Long-lasting antitumor protection by anti-CD20 antibody through cellular immune response

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The anti-CD20 monoclonal antibody (mAb) rituximab has been used successfully for lymphoma therapy for more than 10 years. Although several direct mechanisms by which anti-CD20 mAbs act have been characterized in vitro, their specific role in clinical efficacy is still debated. Little is known about the possible antitumor immune response that they may induce in patients, despite clinical data suggesting a “vaccinal” effect. We show here that an initial treatment with anti-CD20 induces protection against human CD20-expressing tumor cells and allows immunocompetent mice to survive tumor challenge. This long-lasting protection requires the presence of the Fc portion of the anti-CD20 mAb and is achieved through the induction of a cellular immune response. Only CD4+ cells were needed at the beginning of the treatment, but both CD4+ and CD8+ cells were required after tumor challenge to achieve protection. Finally, we show that interleukin-2 treatment, given after tumor challenge, improves the overall survival rate, compared with that obtained by anti-CD20 treatment alone. These findings demonstrate that anti-CD20 mAbs exert therapeutic effects through the induction of an adaptive cellular immune response, aside from any direct mechanisms involving effectors from innate immunity. (Blood. 2010;116(6):926-934)

Introduction

The anti-CD20 monoclonal antibody (mAb) rituximab has revolutionized the management of patients with follicular lymphoma (FL) and aggressive B-cell non-Hodgkin lymphoma (B-NHL) over the past decade.1-4 Other anti-CD20 mAbs, engineered to improve their effector functions, are currently being tested in clinical trials and may prove to be more effective therapeutic antibodies.5-8 This engineering has been based on studies that deciphered the mechanisms of action of these mAbs,9,11 principally programmed cell death induction, complement-dependent cytotoxicity, antibody-dependent cell cytotoxicity, and phagocytosis of mAb-opsonized target cells through receptors for the Fc portion of IgG (FcγRs).

In contrast, the long-term effects of anti-CD20 therapy on the host adaptive response have received little attention. Early in vitro studies indicated that rituximab promotes uptake of lymphoma cell-derived antigens by dendritic cells, thus generating tumor-specific cytotoxic T lymphocytes.12,13 A recent work showed that anti-CD20 mAb therapy synergizes with DC-based vaccination for long-lasting protection of mice with established CD20+ tumors.14 Two clinical observations suggest that rituximab therapy elicits a “vaccinal” effect that might prolong patient survival.9 First, time to progression in B-NHL patients who responded to rituximab and then relapsed (therefore, already a highly selected group) is longer after this second course of rituximab than after the first,15 possibly because the second course reduces the tumor load to a level that the patient’s immune system can control by reactivating adaptive immune effectors and blocking immune suppression. Second, the clinical response rate in rituximab-responding B-NHL patients is higher when rituximab is administered as maintenance therapy.16 We hypothesize, therefore, that a longer period of tumor antigen exposure to immune effectors during maintenance therapy induces antitumor adaptive immune responses sufficient to increase the clinical response rate but not strong enough to ensure full protection from relapse.

Interestingly, the phase 2 clinical studies evaluating the combination of immunomodulatory cytokines with rituximab for treating B-NHL patients also suggest an anti-CD20 vaccinal effect.17,18 Combining interferon-α with rituximab in patients with indolent lymphoma seems to improve both the quality and duration of the clinical response.17 Similarly, combining granulocyte-macrophage colony-stimulating factor (GM-CSF) with rituximab for patients with relapsing or progressive FL produces high clinical response rates and a tolerable safety profile;18 these authors suggest that the improved efficacy over rituximab alone is related to the increased number of cells expressing FcγRs (ie, monocytes, granulocytes, and dendritic cells).19 Mouse models have also demonstrated the role of FcγR-expressing cells in the clinical efficacy of IgG therapeutic antibodies. Studies of tumor-bearing mice show the importance of FcγRs in the DC-dependent induction of antitumor immune responses by immune complexes.19,20 Moreover, FcγR polymorphisms affect human clinical response to rituximab.21-23 This better clinical efficacy may be linked to enhanced antigen presentation by FcγR-expressing antigen-presenting cells.

A murine virus-induced tumor model has demonstrated the effect of mAb therapy on the host immune system by showing that the antimitastatic action of a mAb directed against the Friend leukemia virus envelope gp70 antigen depends on both the antibody Fc region and CD4+ cells.24 Furthermore, anti-gp70–treated mice produce antitumor antibodies that elicit tumor protection in naïve recipient mice. The involvement of these mechanisms in the therapeutic activity of anti-CD20 mAb in vivo nonetheless remains unclear. We therefore investigated whether an anti-CD20 mAb induction regimen could trigger an adaptive antitumor...
immune response in immunocompetent mice. We show here that anti–human CD20 (huCD20) therapy provided long-lasting protection against huCD20+ tumor cells, allowing mice to survive to a subsequent tumor challenge. This protective Fc-dependent effect was achieved by inducing a cellular immune response that required the presence of both CD4+ and CD8+ cells, although only CD4+ cells were needed after the initial anti-huCD20 regimen. We also show that interleukin-2 (IL-2) therapy, administered after the second tumor injection, enhanced the overall survival rate of the mAb-treated mice after this injection.

Methods

Mice

Eight-week-old female C57Bl/6 mice were purchased from Charles River Laboratories. Eight-week-old female CD8 KO C57Bl/6 mice were obtained from Dr Jean-Charles Guéry (Inserm U563, Toulouse, France).

Cells

The mouse thymoma EL4 cells (EL4-wt) were purchased at ATCC. EL4 cells expressing human CD20 (EL4-huCD20) were kindly provided by Pr Hervé Watier (Françoise Rabelais University, Tours, France). The CAT-13.6E12 hybridoma cell line was purchased from the DSMZ. This hybridoma cell line, established by Dr Martin Hadam (Kinderklinik-Medizinische Hochschule, Hanover, Germany), produces a typical type I anti-CD20 mAb. These cells were used to produce the anti-huCD20 CAT-13 IgG2a mAb. The stability and level of huCD20 expression on EL4-huCD20 cells were assessed by indirect immunofluorescence assays (FACSCalibur; BD Biosciences) with the CAT-13 mAb and phycoerythrin (PE)–conjugated F(ab')2 goat anti–mouse (GAM) IgG (Fc-specific; Southern Biotechnology).

Antibody and F(ab')2 fragment production and purification

CAT-13.6E12 culture supernatants were obtained by centrifugation at 270g and cleared by centrifugation at 4800g for 10 minutes at 4°C followed by 0.22-µm filtration (Millipore). CAT-13 mAb was purified by Protein-A affinity chromatography (rProtein A Sepharose fast flow; GE Healthcare). CAT-13 F(ab')2 fragments were produced by pepsin digestion of the CAT-13 mAb, with immobilized pepsin (Pierce Chemical) according to the manufacturer's recommendations. Fragments containing Fc and trace amounts of undigested IgG were then removed by adsorption onto Protein A–coupled beads (rProtein A Sepharose fast flow; GE Healthcare). The purity of the CAT-13 F(ab')2 fragments and the mAb was then assessed by 10% sodium dodecyl sulfate–polyacrylamide gel electrophoresis (Invitrogen). The binding of CAT-13 mAb and F(ab')2 fragments to EL4-huCD20 cells was measured by indirect immunofluorescence assays (FACSCalibur; BD Biosciences) with PE-conjugated F(ab')2; GAM IgG (Fc-specific; Beckman Coulter), PE-conjugated F(ab')2; GAM IgG (Fc-specific; Southern Biotechnology), or fluorescence isothiocyanate–conjugated F(ab')2; GAM IgG subclasses (Fc-specific; IgG1, IgG2a, IgG2b, and IgG3; Southern Biotechnology).

Cytotoxicity assay

The anti–EL4-huCD20 cytotoxicity of spleen cells from naive and challenged CAT-13–treated mice (n = 3 in each group) was assessed by a standard 51Cr release assay: 5 × 104 EL4-huCD20 cells were labeled after incubation for 1 hour at 37°C with 37 µCi of 51Cr (PerkinElmer Life and Analytical Sciences). Spleen cells from challenged mice were isolated 3 days after the challenge. Cells were stimulated with mitomycin C (Sigma-Aldrich)–treated target EL4-huCD20 cells (5 × 103/well) at a 50:1 responder/stimulator ratio for 4 days in the presence of 100 IU/mL of human IL-2 (Roussel-Uclaf) to increase the frequency of tumor-specific cytotoxic T cells among whole spleen cells. In vitro–stimulated cells were then mixed with 3Cr-labeled EL4-huCD20 cells (5 × 103/well) at effectorsratios of 50:1, 100:1, and 200:1 and incubated for 4 hours at 37°C. The percentage of specific lysis was calculated with this formula:

\[
\text{specific lysis} = \frac{cpm_{	ext{max}} - cpm_{	ext{mean}}}{cpm_{	ext{mean}}} \times 100, \quad \text{wherein } cpm \text{ indicates counts per minute, } exp, \text{ experimental; } min, \text{ spontaneous release; and } max, \text{ maximum release.}
\]

Adoptive transfer experiments

Serum for the passive transfer experiment was collected in sterile conditions from untreated mice 20 days after tumor inoculation and from surviving CAT-13–treated mice 20 days after tumor challenge (day 90). On day −1, pooled serum samples were intravenously (thus passively) transferred from either untreated mice or surviving CAT-13–treated mice into the tail vein of naive recipient mice (200 µL of undiluted serum per mouse). Recipient mice and naive control mice were then inoculated intravenously with 5 × 106 EL4-huCD20 cells (in 200 µL of PBS per mouse) on day 0.

For the spleen cell adoptive transfer experiment, surviving CAT-13–treated mice were killed on day 90, 20 days after tumor challenge. Splenocytes were intravenously injected into the tail vein of naive recipient mice on day −1 (5 × 107 spleen cells in 200 µL of PBS per mouse). Recipient
mice and naive control mice were then inoculated intravenously with $5 \times 10^5$ EL4-huCD20 cells (in 200 μL of PBS per mouse) on day 0.

### Phenotypic analysis by direct immunofluorescence

Spleens from naïve mice, untreated mice (20 days after tumor inoculation), and surviving CAT-13–treated mice (on day 90, 20 days after tumor challenge) were removed in sterile conditions. Spleen cells were incubated for 30 minutes at 4°C with 10 μg/mL rat anti–mouse (RAM) FcyR 2.4G2 mAb. Mouse CD3, CD4, and CD8 were visualized by incubation with PE-conjugated RAM CD3, PE-Texas red-conjugated RAM CD4, and Alexa Fluor 700–conjugated RAM CD8 mAbs (BD Biosciences PharMingen, Invitrogen, and eBioscience, respectively) for 30 minutes at 4°C. Isotype-matched mAbs were used as staining controls. Spleen cells were then washed in PBS containing 2% fetal calf serum and 2mM EDTA and fixed in PBS containing 0.5% formaldehyde. Staining was analyzed with the LSR II cytometer (BD Biosciences).

CD25 expression on EL4-huCD20 cells was assessed by direct immunofluorescence. Cells were isolated from tumor-infiltrated lymph nodes of tumor-bearing mice on day 35, incubated in vitro for 2 days in culture medium, and then overnight in the presence of human IL-2 (50, 500, and 5000 IU/mL). Direct immunofluorescence was performed with allophycocyanin-conjugated RAM CD25 (BD Biosciences PharMingen) and Alexa Fluor 488–conjugated CAT-13 mAb, which allows the detection of EL4-huCD20 cells. Staining was analyzed with the FACSCalibur (BD Biosciences).

### Statistical analysis

The Kruskal-Wallis test was used to determine the significance of differences between sample means. Comparative survival was analyzed with the log-rank statistical test, selected to take into account both survival over the follow-up period and time-to-event (death or sacrifice). For statistical analyses, we used the StatView Version 5.0 software package for Windows (SAS Institute). Data are mean plus or minus SD when indicated.

### Results

#### Anti-huCD20 mAb therapy after tumor inoculation led to long-lasting survival, including after a subsequent tumor challenge

To determine whether initial anti-huCD20 mAb therapy induces an antitumor response that can protect mice after tumor challenge, we injected $5 \times 10^5$ EL4-huCD20 cells intravenously into immunocompetent C57Bl/6 mice\textsuperscript{25} on day 0 and treated them with mouse IgG2a anti-huCD20 mAb CAT-13 (5 200-μg intraperitoneal injections on days 1, 4, 7, 10, and 13). The $5 \times 200$ μg induction regimen produced the highest overall survival rate. Figure 1 depicts a representative experiment. Approximately 80% of the CAT-13–treated mice survived to day 70 after the initial tumor injection (Figure 1A; $P < .001$). Overall survival from 7 independent experiments averaged approximately 60% (Table 1). All untreated mice died before day 50 (Figure 1A).\textsuperscript{25}

Surviving CAT-13–treated mice were intravenously challenged on day 70 by either $5 \times 10^5$ EL4-wt or EL4-huCD20 cells, without any further mAb therapy. Approximately 70% of the surviving CAT-13–treated mice challenged with EL4-huCD20 cells were still alive at least 70 days after this challenge (ie, on day 140; Figure 1B; $P < .001$). In contrast, all surviving CAT-13–treated mice challenged with EL4-wt cells died within 25 days. Survival through day 140 of CAT-13–treated mice injected with EL4-huCD20 cells in 7 independent experiments averaged approximately 40% (Table 1).

The anti-huCD20 induction regimen therefore allowed the long-term survival of mice after injection of huCD20$^+$ tumor cells. This protection allowed the mice to survive a subsequent tumor challenge no. 70 days after a tumor challenge on day 70.

### Table 1. Analysis of CAT-13–treated mice survival after tumor inoculation and tumor challenge

<table>
<thead>
<tr>
<th>Independent experiment no.</th>
<th>CAT-13–treated mice</th>
<th>CAT-13–treated surviving mice challenged at day 70</th>
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<tr>
<td></td>
<td>Survivors/total</td>
<td>Overall survival, percentage</td>
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</tr>
<tr>
<td>Mean ± SD, percentage</td>
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<td>64.2 ± 10.6</td>
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</table>

Data are from 7 independent experiments with or without tumor challenge.

*Number of surviving CAT-13–treated mice 70 days after inoculation of tumor cells on day 0.
†Number of surviving CAT-13–treated mice 70 days after a tumor challenge on day 70.

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challenge with huCD20+ but not huCD20− tumor cells. The protection induced was thus huCD20-specific.

**Xeno-immunization did not produce long-term antitumor protection, which required intact CAT-13 mAb**

Recent studies have shown that dying tumor cells can intrinsically induce an antitumor immune response.26,27 Moreover, the presence of a human CD20 molecule on EL4 cells may trigger xenogenic tumor inoculation on day 0, mice received 5 intraperitoneal injections of CAT-13 F(ab’)2 fragments. As shown in Figure 2C, all F(ab’)2-treated mice died before day 35, as the untreated mice did, thereby indicating that the Fc portion of the CAT-13 antibody is required for achieving a therapeutic effect.

Taken together, these results demonstrate that the therapeutic efficacy of the CAT-13 induction regimen requires intact IgG mAb and that EL4-huCD20 cells alone do not trigger a protective antitumor response.

**Endogenous antibody response against tumor cells**

We then evaluated whether CAT-13 mAb therapy could lead to long-lasting antitumor protection by inducing an endogenous antibody response against tumor cells. First, we used indirect immunofluorescence to analyze the presence of IgM and IgG directed against EL4-wt and EL4-huCD20 cells in the sera of CAT-13–treated mice on days 15, 30, 45, 70, 85, and 100. Of note, at all time points, the sera of CAT-13–treated mice showed only faint IgM binding to both EL4-wt (data not shown) and EL4-huCD20 cells (Figure 3A top panels). Similarly, only very weak IgG binding to EL4-wt cells was detected (data not shown). In contrast, IgG binding to EL4-huCD20 cells was easily detected on days 15, 30, and 45 (Figure 3A middle panels) but decreased over time. Interestingly, the lack of any increase in the antitumor IgG titer after the EL4-huCD20 tumor challenge on day 70 suggests that the IgG binding detected on days 15, 30, and 45 was mostly the result of the presence of CAT-13 IgG2a in the tested sera. To test this hypothesis, we analyzed the subclass specificity of IgG bound to EL4-huCD20 cells. No IgG1, IgG2b, or IgG3 binding to EL4-huCD20 cells could be detected (data not shown). In contrast, marked IgG2a binding to EL4-huCD20 cells was detected on days 15, 30, and 45 (Figure 3A middle panels) and decreased thereafter. It was notably weak on days 85 and 100 (ie, 15 and 30 days after tumor challenge). Taken together, these results demonstrate the absence of any potent IgM and IgG response against EL4-huCD20 cells in CAT-13–treated mice. For additional confirmation of the absence of any antitumor antibody response that could be responsible for the protective effect, we pooled sera from CAT-13–treated surviving mice and used it in a passive immunity transfer experiment (Figure 3B). Recipient mice were intravenously injected in the tail vein on day −1 with 200 μL of pooled sera from either untreated (day 20 sera) or surviving CAT-13–treated mice (day 90 sera; ie, 20 days after the tumor challenge). Recipient mice were then intravenously injected on day 0 with 5 × 10^5 EL4-huCD20 cells. The death of all the animals before day 35 showed that the CAT-13–induced long-term antitumor protection could not be adoptively transferred to naive recipients by a single injection of sera from surviving CAT-13–treated mice.

Together, these results indicate that the long-term antitumor protection induced by CAT-13 mAb therapy was not the result of
the generation of a potent endogenous antibody response directed against the EL4-huCD20 cells.

CAT-13–induced antitumor protection was mediated through the triggering of a cellular immune response

We sought to determine whether the long-lasting CAT-13–induced antitumor protection involves cellular immune effectors. First, we examined whether spleen cells from CAT-13–treated animals exerted any anti–EL4-huCD20 activity in an in vitro cytotoxicity assay. Spleen cells from challenged mice were isolated 3 days after the tumor challenge (day 73), stimulated with mitomycin C–treated target EL4-huCD20 cells for 4 days in the presence of IL-2, and then incubated with 51Cr-labeled EL4-huCD20 cells. Figure 4A shows that stimulated spleen cells from CAT-13–treated challenged mice killed the target EL4-huCD20 cells, a finding that demonstrates that these animal spleens contained antitumor immune effectors. Second, we used splenocytes from surviving CAT-13–treated mice to test adoptive immunity transfer into naive recipients on day 90. Figure 4B shows that the intravenous injection of 5 x 10^7 spleen cells from surviving mice 24 hours before the intravenous injection of EL4-huCD20 cells is sufficient enough to protect recipients from death (P = .03). That is, the long-lasting CAT-13–induced antitumor protection can be adoptively transferred into naive mice through spleen cells.

Because this protection is a durable phenomenon probably related to the generation of a cellular, but not humoral, antitumor immune response, we next evaluated the involvement of T cells. In a first set of experiments, we analyzed by flow cytometry the CD4^+ and CD8^+ T-cell subsets in the spleen and calculated the CD4^+ /CD8^+ T-cell ratio among CD3^+ cells (Figure 4C). Untreated mice injected with EL4-huCD20 cells and surviving CAT-13–treated mice challenged with a second injection of EL4-huCD20 cells were analyzed 20 days after the tumor cell challenge (that is, day 20 for the untreated mice, day 90 for the surviving treated mice). Naive C57Bl/6 mice were used as controls. The CD4^+/CD8^+ T-cell ratio was markedly lower 20 days after EL4-huCD20 cell injection in untreated mice than in both the naive and surviving CAT-13–treated mice (Figure 4C top histograms; P = .01). The lower CD4^+/CD8^+ T-cell ratio in untreated mice paralleled their markedly lower absolute number of CD3^+ CD4^+ but not of CD3^+ CD8^+ cells (Figure 4C bottom histograms; P = .008). Thus, the similarity in both splenic T-cell counts and CD4^+/CD8^+ T-cell ratios in naive and surviving CAT-13–treated mice and their differences from those of untreated tumor-bearing animals suggest that antibody treatment affects the splenic CD4^+ T-cell subset.

To explore the role of CD4^+ cells in antitumor protection, we used a depleting anti-CD4 mAb (OK1.5) to assess the effect of CD4^+ cell depletion in C57Bl/6 mice at different times. In the first experiment, depletion began on day 1, before the EL4-huCD20 cell injection on day 0 and subsequent CAT-13 mAb therapy from days 1 to 13. Figure 5A shows that the therapeutic efficacy of the CAT-13 induction regimen was dramatically impaired when CD4^+...
cells were depleted before therapy began. Only 16% of CD4-depleted mice (2 of 12) survived on day 70 versus 58% (7 of 12) of CD4+ mice (P = .01). This result indicates that CD4+ cells are needed at the beginning of mAb therapy to achieve long-term survival. In a second experiment, depletion began on day 69, the day before the tumor challenge on day 70. All depleted mice died after this tumor challenge, and the mortality observed was similar to that of untreated control mice (Figure 5B). This experiment demonstrates that CD4+ cells are also involved in the survival of mice after tumor challenge.

Because CD8+ T cells are potent tumor cell killers, we also analyzed the capacity of CAT-13 mAb to induce long-lasting antitumor protection in C57Bl/6 mice without CD8+ cells. The identical overall survival in both CD8 KO and wt mice indicates that CD8+ T cells are involved in the CAT-13–induced antitumor protection in vivo.

Figure 4. Spleen cells are involved in the CAT-13–induced antitumor protection. (A) Spleen cells from naive (□, n = 3) or CAT-13–treated challenged (day 73) mice (●, n = 3) were stimulated with mitomycin C-treated target EL4-huCD20 cells at 50:1 responder/stimulator ratio for 4 days in the presence of IL-2. Stimulated cells were then mixed with 111In-labeled EL4-huCD20 cells at effector/target ratios of 50:1, 100:1, and 200:1. Effector values correspond to the number of CD3+CD8+ cells at effector/target ratios of 50:1, 100:1, and 200:1. *P < .01; †††P < .001). (B) Surviving CAT-13–treated mice, isolated 20 days after tumor challenge, were intravenously injected into naive recipient mice on day −1 (●, n = 5). These mice were subsequently intravenously injected on day 0 with 5 × 10^5 EL4-huCD20 cells, as were control naive mice (□, n = 5; log-rank analysis, *P = .03). (C) Spleen cells from untreated or surviving CAT-13–treated mice were analyzed by immunofluorescence 20 days after tumor injection (untreated animals) or tumor challenge (surviving CAT-13–treated animals). Spleen cells from naive mice were also tested. The CD4+/CD8+ T-cell ratio was calculated (top panel; Kruskal-Wallis analysis, †P = .01), and the number of CD4+ and CD8+ cells among 3 × 10^5 CD3+ spleen cells was determined for each mouse (bottom panel; Kruskal-Wallis analysis, ††P = .008).

Figure 5. CD4+ and CD8+ cells are required for the CAT-13–induced antitumor protection in vivo. (A) Naive mice were depleted of CD4+ cells by intraperitoneal injections of the anti-CD4 mAb GK1.5 starting on day −1. Both depleted (○, n = 12) and nondepleted control mice (●, n = 12) were then injected with 5 × 10^5 EL4-huCD20 cells on day 0 and then received CAT-13 mAb therapy. Untreated mice (□, n = 6) were also used as controls (log-rank analysis, †P = .01). (B) Surviving CAT-13–treated mice received intraperitoneal injections of GK1.5 mAb starting on day 69 (●, n = 9) or not (○, n = 9) and were intravenously challenged with 5 × 10^5 EL4-huCD20 cells on day 70. Untreated mice (□, n = 5) were used as controls (log-rank analysis, ††P = .005). (C) Wild-type (n = 7) and CD8 KO C57Bl/6 mice (n = 18) were intravenously injected on day 0 with 5 × 10^5 EL4-huCD20 cells. Wild-type animals received the CAT-13 mAb therapy (○, n = 7). CD8 KO mice were divided into 2 groups. The first group was left untreated and used as a control (○, n = 6). The second group of CD8 KO mice received the CAT-13 mAb therapy (●, n = 18; log-rank analysis, †P = .01; ††P < .001). (D) Both wt (○, n = 4) and CD8 KO (●, n = 10) surviving CAT-13–treated animals were intravenously challenged with 5 × 10^5 EL4-huCD20 cells on day 70. Untreated mice (□, n = 6) were used as controls (log-rank analysis, †††P = .02). Black arrows indicate CAT-13 mAb injections.
that effective mAb therapy did not require CD8+ T cells at treatment onset (Figure 5C; *P < .001). CD8 KO and wt treated mice that survived the initial tumor inoculation were then intravenously challenged on day 70 with $5 \times 10^6$ EL4-huCD20 cells. In contrast to the wt mice, all CD8 KO mice died after the tumor challenge (Figure 5D), at a rate similar to that of wt untreated control mice after the initial tumor injection. This finding shows that CD8+ cells are required for antitumor protection after tumor challenge.

Altogether, these results provide evidence that the CAT-13 mAb induction regimen triggers an antitumor cellular immune response that requires the presence of CD4+ cells at the beginning of mAb therapy and both CD4+ and CD8+ cells after tumor challenge.

### The addition of IL-2 to anti-huCD20 mAb therapy improved overall survival after tumor challenge

Because IL-2 enhances T-cell antitumor activity in mouse and human cancers,28,29 we evaluated the effect of its association with CAT-13 mAb therapy on overall survival. To rule out the hypothesis of a direct effect by IL-2 on tumor cells, we incubated EL4-huCD20 cells in vitro with various concentrations of IL-2 and assessed cell viability and CD25 expression. In no test conditions did we see either enhanced EL4-huCD20 cell death or up-regulation of CD25 expression after incubation (data not shown).

IL-2 was first administered concomitantly with CAT-13 mAb (ie, 5 intraperitoneal injections of CAT-13 mAb from day 1 to day 13 and 2 intraperitoneal injections of 100 000 IU of IL-2 on days 1 and 4). Overall survival was similar in the mice treated with CAT-13 alone and with IL-2 and CAT-13 (Figure 6A). Second, we evaluated the effect of IL-2 when injected into surviving CAT-13–treated mice after the tumor challenge on day 70 (ie, 2 intraperitoneal injections of 100 000 IU of IL-2 on days 71 and 74; Figure 6B). In 3 independent experiments, the overall survival of the CAT-13–treated mice receiving IL-2 was higher than that of animals without IL-2 injections after the tumor challenge (Table 2; *P = .004). Thus, IL-2 therapy improved the overall survival rate of tumor-challenged mice only when it was administered after the tumor challenge. These results support the existence of a T cell–dependent antitumor immune response that is triggered by the CAT-13 induction regimen and can be potentiated by IL-2.

### Discussion

Despite the clinical efficacy of rituximab, a complete and durable response in patients treated with this mAb remains rare.4,30,31 Potential factors thought to influence the efficacy of rituximab in vivo include both host- and tumor-related resistance mechanisms.7,23,33 However, another important factor that might affect this is the capacity of some rituximab-treated patients to develop a long-lasting antitumor immune response.

We showed in this study that an induction regimen with anti-huCD20 mAb protects mice injected with EL4-huCD20 tumor cells from a subsequent tumor challenge (Figure 1). The lack of would be beneficial in the treatment of human malignancies.

### Table 2. Analysis of survival of CAT-13–treated mice after tumor challenge, with or without IL-2 therapy

<table>
<thead>
<tr>
<th>Independent experiment no.</th>
<th>CAT-13–treated surviving mice challenged at day 70</th>
<th>CAT-13–treated surviving mice challenged at day 70 + IL-2</th>
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<td>3</td>
<td>2/5 40.0‡</td>
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<tr>
<td>Mean ± SD, percentage</td>
<td>— 34.2 ± 12.4‡</td>
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</table>

Data are from 3 independent experiments with or without IL-2 treatment after tumor challenge.

— indicates not applicable.

*Number of surviving CAT-13–treated mice 70 days after a tumor challenge on day 70.
†Number of surviving CAT-13–treated mice that received IL-2 therapy on days 71 and 74 and survived 70 days after a tumor challenge on day 70.
‡Kruskal-Wallis statistical test, *P = .004.
contrast, we showed with CD8 KO mice that CD8 perhaps through the activation of innate effectors, support this effective than CD8 effectors, or both. Recent reports that CD4 classic T helper cells that enhance the antitumor activity of innate these CD4 might play an essential role in EL4-huCD20 cell depletion, as previously reported in animal models of mAb-mediated B-cell depletion. In contrast, during the late phase, when the mAb concentration is very low, poorly opsonized tumor cells might not be captured by phagocytes. Instead, antitumor memory CD4+ T cells generated during the initial phase might activate CD8+ tumor-specific cytotoxic T lymphocytes, leading to tumor control or eradication.

In our study, no antigen spread occurred after the anti-huCD20 mAb therapy, as shown when EL4-wt cells were used for the tumor challenge (Figure 1B). A recent report showed that the combined use of GM-CSF and mAb has the potential to overcome tolerance to self-antigens. Early administration of GM-CSF together with anti-CD20 mAb could therefore potentiate antigen-presenting cell activity and enhance T-cell priming, ultimately leading to the breakdown of tolerance. This hypothesis is consistent with the recent observation that the combined use of rituximab and GM-CSF produces high response rates in patients with relapsed or progressive FL. Another clinical study suggests that rituximab with interferon-α2a improves both the quality and duration of the responses in relapsing FL patients. Another approach to strengthening the efficacy of anti-CD20 therapy might be to combine anti-CD20 with cytokines acting on tumor-specific effector cells at a later stage of the therapeutic scheme, once these cells have been recruited and activated after the induction regimen. We demonstrated here that IL-2 treatment increased the overall survival of CAT-13–treated mice when infected after tumor challenge, but not when injected in association with CAT-13 mAb during the induction regimen (Figure 6A-B; Table 2). These data show that IL-2, used after the establishment of the antitumor protection, enhances the overall survival rate. This effect could be the result of a boost of tumor-specific effector T cells induced during the initial phase of mAb therapy. This phenomenon appears somewhat similar to the adjuvant effect of IL-2 in antitumor vaccination. It should be pointed out that IL-2 injection did not lead to a decrease in the overall survival rate at any time of administration. This suggests, in contrast to some other studies, that no regulatory T cell–mediated suppression was induced.

In conclusion, anti-CD20 mAb therapy induces a cellular immune response that leads to long-lasting in vivo antitumor protection. As recently discussed by Weiner et al, the promotion of an endogenous antitumor immune response in mAb-treated cancer patients is becoming a major goal for improving mAb-based therapies. This challenge could be addressed in the near future either by rational molecular engineering of mAbs or by their combined use with immunomodulatory molecules.

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Authorship

Contribution: R.A. and J.-L.T. designed research, analyzed data, and wrote the paper; R.A. and E.G. performed research and collected and analyzed data; and W.H.F. revised the paper.
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