A role for Blimp1 in the transcriptional network controlling natural killer cell maturation

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Natural killer (NK) cells are innate lymphocytes capable of immediate effector functions including cytokine production and cytotoxicity. Compared with B and T cells, the factors that control the peripheral maturation of NK cells are poorly understood. We show that Blimp1, a transcriptional repressor required for the differentiation of plasma cells and short-lived effector T cells, is expressed by NK cells throughout their development. Interleukin 15 (IL-15) is required for the early induction of Blimp1 in NK cells, with expression increasing in the most mature subsets of mouse and human NK cells. We show that Blimp1 is required for NK-cell maturation and homeostasis and for regulating their proliferative potential. It is also essential for high granzyme B expression, but not for most cytokine production and cytotoxicity. Surprisingly, interferon regulatory factor 4 (IRF4) and B-cell lymphoma 6 (Bcl6), 2 transcription factors crucial for the regulation of Blimp1 in B and T cells, are largely dispensable for Blimp1 expression in NK cells. T-bet deficiency, however, leads to attenuated Blimp1 expression. We have identified NK cells as the first hematopoietic cell type in which the IRF4-Blimp1-Bcl6 regulatory axis is not in operation, highlighting the distinct nature of the NK-cell gene-regulatory network. (Blood. 2011;117(6):1869-1879)

Introduction

Natural killer (NK) cells are innate lymphocytes specialized in cytokine production and cytotoxicity toward tumors and virus-infected cells. NK cells develop in the bone marrow from hematopoietic stem cells (HSCs) via lymphoid precursors.1 The earliest committed NK-cell precursors can be isolated from lineage marker–negative fetal thymus or adult bone marrow and express high levels of the interleukin 15 (IL-15)/interleukin 2 receptor (IL-2R) chain (CD122). NK-cell precursors then undergo maturation and functional diversification in the bone marrow, thymus, and a variety of peripheral organs including the liver, spleen, and lymph nodes.2 In the mouse, peripheral maturation in the spleen is characterized by the up-regulation of the markers CD43, Mac-1 (CD11b), CD94, and KLRG1, with concomitant down-regulation of CD27 and ekt.1,3,4 Mature Mac-1high/KLRG1low/CD27low NK cells are the dominant population in nonlymphoid organs (with the exception of the liver).3,7

In contrast to B and T cells, knowledge about the transcriptional circuitries controlling NK-cell development and maturation remains limited. Several factors, including E4BP4,8,9 Tox,10 and Id2,11 are required for the development of NK cells from early progenitors, whereas GATA3 is essential for thymic NK cells12 and modulates the function of mature NK cells.13 A second group of transcription factors, including Ets1,14 E74-like factor 4,15 interferon regulatory factor 2 (IRF2),16 and T-bet,17,18 is more specifically required for the later stages of NK-cell differentiation and function. Interestingly, a number of these factors, including Id2 and T-bet, also have important functions in CD8+ T-cell differentiation.19,20

Blimp1 (also known as Prdm1) is a pleiotropic transcription factor that plays crucial roles in the differentiation of plasma cells and CD8+ effector T cells.21-25 In both processes, Blimp1 is required for terminal differentiation and Blimp1 deficiency leads to loss of functional antibody-secreting cells and short-lived cytotoxic T cells. Moreover, in CD8+ T cells, Blimp1 is required for the optimal expression of cytotoxic molecules such as granzyme B and perforin and for the regulation of chemokine receptors such as CC-chemokine receptor 7 (CCR7), CCR5, and CCR6, but is dispensable for most cytokine production.24,25 Blimp1 is also broadly required for the maintenance of T-cell homeostasis, because mice that lack Blimp1 in all T cells or mice reconstituted with Blimp1-deficient fetal liver cells show an accumulation of activated T cells and develop immunopathology.22,26 In CD4+ T cells, Blimp1 has been suggested to attenuate Th1 differentiation by repressing the genes encoding T-bet and interferon (IFNγ).27 Blimp1 expression within B and T cells is controlled by 2 transcription factors, IRF4 and Bcl6. While Bcl6 and Blimp1 mutually repress the expression of each other in activated B and CD4+ T cells,28 IRF4 directly activates Blimp1 expression.29

We show that Blimp1 is expressed by NK cells and is vital for their peripheral maturation and tissue distribution, as well as for restricting their proliferative potential. In striking contrast to B and T lymphocytes, Blimp1 is induced in immature NK cells in an IL-15–dependent manner and maintained throughout the lineage. Blimp1 expression is increased in the most mature subsets of mouse and human NK cells and is further up-regulated by cytokines such as IL-12 and IL-21. Although Blimp1 is dispensable for most cytokine production and cytotoxicity, it is required...
for maximal granzyme B expression. Blimp1 expression is independent of IRF4 and Bcl6, despite their presence in NK cells, but relies on T-bet, suggesting fundamental differences in the transcriptional network of NK cells compared with other lymphocytes.

Methods

Mice

Blimp1/GFP reporter (Blimp1^{GFP+}) mice were generated and maintained on a C57BL/6 (Ly5.2+) background, as described previously. To generate Blimp1^{Gfp+} or Rag2^{−/−} Blimp1^{Gfp+} mice, fetal liver cells were isolated from E14.5 Blimp1^{Gfp+} or Rag2^{−/−} Blimp1^{Gfp+} embryos and intravenously injected into lethally irradiated Rag2^{−/−} Ly5.1 or Rag2^{−/−} γc^{−/−} recipients, respectively (1-3 × 10^6 fetal liver cells per recipient). Ifnar^{−/−} mice, Bcl6^{−/−} mice, and Tbx21^{−/−} mice have been described previously. Blimp1^{Gfp+} Ly5.1 mixed bone-marrow chimeras were generated by reconstituting lethally irradiated Ly5.1 recipients with a mixture of bone-marrow cells isolated from Blimp1^{GfpB} and wild-type Ly5.1 mice. Chimeric mice were analyzed after a minimum of 6 weeks after reconstitution. To allow for variation in the reconstitution frequency, data were normalized for the NK-cell compartment in the spleen. All mice were maintained on a C57BL/6 background at The Walter and Eliza Hall Institute. Animal experiments were undertaken according to the institution’s animal ethics committee guidelines and approval.

Antibodies and flow cytometry

The following anti–mouse monoclonal antibodies (mAbs) were used for flow cytometric analysis: NK1.1 (PK136), Ly5.2 (104), Ly5.1 (A20), CD27 (LG.7F9/H9253), CD43 (S7), Mac-1 (M1/70), ckit (2B8), T-cell receptor (TCR) β (H57-597), CD8α (53-6-7), CD44 (IM7), CD62L (Mel-14) IFNγ (XMG1.2), Ly49A (YE132), Ly49C (SW5E6), Ly49D (4E5E1), Ly49F (BHIF-719), Ly49H (3D10), and CD94 (18d3). Biotinylated mAbs were revealed with streptavidin Cy5 (Southern Biotech) or streptavidin PerCPCy5.5. Anti–human CD3 (SK7), CD56 (1870 KALLIES et al BLOOD, 10 FEBRUARY 2011) or streptavidin Cy5 (53-6-7), CD44 (IM7), CD62L (Mel-14) IFNγ (XMG1.2), Ly49A (YE132), Ly49C (SW5E6), Ly49D (4E5E1), Ly49F (BHIF-719), Ly49H (3D10), and CD94 (18d3). Biotinylated mAbs were revealed with streptavidin Cy5 (Southern Biotech) or streptavidin PerCPCy5.5. Anti–human CD3 (SK7), CD56 (1870 KALLIES et al BLOOD, 10 FEBRUARY 2011)

Cell isolation and culture

Unless stated otherwise, splenic NK cells were enriched using DX5 magnetic beads (Miltenyi Biotec) and cultured in IL-15 (50 ng/mL; R&D Systems) for 5 days. The cells were then used or recultured in the presence of IL-12 (5 ng/mL), IL-18 (20 ng/mL), or IL-21 (100 ng/mL), all from R&D Systems. For some experiments, IL-15-expanded NK cells were cultured in the presence of 20 μg/mL of anti–NK1.1, anti–NK2G2D, or anti–CD16 (24G.2) or various cytokine concentrations for 2 days, as described previously. The differentiation of NK cells from purified HSCs was carried out using OP9 and OP9-DL1 stromal cells as described previously. Lung lymphocytes were purified from single cell suspensions by centrifugation on Histopaque (1.077 g/mL; Sigma-Aldrich) for 20 minutes at 400g at room temperature.

Intracellular cytokine stains

DX5-enriched NK cells were stained for relevant surface molecules, fixed, permeabilized using the Cytofix/Cytoperm reagent (BD Biosciences), and costained for intracellular cytokines. Intracellular granzyme B content and IFNγ were measured after stimulation of the cells with 5 ng/mL of IL-12, 50 ng/mL of IL-18 or 50 ng/mL of phorbol myristate acetate (PMA), and 500 ng/mL of ionomycin for 5 hours in the presence of GolgiPlug, followed by subsequent surface-marker staining and permeabilization using the Cytofix/Cytoperm reagent (BD Biosciences).

Cytokine bead array

NK cells were expanded in IL-15 for 5 days before being transferred into IL-15, IL-12/IL-18, or IL-15/IL-21. After another 2 days in culture, cell numbers were determined and equal numbers of cells seeded in the same conditions. Supernatants were collected after an additional 16 hours and assayed for cytokines using the Bio-Rad Bioplex cytokine bead assay (Mouse 23-Plex Panel).

PCR analysis

Total RNA was prepared from flow cytometry–purified human NK cells using the RNeasy kit (QIAGEN). cDNA was synthesized from total RNA with random hexamers and SuperScript III reverse transcriptase (Invitrogen). Primer sequences were as follows: BLIMP1: 5'–GATGGCGGATGACTCTGTTG-3'; 5'–CTCGGTTGCTTTAGACTGCT-3'; HPRT: 5'-GTGATGAAAGGATGG-GAGG-3'; and 5'-ACTGCGTCCAAGGAAAG-3'.

Cytotoxicity assays

The cytotoxicity of NK cells was assessed using RMAS lymphoma cells transduced with either the control murine stem cell leukemia virus vector (RMAS-MSCV) or the MSCV vector expressing Rae1β (RMAS-Rae1b), as we have described previously.

In vivo tumor model

Groups of 4-7 Rag2^{−/−} Blimp1^{Gfp+} or Rag2^{−/−} Blimp1^{GfpB} mice were inoculated subcutaneously with RMAS-S tumor cells at the indicated doses, as described previously. Tumor growth was examined every second day. Recorded data are the means ± SEM of mice in each group.

Western blotting

Proteins were extracted with radioimmunoprecipitation assay buffer (2 × 10^7 cells/mL) and subjected to Western blotting according to standard techniques. The monoclonal antibody to Blimp1 (5E7) was described previously. Rat anti–mouse IRF4 (3E4) and Bcl6 (7D1) monoclonal antibodies were raised against a glutathione S-transferase fusion protein containing the carboxy-terminal 65-amino acid sequence of IRF4 and amino acids #261-386 of Bcl6, respectively, and were a gift from Lynn Corcoran (The Walter and Eliza Hall Institute of Medical Research, Parkville, Australia). Anti-β-actin (Santa Cruz Biotechnology) was used as a loading control.

Proliferation assay

Sorted TCRβ^{−/−} NK1.1^{CD49b+} NK cells were cultured for 2 days in a range of IL-15 concentrations before being pulsed with 1μCi/well of ^3H-thymidine for the last 8 hours of culture. Cells were harvested and ^3H-thymidine incorporation determined by scintillation counting.

In vivo proliferation assay

Purified wild-type (Ly5.1+) or Blimp1^{GfpB} (Ly5.2+) TCRβ^{−/−} NK1.1^{CD49b+} NK cells were cultured in IL-15 for 7 days before being mixed at a 1:1 ratio (total 2 × 10^6 cells), injected into Rag2^{−/−} γc^{−/−} hosts, and analyzed after 6 days by flow cytometry.

Statistics

Data were analyzed using a 2-tailed Student t test (paired or unpaired as appropriate). A P value less than .05 was considered significant.

Results

Blimp1 is constitutively expressed in NK cells and is up-regulated during maturation

To determine whether Blimp1 is expressed by NK cells, we made use of Blimp1/GFP reporter mice (Blimp1^{Gfp+}). Flow
cytometric analysis of NK cells revealed GFP fluorescence that was similar to that observed for the corresponding effector T-cell populations of the same mouse, indicating that Blimp1 was expressed in NK cells (Figure 1A-B). Blimp1/GFP was detected in all NK cells independent of their localization, including NK cells from the bone marrow, lymph nodes, spleen, blood, liver, and lung (Figure 1 and data not shown). In B and T cells, Blimp1 expression was closely linked to terminal differentiation of effector cells. NK cells undergo significant phenotypic and functional changes during their maturation in the periphery, and we speculated that Blimp1 might be up-regulated during peripheral NK-cell differentiation. Consistent with this idea, Blimp1/GFP was highest in the Mac-1-high population, the most mature NK-cell subset,\(^1\) independent of their anatomical location (Figure 1C). In agreement with this finding and with data recently published,\(^2\) mature human CD16\(^+\) NK cells exhibited distinctly higher BLIMP1 expression than their more immature CD16\(^-\) counterparts, suggesting that BLIMP1 is also up-regulated during maturation of human NK cells (Figure 1D).

Cytokines have been shown to be important modulators of NK-cell activation, function, and maturation.\(^1\) In agreement with this notion, we found that IL-12, a combination of IL-12 and IL-18, and IL-21, cytokines known to promote NK-cell differentiation,
were potent in inducing Blimp1 expression in NK cells higher than the constitutive level maintained by IL-15 alone (Figure 1E-F). Whereas these cytokines induced equivalent Blimp1/GFP over a broad range of concentrations, cross-linking of NK-cell–activating receptors did not result in any substantial up-regulation of Blimp1/GFP (supplemental Figure 1, available on the Blood Web site; see the Supplemental Materials link at the top of the online article). Interestingly, murine NK cells predominantly expressed the full-length Blimp1 protein (Figure 1F), unlike human NK cells, which were recently shown to express 3 distinct isoforms of BLIMP1.38

**Blimp1 expression in NK cells is IL-15–dependent**

Our data so far indicate that NK cells constitutively express Blimp1, which can be further up-regulated during maturation. This is strikingly different from the situation in B and T cells, in which the Blimp1 protein is restricted to terminally differentiating effector cells.23 To determine at what stage Blimp1 expression is initiated in the NK-cell lineage, we sorted HSC-enriched populations from Blimp1<sup>gfp</sup> and control mice and cultured them on OP9 cells in the presence of IL-15 to induce NK-cell development, or on OP9-DL1 cells in the presence of IL-7 to induce T-cell development.35 HSCs did not express Blimp1 (data not shown); however, when cultured under NK-cell–inducing conditions (OP9/IL-15), they gave rise to Blimp1/GFP<sup>+</sup>NK1.1<sup>+</sup>CD122<sup>+</sup> cells (Figure 2A R1). In contrast, the NK1.1<sup>+</sup>CD122<sup>+</sup> NK-cell progenitors (R2) were Blimp1/GFP negative in the same culture, as were NK1.1<sup>+</sup>CD122<sup>+</sup> cells (R3). Transient NK-cell differentiation is also observed when HSCs are cultured in conditions favorable for T-cell differentiation (OP9-DL1/IL-7).35 The NK1.1<sup>+</sup>CD122<sup>+</sup> NK cells induced in these conditions were Blimp1/GFP negative (Figure 2B R1 and data not shown), suggesting that IL-15 signals are required for the induction of Blimp1 expression.

**Blimp1 is required for NK-cell maturation and homeostasis**

To determine the functional relevance of our findings, we generated chimeric mice using lethally irradiated Rag<sup>2<sup>−/−</sup></sup>γ<sup>c<sup>−/−</sup></sup> mice reconstituted with Blimp1<sup>+/gfp</sup> fetal liver cells (termed Blimp1<sup>+/gfp</sup> or Blimp1-deficient mice thereafter) or with control cells. As described previously, this approach results in mice lacking functional Blimp1 in all hematopoietic cells including NK cells.30 This strategy was necessary because Blimp1 deficiency is embryonic lethal30 and a reliable Cre-lox system for the conditional ablation of genes in NK cells is lacking. Examination of Blimp1<sup>+/gfp</sup> and control mice revealed that while the proportion of Blimp1-deficient NK cells was increased in the bone marrow and reduced in the spleen compared with heterozygous or wild-type controls (Figure 3A), the absolute numbers of splenic NK cells were largely unchanged (1.78 ± 0.3 × 10<sup>6</sup> cells in +/+ , 1.72 ± 0.6 × 10<sup>6</sup> in Blimp1<sup>+/gfp</sup>). Therefore, Blimp1 was not required for the generation of normal numbers of NK cells.

Because a deficiency of Blimp1 in the hematopoietic system leads to immunopathy associated with an accumulation of activated T cells and an altered cytokine environment,22,26 we wished to examine Blimp1-deficient NK cells in the absence of other lymphocytes. To do this, we generated chimeric mice from Rag<sup>2<sup>−/−</sup></sup>Blimp1<sup>+/gfp</sup> fetal liver cells, which, due to the lack of T and B cells, remained healthy and showed no immunopathology (data not shown). High cell-surface expression of Mac-1 and CD43 are commonly used to identify the most mature mouse NK cells4 and, interestingly, the expression of both molecules was markedly reduced on splenic NK cells in the absence of Blimp1 (Figure 3B left). More recently, CD27 in conjunction with KLRG1 and Mac-1 was used to describe NK-cell maturation. Spleen CD27<sup>+</sup> NK cells first up-regulate Mac-1 before differentiating through Mac-1<sup>hi</sup>CD27<sup>hi</sup>KLAR1<sup>−</sup> and Mac-1<sup>hi</sup>CD27<sup>−</sup>KLAR1<sup>+</sup> stages.5,6,7 CD27 expression was uniformly high in the bone marrow of Blimp1-deficient and control mice, whereas Blimp1-deficient spleens showed a reduction in the frequency of the Mac-1<sup>hi</sup>CD27<sup>−</sup>KLAR1<sup>+</sup> mature NK cells (Figure 3B-C), with the proportion of KLRG1<sup>+</sup> splenic NK cells decreasing from 57.2 ± 4.7% in Blimp1<sup>+/+</sup> to 28.7 ± 3.8% in Blimp1<sup>+/gfp</sup> mice (P = .002, each n = 5), indicating that Blimp1 was required for the peripheral maturation of NK cells.
To rigorously test if the maturation phenotype and the altered homeostasis of Blimp1-deficient NK cells was indeed intrinsic to NK cells or was a result of an altered cellular composition or cytokine environment in Blimp1-deficient mice, we generated chimeric mice by reconstituting lethally irradiated Ly5.1 mice with a mixture of congenically marked bone marrow from wild-type (Ly5.1+/H11001) and Blimp1gfp/gfp (Ly5.2+/H11001) mice. We have shown before that this strategy significantly delays the onset of pathology caused by Blimp1-deficient T cells, thereby allowing a comparison of the homeostasis and tissue distribution of Blimp1-deficient and control NK cells under the same conditions. Examination of NK cells in various organs of these mice revealed that Blimp1-deficient NK cells were present at significantly increased frequencies in bone marrow and lymph nodes but were dramatically reduced in livers and lungs compared with their wild-type competitors (Figure 3D). Blimp1-deficient NK cells showed a normal repertoire of NK-cell receptors (supplemental Figure 2) while maintaining their immature KLRG1+/H11002 phenotype (Figure 3E), indicating that the requirement of Blimp1 for the correct tissue localization and peripheral maturation of NK cells was cell intrinsic.
Blimp1 is dispensable for most NK-cell cytokine production and cytotoxicity

To test the functional properties of Blimp1-deficient NK cells, we first examined granzyme B and IFNγ levels in NK cells from mixed bone-marrow chimeric mice (Blimp1<sup>gfp/gfp</sup>:Ly5.1). Similar to our findings in CD8<sup>+</sup> effector T cells, the level of granzyme B expression was significantly decreased in Blimp1-deficient NK cells (Figure 4A). In contrast, the level of IFNγ produced by Blimp1<sup>gfp/gfp</sup> NK cells was only mildly reduced compared with control cells after activation with IL-12 and IL-18, and was unchanged after strong ex vivo stimulation using PMA and ionomycin (Figure 4B). This was surprising and in contrast to data obtained from mouse CD4<sup>+</sup>T cells and human NK cells, in which Blimp1 was found to directly repress Ifng<sup>27</sup>; however, these data agree with our findings in CD8<sup>+</sup> effector T cells, which in the absence of Blimp1 display unaltered IFNγ secretion.<sup>24</sup>

Because IL-12 and IL-21 have been shown to induce the functional maturation of NK cells and also to up-regulate Blimp1, we cultured Blimp1<sup>gfp/gfp</sup> and wild-type NK cells with these factors and measured NK-cell functions such as cytokine secretion and cytotoxicity. Surprisingly, in all conditions tested, Blimp1-deficient NK cells secreted similar amounts of IFNγ and IL-10 (Figure 4C) and showed a similar ability to lyse target cells (Figure 4D) compared with the wild-type controls, suggesting that Blimp1 is not required for most NK-cell effector functions.

Blimp1 controls the proliferative potential of NK cells

Given the crucial roles of Blimp1 in late B- and T-cell differentiation, the finding that NK cells were broadly functional in the absence of Blimp1 was intriguing and led us to test NK-cell functions in vivo. To this end, we injected RMAS lymphoma cells subcutaneously into Rag2<sup>−/−</sup>Blimp1<sup>gfp/gfp</sup> and control Rag2<sup>−/−</sup> mice and measured tumor burden. Interestingly, Blimp1-deficient NK cells were superior to their wild-type counterparts in their ability to control the growth of the tumor cells in vivo (Figure 5A). Because the cytotoxic function of freshly isolated NK cells per se did not seem to be altered (Figure 5B), this result suggested that NK cells in the absence of Blimp1 showed enhanced proliferation, survival, or recruitment to the site of tumor growth. To determine...
indicating that both factors were dispensable for Blimp1/GFP in the absence of either IRF4 or Bcl6 (Figure 6B), compartment. Surprisingly, NK cells expressed normal amounts of ground and monitored Blimp1/GFP expression in the NK-cell

Two transcription factors that directly regulate expression in NK cells is independent of IRF4 and Bcl6

Blimp1 regulates NK-cell homeostasis and function in vivo by promoting their differentiation and restricting their proliferation.

**Blimp1 expression in NK cells requires T-bet**

The maturation defect we have found in Blimp1-deficient NK cells resembles those previously reported for mice deficient in T-bet (encoded by Tbx21). Moreover, Blimp1 has been shown to directly repress Tbx21 in CD4+ T cells and to modulate the expression of a related T-box transcription factor, Eomesodermin (Eomes), in CD8+ T cells. Thus, it was possible that the defective NK-cell maturation in the absence of Blimp1 was due to altered Tbx21 or Eomes expression. However, expression of both factors in Blimp1-deficient and control NK cells was similar, with only a relatively minor increase in Eomes in Blimp1-deficient NK cells (mean fluorescence index 7602 in Blimp1gfp/gfp; P = .008), suggesting that Eomes may compensate for some T-bet functions.

We also determined whether Blimp1 expression in NK cells depends on T-bet. Intriguingly, Tbx21-/- Blimp1gfp/gfp NK cells expressed a markedly reduced amount of Blimp1/GFP (Figure 7B), suggesting that T-bet acts upstream of Blimp1 in NK cells. In agreement with earlier reports, T-bet-deficient NK cells showed a distinct block in peripheral maturation at the
stage that was similar to the defects observed in Blimp1-deficient NK cells (Figure 7C). The notion that in NK cells, T-bet but not Bcl6 acts in the same transcriptional network as Blimp1 was further supported by the finding that both T-bet– and Blimp1-deficient NK cells expressed high amounts of ckit, a marker of immature NK cells,4 and showed a similar reduction of granzyme B expression, whereas Bcl6-deficient NK cells were indistinguishable from wild-type cells (Figure 7D). Interestingly, neither T-bet nor IRF4 was required for the IL-12– or IL-21–induced up-regulation of Blimp1 in cultured NK cells, further indicating that the expression of Blimp1 in NK cells differs fundamentally from that in T or B cells (supplemental Figure 4).

Discussion

In this study, we identify Blimp1 as an important player in the transcriptional network governing NK-cell differentiation. Our data demonstrate that NK cells express Blimp1 constitutively from a very early point in their development. We further show that Blimp1 levels increase during functional maturation of NK cells and that Blimp1 controls the differentiation and homeostasis of NK cells. Surprisingly, Blimp1 expression in NK cells is independent of IRF4 and Bcl6, 2 well-characterized regulators of Blimp1 in other hematopoietic lineages.

Constitutive expression of Blimp1 is a unique feature of NK cells among the hematopoietic lineages and is in sharp contrast to B cells and CD8+ T cells, in which Blimp1 expression marks terminally differentiated effector cells, including plasma cells and short-lived cytotoxic effector T cells.20 The unique expression pattern of Blimp1 may be reflective of the innate nature of NK cells, which show effector functions such as cytotoxicity and cytokine production throughout their development without the need for extensive proliferation and discrete peripheral differentiation stimuli.

NK cells crucially depend on IL-15, which controls their proliferation and survival in the periphery.11,22 Our results indicate that IL-15 signals are also required for the induction of Blimp1 in early NK-cell development. This again contrasts with findings in CD8+ T cells, in which IL-15 down-regulated Blimp1 expression.43 Interestingly, however, the constitutive Blimp1 expression in NK cells is paralleled in memory CD8+ T cells,24 and both NK cells and CD8 memory T cells require IL-15 for their survival and turnover.44 Thus, IL-15 may act as a modulator of Blimp1 expression in NK cells and memory CD8+ T cells.
that balances Blimp1 effects on cytotoxicity with those that limit the proliferation and survival of effector cells. The baseline level of Blimp1 was also increased by IL-12, IL-18, and IL-21, cytokines that induce NK-cell maturation. This is again analogous to other lymphocytes in which several cytokines, including IL-21, promote Blimp1 expression.\textsuperscript{29} Interestingly, activation of NK cells through antibody-mediated cross-linking of activating receptors did not significantly enhance Blimp1 expression, suggesting that these receptors function in a manner similar
Peripheral maturation of NK cells is known to be a multistep process. The most immature NK cells in the mouse spleen express the marker combination Mac-1^{hi}CD27^{−}KLRG1^{+}. During maturation, these cells first lose KLRG1 expression before up-regulating KLRG1 and silencing CD27 to undergo their final maturation into Mac-1^{hi}CD27^{−}KLRG1^{+} cells. Blimp1-deficient NK cells had altered expression of several of these maturation markers, including a lower amount of Mac-1, and were impaired in the transition from the CD27^{−}KLRG1^{−} to the CD27^{−}KLRG1^{+} stage. This was in agreement with the hyperresponsiveness of Blimp1-deficient NK cells to IL-15 and their enhanced tumor clearance capacity, because it is known that the CD27^{−}KLRG1^{+} subset of wild-type NK cells has the greatest proliferative potential and superior effector potential. In addition to its role in peripheral NK-cell maturation, Blimp1 was also required to maintain the appropriate homeostatic numbers of NK cells in nonlymphoid organs such as the lung and liver. Failure to populate the lung may be a consequence of the splenic maturation defect, because CD27^{−}KLRG1^{+} NK cells predominate in the lungs of wild-type mice; however, Blimp1 is also known to regulate the expression of several chemokine receptors in CD8^{+} T cells, including CCR5, CCR6, and CCR7, the modulation of which is important for tissue homing. Further analysis is required to investigate the Blimp1-dependent factors that control NK-cell homing to nonlymphoid tissues.

Several transcription factors are known to control the peripheral maturation of NK cells, including GATA-3,13 Irf2,16,44 and T-bet.17,18 Of these, the T-bet-deficient phenotype most resembles that of Blimp1 mutant NK cells in that they are blocked at a CD27^{−}KLRG1^{+} state while having relatively normal expression of NK-cell receptors such as those in the Ly49 family (17 and Figure 7). In keeping with the similarities of the phenotypes, Blimp1 expression was markedly reduced in the absence of T-bet, suggesting that in contrast to CD4^{+} T cells, in which Tbx21 may be directly repressed by Blimp1, T-bet was required for Blimp1 expression in NK cells. Interestingly, Tbet and Blimp1 are both necessary for normal granzyme B production by both NK cells and CD8^{+} T cells. The functions of these transcription factors, however, are not identical, because a deficiency in T-bet leads to a more severe block in NK-cell maturation and plays a modest role in NK-cell cytotoxicity and IFNγ production, and we found no role for Blimp1 in these effector functions. This was the case despite the reduced expression of granzyme B, which contributes to target killing but is not essential for this process.

During the revision of this manuscript, a study describing BLIMP1 expression and function human NK cells was published. This study showed that, in contrast to the mouse homolog, human BLIMP1 is present in 3 isoforms in NK cells. In agreement with our data, Smith et al also found that human BLIMP1 is induced by cytokines and is present in the CD16^{+} NK-cell subset in the peripheral blood. However, in contrast to our data, Smith et al did not observe constitutive expression of BLIMP1 in human NK cells. Whether this was due to a lower sensitivity in the detection method or represents a real biologic difference is presently unclear and requires further study. Smith et al also found a role for BLIMP1 in limiting the production of effector cytokines such as IFNγ and tumor necrosis factor in human NK cells through direct binding to both loci. This finding agrees with the proposed role of Blimp1 in repressing Ifng in mouse CD4^{+} T cells, but contrasts with our finding of normal IFNγ expression in mouse NK cells in the absence of Blimp1. Independence of IFNγ expression from Blimp1 was also found in CD8^{+} T cells after viral infection, suggesting that the regulation of Ifng by Blimp1 displays cell-type and species specificity, potentially at the level of the chromatin structure of the Ifng locus. It is also important to note that in human NK cells and mouse CD4^{+} T cells, Blimp1 only modulates expression of Ifng, indicating that other factors are also involved in controlling the effector cytokine response.

Our analysis of transcription factors that are intimately linked to the regulation of Blimp1 expression in B and T cells revealed that the transcriptional control of Blimp1 in NK cells is distinct from other lymphocyte lineages. IRF4, which binds at multiple places in the Blimp1 locus and is essential for Blimp1 induction in activated T cells and plasma cells, was dispensable for Blimp1 expression in NK cells. Another transcription factor implicated in Blimp1 regulation, Bcl6, was coexpressed with Blimp1 in NK cells, and deficiency of Bcl6 led only to a subtle increase in the Blimp1 and granzyme B levels. The coexpression of these 2 factors is surprising, because Bcl6 and Blimp1 have been shown to antagonize each other’s expression in all hematopoietic cell types yet examined, including germinal center B cells, follicular helper T cells, and, most recently, osteoclasts. Therefore, our data indicate that the regulation of Blimp1 expression is fundamentally different in NK cells compared with B and T lymphocytes. While neither IRF4 nor Bcl6 was required for Blimp1 expression, loss of T-bet led to attenuated Blimp1 expression and abrogated NK-cell maturation. This also highlights the unique nature of the genetic network controlling the NK-cell lineage, because Blimp1 and T-bet are independently regulated in differentiating CD8^{+} effector and memory T cells, while in CD4^{+} T cells Blimp1 is proposed to act upstream of T-bet. Remarkably, neither T-bet nor IRF4 was required for cytokine-mediated up-regulation of Blimp1. T-bet is known to up-regulate the IL12rb gene and could mediate the IL-12 induction of Blimp1 expression, whereas IRF4 is essential for the IL-21-induced Blimp1 expression in B and T cells; however, our loss-of-function experiments showed that these factors were dispensable for the up-regulation of Blimp1 in NK cells mediated by IL-12 and IL-21, respectively. One explanation for the T-bet–independent expression of Blimp1 in the presence of IL-12 is that Eomes levels were increased in Tbx21^{−/−} NK cells and thus potentially compensated for the loss of T-bet. Further studies are now required to investigate how Blimp1 is regulated in NK cells. In particular, it will be important to determine how the multiple signals that mature NK cells integrate, induce, and fine-tune Blimp1 expression, ultimately promoting NK-cell maturation and tissue homeostasis.

Acknowledgments

We thank Y. Hayakawa, R. Thong, S. Sterle, J. Brady, and K. Elder for technical assistance; T. Mak for providing the Ifng^{−/−} mice; L. Corcoran for antibodies; and C. Vinuessa for the Bcl6^{−/−} mice.

This work was supported by grants from the National Health and Medical Research Council (NHMRC) of Australia (to A.K., S.C., D.M.T., M.J.S., and S.L.N.); A.K. was supported by an Australian Research Council (ARC) Future Fellowship and a Leukemia & Lymphoma Society Fellowship; S.C. by an ARC Discovery Fellowship; N.D.H. by a Human Frontiers Science Program Long-term Fellowship; M.J.S. by an NHMRC Australia Fellowship; D.M.T. by NHMRC fellowships; and S.L.N. by a Pfizer Australia Research Fellowship.

Authorship

Contribution: A.K., S.C., N.D.H., N.J.B., and M.J.S. designed and performed the research; A.K., M.J.S., D.M.T., and S.L.N. analyzed the data; and A.K. and S.L.N. wrote the paper.

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Conflict-of-interest declaration: The authors declare no competing financial interests.

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References


A role for Blimp1 in the transcriptional network controlling natural killer cell maturation

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