complementing the known mutational inactivation of S1PR2 in GC-DLBCL with a new mechanism of S1PR2 silencing involving transcriptional suppression by FOXP1 in ABC-DLBCL and providing direct evidence of its role as tumor suppressor in this neoplasm. Additionally, the results, which link FOXP1 to changes in the levels of S1PR2 that occur during B-cell maturation in GCs, further our understanding of the regulation of S1PR2 expression, of which very little is known, not only in DLBCL B cells, but also in normal B cells. The authors show indeed that low FOXP1 correlates with high S1PR2 levels in centrocytes and centroblasts, which rely on this receptor to return to the follicle center during affinity maturation and class switch, whereas S1PR2 is silenced in the FOXP1-high naive and memory B cells.

The results presented in this report have implications both for the treatment and for the molecular classification of ABC-DLBCL (see figure). Although the pharmacologic targeting of Akt has been proposed as a strategy to limit tumor growth in GCB-DLBCL,4 the finding by Flori et al1 that S1PR2-mediated apoptosis is Akt-dependent in DLBCL cells suggests that this approach might not be suitable for ABC-DLBCL and underscores the importance of elucidating the mechanisms linking S1PR2 deficiency to cell survival in these cells. At variance, genetic or RNAi-mediated depletion of Gα13 or p115Rho/ARHGEF1 recapitulates the effects of S1PR2 deficiency, enhancing B-cell survival even in the FoxP1-high S1PR2-low DLBCL cells which are not sensitive to Akt inhibition,1,3 suggesting that Rho mimetics may represent an attractive therapeutic approach for both GCB-DLBCL and ABC-DLBCL. FOXP1 and its regulators also emerge as interesting alternative targets, as FOXP1 has been implicated in the transcriptional silencing not only of S1PR2 but also of several genes that contribute to cell survival and immune surveillance in ABC-DLBCL.5,6 It is noteworthy that the prosurvival effects of FOXP1 overexpression in ABC-DLBCL rely on the nuclear factor-κB pathway,7 which is constitutively active in this disease presentation10 and for which pharmacologic inhibitors such as bortezomib are available. It will be interesting to characterize the effects of these inhibitors on the genes regulated by FOXP1, including S1PR2. Finally, the robust prognostic value of the combination of low FOXP1 with high S1PR2 expression as a positive predictor of survival in DLBCL patients under CHOP (cyclophosphamide, hydroxydaunorubicin, vincristine, and prednisone) or rituximab-CHOP therapy1 may provide a new predictive biomarker for treatment stratification.

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Comment on Shen et al, page 1449

Release the hounds: virotherapy with immunotherapy

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In this issue of Blood, Shen et al demonstrate that the vesicular stomatitis virus (VSV)–murine interferon B (IFNb)–sodium iodide symporter (NIS) (VSV-mIFNb-NIS) oncolytic virus has significant antileukemia activity, which is enhanced when combined with an anti–programmed death-1 (PD-L1) antibody.1

Approximately 30% to 40% of patients with acute myeloid leukemia (AML) may be refractory to initial therapy, and a majority of patients who achieve a first complete remission will experience relapse.2 Relapsed and refractory AML is characterized by chemotherapy-resistant disease for which newer modalities of therapy are critically needed. Targeted small-molecule inhibitors, monoclonal antibodies, and cellular therapies are currently in active development. Oncolytic virotherapy is an emerging therapeutic modality that uses live, replicating viruses to target, infect, and kill cancer cells in vivo. This strategy of a systemically administered, self-replicating vehicle of treatment in “one shot” is appealing in a systemic disease like AML, where the dividing malignant cells are disseminated both intravascularly and in diverse tissues. The ability of oncolytic viruses to repeatedly infect cancer cells also carries the potential for eradicating minimal residual disease.

Oncolytic viruses are engineered to be nonpathogenic to normal cells by removing virulence factors, while still maintaining their tropism to cancer cells.3 Defects in innate immune signaling within tumors may actually provide an ideal setting for viral infection and replication. Cytotoxicity using oncolytic
viruses is thought to occur via several mechanisms, including: (1) direct lysis and cell death after synthesis of new viral particles, and (2) stimulation of the host cytotoxic immune response with direct cell priming and epitope spread after the local cellular lysis. Several phase 1/2 clinical trials using oncolytic viruses have been conducted in patients with solid and hematopoietic malignancies, validating this as a viable approach. Recently, a phase 3 trial demonstrated improved responses and prolonged overall survival in patients with advanced melanoma treated with an oncolytic, modified herpes simplex virus, talimogene laherparepvec. The current study uses an engineered strain of VSV, a single-stranded RNA virus that is minimally pathogenic in humans and has demonstrated oncolytic activity. The VSV-mIFNβ-NIS construct is engineered to include a gene expressing IFNβ that enhances tumor cell selectivity, as well as the NIS transgene, which encodes a sodium-iodide symporter and allows noninvasive in vivo imaging. The local expression of IFNβ can also promote local innate cellular immune responses, potentiating antitumor activity. This strain of VSV has been previously used by the authors, demonstrating potent cytotoxicity in a myeloma model.

The host immune response plays an important role in the surveillance and eradication of cancer. The propagation and spread of malignant cells is often marked by a permissive, immunosuppressive tumor microenvironment that impedes host immune response. Recently, some of these mechanisms of immune evasion have been elucidated. Upregulation of PD-L1 on tumor or associated stromal cells can interact with the programmed death-1 (PD-1) receptor on infiltrating cytotoxic T cells, abrogate their antitumor response, and “keep them at bay.” Disrupting this immune checkpoint interaction could reverse tumor-mediated immunosuppression and augment antitumor T-cell responses. Monoclonal antibodies to PD-1 and PD-L1 have been developed to block this interaction and have demonstrated significant, durable clinical responses in patients with solid and hematopoietic malignancies. The current study aims to exploit the activity of immune checkpoint inhibitors to potentiate the local host immune response after viral infection and increase antileukemia activity. Viral-mediated lysis with epitope spread could create an “antigen-rich” environment in a disease like AML that is not known to have a large number of neoantigens. This, coupled with an activated local immune response, could be a compelling approach.

Using syngeneic, immune-competent mouse models, Shen and colleagues demonstrate selective virus infectivity of tumor cells after systemic administration, dose-dependent antitumor activity, and enhancement of antileukemia activity with the addition of an anti–PD-L1 antibody. In
a model of disseminated AML, the virus was effective at reducing AML burden from blood, bone marrow, liver, and spleen after systemic administration, particularly when combined with the anti–PD-L1 antibody. This translated into a significant survival benefit for virus-treated mice compared with controls—an effect that was again enhanced by the addition of the anti–PD-L1 antibody. Depletion of mouse natural killer cells and CD8+ T cells, but not CD4+ T cells, negated the survival benefit, suggesting that these immune cells were important for the antileukemia response. When the mice tissues were examined at necropsy, the authors were able to isolate both VSV-specific and leukemia-specific cytotoxic T cells, with the highest levels found in the combination-treated cohort. Together, these observations provide evidence that the viral infection alone and in combination with the anti–PD-L1 antibody is functioning also as an immunotherapeutic approach. Finally, the authors performed ex vivo analysis of primary patient samples and demonstrated high infectivity with VSV among patients with chronic myelomonocytic leukemia and monocytic AML.

This preclinical study outlines a very novel approach to AML treatment, combining an oncolytic virus with immune checkpoint inhibition, revealing virotherapy as an immunotherapy (see figure). Such a strategy could expand the spectrum of malignancies that may be amenable to immunotherapy outside of those that are most immunogenic. These results need to be confirmed in future studies, but several challenges remain. Systemic virus administration in patients with AML would require achievement of large viral titers and overcoming the host serum and immune factors that could neutralize the virus. Clearance and sequestration by the liver and spleen, although not observed in the mouse model, are important obstacles with systemic administration in humans.5 As evidenced in the ex vivo patient samples, the virus robustly infected only a subset of cases. Therefore, choosing the right virus and appropriately targeting it to the malignant leukemia cells will remain an ongoing challenge. Because the immune response appears necessary in the antileukemia activity, how this approach fares in previously treated patients who have received chemotherapy and may have a depleted immune repertoire remains to be seen. These and other questions will need to be addressed, but the findings here provide an interesting way to refocus an antileukemia immune response.

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Comment on Huang et al, page 1459

Arf6 arbitrates fibrinogen endocytosis

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In this issue ofBlood, in a departure from studies of classic platelet function, Huang et al turn their attention to endocytosis and show that adenosine 5’-diphosphate–ribosylation factor 6 (Arf6) plays a key role in fibrinogen engulfment.1 Although platelets are known to bind, absorb, and load their granules with plasma proteins, this report is one of the first to explore mechanisms that control endocytosis in this anucleate cell. Huang et al demonstrate that Arf6-dependent endocytosis is restricted to fibrinogen, implying that Arf6 also modulates trafficking of αIIbβ3 integrins in platelets. Consistent with this notion, deletion of Arf6 in platelets enhances spreading on fibrinogen and accelerates clot retraction (see figure). However, activation of surface αIIbβ3 is unaffected, and Arf6 deficiency does not alter thrombosis in vivo. These incongruous results point toward the complexity of anucleate platelets and the need for more detailed studies to understand intracellular trafficking, recycling, and endocytosis in platelets and their precursors.

Arf6 is a small GTPase that localizes to membranes and endosomal compartments.2 In nucleated cells, Arf6 regulates endocytic membrane trafficking and thereby impacts cell motility, cell division, and lipid homeostasis. Arf6 has also been linked to actin remodeling, which may explain why genetic disruption and pharmacologic inhibition of Arf6 in mouse and human platelets, respectively, affects platelet spreading. Like other small GTPases, Arf6 cycles between an active GTP-bound and inactive GDP-bound conformation. When cycling between these 2 states, Arf6 facilitates ligand internalization at cell surfaces, endosomal recycling, and fusion of endosomal membranes with plasma membranes. Previous studies from Whiteheart’s group3,4 showed that Arf6 rapidly converts from a GTP- to GDP-bound state in activated human platelets and that
Release the hounds: virotherapy with immunotherapy

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