PLATELETS AND THROMBOPOIESIS

A novel and essential role for FcγRIIa in cancer cell–induced platelet activation

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Key Points

- The immune receptor FcγRIIa is a key mediator of tumor cell activation of platelets in the circulation.
- Secretion of adenosine 5’-diphosphate from dense granules is the primary response of platelets to activation by tumor cells.

Platelets play a role in cancer by acting as a dynamic reservoir of effectors that facilitate tumor vascularization, growth, and metastasis. However, little information is available about the mechanism of tumor cell–induced platelet secretion (TCIPS) or the molecular machinery by which effector molecules are released from platelets. Here we demonstrate that tumor cells directly induce platelet secretion. Preincubation of platelets with human colon cancer (Caco-2), prostate cancer (PC3M-luc), or breast cancer cells (MDA-MB-231;MCF-7) resulted in a marked dose-dependent secretion of dense granules. Importantly, TCIPS preceded aggregation which always displayed a characteristic lag time. We investigated the role of platelet receptors and downstream molecules in TCIPS. The most potent modulators of TCIPS were the pharmacologic antagonists of Syk kinase, phospholipase C and protein kinase C, all downstream mediators of the immunoreceptor tyrosine-based activation motif (ITAM) cascade in platelets. Supporting this, we demonstrated a central role for the immune Fcγ receptor IIa (FcγRIIa) in mediating platelet-tumor cell cross-talk. In conclusion, we demonstrate that cancer cells can promote platelet dense-granule secretion, which is required to augment platelet aggregation. In addition, we show a novel essential role for FcγRIIa in prostate cancer cell–induced platelet activation opening the opportunity to develop novel antimetastatic therapies. (Blood. 2014;123(2):249-260)

Introduction

There is increasing evidence supporting an association between cancer growth, metastasis, and platelet secretion. An association between thrombocytosis and cancer death was first reported in 1872 by Reiss and colleagues and brought to our attention again in 1964. Boneu et al, in 1984, first reported the presence of degranulated platelets, characterized by low levels of adenosine 5’-diphosphate (ADP) and serotonin, in patients with malignant solid tumors, while others reported an inverse relationship between cancer prognosis and platelet count in patients. Later, Karpatkin and Pearlstein demonstrated that many tumor cells could directly activate platelets.

Platelet depletion in tumor-bearing mice triggers intratumor hemorrhage which is rescued by platelet secretome, demonstrating a requirement for platelet secretion to stabilize angio genesis in tumor metastases. Indeed, platelet α-granules contain large reservoirs of cytokines and growth factors, including transforming growth factor β (TGF-β) and vascular endothelial growth factor (VEGF), which are selectively released upon platelet activation. Consistent with a role of platelet secretion in tumor metastasis, TGF-β derived from platelet α-granules has been shown to profoundly impact tumor metastasis and survival by enhancing an epithelial mesenchymal-like transition and suppressing tumor cell natural killer–mediated lysis, respectively. Moreover, platelet secretion in a tumorigenic microenvironment seems to differ materially from wound-induced platelet secretion. Battinelli and colleagues demonstrated the capability of cancer cells to provoke the preferential release of proangiogenic mediators from platelet α-granules, creating a dynamic microenvironment favorable for their growth and survival. Although considerable progress has been made in understanding the role that platelet secretion plays in tumor metastasis, very little is known about the ability of cancer cells to induce platelet granule release. Moreover, the precise molecular mediators of this phenomenon remain unknown.

In the present study, we had 2 specific objectives. First, we aimed to investigate the ability of cancer cells to elicit platelet secretion using a pair of human prostate cancer cell lines: PC3 and its highly metastatic variant PC3M; a pair of breast cancer cell lines, MCF-7 (low metastatic) and MDA-MB-231 (high metastatic); and colorectal cancer cells Caco-2, whose ability to induce platelet activation in vitro has been well characterized. Second, we sought to elucidate the molecular pathway(s) that mediate platelet activation in response to metastatic prostate cancer cells, PC3M, using a pharmacologic approach. In a context of molecular divergence, we propose a novel role for platelet Fcγ receptor IIa (FcγRIIa) and downstream molecules as key elements in platelet activation induced by metastatic prostate cancer cells.
Material and methods

Reagents
All reagents were purchased from Sigma-Aldrich unless otherwise indicated. Thrombin receptor-activating peptide (TRAP) was obtained from Bachem. CHRONO-LUME reagent and the ristocetin cofactor assay were purchased from Chrono-log (Labmedics Limited). SQ29548 was obtained from Enzo Life Sciences. Aspirin, GF-109203X, AAOCCF3, LY294, PF228, PF2, piceatannol, U73122, and wortmannin were purchased from Tocris Bioscience. Abciximab (ReoPro) was provided from Centocor/Lilly. The monoclonal antibody (mAb) IV.3 Fab fragment against the FcγRIIa was kindly provided by Prof P. Newman (BloodCenter Wisconsin).

Tumor cell culture
Caco-2 cells, a human epithelial colorectal adenocarcinoma cell line, PC3M-luc and PC3, and metastatic and nonmetastatic human prostate cancer cells were a generous gift from Prof Marek W. Radomsky (Trinity College Dublin [TCD], Dublin, Ireland) and Prof Judy Harney (Royal College of Surgeons in Ireland [RCSI], Dublin, Ireland), respectively. MCF-7 and MDA-MB-231 breast cancer cells were kindly provided by Dr Ann Hopkins (Beaumont Hospital, Dublin, Ireland). Caco-2 cells were grown as previously described. PC3M-luc and MCF-7 and MDA-MB-231 breast cancer cells were kindly provided by Dr Ann Hopkins (Beaumont Hospital, Dublin, Ireland). Caco-2 cells were grown as previously described.15 PC3M-luc and MCF-7 were grown in modified Eagle medium (MEM) supplemented with 10% of fetal bovine serum (FBS). PC3 were grown in RPMI 1640 with 10% FBS. MDA-MB-231 were grown in Dulbecco MEM with 10% FBS. When confluent, cells were detached using Accutase (Sigma-Aldrich).

Preparation of human and mouse washed platelets
Human blood was drawn on the day of the experiment from healthy, drug-free volunteers into 1:9 (v/v) sodium citrate anticoagulant (3.15%) with informed consent, in accordance with the Declaration of Helsinki. Ethical approval was obtained from the RCSI ethics committee. Washed platelet (WP) suspension (300 × 10^3 platelets per μL) was prepared as previously described. Mouse WPs were drawn and prepared as previously described.

Platelet aggregation assay
Platelet aggregation was monitored at 37°C with continuous stirring in a PAP-4.8 aggregometer (Bio-Data Corporation). All aggregation experiments were measured against Tyrode buffer as the blank. Platelets (300 × 10^3 platelets per μL) were aggregated in the presence of cancer cells for 30 minutes or twice the lag time taken for the onset of aggregation response, whichever was longest. Where inhibitors were used, platelets were preincubated for 15 minutes at 37°C, 300 rpm, prior to commencement of aggregation. The data are expressed either as the percentage of aggregation or as the area under the aggregation curve (AUC). Data are normalized to the maximal aggregation achieved with each cell line used. Adhered platelets were labeled with a fluoresceinated (fluorescein isothiocyanate) antibody directed against glycoprotein IIb (GpIIb) (CD41a) for 15 minutes at room temperature. Fluorescence was determined as outlined in the previous paragraph.

Statistical analysis
Data were analyzed using GraphPad Prism 4.0 software. Results are expressed as the mean ± standard error (SEM). Analysis of means was performed with a 2-tailed Student t test. Differences were considered significant at P < .05.

Results

Platelet aggregation assay
The ability of cancer cells to induce platelet aggregation was analyzed using Caco-2 human colorectal cancer cells, and 2 variants of a human prostate carcinoma: the metastatic PC3M-luc cells and their nonmetastatic variant subline, PC3. Representative results for the aggregation of washed human platelets by cancer cells are shown in Figure 1A-B. A dose-dependent platelet aggregation occurred in presence of both Caco-2 and PC3M-luc cells. Prostate cancer cells (PC3M-luc) failed to induce aggregation at lower concentration (0.1-10 × 10^3 cells per mL) while colon cancer cells were able to stimulate powerful aggregation at all the concentrations tested. However, the maximum aggregation was identical at the highest cell concentration used (100 × 10^3 cells per mL) in contrast, the nonmetastatic PC3 cells were incapable of triggering platelet aggregation (Figure 1B).

Interestingly, a sustained lag time was observed prior to the onset of a brisk aggregation in the presence of both metastatic cancer cell lines. This lag time decreased as the concentration of cancer cells increased (Figure 1C). The lag time was signiﬁcantly longer compared with that recorded with Caco-2 cells (mean ± SEM: 16.5 ± 1.8 minutes vs 3.3 ± 0.3 minutes; **P < .01, n = 6; 100 × 10^3 cells per mL).

To address whether platelets could bind directly to the cancer cells, we quantitatively evaluated the association of the platelet-specific protein, CD41a, with the cancer cells following coincubation.
cytometric analysis showed that PC3M, PC3, and Caco-2 cells are negative for CD41a expression. However, CD41a expression is associated with Caco-2 and PC3M cells, but not PC3, following preincubation with platelets (Figure 1D), providing compelling evidence that platelet aggregation-inducing cancer cells are physically interacting with platelets.
To verify that a physiological aggregation was occurring, the ristocetin cofactor assay was performed and PC3M-luc cells were assessed for their ability to agglutinate lyophilized platelets in the presence of von Willebrand factor (VWF)–rich plasma. Importantly, no agglutination could be observed when lyophilized platelets were stimulated with prostate cancer cells (100 × 10^3 cells per mL) or TRAP (10 μM) over a period of 30 minutes in the presence of VWF (supplemental Figure 1A, available on the Blood Web site). Altogether, these results confirmed that physiological aggregation, and not passive agglutination was being observed in response to cancer cells.

**Tumor cell–induced physiological platelet secretion**

Because it has been previously shown that platelets are an important source of bioavailable VEGF and TGFβ, and because evidence has been provided that platelet secretion is crucial for platelet regulation of tumor homeostasis, the capability of tumor cells to induce platelet secretion in vitro was explored using a luminometric assay normally used to detect the platelet ATP/ADP secretion (PAS) by dense granules. TRAP (10 μM) was used as positive control. No ATP/ADP was detected following incubation of cancer cells with Tyrode buffer in the absence of platelets. As evident from Figure 2, cancer cells caused a dose-dependent release of significant amounts of ATP/ADP from platelet-dense granules, with a greater release in presence of Caco-2 (Figure 2A) than PC3M-luc cells (Figure 2B), confirming the results obtained in aggregometry (Figure 1B). The nonmetastatic PC3 cell variant failed to induce platelet-dense granule secretion. Similar results reflecting an association between the metastatic potential and the ability to elicit platelet secretion and aggregation were observed using a pair of breast cancer cells (MCF-7 and MDA-MB-231) that differ in their metastatic ability (supplemental Figure 2). The lag time for tumor cell–induced platelet secretion was assessed in response to the highest dose of cancer cells (100 × 10^3 cells per mL) for several time points (30 seconds–30 minutes). Caco-2 cells promoted an almost instantaneous platelet secretion (Figure 2C). In contrast, the response to PC3M-luc cells was slower and was not apparent until 5 minutes after the coincubation with platelets (Figure 2D). We suggest therefore that tumor cell–induced platelet secretion and aggregation preceded by an ATP/ADP scavenger (Apyrase, 100 μM), a cPLA2 inhibitor, resulted in a small but significant inhibition in the secretion (% MaxPAS; mean ± SEM: 100 vs 78.41 ± 8.7; P < .05, n = 10; Figure 3E) and aggregation response (% MaxAUC; mean ± SEM: 100 vs 55.2 ± 8.9; ***P < .001, n = 9; Figure 3C,E) associated with a delay in the onset of TCIPA when compared with the vehicle control. Finally, inhibition of TXA2 generation by platelets by ASA (20 μM), a selective inhibitor of TXA2 receptor, did not inhibit either tumor cell–induced platelet aggregation or secretion. Pretreatment of platelets with AACOCF3 (10 μM), a cPLA2 inhibitor, resulted in a small but significant inhibition in the secretion (% MaxPAS; mean ± SEM: 100 vs 78.41 ± 8.7; P < .05, n = 10; Figure 3E) and aggregation response (% MaxAUC; mean ± SEM: 100 vs 55.2 ± 8.9; ***P < .001, n = 9; Figure 3C,E) associated with a delay in the onset of TCIPA when compared with the vehicle control. Finally, inhibition of TXA2 generation by platelets by ASA (20 μM) failed to induce a significant decrease in PC3M-luc–induced platelet secretion and aggregation (Figure 3E). Overall, this suggests that the thromboxane synthesis pathway is not a critical player in PC3M-cell activation of platelets. For this set of inhibitors, and all subsequent inhibitors, the effects on TRAP-induced platelet secretion and aggregation are shown in supplemental Figure 5.

**P2Y12 and P2Y1 receptors do not contribute to prostate cancer cell–induced platelet secretion**

Because platelet secretion preceded aggregation, we were interested to know whether inhibitors of ADP receptors might have a differential effect on TCIPA/S. In agreement with the literature, the P2Y12 inhibitor (MRS2395, 10 μM) or the ATP/ADP scavenger (Apyrase, 10 U/mL), but not the P2Y1 inhibitor (MRS2179, 20 μM), caused consistent changes in the slope of the aggregation curve and significantly inhibited (% MaxAUC: MRS2395, 100 vs 70.5 ± 6.5; Apyrase, 100 vs 48.2 ± 11.5; *P < .05, n = 3; Figure 4A) the PC3M-luc–induced platelet aggregation response. Thus, TCIPA was sensitive to the ADP-P2Y12–deriving signaling pathway. Surprisingly, platelets preexposed to MRS3295 or MRS2179, before stimulation with PC3M-luc cells, showed a significant enhancement (% MaxPAS: 100 vs 132.3 ± 12.3; 100 vs 132.1 ± 5.6; **P < .01, n = 5; Figure 4A) of TCIPA. These findings suggest that ADP, secreted by platelets in response to PC3M-luc cells, triggers a...
positive feedback loop of activation and dense-granule secretion, culminating in full platelet aggregation.

**Integrin αIIbβ3 is required for PC3M-luc cell–induced platelet response**

There is strong evidence supporting a role for platelet adhesion molecules, such as αIIbβ3, in tumor cell–induced platelet activation.30-32 Abciximab (20 μg/mL), a specific monoclonal blocking antibody targeting αIIbβ3, significantly inhibited platelet aggregation and secretion (% MaxAUC: 100 vs 51.0 ± 10.90; **P < .01, n = 6; % MaxPAS: 100 vs 74.2 ± 9.6; *P < .05, n = 6) upon exposure to metastatic prostate cancer cells (Figure 4B-C). Moreover, changes in the slope of the aggregation curve and a remarkable delay in the onset of aggregation in the treated sample were observed. Similar results were achieved with CD41/SZ22, an αIIbβ3 mAb, when tested...
in platelet-dense granule secretion. Further confirmation of αIIbβ3 activation was provided by western blotting analysis. We show that platelet integrin αIIbβ3 is phosphorylated in response to PC3M-luc (Figure 4D). This result reinforces the hypothesis that αIIbβ3 is a crucial molecular mediator of platelet activation upon cancer cell stimulus.33

Intracellular molecular mediators of TCIPA/S are divergently activated: role of PI3K, Src, and FAK

Phosphoinositide 3-kinase (PI3K) and Src kinase are downstream to a broad range of platelet and tumor cell pathways, whereas focal adhesion kinase (FAK) plays a key role in mediating downstream signals from integrins and growth factor receptors.34-37 However, the role of these intracellular platelet molecular mediators in the tumor cell–elicited signaling cascade remains unknown. As detailed in Figure 5, platelet aggregation induced by PC3M-luc was significantly (% MaxAUC: 100 vs 68.4 ± 8.6; ***P < .001, n = 5; 100 vs 80 ± 4.56; *P < .05, n = 4) affected by both wortmannin (100 nM) and LY294002 (20 μM), 2 unrelated PI3K inhibitors. In contrast, pharmacologic inhibition of PI3K did not impair TCIPS (Figure 5B).

The role of Src kinase in PC3M-luc activation of platelets is detailed in Figure 5. Whereas PP2 (25 μM), a broad-range inhibitor of Src-family kinases, did not alter the extent of TCIPS, it markedly reduced TCIPA (% MaxAUC: 100 vs 68.4 ± 8.6; ***P < .001). In contrast, FAK blockade, achieved using PF228, resulted in a dramatic reduction of TCIPA (% MaxAUC: 100 vs 7.81 ± 0.7; ***P < .0001).

Figure 3. PAR-1 and thromboxane receptors do not mediate TCIPA or TCIPS. (A-C) Representative aggregation traces showing the effect of the reported inhibitors on TRAP (10 μM; A)– or PC3M-luc (100 × 10^3 cells per mL; B-C)–induced platelet aggregation. (D-E) Bar graph comparing the effect of the indicated inhibitors on platelet aggregation (n = 3/9) and secretion (n = 6/10) elicited by TRAP or PC3M-luc. Results were normalized for each donor relative to the maximal response seen in TRAP- or PC3M-luc–stimulated platelets (100%) in presence of the vehicle solution (data are expressed as mean ± SEM; *P < .05; ***P < .001).
n = 4; Figure 5A-B) accentuated by a significant inhibition of TCIPS (% MaxPAS: 100 vs 66.1 ± 9.1; *P < .05, n = 7). Collectively, these results identify crucial intracellular molecular mediators of TCIPA/S, providing evidence for functional divergence in the signaling events involved in the activation of platelets upon exposure to metastatic prostate cancer cells. It is noteworthy that the pro
c
file of the pathways inhibited in our studies strongly overlap with pathways known to be
involved in accessory signaling pathways that use immunoreceptor
tyrosine-based activation motif (ITAM).

Dependence of cancer cell–induced platelet response on the
FCγRIIA signaling pathway
Considering the role that platelets play in the innate immune response38 and thinking of cancer cells as ultimate parasites of the
bloodstream, we next investigated the possible involvement of the
immune FCγRIIA, an ITAM-containing platelet receptor, in platelet
activation induced by PC3M-luc. Washed platelets were preincu-
bated with 10 μg/mL of the Fab fragments of the FCγRIIA blocking
antibody IV.3 before being activated with PC3M-luc cells. The
ability of tumor cells to induce aggregation (% MaxAUC: 100 vs
2.3 ± 1.2; ***P < .0001, n = 3) and dense granule secretion (% MaxPAS: 100 vs 32.9 ± 9.2; **P < .01) was markedly inhibited
by antibody treatment (Figure 6A,C). Moreover, mouse platelets,
which are naturally FCγRIIA negative, did not respond when
stimulated with PC3M cells (Figure 6D). This result therefore
identifies a new platelet receptor that is selectively required in PC3-M
cancer cell–stimulated platelets by a mechanism, to date, com-
pletely unknown. Activation of human platelets by FCγRIIA is
associated with rapid tyrosine phosphorylation and activation of
the nonreceptor tyrosine kinase Syk.39 We compared the effect of
Bay61-3606 (5 μM) and piceatannol (25 μM), 2 unrelated Syk
inhibitors, on TCIPA and TCIPS and demonstrated dramatic inhibition of both responses (\% MaxAUC: Bay-61: 100 vs 12.2 ± 6.8; ***P < .0001; piceatannol: 100 vs 30.7 ± 15.1; *P < .05; n = 6; % MaxPAS: Bay-61: 100 vs 59.1 ± 9.4; **P < .01; piceatannol: 100 vs 20.6 ± 5.8; ***P < .0001; n = 6; Figure 6) strongly supporting our proposal of a central role for FcγRIIa.

Phospholipase Cγ2 (PLCγ2) also lies downstream of the activated FcγRIIa-Syk pathway. The broad-spectrum PLC inhibitor, U73122 (10μM) had a significant effect on tumor cell–induced platelet secretion (\% MaxPAS: 100 vs 75.6 ± 7.9; *P < .05; n = 11) and effectively reduced and delayed TCIPA (\% MaxAUC: 100 vs 40.7 ± 8.5; *P < .0001; n = 9; Figure 6B,E). It is also noteworthy that the intracellular calcium antagonist 1,2-Bis(2-aminophenoxy)ethane-N,N',N'-tetraacetic acid (BAPTA,10 μM) did not affect TCIPS or TCIPA response (n = 7/4; Figure 6F), suggesting a Ca2+-independent platelet activation. The inhibitory properties of BAPTA were verified in collagen-related peptide (CRP)–induced platelet aggregation (supplemental Figure 3F).

The protein kinase C (PKC) family includes many isoforms all differently involved in a number of platelet processes, most importantly aggregation and secretion. The possible role of PKC in PC3M-induced platelet aggregation and secretion was next explored. Human WPs were preincubated with GF109203X (10μM), a potent broad-spectrum PKC inhibitor and a marked decrease in both platelet aggregation and secretion responses to PC3M-luc cells were observed (Figure 6B,E; % MaxAUC: 100 vs 32.5 ± 10.7; ***P < .01; n = 6; % MaxPAS: 100 vs 49.68 ± 11.02; **P < .01; n = 6), confirming the important functional profile of PKC in platelets. A general scheme of platelet-signaling pathways is summarized in Figure 7 showing all potential molecular mediators of TCIPA/S induced by metastatic PC3M prostate cancer cells and highlighting a central role for ITAM-dependent pathway(s).

**Discussion**

Once in the bloodstream, cancer cells perturb the surrounding microenvironment, triggering abnormal platelet responses mediated by direct or indirect interactions. Platelets can be aggregated in vitro and in vivo by various cancer cell lines, a phenomenon described in 1968 as "tumor cell induced platelet aggregation." Many factors are secreted by platelet granules in response to tumor cells, such as TGF-β and VEGF, that support the metastatic cascade and promote tumor vascularization. Kerr and colleagues showed that many tumor-secreted proteins are taken up and stored in platelets and specifically released at the metastatic sites. However, while the ability of cancer cells to aggregate and cluster platelets around the tumor is supported by a large body of experimental and clinical data, the role of cancer cells in promoting platelet secretion of procarcinogenic and thrombotic factors is poorly understood. In the present study, we show that highly metastatic colorectal cancer cells (Caco-2) and a prostate carcinoma cell line (PC3M-luc) stimulate the release of platelet-dense granule contents in a dose-dependent manner. In contrast, cell lines with low metastatic potential including the isogenic PC3 prostate cancer cell line showed a markedly reduced response. A similar pattern of response was observed with a pair of breast cancer cell lines (supplemental Figure 2), emphasizing the hypothesis of a direct association between the metastatic potential and tumor cell–induced platelet aggregation and secretion.

A sustained α-granule release was also observed in response to all the cell lines tested. It is noteworthy that the nonmetastatic prostate cancer PC3 cells activated α-granule secretion with no effect on aggregation and dense granule release suggesting that nonmetastatic cancer cells can activate platelets without stimulating a prothrombotic response. A more precise interrogation of the comparative contents of the bioactive agents in the platelet α-granule releasate may allow a greater understanding of molecular events that ensue following activation of platelets with metastatic or nonmetastatic cells. Evaluation of platelet secretion over time showed that Caco-2 and PC3M-luc cancer cells induced platelet ATP/ADP release earlier than aggregation. The aggregation was characterized by a distinct lag time which correlated with the cell’s ability to induce TCIPS. Moreover, antagonism of platelet P2Y12 receptors with MRS2395 or scavenging of released ADP with apyrase significantly attenuated aggregation, but not secretion responses. Thus, these results strongly imply that cancer cell–induced release of dense granule ADP facilitates the induction of platelet aggregation and suggest that rapid dense-granule secretion is the primary platelet activation pathway in response to cancer cells. If this is indeed the case, then we can expect that all inhibitors of the secretory pathway(s) will also have a downstream impact on the platelet aggregation responses recorded. Thus, agents that inhibit platelet secretion of dense granules will de facto inhibit the dependent response, namely aggregation, which may result in reduced thrombotic complications in cancer patients.

We next explored the activity of a range of pharmacological inhibitors targeted at various platelet receptors and cytosolic signaling proteins. We were interested mainly in agents that inhibited both secretion, which we hypothesized to be a primary response, and aggregation, which would represent a secondary response. In
this screen, inhibitors of integrin αIIbβ3, FAK, Syk kinase, and PKC showed significant effects on both TCIPS and TCIPA. These components have all previously been identified in association with each other in a platelet context.

FcγRIIa is an ITAM-containing receptor used in platelets for immune receptor signaling. It is found in other receptors in platelets, such as the collagen receptor complex GPVI-FcγRIIa chain, and the podoplanin receptor CLEC-2. More recently, a role for the podoplanin-CLEC-2 axis in mediating tumor metastasis has been identified. FcγRIIa directly interacts with Syk, activating PKC either directly or via PLC activation. In our studies, FcγRIIa inhibition with the Fab antibody IV.3 substantially blocked both platelet secretion and aggregation. Similarly, inhibition of the tyrosine kinase Syk reduced both TCIPS and TCIPA. Activation of Syk leads to PLCγ2 phosphorylation and activation, which can positively regulate α- and dense-granule secretion in platelets as well as platelet aggregation. Inhibition of PLC in our studies significantly reduced aggregation and secretion (Figure 6), indicating that an FcγRIIa-Syk-PLCγ signaling pathway is activated. In contrast, we observed that BAPTA, a calcium chelator, has no significant effect on platelet secretion responses, suggesting that calcium is not required. Importantly, inhibitors of PKC (GF109203X) demonstrated a potent dual inhibition of both TCIPS and TCIPA. Together, these data indicate that signaling for PC3M-luc–mediated dense-granule secretion involves FcγRIIa molecular components by engaging downstream signaling mechanisms including Syk and that this event would be sufficient to initiate dense-granule secretion. Further studies will be necessary to identify the PKC isoform(s) activated during PC3M-luc–induced platelet aggregation and secretion.

The discovery of FcγRIIa as an important mediator of both platelet aggregation and secretion in response to PC3M-luc makes it a strong potential target for therapeutic intervention. This raises the possibility of TCIPS as a target for potential intervention in tumor...
angiogenesis and metastasis and in tumor-derived thrombosis. The inhibition of this platelet receptor, although not targeting the tumor cell itself, could potentially reduce metastasis and prevent thrombotic events associated with cancer without compromising the hemostatic functions of platelets.

The nature of the cancer cell ligand that stimulates platelet activation remains to be elucidated. Podoplanin is a recently identified protein found on some cancer types that acts as ligand for CLEC-2, a platelet ITAM-containing receptor. However, podoplanin is not usually found on colon or prostate cancer and we further confirmed...
this by flow cytometry analysis (data not shown). In vivo, it is likely that antitumor antibodies could be inadvertent accessories in platelet responses, binding to FcγRIIa via their Fe portion and resulting in enhanced facilitation of tumor cell transport to a metastatic site instead of the intended immune-mediated elimination of a damaging invader cell. However, in the assays performed in this study, we used washed HPs and washed human prostate cancer cells, so no exogenous antibodies are available for activation of platelets. Therefore, an alternative mode for the FcγRIIa receptor must be operating. Recently, in platelets, FcγRIIa has been identified as a transmembrane receptor responsible for mediating “outside-in” signaling through αgβ3.52 Of note, we have identified that, in addition to the FcγRIIa, Syk, and PKC axis in platelets, inhibitors of αgβ3 had a parallel profile of inhibition of tumor cell–induced platelet activation. Thus, we can propose a collaborative role for the αgβ3 and FcγRIIa receptors in recognition of cancer cells by platelets. Subsequent activation and secretion of granule contents may cause a cycle of platelet recruitment and protection of circulating tumor cells that ultimately facilitates tumor cell survival in the circulation and at metastatic sites.

Finally, it is noteworthy that platelet TxA2 synthesis is a critical component of ADP-induced platelet activation. Aspirin, a potent inhibitor of platelet thromboxane production and a widely used antiplatelet agent, has demonstrable potential as an antineoplastic agent.52 In a large metaanalysis of >17,000 patients, daily usage of low-dose aspirin has been shown to reduce the incidence, growth, and metastasis of a number of cancers in a period of 5 to 6 years.52 This finding provides proof of principle for antiplatelet pharmacological intervention to specifically prevent cancer metastasis. Although our studies failed to demonstrate a significant effect of aspirin on TCIPA and TCIPS, a strong tendency toward inhibition was noted. Our data suggest that antiplatelet treatment with P2Y12 inhibitors may provide a superior inhibition of platelet activation in response to tumor cells and this may have a therapeutic implication.

The cell numbers used in this study are admittedly in excess of numbers that might be found physiologically in circulating blood. Circulating tumor cell counts of >30/mL are routinely observed in prostate cancer patients.53 The detection of circulating tumor cells (CTCs) in the blood and disseminated tumor cells (DTCs) in the bone marrow of cancer patients is associated with a poor prognosis.54 Our study demonstrates that platelet activation can occur after a brief, but targeted, interaction with high numbers of cancer cells in vitro. The in vivo correlate of our study would allow for a longer interaction time with CTCs and a more targeted interaction with DTCs. Thus, we believe that although the numbers of cancer cells used in our study are not physiological, the deductions made from such in vitro studies will have physiological relevance if time and opportunity are taken into account.

In conclusion, this work demonstrates for the first time, the capability of cancer cells to directly promote platelet secretion which is required to augment tumor cell–induced platelet aggregation. In addition, we have identified molecular mediators divergently activated in the aggregation and secretion responses (Figure 7). Furthermore and most importantly, we recognize a novel essential role for FcγRIIa in prostate cancer cell–induced platelet activation opening the door to novel antimitastatic therapies.

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Authorship

Contribution: A.M. and N.M. conceived the study and designed the research; A.M. performed the experiments and wrote the manuscript; D.W. provided clinical advice; N.M. and D.W. reviewed the manuscript; and S.W.K. provided reagents and methodical advice for all signaling components of this study.

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References


22. Kozlowski JM, Fidler IJ, Campbell D, Xu ZL, Szabo I. Syk are essential for activation of mouse platelets by collagen. EMBO J. 1997;16(9):3279-3285.


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