Prolonged half-life and preserved enzymatic properties of factor IX selectively PEGylated on native N-glycans in the activation peptide

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Current management of hemophilia B entails multiple weekly infusions of factor IX (FIX) to prevent bleeding episodes. In an attempt to make a longer acting recombinant FIX (rFIX), we have explored a new releasable protraction concept using the native N-glycans in the activation peptide as sites for attachment of polyethylene glycol (PEG). Release of the activation peptide by physiologic activators converted glycoPEGylated rFIX (N9-GP) to native rFIXa and proceeded with normal kinetics for FXIa, while the $K_m$ for activation by FVIIa–tissue factor (TF) was increased by 2-fold. Consistent with minimal perturbation of rFIX by the attached PEG, N9-GP retained 73%-100% specific activity in plasma and whole-blood–based assays and showed efficacy comparable with rFIX in stopping acute bleeds in hemophilia B mice. In animal models N9-GP exhibited up to 2-fold increased in vivo recovery and a markedly prolonged half-life in mini-pig (76 hours) and hemophilia B dog (113 hours) compared with rFIX (16 hours). The extended circulation time of N9-GP was reflected in prolonged correction of coagulation parameters in hemophilia B dog and duration of effect in hemophilia B mice. Collectively, these results suggest that N9-GP has the potential to offer efficacious prophylactic and acute treatment of hemophilia B patients at a reduced dosing frequency. (Blood. 2011;118(8):2333-2341)

Introduction

Factor IX (FIX) is a vitamin K–dependent glycoprotein and an essential protease of the hemostatic system. The domain organization of FIX is shared with factors VII, X, and protein C and comprises an N-terminal domain rich in $\gamma$-carboxyglutamic acid (Gla), 2 epidermal growth factor-like repeats and a C-terminal trypsin-like protease domain.1 Together they form a 55-kDa single-chain protease precursor circulating in plasma at a concentration of approximately 90nM (5 $\mu$g/mL), defined as 1 IU/mL. FIX is converted to the 2-chain activated form by the tissue factor (TF)–factor VIIa complex or factor Xa (FXa). Activation occurs by limited proteolysis at Arg145 and Arg180 in the protease domain and liberates a 35-amino acid activation peptide that carries the native N-glycans in the activation peptide as sites for attachment of polyethylene glycol (PEG). Release of the activation peptide by physiologic activators converted glycoPEGylated rFIX (N9-GP) to native rFIXa and proceeded with normal kinetics for FXIa, while the $K_m$ for activation by FVIIa–tissue factor (TF) was increased by 2-fold. Consistent with minimal perturbation of rFIX by the attached PEG, N9-GP retained 73%-100% specific activity in plasma and whole-blood–based assays and showed efficacy comparable with rFIX in stopping acute bleeds in hemophilia B mice. In animal models N9-GP exhibited up to 2-fold increased in vivo recovery and a markedly prolonged half-life in mini-pig (76 hours) and hemophilia B dog (113 hours) compared with rFIX (16 hours). The extended circulation time of N9-GP was reflected in prolonged correction of coagulation parameters in hemophilia B dog and duration of effect in hemophilia B mice. Collectively, these results suggest that N9-GP has the potential to offer efficacious prophylactic and acute treatment of hemophilia B patients at a reduced dosing frequency. (Blood. 2011;118(8):2333-2341)

Various approaches have been pursued to extend the half-life of coagulation factors including genetic fusions to albumin and the Fc fragment of IgG as well as chemical modification by hydrophilic polymers such as polyethylene glycol (PEG).12-15 The prolonged circulation time endowed by the latter approach is believed to arise from a reduction in the efficiency of various elimination processes such as renal excretion, receptor mediated uptake, and proteolytic degradation.16 With several PEGylated protein-based therapeutics on the market since 1990, the efficacy and safety of this technology has undergone extensive clinical validation.17 For any protraction strategy applied on FIX, the gain in circulation time should be balanced by the need to preserve biologic activity and normal regulation of the activated form. Being remote from known interaction sites in FIX and present only in the circulating zymogen form, the N-glycans of the activation peptide offer potential acceptor sites for PEGylation that may fulfill these requirements. In...
the present study we demonstrate the feasibility of targeted glycoPEGylation and show that this approach confers a long plasma half-life to FIX in animal models while preserving its biologic activity.

Methods

Production and biochemical characterization of N9-GP

Production and biochemical characterization of glycoPEGylated rFIX (N9-GP) is described in supplemental Methods (available on the Blood Web site; see the Supplemental Materials link at the top of the online article).

Reference FIX

BeneFIX (Pfizer) was used as a source of recombinant FIX (rFIX) for reference purposes. The product was reconstituted according to the manufacturer’s instructions and stored in aliquots at −80°C.

In vitro functional characterization

Kinetics of FIX activation by FXIa or FVIIIa-TF. Activation was initiated by addition of 10pM FXIa (Haematologic Technologies) or 1nM rFVIIIa (Novo Nordisk) and 0.1nM lipitated TF (Innovin; Siemens) to varying concentrations of FIX in 25mM HEPES, 137mM NaCl, 3.5mM KCl, 5mM CaCl2, 0.1% BSA, pH 7.4 at 25°C. Reactions were quenched after 30 minutes by addition of 30nM EDTA and generated FIXa measured at 405 nm in the presence of 31% ethylene glycol and 0.5nM Spectrozyme IXa substrate (American Diagnostica).

Kinetics of FX activation. FX activation experiments were performed as described by Christiansen et al18 (see supplemental Methods for a full description).

Binding of FIX to endothelial cells. Primary HUVECs (Cambrex BioScience) were maintained in EBM-2 medium as described by the manufacturer. Binding experiments were performed in 48-well plates at 95% confluence essentially as described by Cheung et al.19 Briefly, cells were exposed to 125I-FIX (0.5-1nM) and unlabeled competitor in 0.15 mL buffer (10mM HEPES, 150mM NaCl, 4mM KCl, 5mM CaCl2, 0.1% BSA, pH 7.4) at 25°C. Reactions were quenched after 30 minutes by addition of 30nM EDTA and generated FIXa measured at 405 nm in the presence of 31% ethylene glycol and 0.5nM Spectrozyme IXa substrate (American Diagnostica).

FX clot analysis. Coagulant activity was estimated using a 1-stage FIX clotting assay (instrument and reagents were from Instrumentation Laboratories). Test sample was mixed with equal volumes of FIX immunodepleted plasma, activated partial thromboplastin time (aPTT) reagent (SynthAfax), and 25nM CaCl2. The time to clot formation was measured on an ACL9000 instrument using HemosIL reference plasma calibrated against the 3rd WHO international FIX concentrate standard (96/854; National Institute for Biological Standards and Control). Control experiments using N9 and free PEG confirmed that PEG did not interfere with the assay as previously noted for some aPTT reagents.20

Thromboelastography in hemophilia B whole blood. Blood was obtained from 5 moderate and 3 severe HB patients in a study approved by and performed according to guidelines from the Danish National Committee on Biomedical Research Ethics (no. [KF] 01 299489, add. 18 186). For comparative purposes blood was also obtained from 11 healthy volunteers. All participants received oral and written information and signed informed consent before blood sampling. Healthy donors had not taken acetyl salicylic acid for 10 days or other anti-inflammatory drugs for 72 hours. All patients had not received factor concentrate for minimum 72 hours. Blood was sampled in 3.2% citrate and rested 90 minutes before analysis. Equimolar amounts of N9-GP or rFIX (0.08-76nM) were added and clotting initiated by kaolin (Haemroscope Corporation). A normal reference range was generated based on blood from the healthy volunteers. Clot (R) time, maximum thrombus generation (MTG) and maximum amplitude (MA) parameters were used for statistical analysis. Data were fitted to a concentration-response-curve in a nonlinear model. Parameter estimates for efficacy and EC50 were subsequently analyzed on log-scale in a normal linear mixed effects model allowing for random subject-to-subject and random day-to-day variation.

In vivo pharmacokinetics and pharmacodynamics

Animals. HB mice (B6.129P2-F9tm1Dws) were originally obtained from Darrel W. Stafford (University of North Carolina).21 Mice were of both sexes (50:50) and between 12 and 16 weeks old. Male Göttingen mini-pigs were obtained from Ellegaard Göttingen Minipigs A/S. Studies were approved by and performed according to guidelines from the Danish Animal Experiments Council, the Danish Ministry of Justice. Studies in an immunologically tolerant HB dog were conducted at the Chapel Hill colony under the supervision of T.C.N.22 and approved by the Institutional Animal Care and Use Committee at the University of North Carolina. The methods used for tolerization and characterization of the level of transgenic human FIX in the dog are described in supplemental Methods.

Calculation of doses. Doses were calculated based on protein content using the same molecular weight (55 kDa) for rFIX, N9, and N9-GP. Hence, at a given milligram per kilogram dose, equimolar amounts of the molecules were administered.

Quantitative analysis. FX activity and Ag levels were quantitated in citrated plasma samples by FIX clot analysis as described above and by sandwich ELISA using an anti-FIX GlA-domain Ab for capture and a HRP-conjugated sheep anti-FIX detection Ab (Fitzgerald Industries International).

Pharmacokinetic analysis. Unless otherwise stated, pharmacokinetic analysis was carried out by noncompartmental methods (NCA) using WinNonlin software (Pharsight Corporation).

Pharmacokinetics in hemophilia B mice and mini-pigs. N9-GP or rFIX was given intravenously to HB mice as a 1.5 mg/kg bolus injection in the tail vein. Blood samples were collected according to a staggered design consisting of 3 mice per time point and 3 samples per mouse. Six mini-pigs (9.7 ± 1.2 kg) divided in 2 groups received an intravenous bolus injection of 0.2 mg/kg N9-GP or rFIX. Blood samples were collected for PK analysis up to 14 days after administration. Data were fitted to a 2-compartment model and estimated parameters used to simulate elimination profiles in a multiple dose setting using NONMEM software (GloboMax/ICON).

Acute effect in tail bleeding model. Bleeding was induced in HB mice by amputating 4 mm of the tail tip as described previously.23 Briefly, 10 minutes before amputation, the tail was placed in a test tube containing 14 mL of saline (37°C) and 5 minutes later vehicle (buffer), rFIX, or N9-GP were administered randomized and blinded at 10 mL/kg through the carotid artery. The total bleeding time and blood loss (expressed as nanomoles of hemoglobin) were recorded for 30 minutes.

Duration of effect in tail bleeding model. HB mice were administered 0.75 mg/kg rFIX or N9-GP in the tail vein. At timed intervals, bleeding time and blood loss were determined as described for measurement of acute effect.

Duration of effect in ferric chloride-induced carotid artery injury model. Each group of HB mice received 0.75 mg/kg rFIX or N9-GP randomized and blinded through the tail vein, except for mice tested after 5 minutes which were dosed through the jugular vein. At timed intervals after dosing, injury was induced by exposure of the carotid artery to 10% (w/v) ferric chloride for 3 minutes and time to arrest of blood flow measured as described by Möller and Tranholm.24

Pharmacodynamics in hemophilia B dog. A female HB dog (18 kg) immunologically tolerant to human FIX was used in the study.25 rFIX and N9-GP were infused at 0.4 mg/kg. Blood samples for pharmacokinetic (PK) analysis, thromboelastography (TEG; no stabilizer), and whole-blood clotting time (WBCT; no stabilizer) were drawn at regular intervals up to 45 days after administration.

Results

Production and characterization of N9

rFIX (N9) was expressed in CHO cells which are known to support proper maturation and posttranslational modification of
GlycoPEGylation of N9

Selective glycoPEGylation used the substrate promiscuity of ST3GalIII sialyl transferase which allowed for the transfer of CMP-activated sialic acid-6'-40k PEG to terminal galactoses of the N-glycans in the activation peptide of desialylated N9. Remaining exposed galactosyl residues were subsequently sialylated by addition of excess unmodified CMP-sialic acid. By optimizing reaction conditions the conjugation stoichiometry could be controlled to yield a product (N9-GP) containing mainly mono-PEGylated N9 (95%) and a minor proportion of the di-PEG form (5%) as determined by SDS-PAGE and reversed-phase (rp) HPLC (Figure 1C-D). By the latter method the proportion of unPEGylated N9 was estimated to be 1%. The content of activated FIX was measured by the amidolytic method and found to be 0.03% compared with a level of 0.13%-0.18% in rFIX.

Localization of PEG to the activation peptide was confirmed by incubation of N9-GP with FXIa which resulted in quantitative conversion of the single-chain molecule to heavy and light chains with the same electrophoretic mobility as activated N9. Staining for PEG revealed 2 bands with the same relative intensities as unactivated N9-GP but lower apparent molecular weights, which were identified by N-terminal sequencing as the liberated activation peptide (Figure 1C). Further mapping used a markedly prolonged elution time of PEG by rpHPLC which allowed for the isolation of PEGylated peptides after digestion of N9-GP with trypsin and endoproteinase Glu-C to separate the N-glycans on different peptides. N-terminal sequencing of the isolated PEGylated fragments showed the PEG to be almost equally distributed among peptides containing either the Asn157 (55%) or Asn167 (45%) N-glycan acceptor site (see Figure 1B). No other PEGylated peptides were identified.

In vitro functional properties of N9-GP

To address the effects of glycoPEGylation on the functional properties of N9, we examined individually the reactions in the coagulation process relevant to the activation or subsequent proteolytic action of N9-GP. The kinetics of FIX activation catalyzed by FXIa or the FVIIa-TF complex was measured in amidolytic assays. As shown in Figure 2A, the activation of N9 and N9-GP, tenase complexes were assembled on a phospholipid surface by combining the respective FXIa-activated forms and thrombin-activated FVIII. Conversion of FIX to FXa by the complexes was measured from the generated amidolytic activity. By varying the concentration of FVIIIa in the assay, the kinetically derived apparent dissociation constants ($K_{\text{a}}$) for the interaction with FVIIIa- phospholipid were estimated and found to be similar for activated N9 (1.14 ± 0.03nM) and N9-GP.
The effect of glycoPEGylation on the binding to endothelial cells (HUVECs) was investigated in vitro. In accordance with previous reports, rFIX and N9 were both capable of competitively displacing $^{125}$I-labeled rFIX with low-nanomolar $K_i$ values of 1.6 and 2.4nM, respectively (Figure 3A). Specificity was confirmed by the ability of an anti-FIX Gla-domain Ab to abrogate binding, whereas high concentrations of FX or the LRP scavenger receptor antagonist RAP (receptor-associated protein) had no effect (Figure 3B). N9-GP also competitively displaced $^{125}$I-rFIX. However, 20-fold higher concentrations were required corresponding to a $K_i$ of 48nM (Figure 3A).

**Activity in plasma and whole-blood–based assays**

Having established the basic molecular properties of N9-GP, we next assessed its ability to support coagulation in plasma and whole-blood–based assays. In the 1-stage aPTT assay, the specific activity of N9-GP was 190 ± 10 IU/mg, that is, slightly lower (73%) than that measured for N9 (260 ± 28 IU/mg). To further assess the activity of N9-GP, clot formation was examined in whole blood obtained from 8 HB patients with the moderate (5) and severe (3) phenotype. In this and the subsequent studies, rFIX was included for comparison. Clot formation was initiated by addition of kaolin and monitored by TEG. Kaolin is considered an intrinsic pathway activator. However, a contribution from the extrinsic FVIIa-TF-driven pathway was evident because addition of an inhibitory TF Ab to FIX-depleted blood resulted in a 30% increase of the R time. Representative TEG profiles from a severe and a moderate patient are shown in Figure 4A and B before and after addition of 28nM N9-GP or rFIX (about 30% of the normal level) to approximate the plasma level in HB patients receiving a standard prophylactic dose. Baseline TEG profiles were found to differ somewhat between patients; however, all were outside the normal range and thus displayed markedly delayed coagulation. Spiking of blood with increasing levels (0.08-76nM) of N9-GP or rFIX led to a concentration-dependent decrease of the R-time and increase in MTG which entered the normal ranges at the highest concentrations (Figure 4C-D). At this concentration, no significant differences in clotting parameters between rFIX and N9-GP were observed indicating similar efficacies. Likewise, the ratio of EC50 values of N9-GP and rFIX estimated based on R time, MTG, and MA values did not differ significantly from unity indicating comparable potencies (supplemental Table 1).

**Figure 3. Effect of glycoPEGylation on the binding of N9 to endothelial cells.**

(A) Competitive displacement of $1nM^{125}$I-rFIX with increasing concentrations of unlabeled rFIX, N9, or N9-GP (0.6-600nM). Monolayers of HUVEC cells were incubated with compound for 3 hours at 4°C before counting of bound radioactivity. Binding of rFIX was analyzed according to a homologous 1-site binding model and the estimated apparent $K_i$ of 1.6nM used to derive $K_i$ values for N9 (2.4nM) and N9-GP (49nM). (B) HUVECs were incubated with 0.5nM $^{125}$I-rFIX with or without molar excess of anti-FIX Gla-domain Ab (Gla-Ab; 33nM), FX (400nM), or RAP (500nM) for 3 hours at 4°C. All data are shown as mean ± SD (n = 4).
Acute and prolonged hemostatic effect in hemophilia B mice

The hemostatic potentials of N9-GP and rFIX were further compared in vivo in HB mice where bleeding was induced by tail-tip transaction 5 minutes after dosing. In vehicle-treated mice bleeding time and blood loss were significantly increased compared with normal C57BL mice. Both compounds demonstrated a clear dose-response with no effect at the lowest dose of 0.01 mg/kg (corresponding to 1.9 IU/kg N9-GP based on the aPTT clot activity) and normalization of both parameters at 0.4-0.75 mg/kg (76-143 IU/kg N9-GP; Figure 5A-B). Consistent with the results obtained in whole blood, the dose-response relationship for the 2 compounds did not differ significantly. The plasma level of FIX was determined at the end of each 30-minute observation period and found to be proportional to the dose given but, interestingly, on average 2.1 times higher for each 30-minute observation period and found to be proportional to the dose given but, interestingly, on average 2.1 times higher for N9-GP than rFIX at equimolar doses (Figure 5C). The higher in vivo recovery of N9-GP was confirmed in HB mice administered N9-GP or rFIX but not subjected to injury (Table 2). Measurement of plasma levels in these mice up to 1 week after infusion furthermore demonstrated a prolonged pharmacokinetic profile of N9-GP characterized by a half-life (T½) of 41 hours compared with 17 hours for rFIX based on ELISA (Figure 6A, Table 2). Superimposable profiles were obtained by measurement of clot activities indicating that N9-GP retained functionality in circulation (Figure 6A).

The longer circulation time of N9-GP compared with rFIX was reflected in a prolonged duration of effect in the tail-bleeding model. After infusion of 0.75 mg/kg N9-GP (143 IU/kg) or rFIX (188 IU/kg) bleeding was normalized acutely. In addition, N9-GP maintained blood loss in the normal nonhemophilic range until day 2 followed by a gradual return to the hemophilic baseline at day 5. In mice receiving rFIX, blood loss was already significantly elevated at day 2 and back at baseline at day 3 (Figure 6B).

The hemostatic protection provided by N9-GP and rFIX was also compared in the ferric chloride–induced carotid artery injury model in HB mice.24 Injury was induced up to 5 days after administration of 0.75 mg/kg and the time to occlusion recorded. Except for the first time point at 5 minutes, the average occlusion time in mice receiving N9-GP was consistently shorter than in mice treated with rFIX (Figure 6C). This difference was also reflected in the fraction of animals which developed stable occluding thrombi within 25 minutes after injury. One to 4 days after dosing occlusion was observed in 90%-100% of N9-GP treated mice, whereas only 70% of mice receiving rFIX occluded at day 1 and 44% at day 4. Five days after dosing 56% of N9-GP mice still occluded compared with 13% of mice treated with rFIX.

Pharmacokinetics and pharmacodynamics in hemophilia B dog

The pharmacodynamics of N9-GP was investigated in a single HB dog tolerized to human FIX by neonatal retrovial gene transfer25 which allowed for inclusion of rFIX in a crossover design. N9-GP
and rFIX were both given at 0.4 mg/kg corresponding to 76 and 100 IU/kg, respectively. As shown in Figure 7A, plasma clearance of N9-GP was greatly reduced and the plasma concentration did not reach the 1% level (50 ng/mL) until 40 days after administration compared with 8 days for rFIX. This difference was reflected in a functional half-life of N9-GP (113 hours) that was 7 times longer than rFIX (16 hours) and a clearance rate that was reduced 16-fold from 9.8 to 0.6 mL/kg/h (Table 2). Profiles determined by ELISA and clotting activity were super-imposable until day 16 at which the detection limit of the functional assay was reached providing further evidence of preserved functionality of N9-GP during the extended residence in circulation.

![Figure 6. Pharmacokinetics and duration of effect in hemophilia B mice.](image)

(A) N9-GP (circles) or rFIX (squares) was administered at 1.5 mg/kg. Blood samples were analyzed for FIX clot activity (closed symbols) and Ag (open symbols) and are shown as mean ± SD (n = 3). (B) Duration of effect of 0.75 mg/kg N9-GP (143 IU/kg) and rFIX (188 IU/kg) in the tail bleeding model. Blood loss was recorded for 30 minutes at the indicated time points starting at 5 minutes postadministration. Blood loss in normal (C57BL) mice and vehicle-treated HB mice are included for comparison (—). Results are shown as mean ± SEM (n = 8-16/group). *P < .02 and **P < .01 are relative to rFIX using 1-way ANOVA and Bonferroni posttest. (C) Duration of effect in the ferric chloride–induced carotid artery injury model. The occlusion time was determined at indicated intervals after infusion of 0.75 mg/kg N9-GP or rFIX with the first time point after 5 minutes. Occlusion was defined as complete lack of blood flow for >5 minutes. If occlusion did not occur within the 25-minute observation period, as was the case in vehicle-treated mice, the occlusion time was reported as 25 minutes (—). Results are shown as mean ± SEM (n = 6-11/group); *P < .05 and **P < .01 are relative to rFIX using the Kruskal-Wallis test and the Dunn posttest.

![Figure 7. Pharmacokinetics and pharmacodynamics in hemophilia B dog and mini-pig.](image)

A tolerized HB dog received successive infusions of 0.4 mg/kg rFIX (100 IU/kg) and N9-GP (76 IU/kg) separated by a washout period. Blood samples were collected at indicated time points to measure (A) FIX clot activity (closed symbols) and Ag levels (open symbols) and to evaluate the ex vivo hemostatic potential by (B) WBCT and (C-D) TEG. The standard WBCT range in nonhemophilic dogs is indicated. For the TEG experiments, coagulation was triggered with kaolin. (C) Representative TEG traces recorded pre-dose (gray), at 5 minutes and 3, 19, 26, 31, 40, and 43 days (from left to right in black) after administration of N9-GP. (D) MTG values at indicated time points after administration. (E) Plasma concentrations of rFIX and N9-GP in mini-pig (3 per group) shown as individual Ag measurements after administration of 0.2 mg/kg (50 and 38 IU/kg, respectively). Solid lines represent best fit to a 2-compartment model. (F) Fitted parameters were used to simulate pharmacokinetic profiles in a multiple dose setting. Simulations were performed with N9-GP (black line) dosed every 7 (solid) or 14 (stippled) days and rFIX (gray line) dosed every 2.5 days, all at 0.2 mg/kg. Horizontal stippled lines represent 100% (top) and 1% (bottom) levels, respectively.
WBCT is a marker for hemostatic efficacy in HB dogs and was used to evaluate the pharmacodynamics of N9-GP. Because of low endogenous levels of FIX in the tolerized dog as a result of neonatal gene transfer (<0.1% of the normal level), the WBCT pre-dose value (22 minutes) was shorter than for naive HB dogs (>60 minutes) but sufficiently above the normal range (10-15 minutes) to allow for detection of pharmacologic effect. Normalization of the WBCT was observed 5 minutes after administration of rFIX and lasted for 4 days followed by a gradual return to pre-dose levels at day 12 (Figure 7B). A similar immediate correction of the clotting time was observed with N9-GP. However, the WBCT remained in the normal range for 12-19 days and then slowly increased toward the pre-dose level at day 45 in accordance with its reduced elimination kinetics (Figure 7B). A similar pattern was observed when blood samples were analyzed by TEG which is more sensitive to trace amounts of FIX. Here, clot formation was corrected until approximately day 26 followed by a return of the R time (data not shown) and MTG to baseline values (Figure 7C-D).

Multiple dose simulations
To further verify the pharmacokinetics of N9-GP in a larger animal model and to generate a dataset sufficient for reliable pharmacokinetic modeling, clearance studies were performed in mini-pigs. As shown in Figure 7E, the variation between animals (n=3) was small and the elimination profiles for N9-GP and rFIX declined in a biphasic manner that was well-described by a 2-compartment model. The estimated terminal half-life of N9-GP was 76 hours and thus significantly longer than rFIX (16 hours). Based on the fitted parameters, plasma profiles were simulated in a multiple dose setting assuming a fixed dose level of 0.2 mg/kg given at regular intervals. This amount corresponds to 38 and 50 IU/kg N9-GP and rFIX, respectively, and represents a typical prophylactic dose. For rFIX, a trough of 1% of the normal level was observed 2 days after dosing, which mirrors the typical 2-3 times weekly dosing regimen in HB patients. In comparison, the plasma level of N9-GP was 4% after 1 week and 1% 2 weeks after administration (Figure 7F).

Discussion
PEGylation is a well-established strategy for extending the plasma half-life of proteins. The application of classic nonspecific means of attachment are, however, often associated with an unwanted loss of activity. To circumvent this problem, we employed enzymatic glyco-conjugation to direct PEGylation to the native N-glycans of FIX which are attached to the activation peptide and remote from known interaction sites of functional importance. The attached PEG moiety is thus present only on the circulating zymogen form and removed on its activation. Confinement of PEG to the activation peptide was confirmed by peptide mapping and corroborated by functional characterization demonstrating unaltered enzymatic properties of the activated forms of N9-GP and N9 (Figure 2C-D) including normal reactivity toward the principal plasma inhibitor antithrombin III (L.C.P., unpublished results, February 10, 2011). These results indicate that, once activated, N9-GP retains full activity and is subjected to normal physiologic regulation.

The ability of FIX to exert its function is dependent on its conversion to the activated form. FXIa and FVIIa-TF both recognize FIX by extended interactions characterized by $K_a$ values in the low nanomolar range (Table 1), which could compensate for steric hindrance by the PEG. Indeed, activation of N9 by FXIa was found to be insensitive to glycoPEGylation, while the membrane-dependent activation by the FVIIa-TF was reduced by approximately 50% primarily as a result of an increased $K_a$ (Figure 2A-B). Interestingly, no significant differences in the efficacy and potency of N9-GP and rFIX were observed in whole blood from HB patients (Figure 4) or in the tail-bleeding model in HB mice (Figure 5) although both models exhibit TF dependency. Thus, despite the slightly slower rate of activation, N9-GP appears to retain near normal specific activity and the ability to stop bleeds with the same efficiency as rFIX suggests that it may be efficacious for acute treatment of bleeding episodes.

The mechanisms by which FIX is cleared from the circulation have been addressed in a few studies. High-affinity binding sites associated with the vascular endothelium have been implicated in the recovery and initial disappearance of FIX. In addition, uptake by the vascular endothelium or other cell types expressing the neonatal Fc receptor (FcRn) has been demonstrated by genetic fusion of FIX to the Fc fragment of IgG. Using this concept, Peters et al obtained a 3- to 4-fold extension of FIX half-life in mice that was dependent on endogenous FcRn expression, and similar half-lives of approximately 47 hours were determined in monkey and HB dog. The pharmacokinetic studies conducted in the present work are in accord with published results on rFIX and furthermore demonstrate a significant beneficial effect of glycoPEGylation on the circulation time of FIX. The half-life of N9-GP was found to increase according to body weight from 41 hours in mice to 76 and 113 hours in mini-pig and dog, respectively (Table 2). The differential impact of PEG and Fc-fusion on FIX half-life in larger animal models suggests that multiple pathways contribute to the clearance of FIX and that PEG may act by distinct or additional retention mechanisms than previously used to prolong FIX survival in the circulation. Importantly, the longer circulation time of N9-GP compared with rFIX was evident as an extended hemostatic protection in HB mice and normalization of coagulation parameters in HB dogs (Figures 6, 7B-D). Accordingly, several lines of evidence suggest that N9-GP may support a considerably longer dosing interval than feasible with existing therapy, a conclusion which is further supported by multiple dose simulations using the plasma profiles obtained in mini-pig as a model for human pharmacokinetics. At a standard prophylaxis dose, the predicted 1% trough level of N9-GP was not reached until 2 weeks after administration compared with 2 days for rFIX (Figure 7F).

The lower in vivo recovery of rFIX compared with plasma-derived products is well documented in humans and animal models and is compensated for in current dosage recommendations. The difference has been ascribed to variations in posttranslational modifications although no definitive results have been presented. Interestingly, an up to 2-fold higher in vivo recovery of N9-GP compared with rFIX was found across species (Table 2). Considering the structural similarity of N9 and rFIX, this difference is most likely related to effects of carbohydrate modification, that is, PEGylation or the subsequent capping with sialic acid. Incomplete sialylation of the N-linked carbohydrates may target FIX for hepatic degradation and has been proposed as one cause of the lower recovery. The fact that N9 is capped with sialic acid as the last step in the glycoPEGylation process could protect N9-GP from this pathway and consequently increase recovery. Another contributing factor may relate to the presence of high-affinity binding sites for FIX on collagen IV associated with the vascular endothelium. Cheung et al localized the binding site on FIX to the Glu domain and a FIX variant (K5A) devoid of collagen IV binding was shown by Gui et al to exhibit increased recovery and accelerated plasma clearance in mice. Endothelial cell binding
by N9-GP was addressed in vitro and found to be reduced by approximately 20-fold ($K$, 48nM) compared with N9 and rFIX which bound with similar high affinities (Figure 3). As the plasma levels achieved after infusion are in the range of the estimated dissociation constant for N9-GP, it is conceivable that the reduced affinity would allow for a greater fraction of N9-GP to remain in circulation in the initial phase after administration. Pre-occupancy of vascular binding sites by endogenous FIX may reduce the contribution of this pathway to initial FIX disappearance and is consistent with the only marginally higher in vivo recovery of N9-GP compared with rFIX in nonhemophilic mini-pigs in contrast to HB mice and dogs (Table 2). A recent study suggested that lack of collagen binding was associated with a partial hemostatic defect in models involving exposure of vascular collagen including ferric chloride and tail transection models. It is presently not clear whether the preserved efficacy of N9-GP in the same models is explained by adequate residual affinity of N9-GP for collagen or by the higher in vivo recovery which may compensate for a reduced accumulation at vascular sites.

In summary, a novel releasable protraction concept has been developed that uses the ability to selectively target naturally occurring carbohydrates in the activation peptide of FIX without the need for amino acid modifications. This approach offers the additional advantages of retained biologic activity and normal physiologic activation of the activated form, while providing greatly prolonged pharmacokinetics and pharmacodynamics in several animal models. Clinical studies will show whether these properties translate into improved on demand treatment and effective bleeding prophylaxis in HB patients with a once-weekly or less frequent dosing schedule.

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Authorship

Contribution: H.O. and M.T. designed and performed research, analyzed data, and wrote the manuscript; J.R.B., L.H., L.C.P., A.A.P., T.E., F.M., M.B.H., P.K.H., T.N.K., J.M.P., M.D.A., H. Ahmadian, K.W.B., and M.L.S.C. designed and performed research, and analyzed data; M.E., B.B.S., H. Agersø, and S.E.B. designed research and analyzed data; K.K. provided clinical input and wrote the manuscript; and T.C.N. designed and performed research.


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References


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