To the editor:

**Cytopenia levels for aiding establishment of the diagnosis of myelodysplastic syndromes**


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The recent article by Arber et al1 detailing the 2016 revision of the World Health Organization (WHO) classification of myeloid malignancies and AML was timely and germane. Regarding myelodysplastic syndromes (MDS), the authors indicate diagnostic criteria that include levels of dysplasia and cytopenias. They further indicated that ethnic variation should be taken into consideration in patients with borderline levels of dysplasia and cytopenias. They also indicated that ethnic variation may be a sine qua non for any MDS diagnosis,1,2 and some patients with ICUS may eventuate into nonhematopoietic/reactive disorders such as immune dysregulation, whereas IDUS is a morphological alteration with many potential causes that do not necessarily influence hematopoiesis in terms of the number of generated cells. These entities have been reviewed in the current National Comprehensive Cancer Network MDS Practice Guidelines 1.2017.10

However, although the WHO perspective indicates that “cytopenia is a sine qua non for any MDS diagnosis,”11 the recommended threshold levels of cytopenias it proposes for this purpose are those previously reported in the International Prognostic Scoring System (IPSS) risk stratification categorization that were used for prognostic but not diagnostic purposes (hemoglobin [Hb] 10 g/dL, absolute neutrophil count [ANC] 1.8 × 10^9/L, platelets 100 × 10^9/L).11 Table 1 provides data from the International Working Group for Prognosis in MDS (IWG-PM) database that was used to generate the Revised-IPSS12; if these cytopenia levels were used to diagnose MDS, 18% of MDS patients and 23% of those with <5% marrow blasts would lack any

### Table 1. Cytopenias in MDS

<table>
<thead>
<tr>
<th>Marrow blasts</th>
<th>None, n</th>
<th>None, %</th>
<th>1, n</th>
<th>1, %</th>
<th>2, n</th>
<th>2, %</th>
<th>3, n</th>
<th>3, %</th>
<th>Total</th>
</tr>
</thead>
<tbody>
<tr>
<td><strong>Less than normal</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;5%</td>
<td>106</td>
<td>2.3</td>
<td>1946</td>
<td>43</td>
<td>1543</td>
<td>34</td>
<td>950</td>
<td>21</td>
<td>4545</td>
</tr>
<tr>
<td>≥5%</td>
<td>19</td>
<td>0.8</td>
<td>421</td>
<td>17</td>
<td>927</td>
<td>38</td>
<td>1100</td>
<td>45</td>
<td>2467</td>
</tr>
<tr>
<td>Total</td>
<td>125</td>
<td>1.8</td>
<td>2367</td>
<td>34</td>
<td>2470</td>
<td>35</td>
<td>2050</td>
<td>29</td>
<td>7012</td>
</tr>
<tr>
<td><strong>Less than normal, without CMML</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;5%</td>
<td>73</td>
<td>1.7</td>
<td>1814</td>
<td>43</td>
<td>1395</td>
<td>33</td>
<td>912</td>
<td>22</td>
<td>4194</td>
</tr>
<tr>
<td>≥5%</td>
<td>8</td>
<td>0.4</td>
<td>318</td>
<td>15</td>
<td>792</td>
<td>37</td>
<td>1047</td>
<td>48</td>
<td>2165</td>
</tr>
<tr>
<td>Total</td>
<td>81</td>
<td>1.3</td>
<td>2132</td>
<td>34</td>
<td>2187</td>
<td>34</td>
<td>1959</td>
<td>31</td>
<td>6359</td>
</tr>
<tr>
<td><strong>WHO categorization</strong></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>&lt;5%</td>
<td>1040</td>
<td>23</td>
<td>1988</td>
<td>44</td>
<td>1064</td>
<td>23</td>
<td>453</td>
<td>10</td>
<td>4545</td>
</tr>
<tr>
<td>≥5%</td>
<td>197</td>
<td>8</td>
<td>776</td>
<td>32</td>
<td>922</td>
<td>37</td>
<td>572</td>
<td>23</td>
<td>2467</td>
</tr>
<tr>
<td>Total</td>
<td>1237</td>
<td>18</td>
<td>2764</td>
<td>39</td>
<td>1896</td>
<td>28</td>
<td>1025</td>
<td>15</td>
<td>7012</td>
</tr>
</tbody>
</table>

Percent values rounded off except for values <3%. Data obtained from Greenberg et al.12
CMML, chronic myelomonocytic leukemia.

1Standard values: Hb <13 g/dL (males), <12 g/dL (females), ANC <1.8 × 10^9/L, platelets <150 × 10^9/L.
2IPSS values: Hb <10 g/dL, ANC <1.8 × 10^9/L, platelets <100 × 10^9/L.
cytopenia and thus would not be classifiable as MDS. Using standard laboratory values for cytopenias (Hb <13 g/dL [males], <12 g/dL [females], ANC <1.8 × 10^9/L, platelets <150 × 10^9/L), the data demonstrated that only 1.8% of patients evaluated in that study of 7012 MDS subjects would lack a cytopenia (1.3% of patients when nonproliferative chronic myelomonocytic leukemia patients were excluded). Of note, and relevant predominantly for patients with low marrow blast counts in the IWG-PM cohort, the patient’s blood counts also needed to demonstrate ≥2 months of stable disease as a potential means of excluding other causes for the cytopenias.

Regarding our main point, it is of relevance that the MDS database (n = 816) used to generate the IPSS (Table 1) similarly demonstrated that 19% of these patients lacked a cytopenia if defined by the prognostic level cutpoints used by the WHO and also incorrectly would not have been considered to have MDS. Similar findings were found in an independent study using these cytopenic cutpoints. Prior investigations have demonstrated ethnic-, age-, and altitude-related differences in normal Hb levels; ethnic-, age-, and sex-related differences in platelet levels; and ethnic- and sex-related differences in platelet and white counts. Thus, being cognizant of these conditional blood count variations, we recommend that standard hematologic values be used to define cytopenias in MDS and believe a modification of the WHO definition of cytopenias as 1 of the criteria (in addition to definitive morphologic and/or cytogenetic findings) to diagnose MDS would be valuable and most accurate.

**Contribution:** P.L.G. and H.T. designed the study, analyzed the data, and wrote the report; A.A.v.d.L., D.H., J.M.B., P.F., M.C., and U.G. critically reviewed and modified the manuscript and contributed data to the IWG-PM database; and the remaining authors contributed data to the IWG-PM database and critically reviewed and approved the manuscript.

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**References**


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**To the editor:**

**Chronic neutrophilic leukemia in a child with a CSF3R T618I germ line mutation**

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Chronic neutrophilic leukemia (CNL) is a rare myeloproliferative neoplasm characterized by sustained elevation of neutrophil count, splenomegaly, and poor prognosis. Activating mutations in the colony-stimulating factor 3 receptor (CSF3R), also known as the granulocyte colony-stimulating factor (G-CSF) receptor, have recently been identified in 80% of patients studied with CNL. The most common mutation is T618I, which renders the receptor ligand independent through constitutive JAK/STAT activation.1,3 The strong association of activating CSF3R mutations with CNL has led to the addition of a CSF3R T618I mutation or other activating CSF3R mutation to the diagnostic criteria for CNL in the 2016 revision to the World Health Organization (WHO) classification of myeloid neoplasms and acute leukemia.4

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