Effect of Indole Compounds on Vitamin B\textsubscript{12} Utilization

By Jerry Drexler

A definite relationship has never been established between indole metabolism and the megaloblastic anemias. Rhoads and associates\textsuperscript{1,2} were able to produce a hemolytic-like anemia with many features of pernicious anemia, in dogs on vitamin B deficient diets, by the oral administration of pure indole. Interestingly, the relief of this anemia by oral feeding of liver extract was accompanied by a reticulocytosis.\textsuperscript{2} In dogs on well balanced diets, the administration of a similar amount of pure indole caused a well defined increase in bilirubin excretion, and massive doses of indole caused a severe anemia. Tönnis and Horster, cited by Rhoads and Miller,\textsuperscript{2} reported the occurrence of indicanuria and anemia in dogs with surgically formed, inactive, open jejunal segments, and the relief of the anemia after the administration of liver extract. These investigations suggested the possibility that indole compounds might produce anemia by affecting the utilization of some hematopoietic factor.

The purpose of the present study is to describe the effect of indole compounds on the utilization of vitamin B\textsubscript{12} by a vitamin-dependent microorganism.

Materials and Methods

The indole compounds were selected from the serotonin (5-hydroxytryptamine) pathway of tryptophan metabolism, since these compounds may be present in the gastrointestinal tract where vitamin B\textsubscript{12} normally is absorbed. A mutant strain of Escherichia coli, which is dependent upon vitamin B\textsubscript{12} for growth, was used in the microbiologic assays. The inherent dangers of such an organism for assay of impure vitamin B\textsubscript{12} preparations have been stressed,\textsuperscript{3} but for assay of purified preparations, the E. coli tube method, as modified by Glick,\textsuperscript{4} is reliable and easily and rapidly performed.

The assay medium deficient in vitamin B\textsubscript{12} was prepared according to the method of Burkholder.\textsuperscript{4} One ml. of this double strength medium was used in each tube of the eight series of assays. The volume of the contents of each tube was increased to 2 ml. by the addition of distilled water (series 1 and 2), vitamin B\textsubscript{12} and distilled water (series 3), or vitamin B\textsubscript{12} and indole compound (series 4 through 8) (table 1). The distilled water and vitamin B\textsubscript{12} were added to the assay medium in cotton-plugged test tubes which were then autoclaved at 115 C. for five minutes. The standard vitamin B\textsubscript{12} solution contained 0.5 millimicrogram B\textsubscript{12} per milliliter. The indoles were prepared in a concentration of 1 mg. per milliliter.

All experiments were performed in triplicate. Except for series 1, each tube was inoculated with one drop of a freshly prepared saline suspension of washed E. coli,

From the Department of Clinical Pathology, The Cleveland Clinic Foundation, and The Frank E. Bunts Educational Institute, Cleveland, Ohio.

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TABLE 1.—Ingredients of Each Tube in Eight Series of Assay Tubes

<table>
<thead>
<tr>
<th>Series no.</th>
<th>Media (ml.)</th>
<th>5-Hydroxytryptophan (mg.)</th>
<th>5-Hydroxytryptamine* (mg.)</th>
<th>5-Hydroxyindoleacetic acid (mg.)</th>
<th>Indole (ml.)</th>
<th>Water (ml.)</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>1.0</td>
</tr>
<tr>
<td>2</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.5</td>
<td>0.5</td>
</tr>
<tr>
<td>3</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.5</td>
</tr>
<tr>
<td>4</td>
<td>1.0</td>
<td>0.5</td>
<td>0.5</td>
<td>-</td>
<td>-</td>
<td>0.5</td>
</tr>
<tr>
<td>5</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.5</td>
</tr>
<tr>
<td>6</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>0.5</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>7</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>0.5</td>
<td>-</td>
<td>-</td>
</tr>
<tr>
<td>8</td>
<td>1.0</td>
<td>-</td>
<td>-</td>
<td>-</td>
<td>0.5</td>
<td>-</td>
</tr>
</tbody>
</table>

*5-hydroxytryptamine supplied by Abbott Laboratories.
†5-hydroxyindoleacetic acid supplied by Dr. M. Speeter, Research Laboratories of the Upjohn Company, Kalamazoo, Michigan.

strain 113-3 (A.T.C.C. 11505).* The tubes were shaken for 18 hours; the ingredients were then diluted to 5 ml. with distilled water, and the final turbidity, indicating the growth of the microorganism, was measured by a Klett immersion colorimeter.

Two of the tested indole compounds were found to inhibit the growth of the microorganism. Consequently, experiments using procedures similar to those described were performed in order to elucidate the mechanism of this inhibition. The results of these latter experiments are reported later.

RESULTS

The results (table 2) of these experiments demonstrate the ability of indoleacetic acid and pure indole to inhibit the growth of the vitamin B₁₂ dependent microorganism.

The standard growth curve for E. coli, strain 113-3, was depressed by the addition of indole (fig. 1). The inhibitory effect on the growth of the microorganism, which resulted from increasing the concentration of indoleacetic acid, is shown in figure 2. Although concentrations of the indole compound may appear high, a definite inhibitory effect can be seen after the addition of only 50 micrograms of indoleacetic acid.

Since an excess of vitamin B₁₂ was found to overcome the inhibitory action of the indole (fig. 3) and since several non-B₁₂ dependent E. coli organisms were not affected by a similar amount (0.5 mg.) of indole or indoleacetic acid, the indole compounds appeared to act by preventing utilization of vitamin B₁₂. As measured by radioactive B₁₂ studies,† the uptake of vitamin B₁₂ from the medium was actually only slightly impaired by indoleacetic acid. The addition of excess tryptophan, or the removal of tryptophan from the Burkholder medium, did not affect the inhibition. Cobalt, copper, and iron

*Supplied by Dr. E. L. R. Stokstad, American Cyanamid Co., Research Division, Lederle Laboratories Pearl River, N. Y.
†The radioactive material used in this study was supplied by Abbott Laboratories on authorization of the Isotopes Division, U. S. Atomic Energy Commission, Oak Ridge, Tennessee.
### EFFECT OF INDOLE COMPOUNDS ON VITAMIN B₁₂ UTILIZATION

#### Table 2.—Turbidity Readings* in Eight Experiments, Each Performed in Triplicate

<table>
<thead>
<tr>
<th>Series no.</th>
<th>Assay solution</th>
<th>Turbidity readings</th>
<th>Average of turbidity readings</th>
</tr>
</thead>
<tbody>
<tr>
<td>1</td>
<td>Uninoculated media</td>
<td>1 0 0</td>
<td>0.3</td>
</tr>
<tr>
<td>2</td>
<td>Inoculated media</td>
<td>6 4 4</td>
<td>4.7</td>
</tr>
<tr>
<td>3</td>
<td>B₁₂ control</td>
<td>106 96 96</td>
<td>99.3</td>
</tr>
<tr>
<td>4</td>
<td>B₁₂ plus 5-hydroxytryptophan</td>
<td>97 91 105</td>
<td>97.7</td>
</tr>
<tr>
<td>5</td>
<td>B₁₂ plus 5-hydroxytryptamine</td>
<td>112 94 84</td>
<td>96.7</td>
</tr>
<tr>
<td>6</td>
<td>B₁₂ plus 5-hydroxyindoleacetic acid</td>
<td>107 105 94</td>
<td>102.0</td>
</tr>
<tr>
<td>7</td>
<td>B₁₂ plus indoleacetic acid</td>
<td>56 34 47</td>
<td>45.7</td>
</tr>
<tr>
<td>8</td>
<td>B₁₂ plus pure indole</td>
<td>23 27 25</td>
<td>25.0</td>
</tr>
</tbody>
</table>

* Measured by Klett immersion colorimeter.

#### Diagram

**Figure 1.**—Effect of pure indole on standard growth-curve of E. Coli, 113–3.
Fig. 2.—Effect of indoleacetic acid on maximum growth of E. Coli, 113-3.

Fig. 3.—Effect of excess vitamin B$_{12}$ on indoleacetic acid inhibition.
also did not check the inhibitory effect. Autoclaving the indole compounds with the vitamin B₁₂, which usually frees vitamin B₁₂ from binding complexes, or adding potassium cyanide which breaks peptide linkages with B₁₂, did not alter the inhibition. Incidentally, hydroxylamine, a known decarboxylase inhibitor, caused complete inhibition of the vitamin B₁₂ dependent microorganism, but did not affect the growth of a non-B₁₂ dependent organism.

Only a fresh preparation of indole was effective, whereas indoleacetic acid maintained its inhibitory powers for weeks. The failure of the 5-hydroxyindoleacetic acid to inhibit the growth of *E. coli* in the present experiment may be due in part to the lability of the indole compound to oxidation. On the other hand, two 5-benzyl serotonin-like compounds* in a concentration of 1 mg. per millimeter also inhibited the growth of the *E. coli* mutant by 90 per cent, but tryptamine hydrochloride failed to do so. Skatole in a similar concentration produced complete inhibition of growth. Concomitant studies revealed no significant change in pH to account for any of the results.

**Discussion**

The present study confirms the ability of several indole compounds to prevent the utilization of vitamin B₁₂ by the test microorganism. That this is a specific effect on the vitamin B₁₂ mechanism and not a general effect on microorganisms is established by the reversal of the inhibition with added B₁₂, and the inability of the indole compounds to affect the growth of several other *E. coli* microorganisms. Whether the indole compounds act by binding the vitamin, by blocking some enzymatic system, or by some other mechanism, is not known. Unsaturated indoles such as indole acrylic acid have been reported to inhibit the growth of *E. coli*, but this process is reversed by tryptophan.* Since tryptophan failed to affect the inhibition in the present study, a similar mechanism does not appear to be involved. Also, radioactive vitamin B₁₂ dialysis studies, electrophoretic and chromatographic procedures failed to demonstrate the formation of a vitamin B₁₂ indole complex. Since the inhibition of B₁₂ utilization was reversed by excess B₁₂, the indole compounds would appear to act by a competitive inhibition of some enzymatic system necessary for the utilization of vitamin B₁₂. Saxena and Agarwala have shown that phenylhydrazine and heavy metals like molybdenum and tungsten inhibit the *E. coli* mutant, and vitamin B₁₂ reverses this inhibition. The particular enzymatic system affected is unknown. Perhaps a similar mechanism is involved in the present experiments.

Watson and Witts* attribute the macrocytic anemia associated with intestinal strictures and anastomoses to a change in the bacterial flora of the small intestine which interferes with the formation or utilization of hematopoietic material. Coliform microorganisms are known to produce indole; in the present experiment, indoles have been shown to inhibit the utilization of vitamin B₁₂, an essential hematopoietic material. Thus, an indole product of gastrointestinal microorganisms could conceivably be the toxic factor in the mega-

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*5-benzyl-3-β-N-benzyl-N-methylaminoethyl indole hydrochloride and 5-benzyl-3-β-dimethylaminoethyl-indole hydrochloride.
loblastic anemia of intestinal blind loops. Antibiotics prevent the formation of this factor and the development of anemia. Perhaps a similar compound in the human intestine, eliminated by sterilization of the gastrointestinal flora, might be the cause of the slight hematologic response to orally administered aureomycin noted by Lichtman, Ginsberg, and Watson in several patients with pernicious anemia. Similarly, the anemia produced by Rhoads and associates, with indole feeding in dogs, is likely due to interference with vitamin $B_{12}$ utilization. The role of tryptophan and indole metabolism in normal absorption and utilization of vitamin $B_{12}$ needs further elucidation.

**SUMMARY**

Indole and indoleacetic acid are shown to inhibit the growth of a vitamin $B_{12}$ dependent microorganism, a mutant strain of *Escherichia coli*. It is suggested that the mechanism of that inhibition may be a competitive inhibition of some enzymatic system necessary for the utilization of vitamin $B_{12}$.

**SUMMARIO IN INTERLINGUA**

Es monstrate que indol e acido indolo-acetic inhibi le crescentia de un microorganismo, un racia mutante de *Escherichia coli* que depende de vitamina $B_{12}$. Es formulate le these que le mechanismo de ille inhibition es possibilemente indirecte e consiste in le inhibition de un sistema enzymatic que es indispensabile in le utilisation de vitamina $B_{12}$.

**REFERENCES**

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JERRY DREXLER

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