. The beads were then washed five times with buffer A, two times with buffer B supplemented with 0.2 mol/L NaCl, and two times with buffer C supplemented with 0.35 mol/L NaCl. Fifty microliters of 2% sodium dodecyl sulfate (SDS) and 5% β-mercaptoethanol were added to each pellet and, after boiling for 5 minutes, the solutions were analyzed in a 7.5% sodium dodecyl sulfate (SDS)-polyacrylamide gel according to the method of Laemmli. Gels were then fixed, dried, and autoradiographed.

Platelet eluates. Either elution of PAIgG was performed according to the method of Helmerhorst et al. The direct eluates were then tested for antiplatelet and anti-HIV activity in different techniques in comparison with the last platelet washing sample to detect possible contamination of the eluates by serum. Results were interpreted only if the last platelet washing sample was negative. Negative controls included direct eluates of platelets from normal individuals and HIV-negative patients with ITP.

HIV antibody determination. HIV antibodies were detected using a commercially available enzyme-linked immunosorbent assay (ELISA) kit (Abbott Laboratories, Paris, France) and by Western blot inmaunoassay (Dupont De Nemours, Les Ulis, or Diagnostics Pasteur, Marnes-La-Coquette, France). The ELISA kit contained mainly GP41 and GP24 recombinant HIV-1 proteins, with small amounts of GP160 and GP55 of HIV-1 and GP41 of HIV-2. The ELISA was considered positive if the ratio of sample absorbance to positive control absorbance was greater than or equal to the calculated optical density (OD) cutoff value (the cutoff value was calculated according to the manufacturer’s recommendations using the positive and positive controls included in the commercial kit). In the HIV1 Western blot assay, eluates (diluted 1:40) were incubated with the test strips for 2 hours at room temperature and the bands were disclosed by an alkaline phosphatase-conjugated goat anti-human serum (Diagnostics Pasteur) or by a series of reactions with goat antihuman IgG conjugated with biotin, avidin conjugated with horseradish peroxidase (HRP), and the HRP substrate 4-chloro-1-naphthol (Dupont). The result was considered positive in these experiments when at least one band was visualized.

HIV-infected cell lines. HIV antibodies from patient Bou platelet direct eluates were detected by indirect immunofluorescence on methanol-permeabiled (1 minute at 22°C) cyto centrifuged U937 and CEM cell lines infected by the HTLV-IIIB and Bru-HIV isolates, respectively.

Detection of HIV antigens on patient platelets. HIV antigens were sought on the patient’s platelets by immunoblotting, immuno precipitation, and the indirect immunofluorescence procedure, using the following HIV MoAbs: anti-p24, anti-p17, and anti-Gpl20 (provided by Dr R.M. Vazeux, Institut Pasteur, Paris, France). Two additional antibodies were used in the immunofluorescence technique, one directed against p25, p34, p55, and p65, (Genzyme, Boston, MA) and the second recognizing GP160/120 (Biosoft, Paris, France).

Immunoglobulin and F(ab')2 preparations. IgG was purified from patient Bou serum by means of the staphylococcal protein A- Sepharose CL4B technique (Pharmacia, Paris, France). Two additional antibodies were used in the immunofluorescence technique, one directed against p25, p34, p55, and p65, (Genzyme, Boston, MA) and the second recognizing GP160/120 (Biosoft, Paris, France).

Binding and elution experiments performed with patient sera. Washed platelets (10^9) harvested from healthy blood group O donors or GT patients were suspended in 500 μL PBS containing 3 mmol/L EDTA and 0.25% bovine serum albumin (BSA) and then incubated with 500 μL patient serum or plasma for 2 hours at room temperature. The mixture was subsequently washed in 20 mL of the same buffer four times with repeated centrifugation. The platelets were resuspended in 500 μL buffer, and 1 mL diethyl ether was added. The samples were then shaken for 2 minutes, incubated for 30 minutes at 37°C, and spun for 10 minutes at 3,000 g. The indirect eluates (aerial solution) were tested in parallel with the last washing fluid for antiplatelet and anti-HIV activity. Results were interpreted only if the last platelet washing sample was negative.

Binding and elution experiments performed with eluates, sera, and F(ab')2 fragments on recombinant GP160 and purified platelet GP120/120. Two recombinant GP160s (rGP160) of the HIV1 envelope (rGP160-1, rGP160-2), supplied by Drs M. Girard and M. Kaczorek (Pasteur Merieux, Val de Reuil, France) and by Repligen (Cambridge, MA), respectively, were studied. rGP160-1 is secreted into the culture medium of baby hamster kidney (BHK 21) cells infected by the recombinant vaccinia virus VV 1163.38 Amino acid sequencing has shown that rGP160-1 lacks 22 amino acids at position 696 to 717. 38 rGP160-2 (derived from the IIIB isolate of HIV-1) is expressed as a full-length protein in insect cells by using a baculovirus expression vector and is neither processed nor secreted.29,30 Four hundred micrograms of the rGP160 was coupled to 0.5 mL Affi-gel 10 according to the manufacturer’s directives (BioRad, Paris, France).

Platelet GPIb/IIIa was purified from human platelets as described by Fitzgerald et al.11 with modifications. The platelet lysate was loaded onto a Concanavalin A-ultrogel column. Fixed materials were eluted by 0.2 mol/L methyl-o-glucopyranoside (Serva, St
Germain en Laye, France), and then dialyzed against TBS containing 0.1% Triton-X 100. Further purification was performed with insoluble anti-GPIb/IIa MoAb (SZ21) (Immunotech, Marseille, France) coupled to a column of Affi-gel 10 (1 mg antibody/1 mL Affi-gel 10). GPIb/IIa complex was eluted from the column with 0.1 mol/L glycine, 0.1 mol/L NaCl, and 0.1% Triton-X 100, pH 2.8, and the eluted fraction was immediately neutralized with 1 mol/L Tris. The purified GPIb/IIa was greater than 90% homogeneous as analyzed by SDS-PAGE. The only slight contaminant observed was the fibrinogen that was probably copurified with the GPIb/IIa. In addition, in an ELISA, the GPIb/IIa preparation failed to react with HLA class I, CD42b, CDw49b, and CD62 MoAbs coupled to 0.5 mL Affi-gel 10 according to the manufacturer’s directives (BioRad, Paris, France).

Absorption experiments with eluates, sera or F(ab')2 fragments were performed using Affi-gel 10 coupled to rGP160 or to purified GPIb/IIa. After several washes, absorbed materials were eluted by addition of an aqueous buffer containing 150 mmol/L H3PO4/100 mmol/L NaCl/1.5% BSA, pH 2.8, for 10 minutes at 4°C. The acidified suspension was centrifuged and then neutralized with 1 mol/L Tris. Eluted materials were tested for antiplatelet and anti-HIV activity in comparison with the last GP160 or GPIb/IIa Affi-gel column washing samples. Results were interpreted only if the last platelet washing sample was negative.

RESULTS

Platelet autoantibody determination. In accordance with a previous report,18 PAIgG levels were increased in 20 (80%) of the 25 patients with HIV-related ITP studied. For 9 of these 20 patients (45%), the direct eluate contained platelet antibodies that recognized in the PIIFT all the platelet panel except platelets from ABO compatible type I GT patients. Five of these positive eluates, including the three reported in our previous work,18 precipitated platelet GPIb/IIa (partially illustrated in Fig 3). In contrast, a weak increase in PAIgG was observed in only 2 of the 11 nonthrombocytopenic patients (18%) and was never elutable. Likewise, sera of 25 of the 28 thrombocytopenic but only 3 of the 11 nonthrombocytopenic patients were positive in the PIIFT.

Presence of HIV antibodies on platelets from HIV-infected thrombocytopenic patients. Direct platelet eluates from HIV-infected thrombocytopenic (n = 25) and nonthrombocytopenic patients (n = 11) were tested using ELISA and Western blot procedures for the presence of HIV antibodies (Fig 1). In the ELISA, 21 of the 25 eluates (84%) from thrombocytopenic patients were positive, whereas only 3 of the 11 eluates (27%) from nonthrombocytopenic patients were weakly positive. Furthermore, all 25 direct eluates from HIV-infected ITP patients were positive in the HIV Western blot immunoassay, whereas only four of the nonthrombocytopenic patients were positive (including two who were very weakly positive), and seven were negative. As shown in Fig 1, all 29 eluates positive in HIV-Western blot recognized HIV GP160/120, and 12 also recognized other proteins encoded by the Gag or Pol genes (p17, p24, p31, p53, p68).

Viral antigens on platelets of HIV-infected ITP patients. Immunoblotting and immunoprecipitation procedures with anti-HIV MoAbs were used to detect the presence of HIV antigens on HIV-infected ITP patient platelet extracts. Negative results were obtained for all four patients studied. Similar negative results were obtained with three other platelet samples in the indirect immunofluorescence test. These results do not strictly eliminate the presence of HIV antigens on the platelet membrane in these patients.

Fig 1. Analysis of direct and indirect platelet eluates by means of HIV-Western blot immunoassay. Direct and indirect platelet eluates were obtained by ether elution from HIV-infected patient platelets and from normal platelets first incubated with patient sera, respectively, and were analyzed by commercially available HIV1-Western blot kits (Dupont or Diagnostics-Pasteur). According to the manufacturer’s instructions, the markers (left) correspond to the major HIV proteins. Positive control (1), and representative aspects of immunoblots obtained with direct eluates from HIV-infected ITP patients (2-5). Only anti-GP160/120 was observed in 14 of 25 eluates (6,7). Positive immunoblots obtained with direct eluates from HIV-infected nonthrombocytopenic patients. In these cases, negative results were obtained in 7 of 11 eluates (8-10). Pattern of immunoblots obtained with indirect eluates from HIV-infected ITP patients. Only anti-GP160/120 was observed in 12 of 14 indirect eluates (11). The weakly positive immunoblots were obtained with indirect eluates from HIV-infected nonthrombocytopenic patients. Negative results were obtained in six of the eight nonthrombocytopenic patients studied. Direct and indirect eluates of patient Bou (2 and 8, respectively). All direct and indirect eluates were used at a dilution of 1:40 in these experiments.
however, owing to the limited sensitivity of the methods used.

**Binding of serum HIV antibodies to normal platelets.** Because HIV antibodies were present but HIV antigens were not detectable on the membrane of platelets from the HIV-infected ITP patients, we determined whether HIV antibodies could bind to platelets from uninfected healthy blood donors. Indirect platelet eluates were thus obtained by ether elution after incubation of patient sera with normal platelets. As shown in Fig 1, all 14 indirect platelet eluates obtained with serum from HIV-infected ITP patients were positive in the HIV-Western blot immunobassay. All of these eluates recognized GP160/120, and two also recognized other viral proteins, ie, p17, p24, p31, p53, p68. In the HIV-ELISA, 13 of these eluates were negative, whereas one (the sample showing antiplatelet activity in the PIIFT, ie, patient Bou [described below]) was strongly positive (four times the cutoff value). In contrast, only two of eight eluates obtained with serum from HIV-infected non-ITP patients were weakly positive in the Western blot immunobassay (recognizing GP160/120), whereas all were negative in the HIV-ELISA and the PIIFT. The discrepancies between the results obtained in the HIV Western blot immunobassay and the HIV-ELISA with both the indirect and direct eluates may have been due to the presence of only trace amounts of GP160/120 in the ELISA kit, since GP160/120 antibodies were predominant in the eluates. To determine whether this ability of HIV antibodies to bind to platelets from HIV-negative patients was specific (ie, due to the presence of cross-reactive antigens between platelets and virus) or nonspecific (ie, Fc binding), we focused our study on patient Bou, who showed the highest serum titer of antiplatelet and anti-HIV activities able to bind to normal platelets.

**Characterization of PAIgG from patient Bou.** The PAIgG eluted from patient (auto) and normal (allo) platelets preincubated with patient serum were strongly reactive in the PIIFT with platelets from healthy blood group O donors (Table 1). In contrast, they did not recognize ABO-compatible platelets from two GT patients with no detectable GPIb/IIIa in immunochromed analysis, strongly suggesting anti-GPIb and/or anti-GPIIa specificity of the platelet antibody. Furthermore, the direct Bou platelet eluate immunoprecipitated a band of 97 Kd characterized as GPIIa (see Fig 3). As we have reported for other patients,18 the sera of the patients in this study could not precipitate platelet GPIb/IIIa. The direct and indirect eluates also contained HIV antibodies (described above) detectable in the ELISA and HIV-Western blot immunobassay (Table 1, Fig 2). In this latter assay, only GPIb/120 antibodies were detectable (Fig 1, strips 2 and 8, Table 1). To determine if two distinct antibodies were present in these eluates, we performed absorption/elution experiments with ABO-compatible platelets from two GT patients. Neither the anti-HIV nor antiplatelet activity present in these eluates, we performed absorption/elution experiments with normal healthy blood-donor platelets (described above), suggested that both patient Bou HIV and platelet antibodies bound to GPIb/IIIa (Fig 2 and Table 1).

The presence of cross-reactive antibody between platelet GPIb/IIIa and HIV GP160 in patient Bou serum and the anti-GPIb/IIIa specificity of Bou platelet antibody was confirmed by binding/elution experiments with patient Bou serum on insoluble purified GPIb/IIIa. As shown in Table 1, the antibodies eluted from purified immobilized GPIb/ IIIa recognized healthy blood-donor ABO-compatible platelets in the PIIFT and HIV-Gp160 in the HIV-Western blot immunobassay. The anti-GPIb/IIIa specificity of the platelet antibody was confirmed by an immunoprecipitation procedure, because, when incubated with 125I-labeled platelets, the eluate obtained from purified immobilized GPIb/ IIIa incubated with patient serum (as well as with direct platelet eluate) precipitated a band of 97 Kd in reducing conditions which comigrated with the GPIIa precipitated by an PLA1 antibody or a GPIIa MoAb (SZ21) (Fig 3). Controls of these absorption-elution experiments included serum of a nonthrombocytopenic HIV-infected patient (that did not recognize the immobilized GPIIIa) and polyclonal GPIb/IIIa antibodies from an HIV-negative GT patient26 (The eluate obtained from the immobilized GPIIIa column after incubation with this latter serum contained only GPIb/IIIa antibody).

To determine whether binding of the cross-reactive platelet antibody to normal platelets or to purified GPIb/ IIIa was specific, the same experiments were performed with the F(ab')2 fragment of purified serum Bou IgG. As shown in Fig 2, similar results to those observed with patient serum were obtained with this material. Thus, both the platelet and HIV GP160 antibody activities present in patient Bou F(ab')2 fragments were capable of binding to

<table>
<thead>
<tr>
<th>Characteristic</th>
<th>Patients' Sera</th>
<th>Antibodies Eluted From Normal Platelets After Incubation With Patients' Sera (direct eluate)</th>
<th>Antibodies Eluted From Normal Platelets After Incubation With Patients' Sera (indirect eluate)</th>
<th>Antibodies Eluted From Purified GPIb/IIIa After Incubation With Patients' Sera</th>
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</thead>
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<tr>
<td>IF on normal platelets</td>
<td>++ to ++++</td>
<td>+++</td>
<td>++</td>
<td>++</td>
</tr>
<tr>
<td>IF on GT platelets</td>
<td>− to +</td>
<td>−</td>
<td>−</td>
<td>ND</td>
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<td>Platelet immunoprecipitation</td>
<td>Negative</td>
<td>GPIb/IIIa</td>
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<td>HIV-ELISA (cutoff = 0.3 OD)</td>
<td>&gt;2</td>
<td>1.54 ± 0.30</td>
<td>1.29 ± 0.44</td>
<td>ND</td>
</tr>
<tr>
<td>HIV-Western blot assay</td>
<td>All HIV-proteins and glycoproteins</td>
<td>HIV-GP160/120</td>
<td>HIV-GP160/120</td>
<td>HIV-GP160/120</td>
</tr>
</tbody>
</table>

Platelet membrane fluorescence was graded as negative (−), weakly positive (+), or positive (++ to ++++) in comparison with results obtained using a negative control (serum from a non-transfused healthy male blood group AB donor), and a positive control (pool of sera from multitransfused immunized patients).
normal healthy blood-donor platelets but not to GT platelets. Likewise, binding/elution experiments with this material on insoluble purified GPIIb/IIIa demonstrated that GPIIb/IIIa antibody was entirely absorbed on the column and that the GPIIb/IIIa column eluate contained HIV GP160 antibody (data not shown). Furthermore, in these experiments, F(ab')2 binding could be demonstrated only with an anti-light chain but not with an antiglobulin directed against the Fc portion of the γ chain, ruling out binding by residual IgG contaminating the F(ab')2 preparation. Overall, these results demonstrated the presence in patient Bou of GP160/120 antibodies able to bind specifically to platelet GPIIb/IIIa.

Binding of the cross-reactive antibody to r-GP160 of the HIV-1 envelope GP. To characterize the cross-reactive epitope between HIV-GP160/120 and platelet GPIIb/IIIa, several experiments were conducted (Table 2). Direct Bou platelet eluate did not recognize the cell membrane of HIV-1–infected cell lines (U937-HTLVIIL and CEM-Bru), whereas intracytoplasmic staining was observed when infected cells were permeabilized with methanol and Triton X-100, suggesting that the eluate contained HIV antibodies probably directed against the native immature GP160.

Binding and elution experiments were performed with patient Bou platelet eluates on the two immobilized r-GP160s of the HIV envelope. Nonabsorbed and eluted materials from the column were tested for anti-HIV and antiplatelet activities. As shown in Table 2, neither the antiplatelet nor anti-HIV activities present in the Bou eluate bound to r-GP160-1, whereas they bound almost completely to r-GP160-2. The same results were obtained with the patient Bou serum F(ab')2 fragments. These results suggest either that the cross-reactive epitope recognized was located on the deleted portion of r-GP160-1 (present on r-GP160-2) or that it is dependent on the glycosylation or conformational structure of the GP.
Karpatkin et al suggested that both autoantibody fixation and nonspecific immune complex deposition may be responsible for the thrombocytopenia.\cite{6,12,14}

The present findings concerning 28 HIV-infected ITP patients confirm our previous results.\cite{18} IgG with antiplatelet activity was observed in the serum of 89% of the patients, PAIgG levels were increased in 80%, and 9 of 20 direct platelet eluates contained platelet antibodies sometimes immunochemically characterized as anti-GPIIb/IIIa. In addition, we noted that in one of these patients (patient Bou), serum antiplatelet IgG specifically bound to normal platelets and purified immobilized GpIIB/IIIa through the F(ab')2 portion. These results argue for an autoimmune cause of the thrombocytopenia.

We also observed HIV antibodies (mainly anti-GP160/120) in the direct eluates of the patients' platelets as previously reported.\cite{14,33} Apparently because HIV antibodies bind in vivo to the platelet membrane and not because a granule-incorporated plasma proteins are extracted,\cite{35} since GP160/120 antibodies were strongly predominant and were absorbed by or eluted from normal platelets but not from GT platelets. This binding was not explained by the presence of HIV antigens on the platelet membrane because viral antigens were not detectable in various immunologic tests and especially because the anti-HIV activity also bound to platelets from HIV-negative subjects. The absence of viral antigens or proviral DNA on HIV-ITP platelet extracts has been reported by other investigators.\cite{14}

Furthermore, binding of HIV antibodies to the platelet surface was specific in the case of patient Bou and occurred through the F(ab')2 portion. Indirect evidence for the responsibility of this anti-HIV activity in immune platelet destruction is the finding that all direct and indirect platelet eluates from thrombocytopenic patients, but only a few of those obtained from nonthrombocytopenic patients, contained anti-GP160/120 antibodies.

The specific binding of both anti-HIV and antiplatelet activities to normal platelets and purified GpIIB/IIIa suggested the presence of an antibody that recognizes both HIV and platelet antigens. Shared antigenicity between an infectious agent and normal tissue as a mechanism of host damage has been demonstrated in parasitic, bacterial,\cite{35,37} and, more recently, viral systems.\cite{38,41} To test this hypothesis, we focused our study on patient Bou, who showed the highest titer of antiplatelet (anti-GP1IB/IIIa) and anti-HIV (anti-GP160/120) activities that specifically bound to normal platelets and purified GpIIB/IIIa. Results of the absorption-elution experiments on purified immobilized GpIIB/IIIa, which bound both patient Bou GpIIB/IIIa and HIV GP160 antibody, as well as the absence of binding of either activity to platelets from GT patients clearly demonstrated the presence in the serum or eluates of a single antibody with both activities. This antibody recognized only the native immature GP160 in HIV-infected cell lines. To confirm cross-reactivity, we performed cross-absorption experiments with two r-GP160s and direct and indirect Bou eluates, as well as serum F(ab')2 fragments. The cross-reactive antibody could be absorbed on r-GP160-2 (expressed in baculovirus-infected insect cells) but not on r-GP160-1 (expressed in vaccinia virus-infected BHK 21 cells). Then, we believe, the absence of recognition of r-GP160-1 would result from abnormal glycosylation leading to conformational changes and rendering some epitopes inaccessible.

Therefore, we clearly demonstrated in patient Bou the presence of a platelet-bindable antibody that specifically recognizes an epitope (not precisely identified) shared by HIV GP160/120 and platelet GpIIB/IIIa, suggesting that homologies exist between the two GPs. Perhaps only small numbers of HIV-infected patients develop such antibodies at a titer sufficient to lead to platelet destruction. Alternatively, HIV antibodies may also occasionally bind to undetectable viral antigens on the platelet membrane, because we\cite{43} and other investigators have demonstrated the presence of HIV transcripts and, occasionally, HIV proteins in megakaryocytes from HIV-infected thrombocytopenic patients. However, the presence of both GP160 and GpIIB/IIIa antibodies in at least 5 of the 20 direct platelet eluates studied argues for the frequent responsibility of cross-reactive antibodies in the immune platelet destruction observed in HIV-infected patients. Finally, if the hypothesis of cross-reactivity is confirmed, the antigenic structure involved will be of importance because it will have to be taken into account in treatment of such patients and in production of vaccines based on HIV envelope components.

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We thank Professor M. Seligmann (Hôpital Saint Louis, Paris, France) for invaluable help, Drs M.L. Dubreuil and A. Sobel for referring patients, Dr L. Desforges (Département de Virologie, Hôpital Henri Mondor, Créteil, France) for providing the infected CEM cell line, Dr C. Dessaint for performing the ELISA test, J.-M. Massé (INSERM U 91, Créteil, France) for photographic assistance, B. Bürglen for technical assistance, C. Marie-Joseph for typing the manuscript, and D. Young for help with the English translation.

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**Table 2. Binding of Patient Bou Cross- Reactive Antibody to r-GP160s of the HIV1 Envelope**

<table>
<thead>
<tr>
<th>Assay</th>
<th>Platelet Eluates (direct of indirect)</th>
<th>Unbound Material After Absorption of Platelet Eluates on r-GP160(1)*</th>
<th>Unbound Material After Absorption of Platelet Eluates on r-GP160(2)*</th>
</tr>
</thead>
<tbody>
<tr>
<td>PIIFT</td>
<td>+++</td>
<td>+</td>
<td>+</td>
</tr>
<tr>
<td>HIV-ELISA (cutoff = 0.28)</td>
<td>1.3 ± 0.4</td>
<td>1.0 ± 0.1</td>
<td>0</td>
</tr>
<tr>
<td>HIV-Western blot</td>
<td>GP160/120</td>
<td>GP160/120</td>
<td>GP160/120</td>
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<tr>
<td></td>
<td>(+)</td>
<td>(++)</td>
<td>(+)</td>
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</table>

*PIIFT, platelet indirect immunofluorescence test on normal platelets. Platelet membrane fluorescence was graded as negative (−), weakly positive (+), or positive (+ + + +) in comparison with results obtained using negative control (serum from a nontransfused healthy male blood group AB donor), and a positive control (pool of sera from multitransfused immunized patients). Direct eluate, antibodies eluted from patient Bou's own platelets; indirect eluate, antibody eluted from normal platelets preincubated with patient Bou serum.

**r-GP160 (1)** and (2), two different recombinant GP160 of the HIV1 envelope (described in the Materials and Methods section).
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Presence of cross-reactive antibody between human immunodeficiency virus (HIV) and platelet glycoproteins in HIV-related immune thrombocytopenic purpura

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