Plasma Antigen Levels of the Lipoprotein-Associated Coagulation Inhibitor in Patient Samples

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Human plasma contains an inhibitor of tissue factor-initiated coagulation known as the lipoprotein-associated coagulation inhibitor (LACI) or also known as the extrinsic pathway inhibitor (EPI). A competitive fluorescent immunoassay was developed to measure the plasma concentration of LACI in samples from normal individuals and patients with a variety of diseases. The LACI concentration in an adult control population varied from 60% to 160% of the mean with a median value corresponding to 89 ng/mL or 2.25 nmol/L. Plasma LACI levels were not decreased in patients with severe chronic hepatic failure, warfarin therapy, primary pulmonary hypertension, thrombosis, or the lupus anticoagulant. Plasma LACI antigen was decreased in some, but not all patients with gram-negative bacteremia and evidence for disseminated intravascular coagulation. Plasma LACI levels were elevated in women undergoing the early stages of labor (29%), in patients receiving intravenous tissue-type plasminogen activator (45%), and in patients receiving intravenous heparin (375%). A radioligand blot of the pre- and post-heparin plasma samples shows the increase to be in a 40-Kd form of LACI. Very low levels of plasma LACI antigen were found in patients with homozygous abetalipoproteinemia and hypobetalipoproteinemia, diseases associated with low plasma levels of apolipoprotein B containing lipoproteins. Following the injection of heparin into one patient with homozygous abetalipoproteinemia, the plasma LACI antigen level increased to a level comparable with that in normal individuals after heparin treatment.

I NCREASING EVIDENCE suggests that tissue factor provides the initial trigger for blood coagulation in normal hemostasis and likely plays an important role in the pathologic thrombosis associated with atherosclerosis, inflammation, and cancer. That plasma contains an inhibitor of tissue factor-initiated coagulation was demonstrated in 1947 by investigators who discovered that preincubation of tissue extracts with plasma protected mice from the lethal disseminated intravascular coagulation caused by injection of the tissue extracts. This inhibitor is associated with lipoproteins in plasma and requires the presence of factor Xa to inhibit the tissue factor/factor VII(a) complex.

Our group has named this inhibitor the lipoprotein-associated coagulation inhibitor (LACI) and it is also known as the extrinsic pathway inhibitor (EPI). LACI has been purified from HepG2 cells, lipoproteins, and post-heparin plasma and its cDNA cloned from fetal liver and placental cDNA libraries.

Several groups have studied plasma LACI levels using functional assays. One report claimed that LACI levels were uniformly decreased in patients with disseminated intravascular coagulation (DIC), suggesting that LACI was consumed in a clinical situation in which tissue factor would be exposed to blood. LACI levels were not decreased in patients with severe hepatocellular disease, which was interpreted as indicating that the liver is not the main source of LACI in vivo. However, other investigators found normal to elevated levels of LACI in DIC and low levels of LACI in some cases of severe acute hepatic failure. LACI levels were normal in patients with the lupus anticoagulant and on warfarin therapy. LACI levels were modestly elevated in pregnancy and cancer, and acute coronary disease. The most remarkable finding was that plasma LACI levels increased twofold following an intravenous dose of 7,500 U of heparin. The source of the heparin-releasable LACI (HRL) is not known; however, the authors hypothesized that a pool of LACI is associated with endothelial cell glycosaminoglycans in vivo. There are no reports of LACI deficiency associated with thrombosis.

We developed an immunoassay to measure LACI antigen in a wide variety of patient samples. The assay is rapid, simple, and reproducible and allows for the screening of large numbers of clinical samples. We sought to confirm and extend the results found by others with functional assays, with particular emphasis on studying patients with thrombosis, lipid abnormalities, and diseases in which tissue factor may be exposed to blood.

MATERIALS AND METHODS

Materials. Pooled human plasma was obtained from George King Biomedical (Overland Park, KS). Carboxyl polystyrene latex particles were from Interfacial Dynamics Corp (Portland, OR) and fluorescein-5-isothiocyanate (FITC) was obtained from Molecular Probes, Inc (Eugene, OR). EDAC (1-ethyl-3-[3-dimethylaminopropyl] carbodiimide hydrochloride) was obtained from Sigma Chemical Co (St Louis, MO) as were affinity isolated antibodies to mouse IgG (whole molecule), absorbed with human serum proteins. Bovine serum albumin (BSA), fraction V, was obtained from Miles Scientific (Naperville, IL) and Biogel P-2 from Biorad (Richmond, CA). Porcine intestinal heparin for intravenous injection was obtained from Elkins-Sinn, Inc (Cherry Hill, NJ) and dermatan sulfate for intravenous injection was obtained from Mediolanum (Milan, Italy). All other reagents were from Sigma or Fisher (Springfield, NJ).

Methods. HepG2 LACI, bovine factor Xa, and a rabbit anti-LACI (HepG2) serum were prepared as previously described. Patient samples were provided from the following sources:

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Red Cross Blood donors, St Louis Bi-State Chapter of the American Red Cross; patients with thrombosis, Dr William Haire (University of Nebraska); patients with primary pulmonary hypertension, Dr Stewart Rich (University of Chicago); patients receiving tissue-type plasminogen activator (tPA), Dr Paul Eisenberg (Washington University); patients with homoygous abetalipoproteinemia, hypobetalipoproteinemia, apolipoprotein E deficiency, and Tangier disease, Dr Stephen M. Prescott (one abeta, one Tangier) (University of Utah), Dr James Keating (one abeta) (Children's Hospital, St Louis), and Drs Daniel J. Rader and H. Bryan Brewer, Jr (three Tangier, one Tangier with heparin, four abetas, one abeta with heparin, two hypobetas, one apo E deficiency before and after heparin) (National Institutes of Health). All other samples were obtained from the chemistry or hematology laboratories at the Jewish Hospital of St Louis or Barnes Hospital, St Louis.

**Assays.** LAC1 functional assays (tissue factor-inhibition assay) were performed as described. The assays for fibrin degradation products (FDP) were performed in the clinical coagulation laboratory at Barnes Hospital and used latex particles coated with antibody to human fibrinogen. A positive result is seen as agglutination of the particles in the presence of diluted serum samples. Samples were considered positive at an FDP concentration of greater than 8 μg/mL.

LAC1 immunoassay. A competitive florescent immunoassay was constructed in which an anti-LAC1 rabbit serum is used to bind LAC1, goat antirabbit IgG covalently linked to latex particles is used to capture the rabbit IgG, and FITC-LAC1 is added to quantitate the number of free anti-LAC1 IgG sites remaining. To an Eppendorf tube is added 200 μL of the plasma sample to be tested (diluted 1/4 or greater in TBSA [50 mmol/L tris-HCl, pH 7.5/100 mmol/L NaCl/0.1% BSA]) and 200 μL of anti-LAC1 rabbit serum (diluted 1/350 in TBSA). After an overnight incubation, four 40-μL aliquots of the mix are added to wells of a 96-well assay plate (Baxter-Pandex Laboratories, Inc, Mundelein, IL) that already contain 20 μL of goat antirabbit IgG linked to latex beads (resuspended at 0.5% wt/vol in TBSA). After 30 minutes of incubation at room temperature, 20 μL of FITC-LAC1 (5 μg/mL in TBSA) are added, and 6 minutes later a vacuum applied to the assay plate separating the solution from the latex particles. The particles are washed once for 2 minutes with TS (50 mmol/L tris-HCl, pH 7.5/100 mmol/L NaCl) and the amount of bound fluorescence quantitated with a Screen Machine (Baxter-Pandex). Standard curves were constructed using dilutions of pooled human plasma (George King Biomedical) in TBSA.

Immunoprecipitations, immunoblots, and radioligand blots were performed as previously described.

**Covalent linkage of antirabbit IgG to latex particles.** Five milligrams of affinity-purified goat antirabbit IgG were added to 6 mL carboxylated latex particles (4.2% wt/vol) and 0.1 mol/L sodium citrate, pH 5.0, was added to a final volume of 50 mL. After gentle

![Fig 1](representative-standard-curve-for-the-LAC1-immunoassay.png)

**Fig 1.** Representative standard curve for the LAC1 immunoassay. Standard curve was constructed using dilutions of pooled human plasma (George King). The precision of the assay was estimated by testing a normal plasma in quadruplicate 12 times.

![Fig 2](correlation-of-LAC1-immunoassay-to-LAC1-functional-assay.png)

**Fig 2.** Correlation of LAC1 immunoassay to LAC1 functional assay (tissue factor-inhibition assay). Serum samples from 43 Red Cross blood donors were tested for LAC1 antigen and LAC1 function (tissue factor inhibition) and results compared as shown.

![Fig 3](concentration-of-LAC1-antigen-in-a-control-population.png)

**Fig 3.** Concentration of LAC1 antigen in a control population. The control population consisted of Red Cross blood donors, visitors to a health fair, and medical students.
rocking at room temperature for 1 hour, 50 mg solid EDAC [1-ethyl-3-(2-dimethylaminopropyl)carbodiimide hydrochloride] was added and the mixture rocked gently at room temperature for 1 more hour. The particles were pelleted by centrifugation at 3,000g and resuspended in a final volume of 50 mL TBSA. Following one more TBSA wash, the particles were resuspended in a final volume of 50 mL TBSA and stored at 4°C.

**Fluorescein labeling HepG2 LACI.** HepG2 LAC1 (0.1 mg) was dialyzed overnight into HS (0.05 mol/L HEPES, pH 7.4/0.1 mol/L NaCl)/0.05% p-octylglucoside) and diluted to a final volume of 800 μL. To the LAC1 was added 200 μL of FITC at 1 mg/mL in 0.5 mol/L sodium carbonate, pH 9.5. This mixture was rocked at room temperature for 1 hour and free florescein was separated from FITC-LAC1 by chromatography on a Biogel P-2 column (Pharmacia). Fractions were assayed for protein and FITC by absorbance at 280 and 495 nm. The FITC-LAC1 fractions were pooled and stored at 4°C in HS/0.05% p-octylglucoside.

**RESULTS**

**Standardization of immunoassay.** An example of a standard curve for the immunoassay is shown in Fig 1. The coefficient of variation for a control plasma, assayed in quadruplicate, on a single plate on a single day was 2.3%. Six standard curves, using serial dilutions of different control plasmas, were parallel to a standard curve using dilutions of pooled human plasma (data not shown). Using serum samples from 43 adult Red Cross blood donors, LAC1 levels as determined by immunoassay were shown to correlate with LAC1 levels as determined by functional assay (tissue factor-inhibition assay), with an r value of .89 (Fig 2).

**Population distribution.** The concentration of LAC1 antigen in a control population is shown plotted on a frequency histogram in Fig 3. The control population consisted of Red Cross blood donors, visitors to a health fair, and medical students. The 95% range of plasma LAC1 antigen extended from 60% to 160% of the mean, with the

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**Fig 4.** Composite results measuring LAC1 antigen in a variety of patient samples. Bar represents the 95% range. (×) The mean and number of patients in each group in parenthesis.

**Fig 5.** LAC1 antigen in patients with gram-negative bacteremia with (greater than 8 mg/mL) and without FDP. (---) Mean for each group.

**Fig 6.** Mean serial levels of plasma LAC1 antigen (-•-) and hematocrit (-○-) in patients undergoing hip (N = 17) and knee (N = 9) replacement surgery. Patients were transfused an average of 2.5 U of packed red blood cells on the day of surgery and 0.75 U of packed red blood cells on postoperative day 1. None of the patients received plasma or heparin.
mean level corresponding to 89 ng/mL of LACI or 2.25 nmol/L.

Patient samples. The plasma LACI antigen levels in a variety of patient diagnostic groups are shown in Fig 4. Plasma LACI antigen was increased 29% in women undergoing the early stages of labor compared with sex and age matched controls (Fig 4). Plasma LACI antigen was not decreased in patients with severe chronic hepatocellular disease or in patients on warfarin therapy (Fig 4). Plasma LACI antigen was elevated in patients with gram-negative bacteremia, without FDP. However, plasma LACI antigen was decreased in some patients with gram-negative bacteremia and FDP (Figs 4 and 5). A comparison of the individual values shows that the ranges of values in each group are quite wide and overlap considerably (Fig 5). There was no correlation between LACI levels and levels of FDP (not shown).

Plasma LACI antigen decreased postoperatively in patients undergoing total hip and knee replacement surgery (Fig 6); however, the degree of LACI decrease appears to correspond to the degree of postoperative blood loss. Plasma LACI levels were not decreased in patients with the lupus anticoagulant or primary pulmonary hypertension (Fig 4), including several patients with thrombogenic primary pulmonary hypertension.23

Plasma LACI antigen was not decreased in a group of 90 patients with thrombosis (Fig 4). This group included 65 patients with deep venous thrombosis and/or pulmonary emboli, 10 patients with peripheral arterial occlusion, five patients with stroke, and 10 patients with coronary artery disease. Venooclusion increased LACI levels 15% in these patients. The highest levels of plasma LACI antigen were found in patients receiving intravenous heparin. Twenty-three patients undergoing cardiac catheterization received 2,500 U of intravenous heparin during the procedure and had plasma samples drawn before and then 10 to 15 minutes after injection. The increase in plasma LACI following intravenous heparin appears to be quite variable in this group, ranging from 150% to 650% of baseline levels. This finding was confirmed in a more controlled fashion by injecting increasing doses of heparin into seven healthy male volunteers and taking plasma samples 6 minutes after injection (Fig 7). The time course of LACI increase in a normal volunteer following intravenous dose of 7,500 U of heparin (100 U/kg) is shown in Fig 8. In this volunteer, LACI levels peak at 6 to 8 minutes at 950% of the baseline level. A radioligand blot of these plasma samples, probed with I125-factor X, shows the increase to be in a 40-Kd form of LACI (Fig 8, insert). When the same volunteer was
injected with dermatan sulfate (2 mg/kg), plasma LACI antigen did not increase significantly (data not shown). Plasma LACI levels also increased significantly (45%) in patients infused with tPA for acute myocardial infarction (Fig 4). These patients had not received heparin before these samples were taken.

Plasma LACI antigen was elevated a mean of 12% in patients with increased levels of very low-density lipoproteins (VLDL) and low-density lipoproteins (LDL) from a variety of causes, including familial hypercholesterolemia (Fig 4). There was no correlation between the level of plasma LACI antigen and the levels of LDL cholesterol, high-density lipoprotein (HDL) cholesterol, or triglycerides in 50 hospitalized patients (data not shown). The lowest levels of plasma LACI antigen were found in six patients with homozygous abetalipoproteinemia and two patients with hypobetalipoproteinemia (Fig 4), distinct diseases associated with low plasma levels of apolipoprotein B-containing lipoproteins.24 These patients do not suffer from abnormal thrombosis. Four patients with Tangier disease, lacking normal HDL, have normal plasma LACI levels. An immunoblot of a Tangier patients’ lipoproteins show that they have both major forms of plasma LACI (34 and 40 Kd) present in LDL (Fig 9). One patient with homozygous abetalipoproteinemia and one patient with Tangier disease were injected with 60 U/kg of heparin and bled 6 minutes later showing normal increases in plasma LACI antigen (Fig 4). One patient with complete apolipoprotein E deficiency had an elevated baseline level of plasma LACI antigen (221% of pooled plasma) and a supranormal response to intravenous heparin (1412% of pooled plasma) (Fig 4). An immunoblot of this patient’s lipoproteins show a pattern similar to the normal pattern (Fig 9).

DISCUSSION

Several groups have studied plasma LACI levels using functional assays. We describe here the first immunoassay used to measure LACI antigen in patient plasma samples. The data in this study confirm results from other groups studying patients with liver disease, on warfarin therapy, with the lupus anticoagulant,14 and with pregnant women.17,18

Is LACI consumed in DIC? As opposed to groups who found uniformly low,14 or normal to high15,16 levels of LACI in DIC, we found tremendous variation in LACI levels in patients with gram-negative bacteremia and FDP. Although it is tempting to attribute the lowest LACI levels in this group to consumption of LACI, we believe that cannot be done. These patients were clinically very complicated, often hypotensive, receiving multiple blood products, and with multi-organ failure. How LACI levels would be affected by these clinical variables, in the presence or absence of DIC, is not known.

Is LACI synthesized by the liver in vivo? That LACI levels are normal in patients with severe chronic hepatic failure does not conclusively answer this question. Patients with hepatic failure both synthesize and clear lipoproteins at a rate that is lower than normal. Abnormal LACI levels in some patients with lipid disorders suggest that plasma LACI levels are related to lipoprotein metabolism. One recent study found low levels of LACI activity and messenger RNA (mRNA) in primary hepatocyte cultures25; however, it is not clear that this accurately reflects in vivo activity.

Is LACI a physiologically important natural anticoagulant? We had hoped to answer this question by finding a patient or family in which LACI deficiency was associated with thrombosis. None of the 90 individuals with thrombosis tested in this study had a low level of plasma LACI antigen. We are now searching for plasma samples from patients with strong family histories of thrombosis, which were not present in the patients tested thus far.
Sandset et al, using a functional assay, had previously reported that LACI levels increase twofold following an intravenous dose of 7,500 U of heparin.21 We have found significantly greater increases in plasma LACI antigen following heparin infusion. Although the source of HRL is not known, Sandset et al hypothesized that a pool of LACI exists in vivo associated with vascular endothelial glycosaminoglycans. We have recently purified heparin-releasable LACI from post-heparin plasma and found that it is not tightly associated with lipoproteins, and it binds heparin-agarose with high affinity.22

The source and mechanism of LACI increase following intravenous tPA is also not known. Saksela and Rifkin have reported that plasmin proteolytically cleaves endothelial cell heparan sulfate glycosaminoglycans, thereby releasing complexes of partly degraded heparan sulfate and basic fibroblast growth factor.23 If LACI were anchored to endothelial cell heparan sulfate proteoglycans in vivo, then LACI could theoretically be released by a similar mechanism.

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