is a necessary adjunct to prolonged, life-saving antileukemia therapy.

Conflict-of-interest disclosure: The author declares no competing financial interests.

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Sleeping Beauty: does ETP-ALL awaken later?

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In the hope of identifying key events that drive malignant transformation, researchers have been aiming to compare tumor cells with the corresponding pre-transformed cell population of origin. This approach is attractive because, once established, such key events can then become the focus of new therapies that interfere with the transformation process as it happens, thus preventing it.

Comment on Barquam-Vrieze et al, page 4646

In this issue of Blood, Berquam-Vrieze et al directly show that the biologic state at which a population of cells first experiences oncogenic mutations strongly influences the type of driver mutations selected for in the resulting tumor in a mouse model of T-cell acute lymphoblastic leukemia (T-ALL). T-ALL is a malignant clonal expansion of immature T cells that accounts for 10% to 15% of pediatric and 25% of adult ALL cases. While 80% of patients with T-ALL can currently be cured with intensive chemotherapy, a subset of cases has a poor prognosis. A recent study found early T-cell precursor (ETP) leukemia to be a high-risk subset of T-ALL characterized by an immature surface immune-phenotype, increased genomic instability, a high frequency of remission failure or hematologic relapse, and a distinct gene expression profile. Because of its ETP-like immune-phenotype and gene expression profile, ETP-ALL was proposed to originate from very early thymic immigrants. However, the current work of Berquam-Vrieze and colleagues introduces an alternative scenario.

The authors designed a forward-genetic screen, using a Sleeping-Beauty (SB)-based transposon system that allows Cre-dependent activation of the SB transposase. This allows transposon-induced mutagenesis to be initiated under the control of stage-specific Cre transgenes. Mutagenesis was induced either early in hematopoietic stem cells (Vav-SB) or at 2 subsequent developmental stages of thymocyte development: the CD4+CD8− double-negative (DN) compartment (Lck-SB), and the β-selected, CD4+CD8+ double-positive (DP) compartment (CD4-SB). Tumors develop as a result of transposon insertions into or near tumor suppressors or oncogenes, either activating or disrupting their expression. By mapping common insertion sites (CISs), that is, transposon insertion sites frequently selected for, one can thus identify bona fide driver mutations for T-ALL originating from different developmental states (see figure).

Berquam-Vrieze et al sequenced the insertion sites of 101 lymphomas and deduced a total of 94 driver mutations. As a proof of principle, 75% of these mutations have previously been described to cause cancer. On the other hand, 25% of these candidate driver mutations have no previous entry in the literature linking them to cancer development and thus invite follow-up studies.

Do differences or commonalities exist between the models as well as between mouse and human T-ALL? Interestingly, yes. When transposon-induced mutation was initiated in hematopoietic progenitors, under the control of Vav1-Cre, a large fraction (72%) of the resulting T-cell tumors had mutations in Notch1, a hallmark of human T-ALL. The second most common mutation affected the Ikaros locus encoding for the transcription factors Ikaros, which is a rare target for mutation in human T-ALL. However, Ikaros mutations have frequently been observed to cooperate with Notch1 in murine T-ALL models. Interestingly, loss of Ikaros function was recently...
shown to result in increased activated Notch1 levels through alternative promoter usage in the \textit{Notch1} locus.\textsuperscript{6,7} When transposon-induced mutations were initiated much later during T-cell development, however, Berquam-Vrieze and colleagues note very different mutation profiles in the CD4-Cre–dependent mouse strain:\textit{Notch1} mutations were no longer significantly overrepresented. Instead, the tumors now predominantly harbored mutations of \textit{Myc} and \textit{Stat5b}, often in a mutually exclusive combination with additional CISs affecting either \textit{Gfi1} or \textit{Whsc1}, \textit{Akt2}, \textit{Jak1}, and/or \textit{Sos1}. Inappropriate overexpression of \textit{Myc} drives many cancers and has also been implicated in human T-ALL as a downstream target of Notch1.\textsuperscript{8} \textit{Myc} is also required for mouse T-ALL induced by up-regulation of the Wnt/\beta-catenin signaling pathway.\textsuperscript{9}

The Lck-SB tumors, finally, segregated into 2 categories. In the one group, mutually exclusive \textit{Gfi1} versus \textit{Whsc1} mutations in combination with \textit{Myc} and \textit{Stat5b} were observed similar to CD4-SB tumors. In the other group, tumors carried \textit{Notch1} and \textit{Hes1} mutations similar to the Vav-SB model. This dichotomy might reflect a major change in cell state on completion of \beta-selection at the DN3 stage. Extensive chromatin reorganization associated with pre-TCR expression and signaling might result in very different genetic selection events becoming predominant. And a narrow window of time between Lck-Cre expression and \beta-selection at the DN3 stage might reflect a major change in cell state on completion of \beta-selection at the DN3 stage.

In conclusion, this study by Berquam-Vrieze and colleagues confirms the notion that the biologic state of origin could be reflected in the genetic selection events observed in the resulting tumor. The study further demonstrates the power of the Sleeping Beauty transposon system to derive valuable new animal models that provide insights into the etiology of cancers less likely to be gained from gain or loss of function of single genes in mice and impossible to be gained in humans.

\textbf{Conflict-of-interest disclosure: The authors declare no competing financial interests.}

\section*{REFERENCES}


\section*{MYELOID NEOPLASIA

Comment on Li et al, page 4509

\textbf{TET2: epigenetic safeguard for \textit{HSC}}

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The TET-family enzymes TET1, TET2, and TET3 influence DNA methylation by modifying 5-methylcytosine to 5-hydroxymethylcytosine (5hmC), 5-formylcytosine (5fC), and 5-carboxycytosine (5caC; see figure).\textsuperscript{1-3} Somatic loss-of-function mutations in \textit{TET2} are frequently observed in myeloid neoplasms.\textsuperscript{2,4} In this issue of \textit{Blood}, Li et al demonstrate that ablation of \textit{Tet2} alters the homeostasis and function of hematopoietic stem cells (HSCs) and induces leukemia in mice,\textsuperscript{3} as also reported by other groups.\textsuperscript{6,8}

\textbf{TET1, TET2, and TET3, members of the TET family of Fe\textsuperscript{2+} and \alpha-ketoglutarate–dependent dioxygenases, successively oxidise 5-methylcytosine (5mC) to 5-hydroxymethylcytosine (5hmC), 5-formylcytosine (5fC), and 5-carboxycytosine (5caC; see figure).\textsuperscript{1-3} 5caC can be excised by the base excision repair enzyme thymine–DNA glycosylase (TDG),\textsuperscript{9} suggesting that TET proteins cooperate with TDG to effect DNA demethylation. Thus, TET proteins have the potential to be important regulators of epigenetic status in the cell types in which they are expressed. TET2 is frequently mutated in myeloid malignancies. Microdeletions and copy number–neutral loss of heterozygosity (also called uniparental disomy) are recurrently observed in the \textit{TET2} locus in diverse myeloid malignancies including myelodysplastic syndromes (MDS), myeloproliferative neoplasms (MPNs), chronic myelomonocytic leukemia (CMML), and acute myeloid leukemia (AML). TET2 is expressed, and \textit{5hmC} is readily detectable, in hematopoietic stem/progenitor cells and mature blood subsets.\textsuperscript{2,6} Moreover, leukemia-associated \textit{TET2} mutations have been shown to impair the catalytic activity of TET2 and diminish \textit{5hmC} levels in cells from patients with MDS/MPN/CMML and AML, and shRNA-mediated depletion of TET2 in hematopoietic stem cells (HSCs) resulted in skewing toward myeloid differentiation.\textsuperscript{2}
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