A New Glucose-6-Phosphate Dehydrogenase Deficient Variant in a Patient With Chediak-Higashi Syndrome

By Josef T. Prchal, William M. Crist, Ahmad Malluh, Alena Vitek, W. Newlon Tauxe, and Andrew J. Carroll

A description is given of enzymatic characteristics of a new glucose-6-phosphate dehydrogenase variant (G-6-PD) found in 2 male members of an inbred Pakistani family. Properties of this enzyme differ in respect to electrophoretic mobility, utilization of deamino NADP, and $K_m$ for both G-6-PD and NADP from the variant reported to be common in Pakistan. The propositus, in addition to G-6-PD deficiency, also had Chediak-Higashi syndrome. Some male members of this family are affected by two distinct clinical disorders. One disease is characterized by intermittent jaundice and anemia, often concurrent with infection, compatible with G-6-PD deficiency. The other is characterized by severe, frequent, and occasionally fatal infections that are not necessarily associated with jaundice, a disease pattern compatible with Chediak-Higashi syndrome. A possible additive relationship of G-6-PD deficiency with Chediak-Higashi syndrome in causing frequent infections was examined. Only moderately decreased activity of G-6-PD was found in nonerythroid cells of 2 male children having this G-6-PD deficient variant. Moreover, the study of the patient’s mother revealed no obvious survival disadvantage of her nonerythroid cell clones carrying the G-6-PD-deficient variant as compared to the clones carrying the normal G-6-PD isoenzyme. Ascorbic acid therapy for the brother affected by both of these diseases did not lead to increased hemolysis. No obvious clinical improvement of severity and infection frequency was noted after several months of ascorbic acid therapy.

GLOUCOSE-6-PHOSPHATE DEHYDROGENASE is a polymorphic enzyme with over 100 variants described thus far.

Some of the deficient variants appear to be particularly common within certain racial and ethnic groups. For example, about 2.5% of Pakistanis were found to have G-6-PD deficiency characterized by no definite ill effects; this variant was designated as G-6-PD Campbellopore.

This report describes an inbred Pakistani family with a previously undescribed G-6-PD mutant associated with intermittent hemolytic anemia. One of the affected boys also had Chediak-Higashi syndrome, an autosomal recessive disorder characterized by partial albinism, granular giantism in many cell lines, frequent infections, and often neutropenia.

CASE REPORTS

An 8-mo-old Pakistani male, N.A., was admitted with fever to Children’s Hospital, University of Alabama in Birmingham. His height was in the 50th percentile and weight at the 7th percentile. He was pale and irritable. Multiple skin areas and hair were hypopigmented. His hemoglobin was 6 g/dl and the reticulocyte count varied between 0% and 7%, and his neutrophil count ranged between 90 and 4000/cumm. His lymphocyte and platelet counts were normal. The child was iron deficient with a serum ferritin of 5 ng/ml. He responded to iron therapy with normalization of the hemoglobin, and thereafter his reticulocyte count remained normal. Morphological examination of blood and marrow cells revealed giant granules in neutrophils, lymphocytes, monocytes, and platelets, and their hemopoietic precursors in the marrow. Skin melanocytes contained similar granules. A family history showed that multiple male members on the maternal side of the family were affected by either frequent infections, jaundice, or both (Fig. 1). The parents were first cousins and consanguinity had also occurred in previous generations. The patient’s mother had aborted two female fetuses. A.S. is 8 yr and 4 mo older than his brother, the propositus. He had a severe hemolytic crisis at the age of 1 yr that required multiple blood transfusions. Both boys were found to have severe G-6-PD deficiency, but the elder had no history of significant infections, nor had he had any morphological evidence of Chediak-Higashi syndrome.

MATERIALS AND METHODS

Biochemical Study

G-6-PD was partially purified and characterized by standard methods. The $K_m$ for NADPH was determined according to Yoshi-da. The erythrocytes, lymphocytes, platelets, and granulocytes were purified and skin fibroblasts were cultured, and G-6-PD electrophoresis

From the Divisions of Hematology, Pediatrics, Nuclear Medicine, and Medical Genetics, University of Alabama in Birmingham, Birmingham, Ala.

Supported in part by NIH Grant EY02432.

Address reprint requests to Josef T. Prchal, M.D., Division of Hematology, University of Alabama in Birmingham, 642 Lyons-Harrison Research, University Station, Birmingham, Ala. 35294.

© 1980 by Grune & Stratton, Inc. 0006-4971/80/5603-0021$01.00/0

From www.bloodjournal.org by guest on October 23, 2017. For personal use only.
Table 1. Kinetic Data of Described G-6-PD Variant

<table>
<thead>
<tr>
<th></th>
<th>Red Cell Enzyme Activity (% of Normal)</th>
<th>Electro-</th>
<th>Kinematic Data</th>
<th>Deamino</th>
<th>Heat</th>
<th>pH</th>
<th>NADPH (μM)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td></td>
<td>Electro-</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>phoretic</td>
<td>2-deoxy</td>
<td>G-6-P</td>
<td>G-6-P</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td>Mobility</td>
<td>NADP</td>
<td>(μM)</td>
<td>(μM)</td>
<td></td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td>(μM)</td>
<td>NADP</td>
<td>Stability</td>
<td>Optimum</td>
<td></td>
</tr>
<tr>
<td></td>
<td></td>
<td></td>
<td></td>
<td>G-6-P</td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>G-6-PD-Birmingham</td>
<td>5%</td>
<td>Phosphate</td>
<td>12</td>
<td>22</td>
<td>&lt;4</td>
<td>256%</td>
<td>Normal</td>
</tr>
<tr>
<td>Normal control</td>
<td>7.69 U/g Hb</td>
<td>Phosphate</td>
<td>41</td>
<td>5</td>
<td>&lt;4</td>
<td>61%</td>
<td>Normal</td>
</tr>
<tr>
<td>Reported range</td>
<td>100%</td>
<td>Phosphate</td>
<td>40-70</td>
<td>3-6</td>
<td>&lt;4</td>
<td>55%-60%</td>
<td>Normal</td>
</tr>
</tbody>
</table>

Informed consent and approval by the Human Use Committee was obtained prior to the study.

Red Cell Survival

Jandl demonstrated that the chromium released from destroyed labeled red cells is not utilized for red cell binding, but is excreted in the urine at a relatively standard daily rate. Shih et al. studied the kinetics of excretion of chromium-51 into the urine and determined that the chromium-51 is quantitatively excreted into the urine after red cell destruction. More recently, these authors have determined the red cell survival as based on urinary excretion of chromium calculated from 3 to 4 24-hr urine collections. We have modified this procedure to study erythrocyte destruction rates in this case. Erythrocytes from approximately 10 ml of whole blood were labeled with chromium and washed 3 times in sterile physiologic saline. An aliquot of the dose was retained as a standard. Thereafter, the urines were collected at 4-24 hr intervals. Each urinary sample was counted for radioactivity along with the dose aliquot, and the creatinine excretion was determined to eliminate possible errors of collection and varying glomerular filtration rate. The radioactivity of each urine sample was expressed as a percentage of the total injected radioactivity; this value was subtracted from 100% radioactivity in sequential fashion; and from the declining slope of the resulting curve, the red cell survival was determined.

RESULTS

Characterization of G-6-PD Variant

The enzymatic properties of the as-yet-undescribed G-6-PD variant are summarized in Table 1. Repeated testing of both brothers' purified erythrocytes revealed activity less than 5% of normal, whereas the mother's activity was 64% of the normal, a level compatible with heterozygosity. The enzyme was electrophoretically fast on starch gel, using phosphate buffer (Fig. 2). With EBT and Tris buffers, electrophoretic mobility was normal. The mobility of this variant on a cellulose-acetate electrophoretic system was also identical to that of a normal "B" enzyme. The Km for NADP was increased (22 μmole), but the Km for G-6-P was decreased (12 μmoles). The rate of utiliza-
with "Cr and washed before injection. No ascorbic acid was added prior to the injection. Giant granules were present and melanin heterozygote). Only slightly decreased intensity of the fast band (G-6-PD-Birmingham) compared to the slower band (G-6-PD type B), using starch gel with phosphate buffer system were found in all cell lysates.

Evaluation of Glutathione Metabolism

Table 3 shows levels of GSH, GSSG, and glutathione reductase in neutrophils and lymphocytes of the propositus N.S. and in a normal control. Normal glutathione reductase activity of granulocytes, when there is no evidence to indicate defective glutathione synthesis, rule out an impaired glutathione metabolism as a cause of infection in N.S.14,15

The Red Cell Survival of the Propositus and Influence of Ascorbic Acid on his Red Cell Survival

The survival of red cells before and after ascorbic acid challenge was estimated by labeling N.S.'s erythrocytes with chromium-51 followed by sequential detection of radioactive chromium in the urine. The results, demonstrated in Fig. 3, reveal that the lifespan of the red cells in the steady state is only slightly decreased. Furthermore, the challenge by 1 mg of oral ascorbic acid produced no significant hemolysis. The study was repeated 1 yr later, carried for longer period of time, and the red cell survival was also performed by more conventional method based on determining blood radioactivity. The repeated study showed similar results (see Fig. 4). Results of identical experiments carried out in two normal controls is shown in Fig. 5. No change of disappearance slopes were observed after challenge with 4 g of ascorbic acid.

DISCUSSION

G-6-PD-Birmingham is a G-6-PD mutant, characterized by normal hemoglobin concentration in the

Table 3. Some Parameters of Glutathione Metabolism of Erythrocytes and Leukocytes in N.S. and Normal Control

<table>
<thead>
<tr>
<th></th>
<th>Erythrocyte GSH (in μmole GSH/g Hb)</th>
<th>Erythrocyte GSSG (in μmole GSSH/g Hb)</th>
<th>Erythrocyte GR (in IU/g Hb)</th>
<th>Lymphocyte GR (in IU/g Hb)</th>
<th>Granulocyte GR (in IU/g Hb)</th>
</tr>
</thead>
<tbody>
<tr>
<td></td>
<td>Without FAD</td>
<td>With FAD</td>
<td>Without FAD</td>
<td>With FAD</td>
<td>Without FAD</td>
</tr>
<tr>
<td>N.S.</td>
<td>7.63</td>
<td>0.0717</td>
<td>7.53</td>
<td>12.1</td>
<td>152.08</td>
</tr>
<tr>
<td>Normal control</td>
<td>7.64</td>
<td>0.0122</td>
<td>6.37</td>
<td>9.61</td>
<td>66.99</td>
</tr>
<tr>
<td>Normal range</td>
<td>6.57 ± 1.04</td>
<td>0.0123 ± 0.0045</td>
<td>7.18 ± 1.09</td>
<td>10.4 ± 1.5</td>
<td>Not determined</td>
</tr>
</tbody>
</table>

Studies of Nonerythroid Cells of G-6-PD Deficient Boys and Heterozygous Mother

Fibroblasts, derived from skin biopsy, were examined by light microscopy. The fibroblasts of the propositus, N.S., were typical of Chediak-Higashi disease. Giant granules were present and melanin absent.13 The fibroblasts of the elder brother, A.S., were normal. Table 2 depicts the activity of G-6-PD in the granulocytes, lymphocytes, platelets, and fibroblasts of both A.S. and N.S. The data reveal that in these cells, the decrease of G-6-PD activity is only mild when compared with erythrocyte G-6-PD activity. Electrophoresis was performed on cell lysates of purified granulocytes, lymphocytes, and platelets of the mother of these G-6-PD-deficient boys (obligate
distinct from G-6-PD-Campbellpore, which has been deceptive. The possible additive effect of this particular variant demonstrates that it has not been previously described. Specifically, it appears to be somewhat analogous to the Birmingham/Chediak Higashi syndrome. The enzymatic characteristics of this variant demonstrate that it has not been previously described. Specifically, it appears to be distinct from G-6-PD-Campbellpore, which has been reported to be very prevalent among the Pakistanis.

A significant G-6-PD deficiency in leukocytes would result in a defective pentose shunt; this would impair the generation of NADPH, which is necessary not only to protect the host against bacteria but also to protect phagocytic cells from the oxidative radicals produced during the process of disposal of infectious agents. This defect could be expected to impair further the antibacterial defense already reduced by Chediak-Higashi disease. G-6-PD deficiency is generally not complicated by infections, and this metabolic defect is detrimental only to the red cells; however, increased susceptibility to infection has been described in a patient with severe leukocytic deficiency of G-6-PD. Our data would indicate that nonerythroid cells in two studied subjects having the 0-6-PD deficiency are not complicated by infections, and this metabolic defect is likely not to affect nonerythroid cells. However, the enzymatic activity detected in vitro does not necessarily correlate with in vivo situations. Complicated interplay with unknown and known activators and inhibitors in the living cell could make such a comparison deceptive. The possible additive effect of this particular G-6-PD deficiency on Chediak-Higashi neutrophils is further suggested by the analysis of the family pedigree, which reveals that the male rather than the female members of this family are afflicted by frequent infections. However, two aborted female fetuses might have developed Chediak-Higashi disease. The in vivo influence of this G-6-PD mutant on the neutrophil lifespan could perhaps best be tested by examining the proportions of normal and G-6-PD-Birmingham neutrophils in a heterozygous female. Thus, the neutrophils with the mutant enzyme might be at a disadvantage in comparison with neutrophils that exhibited normal G-6-PD activity, and the latter would then be expected to predominate. In a somewhat analogous situation, suppression of leukocytes carrying a sex-linked defect of purine metabolism (Lesch-Nyhan syndrome) has indeed been described in a female who was also heterozygous for G-6-PD isoenzymes A and B, as well as for the Lesch-Nyhan syndrome. In this instance, both the erythrocytes and leukocytes displayed a single G-6-PD isoenzyme and normal hypoxanthine guanine phosphoribosyltransferase activity. Presumably, those cells expressing the opposite G-6-PD isoenzyme, as well as Lesch-Nyhan defect, were at a survival disadvantage. The mother of our two deficient boys exhibited a similar proportion of both G-6-PD isoenzymes (G-6-PD-B and G-6-PD-Birmingham) in her neutrophils, which suggested that the neutrophils that expressed the deficient mutant enzyme and the normal neutrophils would have relatively equal chances of survival.

Ascorbic acid has been reported to ameliorate the clinical course of patients with Chediak-Higashi disease. On the other hand, ascorbic acid has been reported to lead to a decrease of erythrocyte GSH.
level in G-6-PD-deficient erythrocytes, to decrease the red cell survival of in vitro treated human G-6-PD-deficient erythrocytes in an animal model, to produce mild hemolysis in G-6-PD deficient subjects, and to lead to severe intravascular hemolysis after intravenous administration in massive doses of 80 g for 2 consecutive days. However, Winterbourn proposed that ascorbate might aid in alleviating G-6-PD deficiency. Challenge with doses of ascorbic acid considered to be therapeutic caused no measureable decrease in life span of red cells in this particular G-6-PD variant. In point of fact, the red cell survival appeared to improve following an ascorbic acid challenge based on the urinary chromium excretion method; however, we did not observe any significant change of red cell survival when measured by more conventional method based on following of blood radioactivity, possibly due to greater scatter of the points. Moreover, the apparent change of the slope of urinary radioactivity may have been caused by more than one red cell population present in the patients with red cell metabolic defect. This possibility is further supported by the fact that the putative beneficial influence of a single dose of ascorbic acid would be expected to be transient following a single dose of ascorbic acid, however, as shown in Fig. 4, there was no change of the slope for 120 hr after ascorbic acid administration. We have not had an opportunity to repeat the study again on this patient without ascorbic acid administration to clarify the effect of the ascorbic acid on this patient's red cell survival. Unfortunately, after receiving 200 mg of ascorbic acid twice daily for 8 mo, the patient has not displayed any obvious clinical benefits with respect to infections similar to experience of others.

We conclude that the frequency and severity of infection in our patient with Chediak-Higashi syndrome cannot be clearly attributed to the possible additive interaction from the concomitant presence of the newly described G-6-PD-deficient variant.

REFERENCES

A new glucose-6-phosphate dehydrogenase deficient variant in a patient with Chediak-Higashi syndrome

JT Prchal, WM Crist, A Malluh, A Vitek, WN Tauxe and AJ Carroll