Correlation of Response of Aplastic Anemia Patients to Antilymphocyte Globulin With In Vitro Lymphocyte Stimulatory Effect: Predictive Value of In Vitro Test for Clinical Response

By Tokuichiro Abe, Hiroshi Matsuoka, Seiji Kojima, Yoshiro Kamachi, Ikuya Tsuge, Yoshihisa Kodera, and Takaharu Matsuyna

Therapy with antilymphocyte globulin (ALG) has been shown to be effective in restoring hematopoiesis to some patients with aplastic anemia. It would be useful to have a method for predicting those likely to be responders versus nonresponders. The mode of immunostimulatory action of ALG is of interest in addition to its immunosuppressive action. We examined in vitro the distribution of the proliferative responses of ALG-stimulated peripheral blood mononuclear cells (PBMCs) obtained from 18 patients with aplastic anemia, eight of whom responded to ALG and 10 who did not. We found a significant difference in the proliferative response of PBMCs obtained from the eight responders versus the 10 nonresponders (P < .01). Two-color flow cytometry analysis of the patients' PBMCs stimulated by ALG in vitro showed that the CD4-positive subsets were activated to a greater extent by ALG than the CD8-positive subsets. Moreover, a positive correlation with the clinical response of patients to ALG with granulocyte-macrophage colony-stimulating factor produced by their PBMCs stimulated by ALG suggests that the immunostimulatory property of ALG has an important role in the treatment of aplastic anemia. Our results suggest that the clinical response to ALG therapy is correlated with its lymphocyte proliferative effect in vitro, and indicates that the assessment of the proliferative response of PBMCs in vitro would be useful in predicting the clinical response to ALG therapy.

© 1991 by The American Society of Hematology.

Although the clinical response varies from 28% to 77%, the administration of antilymphocyte globulin (ALG) has been considered effective in treating severe aplastic anemia. Efforts have been made to find a relationship between those patients who respond to ALG therapy and the results of in vitro tests or some pretreatment clinical variable to predict those likely to respond to ALG. Since the introduction of ALG therapy for aplastic anemia, it is believed that ALG exerts its cytotoxic effect on lymphocytes in a complement-dependent manner in vivo as well as in vitro, and indirectly improves the suppressed hematopoiesis. Although some studies have attempted to clarify the mode of ALG action in this respect, the findings are not in agreement. Recent in vitro observations indicate that ALG can exert a mitogenic effect on the lymphocytes and induce them to produce hematopoietic growth factors. This mechanism for the response to ALG treatment is supported by findings in treating aplastic anemia with recombinant human granulocyte-macrophage colony-stimulating factor (rhGM-CSF) and with rhG-CSF. Assuming that the immunostimulatory effects of ALG lead to an improvement of the bone marrow hypoplasia, we investigated the correlation between the clinical response observed with ALG therapy of aplastic anemia and the lymphocyte stimulatory effect of this agent in vitro.

MATERIALS AND METHODS

Patients. Eighteen patients with aplastic anemia, 10 males and eight females aged 2 to 43 years, were included in the study. Their clinical and laboratory data are summarized in Table 1. Of the 18 patients, 14 had severe aplasia according to the criteria of the International Cooperative Aplastic Anemia Study Group. The cause of the aplasia was acute hepatitis in one patient, but etiology was unknown in 17. The duration between verification of the diagnosis to the initiation of therapy ranged from 2 weeks to 52 months. Nine patients had acute disease (ie, diagnosed within 3 months of initiating the study), while nine had chronic disease. This study was approved by the Review Board of the Nagoya University School of Medicine. Patients were informed that blood samples were obtained for research purposes and that their privacy will be protected.

Therapy. All patients were treated with a combination of ALG (Ahlbulin; Green Cross, Osaka, Japan) and high-dose methylprednisolone with an androgen. ALG was administered at a dose of 30 to 40 mg/kg/d infused intravenously (IV) daily for 5 days. Methylprednisolone was administered daily for 28 days, initially at a high dose administered IV and then tapered off to an oral dose as follows: days 1 through 4, 20 mg/kg IV/d; days 5 through 8, 10 mg/kg IV/d; days 9 through 12, 5 mg/kg IV/d; days 13 through 20, 2 mg/kg per os/d; and days 21 through 28, 1 mg/kg per os/d. All patients were also receiving danazol (5 mg/kg/d) or Mepitiostane (0.3 mg/kg/d), administered orally for 1 year. The hematologic response of each patient was evaluated 3 months after the initiation of treatment. All patients were observed over 12 months. Responders were defined as those patients with an increase of at least two of the following parameters in their peripheral blood cell counts: granulocytes >1.0 x 10^9/L, platelets >50 x 10^9/L, and reticulocytes >60 x 10^9/L. All patients were observed over 12 months.

Preparation of peripheral blood mononuclear cells (PBMCs). Before ALG therapy was initiated, PBMCs from the 18 patients were isolated on Ficol-Paque (Pharmacia, Picataway, NJ) density gradients, and resuspended in RPMI 1640 (GBICO, Grand Island, NY) supplemented with 10% fetal calf serum (FCS, lot 60011; MA Bioproducts, Walkersville, MD), 2 mmol/L L-glutamine, 10 mmol/L N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid (HEPES), 1.2 mg/mL NaHCO₃, 100 U/mL penicillin, and 100 µg/mL streptomycin.

From the Department of Pediatrics, Nagoya University School of Medicine; Division of Hematology/Oncology, Children's Medical Center; and the Department of Medicine, The Japanese Red Cross Nagoya First Hospital, Nagoya, Japan.


Supported by a grant from the Mother and Child's Health Foundation.

Address reprint requests to Tokuichiro Abe, MD, Department of Pediatrics, Nagoya University School of Medicine, Tsuurnai-cho 65, Showa-ku, Nagoya 466, Japan.

The publication costs of this article were defrayed in part by page charge payment. This article must therefore be hereby marked "advertisement" in accordance with 18 U.S.C. section 1734 solely to indicate this fact.

© 1991 by The American Society of Hematology.

0006-4971/91/7710-0021$3.00/0

Blood, Vol 77, No 10 (May 15), 1991; pp 2225-2230

2225
were cultured in RPMI-FCS with ALG (250 kg/mL) or PHA (10 pg/200 pL phytohemagglutinin (PHA-P; Difco, Detroit, MI) under the same conditions as in the case of ALG therapy. There was no significant difference between responders versus nonresponders as to their age, sex, severity of disease, duration to the initiation of therapy, pretreatment peripheral blood counts (granulocytes, reticulocytes, or platelets), or pretreatment blood transfusions.

The severity of aplastic anemia was assessed according to the criteria of the International Cooperative Aplastic Anemia Study Group.6

A total of eight patients (nos. 1 through 8) were responders, while 10 patients (nos. 9 through 18) were nonresponders. There was no significant difference between responders vs nonresponders (P > .1) with respect to age, sex, severity of disease, duration to the initiation of therapy, pretreatment peripheral blood counts (granulocytes, reticulocytes, or platelets), or pretreatment blood transfusions.

*The severity of aplastic anemia was assessed according to the criteria of the International Cooperative Aplastic Anemia Study Group.6

Clinical response. Of the 18 patients, eight (44%) showed a response to ALG therapy and 10 (56%) had no response 3 months after the initiation of treatment. Of the eight responders, six were maintained in remission at 12 months and two relapsed at 12 and 36 months after the initial response, one of whom responded to a second course of ALG therapy. There was no significant difference between responders versus nonresponders as to their age, sex, duration to the initiation of therapy, severity of disease, their pretreatment granulocyte, reticulocyte, or platelet counts (the Student's t-test, P > .1), or pretreatment blood transfusions (over 100 U of packed red blood cells [PRBCs] and platelets, χ² test, P > .1).

Dose-response curves of PBMCs from patients to ALG. PBMCs from the 18 patients were cultured with various concentrations of ALG and the proliferation of cells was analyzed by the incorporation of [3H]thymidine. Dose-response curves of ALG were also determined by liquid scintillation counter. Because of the variability of the biologic assay, PBMCs were also cultured with 2.5 pg/200 pL phytohemagglutinin (PHA-P; Difco, Detroit, MI) under the same conditions as in the case of ALG. The FCS, ALG, and horse serum IgG used in these studies were heat-inactivated at 56°C for 30 minutes.

GM-CSF assay. Supernatants for GM-CSF assay were prepared as follows: peripheral blood mononuclear cells from 14 of 18 patients, including 7 of 8 responders and 7 of 10 nonresponders, were cultured in RPMI-FCS with ALG (250 μg/mL) or PHA (10 μg/mL) in a culture tube (Falcon 3033; Becton Dickinson, Lincoln Park, NJ) at 1 × 10⁶ cells/mL for 48 hours. Cells were subsequently pelleted and the supernatants recovered and stored at −80°C until studied. GM-CSF concentrations in each culture supernatant were determined by enzyme-linked immunosorbent assay (ELISA; Enzyme, Boston, MA). The lowest level of GM-CSF detectable with this ELISA system was 0.001 ng/mL. The spontaneous production of GM-CSF from the patients' PBMCs were tested in parallel.

Two-color flow cytometry analysis of PBMCs cultured with ALG. Surface analysis of the PBMCs, cultured with ALG for 48 hours, was conducted by direct immunofluorescence using monoclonal antibodies (MoAbs). The latter were fluorescein isocyanate (FITC)-conjugated anti-CD3, anti-CD4, and anti-CD8 MoAbs (Coulter Immunology, Hialeah, FL), and phycoerythrin (PE)-conjugated anti-CD25 MoAb (DakoCytomation, Glostrup, Denmark). Cultured cells were incubated with anti-CD25 MoAb and one of the anti-T-cell MoAbs simultaneously at 4°C for 30 minutes. Flow cytometry analysis was performed using a fluorescence-activated cell sorter (EPICS PROFILES; Coulter).

RESULTS

Table 1. Patient Characteristics

<table>
<thead>
<tr>
<th>Patient No.</th>
<th>Age (y)</th>
<th>Sex</th>
<th>Etiology of Aplastic Anemia</th>
<th>Duration of Disease (mo)</th>
<th>Severity of Aplastic Anemia*</th>
<th>Pretreatment Peripheral Blood Counts</th>
<th>Blood Transfusions (U)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td>Neutrophils (× 10⁹/L)</td>
<td>Reticulocytes (× 10⁹/L)</td>
</tr>
<tr>
<td>1</td>
<td>8</td>
<td>M</td>
<td>Unknown</td>
<td>52</td>
<td>Moderate</td>
<td>0.6</td>
<td>44</td>
</tr>
<tr>
<td>2</td>
<td>26</td>
<td>M</td>
<td>Unknown</td>
<td>13</td>
<td>Moderate</td>
<td>0.5</td>
<td>45</td>
</tr>
<tr>
<td>3</td>
<td>3</td>
<td>M</td>
<td>Unknown</td>
<td>1.5</td>
<td>Severe</td>
<td>0.48</td>
<td>80</td>
</tr>
<tr>
<td>4</td>
<td>5</td>
<td>M</td>
<td>Hepatitis-associated</td>
<td>6</td>
<td>Severe</td>
<td>0.24</td>
<td>20</td>
</tr>
<tr>
<td>5</td>
<td>6</td>
<td>F</td>
<td>Unknown</td>
<td>3</td>
<td>Severe</td>
<td>0.22</td>
<td>57</td>
</tr>
<tr>
<td>6</td>
<td>6</td>
<td>M</td>
<td>Unknown</td>
<td>29</td>
<td>Severe</td>
<td>0.48</td>
<td>30</td>
</tr>
<tr>
<td>7</td>
<td>23</td>
<td>F</td>
<td>Unknown</td>
<td>3</td>
<td>Severe</td>
<td>0.48</td>
<td>28</td>
</tr>
<tr>
<td>8</td>
<td>43</td>
<td>M</td>
<td>Unknown</td>
<td>18</td>
<td>Severe</td>
<td>0.45</td>
<td>30</td>
</tr>
<tr>
<td>9</td>
<td>5</td>
<td>M</td>
<td>Unknown</td>
<td>48</td>
<td>Moderate</td>
<td>0.91</td>
<td>66</td>
</tr>
<tr>
<td>10</td>
<td>7</td>
<td>F</td>
<td>Unknown</td>
<td>0.5</td>
<td>Moderate</td>
<td>1.2</td>
<td>50</td>
</tr>
<tr>
<td>11</td>
<td>1</td>
<td>F</td>
<td>Unknown</td>
<td>2</td>
<td></td>
<td>0.074</td>
<td>16</td>
</tr>
<tr>
<td>12</td>
<td>2</td>
<td>F</td>
<td>Unknown</td>
<td>1</td>
<td>Severe</td>
<td>0.047</td>
<td>19</td>
</tr>
<tr>
<td>13</td>
<td>3</td>
<td>F</td>
<td>Unknown</td>
<td>2</td>
<td>Severe</td>
<td>0.74</td>
<td>18</td>
</tr>
<tr>
<td>14</td>
<td>6</td>
<td>F</td>
<td>Unknown</td>
<td>1</td>
<td>Severe</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>15</td>
<td>7</td>
<td>M</td>
<td>Unknown</td>
<td>20</td>
<td>Severe</td>
<td>0.46</td>
<td>16</td>
</tr>
<tr>
<td>16</td>
<td>11</td>
<td>F</td>
<td>Unknown</td>
<td>17</td>
<td>Severe</td>
<td>0.42</td>
<td>120</td>
</tr>
<tr>
<td>17</td>
<td>13</td>
<td>M</td>
<td>Unknown</td>
<td>1</td>
<td>Severe</td>
<td>0</td>
<td>0</td>
</tr>
<tr>
<td>18</td>
<td>22</td>
<td>M</td>
<td>Unknown</td>
<td>32</td>
<td>Severe</td>
<td>1.0</td>
<td>28</td>
</tr>
</tbody>
</table>

A total of eight patients (nos. 1 through 8) were responders, while 10 patients (nos. 9 through 18) were nonresponders. There was no significant difference between responders vs nonresponders (P > .1) with respect to age, sex, severity of disease, duration to the initiation of therapy, pretreatment peripheral blood counts (granulocytes, reticulocytes, or platelets), or pretreatment blood transfusions.

*The severity of aplastic anemia was assessed according to the criteria of the International Cooperative Aplastic Anemia Study Group.6

Analysis of the proliferative capability of the patients' PBMCs, the cells were cultured in RPMI-FCS at 1 × 10⁶ cells per 200 μL in triplicate using U-bottom 96-well plates (Nunc, Roskilde, Denmark) with various concentrations of ALG (lot 113GU) or horse serum IgG (lot 28839; Cappel, West Chester, PA). Plates were settled in a humidified atmosphere containing 5% CO₂ in air at 37°C for 70 hours. Cells were subsequently pulsed for 2 hours with 18.5 kBq of [3H]thymidine and harvested. Radioactivity was determined in a liquid scintillation counter. Because of the variability of the biologic assay, PBMCs were also cultured with 2.5 μg/200 μL phytohemagglutinin (PHA-P; Difco, Detroit, MI) under the same conditions as in the case of ALG. The FCS, ALG, and horse serum IgG used in these studies were heat-inactivated at 56°C for 30 minutes.

GM-CSF assay. Supernatants for GM-CSF assay were prepared as follows: peripheral blood mononuclear cells from 14 of 18 patients, including 7 of 8 responders and 7 of 10 nonresponders, were cultured in RPMI-FCS with ALG (250 μg/mL) or PHA (10 μg/mL) in a culture tube (Falcon 3033; Becton Dickinson, Lincoln Park, NJ) at 1 × 10⁶ cells/mL for 48 hours. Cells were subsequently pelleted and the supernatants recovered and stored at −80°C until studied. GM-CSF concentrations in each culture supernatant were determined by enzyme-linked immunosorbent assay (ELISA; Enzyme, Boston, MA). The lowest level of GM-CSF detectable with this ELISA system was 0.001 ng/mL. The spontaneous production of GM-CSF from the patients' PBMCs were tested in parallel.

Two-color flow cytometry analysis of PBMCs cultured with ALG. Surface analysis of the PBMCs, cultured with ALG for 48 hours, was conducted by direct immunofluorescence using monoclonal antibodies (MoAbs). The latter were fluorescein isocyanate (FITC)-conjugated anti-CD3, anti-CD4, and anti-CD8 MoAbs (Coulter Immunology, Hialeah, FL), and phycoerythrin (PE)-conjugated anti-CD25 MoAb (DakoCytomation, Glostrup, Denmark). Cultured cells were incubated with anti-CD25 MoAb and one of the anti-T-cell MoAbs simultaneously at 4°C for 30 minutes. Flow cytometry analysis was performed using a fluorescence-activated cell sorter (EPICS PROFILES; Coulter).
ALG'S LYMHOSTIMULATORY EFFECT IN AA THERAPY

Fig 1. Dose-response curves of PBMCs from 18 patients with aplastic anemia to ALG (—) and horse IgG (——). PBMCs were cultured with ALG or horse IgG for 72 hours. Proliferation of cells was subsequently assessed by 'H-thymidine incorporation. Results are expressed as Im ± SD (cpm) from each group of patients (responder, [O]; nonresponder, [Q]). Results obtained with these groups differed significantly (P < .01) for ALG (500 μg/mL) when compared by two-way analysis of variance. No statistical difference for horse IgG was found between nonresponders versus responders (P > .1).

Data from the responders and nonresponders are presented in Fig 2. The responders (logarithmic mean [Im] = 11,111, Im + SD = 16,710, Im − SD = 7,387 cpm) differed significantly from the nonresponders (Im = 4,229, Im + SD = 9,356, Im − SD = 1,912 cpm) for ALG (500 μg/mL) according to two-way analysis of variance (P < .01). No significant difference for PHA-P or horse IgG was found between the responders and nonresponders.

GM-CSF assay in culture supernatants from PBMCs stimulated with ALG. The distribution of GM-CSF produced by the ALG-stimulated PBMCs is shown in Fig 3C. The results obtained with the responders (Im = 0.0293, Im + SD = 0.1258, Im − SD = 0.0068 ng/mL) differed significantly from the nonresponders (Im = 0.0014, Im + SD = 0.0025, Im − SD = 0.0008 ng/mL; P < .01). The production of GM-CSF by the PBMCs of all but one responder ranged from 0.012 to 0.3 ng/mL. The single exception was the patient with hepatitis-associated aplastic anemia in whom the ALG-stimulated PBMCs could produce GM-CSF as 0.005 ng/mL among the responders. The concentration of GM-CSF in the supernatant from the PBMCs of five of the seven nonresponders was below 0.001 ng/mL, the lowest limit detected by this assay system. No significant difference for PHA-P was found between the responders and nonresponders.

Surface phenotype of PBMCs stimulated with ALG. An activated subpopulation of T lymphocytes in the PBMCs from three responders was analyzed with anti-CD25 MoAb using two-color flow cytometry after the incubation of PBMCs with ALG for 48 hours. A representative result appears in Fig 4. Before treatment, the expression of CD4, CD8, and CD25 in these patients' fresh PBMCs was examined; the means ± SD were 43.1% ± 5.3%, 41.3% ± 7.6%, and 0.7% ± 0.3%, respectively. Following incubation of PBMCs with ALG, the CD25 antigen was expressed for 13.3% ± 7.5% of the CD3-positive cells, predominantly among the CD4-positive rather than the CD8-positive T cells. The CD4-positive subpopulation was 78.8% ± 8.5%, whereas the CD8-positive cells were 21.0% ± 7.3% of the CD25-positive cells. Nonimmune horse IgG induced little CD25 antigen expression (<1.9% of the CD3-positive cells).

DISCUSSION

Because immunocompetent cells are involved in the pathogenesis of aplastic anemia, ALG is believed to restore the depressed hematopoiesis through a cytotoxic action on such cells. Because it has been difficult to demonstrate that the effectiveness of ALG in treating aplastic anemia is due to its cytotoxicity, an alternative
Fig 3. Distribution of GM-CSF production by PBMCs from 12 patients with aplastic anemia cultured in RPMI-FCS (A), with PHA stimulation (B), or ALG (C). The four patients consisted of one responder and three nonresponders (nos. 6, 9, 14, and 16), and were excluded from the GM-CSF assay because of the marked lymphocytopenia and the difficulty preparing the adequate number of PBMCs in these cases. Each dot represents one patient (responder, [e]; nonresponder, [o]). Im ± SD (nanograms per milliliter) of GM-CSF concentration in culture supernatants are also shown. Results obtained in the responders and nonresponders differed significantly \( P < 0.01 \) when PBMCs were cultured with ALG (C) (responders: \( \text{Im} = 0.0293, \text{Im} + \text{SD} = 0.1258, \text{Im} - \text{SD} = 0.0068 \text{ng/mL}; \) nonresponders: \( \text{Im} = 0.0014, \text{Im} + \text{SD} = 0.0025, \text{Im} - \text{SD} = 0.0008 \text{ng/mL} \). [*] Patient with hepatitis-associated aplastic anemia; (t), moderate cases.

Fig 4. Two-color flow cytometry of PBMCs from patients with aplastic anemia cultured with ALG (A) or horse IgG (B). Intensity of fluorescence (FITC, along the horizontal line; PE, along the vertical line) is expressed in a logarithmic scale. Representative results from three responders are shown (nos. 1, 5, and 6). CD25 antigen is expressed on 10.1% of the CD3-positive cells among the more predominantly CD4- than CD8-positive cells after incubation with ALG (A), in contrast to the scanty expression of CD25 antigen for CD3-positive cells after incubation with nonimmune horse IgG (B).
PBMCs. Other hematopoietic factors such as G-CSF and IL-3 would also be expected to be produced by PBMCs stimulated with ALG.

Our discussion of the lymphocyte stimulatory potencies of ALG does not necessarily disagree with previous reports that suggest the clinical importance of immunosuppressive agents. One such report is the downregulation of γ-interferon production induced by ALG in the PHA-stimulated patients' PBMCs and the correlation of clinical response to ALG with this activity of ALG. The recently reported excellent results with cyclosporine in aplastic anemia seem to suggest that an immunosuppressive mode of action of the therapeutic agent is essential, in comparison to the transient improvement obtained with GM-CSF administration.

Thus, the possibility is far from slight that ALG primarily exerts an immunosuppressive effect and that residual hematopoietic potency is essential for a response to ALG. However, our results with ALG therapy did not show a correlation between the response to ALG and the severity of disease. In addition, in a previous study we failed to detect the suppressor T cells to hematopoietic progenitors by comparing the growth of GM colony-forming cells and burst-forming units before and after treating bone marrow cells with anti–T-cell MoAbs and complement in patients with aplastic anemia. The clinical significance of the direct cytotoxicity of ALG to lymphocytes, especially to the T lymphocytes expected to be suppressive to hematopoiesis, is still controversial. It would be necessary to examine further the immunosuppressive potency of ALG on many other kinds of cells, including various subsets of lymphocytes, monocytes, and stroma cells.

Considering the pathophysiology of aplastic anemia and the mode of action of ALG, a possible therapeutic strategy would be to administer a hematopoietic factor such as GM-CSF, particularly if the clinical response is positively correlated with the in vitro production of the patients' PBMCs induced by ALG, given in combination with immunosuppressive therapy.

Antilymphocyte globulin has two different effects on the cells that are committed to the immune system, immunostimulatory and immunosuppressive, apparently in opposition to each other, but which, acting together, restore the suppressed hematopoiesis in some patients with aplastic anemia. Our results indicate that the clinical response to ALG is correlated with the lymphocyte proliferative response induced by this agent in vitro, and suggests the utility of an in vitro test for predicting the clinical response to ALG therapy.

ACKNOWLEDGMENT

We are obliged to Dr Keizo Horibe, Dr Testushi Yoshikawa, and Dr Kouichi Miyamura for referring patients for study. We thank Prof Kazuyoshi Watanabe and Prof Shimpei Torii for their advice on this manuscript.

REFERENCES


Correlation of response of aplastic anemia patients to antilymphocyte globulin with in vitro lymphocyte stimulatory effect: predictive value of in vitro test for clinical response

T Abe, H Matsuoka, S Kojima, Y Kamachi, I Tsuge, Y Kodera and T Matsuyama