Gastric MALT lymphoma B cells express polyreactive, somatically mutated immunoglobulins

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Introduction

Gastric mucosa-associated lymphoid tissue (MALT) B-cell lymphoma develops in the context of long-term infection with the Gram-negative gastric bacterium Helicobacter pylori.1-3 Persistent infection with H pylori causes chronic gastritis that, in some people, can develop into more organized gastric mucosa-associated lymphoid tissue (MALT) with histologic similarity to the Peyer patches of the small intestine.4 The disease progresses when individual malignant clones grow out, displace the benign lymphoid tissue, and ultimately form the lymphoepithelial lesions that are a hallmark of MALT lymphoma.5,6 In its early stages, gastric MALT lymphoma is believed to be an antigen-dependent disease; H pylori infection is detectable in a large majority of cases.1,3,7 Eradication therapy induces tumor regression in approximately 75% of patients at this stage.8,9 Early-stage low-grade MALT lymphoma is generally considered an indolent tumor due to its slow growth, low proliferation rates, and minimal propensity for spreading. At later stages, however, the tumors can eventually undergo high-grade transformation or acquire one of several known characteristic chromosomal translocations, thereby rendering the lymphoma independent of antigen exposure and refractory to H pylori eradication therapy.

In contrast to cases with chromosomal rearrangements, which grow autonomously due to constitutive activation of the nuclear factor κB (NF-κB) signaling pathway brought about by overexpression of MALT-1, B-cell leukemia 10 (Bcl-10), or production of the Baculovirus IAP repeat-containing 3 (API2)–MALT1 fusion protein (reviewed in Isaacs and Du),4 little is known about the pathogenesis of early MALT lymphoma. Several studies have implicated the abundant population of tumor-infiltrating T cells in providing growth signals to tumor B cells.10-12 In one study, depletion of T cells was shown to abrogate the proliferation of explanted MALT lymphoma cultures.12 We and others have reported that MALT lymphomas express high levels of interleukin-4 (IL-4) and other T helper 2 cytokines in vivo, supporting a role for T helper 2–polarized T-helper responses in early MALT lymphomagenesis.10,11

An alternative possibility is that early stage MALT lymphoma B cells receive signals via antigenic stimulation through their B-cell receptor (BCR), which would lead to NF-κB activation, survival, and proliferation. Indeed, MALT lymphoma cells carry functional, rearranged, and somatically mutated immunoglobulin genes on their surface.13,14 Sequence analysis of the VH genes suggests that the tumor cells have undergone positive selection in germinal centers.14-16 Intracellular variation caused by ongoing somatic mutation and/or replacement of a part of the variable heavy segment (receptor revision) has been reported.15-17 Despite these clear results, the search for a target antigen has proven difficult and has yielded controversial results, either providing evidence for reactivity toward certain structures of normal human tissues (follicular dendritic cells, venules, epithelial cells, connective tissue)18,19 or human immunoglobulin G (IgG),13 or not identifying any target antigens at all.20
We and others have shown earlier that long-term, persistent infection of the BALB/c strain of mice with *H pylori* or its close relatives *Helicobacter felis* and *Helicobacter heilmannii* results in the reproducible induction of lesions that clinically and histologically resemble human MALT lymphoma.10,21-23 These lesions are slow growing, spread only infrequently (usually to the spleen), and rarely transform to high-grade lymphoma.10,22 The tumors further recapitulate the human disease in that they regress in a majority of mice upon eradication of the infection,25 but recur rapidly upon reintroduction of the bacteria, revealing gastric persistence of the tumor clone despite complete histologic remission.10

Using this experimental mouse model as well as human MALT lymphoma biopsy material, we deduced and compared the immunglobulin sequences of 20 murine and 7 human tumors. Based on the monoclonal Ig sequences, we generated a comprehensive panel of recombinant antibodies expressed as soluble IgG of matching heavy and light chains and tested their reactivity toward various self- and foreign antigens. The tumor antibodies, which were functional and somatically hypermutated and in some cases showed evidence of positive and/or negative selection, surprisingly reacted with roughly equal affinity with both gastric self- and *Helicobacter*-derived foreign antigens. This pattern of reactivity is consistent with a diagnosis of polyreactivity, and could further be linked to a strongly biased use of specific V<sub>H</sub> family gene segments in both the murine and the human system.

**Methods**

**Animal experimentation, tumor cell cultures, and immunophenotyping**

Specific pathogen-free female BALB/c mice were infected orally at 6 weeks of age with 3 consecutive doses of approximately 5 x 10<sup>9</sup> *H felis* (CS1, ATCC 49179). All animal experiments were approved by Stanford University’s Institutional Animal Care and Use Committee. Upon killing, the stomachs were removed and opened along the lesser curvature; macroscopically visible tumors were dissected. Single-cell suspensions were generated and cultured in RPMI supplemented with 10% fetal calf serum (FCS) and antibiotics. Where appropriate, 10 μg/ml *Helicobacter* lystate was used. Tumor cell proliferation was quantified by [3H]-thymidine incorporation assay, or by bromodeoxyuridine (BrdU) incorporation followed by fluorescence-activated cell sorting analysis using a fluorescent in situ cell proliferation kit (Roche Diagnostics) The following antibodies were used for immunophenotyping: anti-CD19 (monoclonal antibody [mAb] clone 6D5; Abcam), anti-B220 (mAb clone RA3-6B2, BD Pharmingen), anti-IgM (polyclonal goat, no. 1020; Southern Biotech), anti-IgG (polyclonal goat, no. 1030; Southern Biotech), anti-CD3 (mAb clone 145-2C11; BD Pharmingen), anti-CD11c (mAb clone N418; AbD Serotec), anti–proliferating cell nuclear antigen (PCNA; mAb clone PC10; Zymed Labs), and anti-CD11b (mAb clone M1/70.15; AbD Serotec). Flow cytometry was performed on a CyanADP instrument (Dako).

**Patient material**

Human patient material was obtained from 7 patients with gastric MALT lymphoma who were part of a previously published study conducted at Philipps-University Marburg.24 All tumors were diagnosed as *H pylori*–positive low-grade gastric MALT lymphomas and all were negative for the translocation (t(11;18)(q21;q21). The 5 cases of chronic lymphocytic leukemia (CLL), diagnosed according to standard criteria, were described in previous studies25-26; all 5 tumor-derived IgV<sub>H</sub> sequences had been determined previously to be somatically mutated.

**RNA extraction, V<sub>H</sub> and V<sub>L</sub> gene amplification, and cloning**

RNA was isolated from the murine tumor cell suspensions and from patient biopsy material using the RNeasy kit (QUAGEN) according to the manufacturer’s instructions. cDNA was synthesized using oligo-dT primers and SuperScript III reverse transcriptase (Invitrogen). The human Ig heavy and light chain sequences were obtained using Ig framework-specific primers for all human variable heavy and light chain framework 1 families in combination with primers specific for all human J segments, as specified by Marks et al. The obtained variable heavy and light chain products were cloned into expression vectors in frame with a 5’ leader peptide and a 3’ human IgG1 or kappa light chain constant domain. The amplification and cloning of murine Ig variable heavy and light chain sequences was performed accordingly.

**Sequence analysis**

Sequencing of heavy and light chain variable regions was performed by Microsynth. The obtained sequences were aligned to germline sequences from the IGMT database (international ImMunoGeneTics database, http:// www.imgt.org).24 Tumors were defined as clonal if identical or near identical V<sub>H</sub> sequences were obtained from 2 independent polymerase chain reactions. Somatic hypermutation and intrachromal variation of IgV<sub>H</sub> genes was assessed as described by Lossos et al27; V<sub>H</sub> gene sequences that deviated by more than 2% from the corresponding germline gene sequence were defined as somatically hypermutated. The degree of intrachromal diversity of IgV<sub>H</sub> genes was defined as follows: an unconfirmed mutation is a substitution mutation observed in only one of the V<sub>H</sub> gene clones from the same tumor specimen; a confirmed mutation is a mutation observed in more than one V<sub>H</sub> clone from the same tumor specimen.29 The role of antigen selection in shaping MALT lymphoma immunoglobulins was assessed by calculating and comparing the ratios of replacement (R) to silent (S) mutations in the framework (FR) and complementary determining regions (CDRs) of the somatically mutated Ig heavy chain genes. Positive selection was diagnosed if the R/S ratios of mutations in the CDRs were greater than 2.9; negative selection pressure by antigen was postulated if the R/S ratios in the FR regions were lower than 2.9.30 The probability that an excess or scarcity of replacement mutations in V<sub>H</sub> CDRs or framework regions occurred by chance was calculated by a multinomial distribution model.31 A *P* value less than .05 was considered statistically significant.

**Expression and purification of recombinant antibodies**

293T cells were cultured in Opti-MEM I reduced Serum Media (Invitrogen) supplemented with GlutaMAX (Invitrogen). Cells were transfected by calcium phosphate coprecipitation with equimass amounts of the heavy and light chain vector and the supernatant was harvested after 7 days. Recombinant antibodies were purified from culture supernatants on a 1-mL HiTrap Protein G Sepharose column (Amersham Pharmacia Biotech) and dialyzed against phosphate-buffered saline (PBS).

**Preparation of *H felis* antigens, murine stomach extract, and AGS cell extract**

*H felis* and *H pylori* were grown as described.31 Sonicate was prepared by harvesting cells in PBS and the cells were disrupted by sonication (Bandelin Sonopuls GM 70; Bandelin). For preparation of gastric mucosal extract, the mucosa was scraped from wild-type BALB/c mice and lysed in lysis buffer (10% glycerol, 1% Triton X-100, 100 mM NaCl, 20 mM N-2-hydroxyethylpiperazine-N’-2-ethanesulfonic acid [pH 7.4], and 1mM sodium vana-date). For preparation of human stomach cell extract, human gastric carcinoma cells (AGS) were lysed with the same buffer. The protein content of all antigen preparations was determined by Bicinchoninic acid protein assay (Thermo Scientific) and aliquots were stored at –70°C until used.

**Enzyme-linked immunosorbent assays**

Duplicate wells of Maxisorp microtiter plates (Nunc) were coated with various antigens. Single-stranded DNA (ssDNA) and double-stranded DNA
(dsDNA) were coated at 100 μg/mL in PBS. All other antigens tested were coated at 5 μg/mL each in PBS. After washing with PBS-0.5% (vol/vol) Tween-20, the wells were blocked with 2% bovine serum albumin (BSA) in PBS for 2 hours. All antibodies were incubated overnight at 0.1, 1, and 10 μg/mL at 4°C. After washing, wells were incubated for 1 hour with either peroxidase-conjugated goat anti–mouse IgG (Jackson ImmunoResearch Laboratories) or goat peroxidase-conjugated anti–human IgG (Abcam). Enzyme activity was detected by incubation with tetramethylbenzidine (Sigma-Aldrich). Rheumatoid factor activity of murine and human antibodies was determined using the mouse rheumatoid factor Ig enzyme-linked immunosorbent assay (ELISA) kit (Alpha Diagnostic International) and the Aeskuhsa Human RF-AGM kit (Aesku Diagnostics), respectively. The dissociation constants (K_D) were calculated according to Friguet et al.32

Results

Induction of Helicobacter-dependent gastric MALT lymphoma in a BALB/c mouse model and ex vivo culturing of tumor cell suspensions

To induce the formation of Helicobacter-dependent gastric MALT lymphoma, 18 female BALB/c mice were infected with H. felis for 18 months; 7 additional age-matched females served as uninfected controls. Sixteen of the 18 infected, but none of the uninfected mice, had developed gastric MALT lymphoma at the time of killing. The tumors were macroscopically visible as nodules ranging from 1 to 2 mm in diameter. They were positive for the B-cell marker B220 and had proliferative indices of approximately 30% as determined by PCNA staining (Figure 1A-B). The tumors had formed predominantly at the forestomach/corpus junction. A total of 20 macroscopically discernible tumors were dissected and single-cell suspensions were generated of individual tumors. These were subjected to Ig sequence analysis (all 20 tumors; isolated from 10 individual mice), immunophenotyping (a subset of 14 tumors, from the same 10 mice), and/or ex vivo culturing (9 tumors from 8 mice).

In the tumor cell suspension cultures, an average of 64% of all leukocytes were CD19+ B cells (Figure 1C); most of the remaining leukocytes stained positive for CD3 (Figure 1C). Only a minor fraction was positive for CD11b or CD11c (1.5% on average; data not shown). The suspensions generally contained less than 15% nonleukocyte cells. Although all tumor cell suspensions could be kept alive for 5 days in standard cell culture media, only cultures to which Helicobacter lysate had been added proliferated within this time frame, as determined by [3H]-thymidine (Figure 1D) or BrdU incorporation (Figure 1E). Addition of Helicobacter lysate induced an on average 4-fold increase in [3H]-thymidine incorporation compared with the corresponding unstimulated culture (Figure 1D); BrdU-positive cells increased from approximately 2.5% in unstimulated cultures to approximately 3.8% in stimulated cultures (Figure 1E). Immunophenotyping of BrdU-positive cells further revealed that the proliferating population in the cultures consisted predominantly of CD19+ B cells (average: 83%; Figure 1F), with T cells accounting for the remaining proliferative subset. In conclusion, murine MALT lymphoma B cells not only are antigen dependent in vivo as we and others have shown previously by inducing tumor regression through Helicobacter eradication therapy,10,22 but also retain their dependence on Helicobacter antigen ex vivo.

Immunophenotyping of murine MALT lymphomas

Human MALT lymphomas are known to express surface IgM and pan-B-cell markers (CD19, CD20, CD79a) and the marginal zone markers CD35 and CD21, although are negative for CD5, CD10, CD23, and cyclin D1.33 Because we aimed to analyze the antibody sequences and specificity of our murine tumors, we flow cytometrically determined the surface IgM and IgG expression of 14 tumor cell suspensions (Figure 2). Indeed, the majority of CD19+, B220+ B cells in every tumor showed high expression of IgM (average: 90%); a minority expressed high IgG (average: 1.5%) and the rest expressed only low levels of either Ig (Figure 2).

Sequence analysis of murine MALT lymphoma immunoglobulins

To determine the clonal status of a panel of 20 murine MALT lymphomas, the corresponding cDNAs were generated and subjected to several rounds of cloning and sequencing of the IgV_H genes. (All sequences reported in this paper have been deposited in the GenBank database; accession nos. GQ856044-GQ856073,34) Tumors were pronounced monoclonal if more than half of all obtained IgV_H sequences were identical or near identical in 2 independent rounds of sequencing. Clear monoclonal status was demonstrated for 11 lymphomas (55% of cases), whereas 1 lymphoma appeared biclonal and the remaining 8 tumors were classified as polyclonal. None of the monoclonal sequences harbored internal stop codons, suggesting that murine MALT lymphoma B cells express potentially functional surface IgM.

The 11 clonal IgV_H sequences were further analyzed for evidence of somatic hypermutation and intraclonal variation (Table 1). Germline genes with the highest homology to the consensus tumor IgV_H sequence were identified (supplemental Figure 1, available on the Blood website; see the Supplemental Materials link at the top of the online article); with the exception of 2 cases, all IgV_H genes analyzed displayed somatic mutation, that is, they deviated from the most closely related germline sequence in more than 2% of positions, with the average homology being 92.2% (Table 1, supplemental Figure 1). This result is in line with previous studies reporting somatic hypermutation of human MALT lymphoma IgV_H.14-16 Four IgV_H sequences were found to be positively selected by antigen, that is, the ratios of replacement/silent mutations in their CDRs were higher than would be expected if mutations had occurred by chance alone without selective forces (based on a cutoff ratio of >2.990; Table 1). Six sequences demonstrated the presence of negative selection pressure by antigen, that is, the replacement/silent mutation ratios in their FR regions were lower than expected by chance alone. Two sequences showed evidence of both negative and positive selection. Analysis of antigen selection using the more stringent multimodal model29,31 identified only 3 sequences with significant positive selection and one with significant negative selection (Table 1). Previous studies examining antigenic selection of human IgV_H have mostly reported evidence of positive selective pressure,14-16 suggesting that MALT lymphoma B cells have undergone germinal center reactions and affinity maturation.

To evaluate the presence of ongoing mutation, the 11 clonal sequences were examined by comparing at least 8 and up to 22 molecular clones from each tumor (supplemental Figure 2). Five of the somatically mutated clonal V_H gene isolates did not show intraclonal heterogeneity (Table 1), whereas the remaining 6 tumors, including the 2 with unmutated IgV_H sequences, harbored confirmed mutations and therefore showed evidence of intraclonal variation (Table 1, supplemental Figure 2). The average length of the CDR3 region for the 11 monoclonal tumor immunoglobulins was 12.5 amino acids, which is longer than the average...
length of 8.5 amino acids usually found in a normal murine splenic B-cell repertoire (Table 1). VH gene family use was found to be strongly biased toward V\textsubscript{H}1\textsubscript{n/H11005}7 and V\textsubscript{H}14\textsubscript{n/H11005}4. Of the 12 known murine VH gene families, the VH1 gene family accounts for 28% of the antibody repertoire in BALB/c splenocytes; the VH14 family in contrast is rarely used in BALB/c mice.

Antigen reactivity of recombinant murine MALT lymphoma–derived antibodies

To investigate the antigen specificity of murine MALT lymphoma Ig, the corresponding light chains of 5 of the clonal heavy chains were identified. In cases with ongoing mutation, the IgV\textsubscript{H} sequence most closely matching the consensus sequence was chosen. Paired heavy and light chains were expressed recombinantly in 293T cells and purified by affinity chromatography. We deliberately selected a mixed panel of mutated and unmutated, antigen selected and unselected IgV\textsubscript{H} sequences (mu Abs 2, 3, 8, 10, and 11). A series of ELISAs were performed to systematically screen Ig reactivity toward a broad range of antigens. Possible autoreactivity was assessed using murine stomach extract; antinuclear activity was examined using both ssDNA and dsDNA, and rheumatoid factor activity was assessed by IgG binding; the foreign antigens tested included \textit{H felis} and \textit{Escherichia coli} sonicate and purified \textit{E coli} lipopolysaccharide (LPS). BSA was used as a negative control. The
combined results of all ELISAs revealed quite unequivocally that all antibodies analyzed bound to a surprising variety of antigens in a dose-dependent manner (Figure 3). An overall trend of higher affinity binding toward more complex antigen compositions (extracts of *H. felis* and of murine stomach) was noted; no preference for either *Helicobacter*-only reactivity or autoreactivity-only could be detected in any of the antibodies’ ELISA profiles. A monoclonal, isotype-matched murine control antibody failed to bind any of the antigens tested (Figure 3). Somatically mutated antibodies did not differ from unmutated antibodies, and evidence of antigen selection in the IgVH sequence also did not seem to correlate with an increased overall or antigen-specific affinity.

**Table 1. VH gene analysis of murine gastric MALT lymphoma cases**

<table>
<thead>
<tr>
<th>Case</th>
<th>VH segment</th>
<th>Homology %</th>
<th>Clones sequenced</th>
<th>Somatic* mutation</th>
<th>FR/CDR</th>
<th>R</th>
<th>S</th>
<th>Antigen selection†</th>
<th>P‡</th>
<th>Intraclonal variation</th>
<th>CDR3 length</th>
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<tr>
<td>mu-1</td>
<td>IGHV1-S130*01</td>
<td>93.06</td>
<td>11</td>
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<td>FR</td>
<td>42</td>
<td>1</td>
<td>--</td>
<td>.490</td>
<td>+</td>
<td>11</td>
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<td>mu-2</td>
<td>IGHV14-3*02</td>
<td>92.36</td>
<td>10</td>
<td>+</td>
<td>CDR</td>
<td>29</td>
<td>0</td>
<td>+</td>
<td>.000</td>
<td>--</td>
<td>15</td>
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<td>2</td>
<td>+</td>
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<td>--</td>
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<td>10</td>
<td>--</td>
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<td>0</td>
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<td>8</td>
<td>+</td>
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<td>+</td>
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<td>.156</td>
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MALT indicates mucosa-associated lymphoid tissue; FR, framework region; CDR, complementary determining region; R, replacement; S, silent; and mu, murine.

*VH gene sequences deviating more than 2% from the corresponding germline gene were defined as somatically mutated.

†Presence or absence of positive selection by antigen in the CDR is denoted by + and --, respectively. Presence or absence of negative selection by antigen in the FR is denoted by + and --, respectively, based on the cutoff ratio of 2.9 for replacement to silent mutations.

‡The *P* value was calculated based on the multinomial distribution model and is the probability that excess (for CDR) or scarcity (for FR) of mutations occurred by chance.
Taken together, our results support a diagnosis of polyreactivity. To confirm the specificity of the assay, we performed a series of competitive inhibition ELISAs (Figure 4A-C). To this end, binding of the antibody of interest to one of its targets (immobilized on the ELISA plate) was competitively blocked with increasing concentrations of soluble target of the same or an alternative specificity. In all combinations tested, the soluble-phase antigens were able to competitively block binding to the immobilized target in a dose-dependent manner (Figure 4A-C). As would be expected, high-affinity soluble antigens blocked binding to low-affinity immobilized targets with a higher efficiency than vice versa (Figure 4A-C). BSA, which was used as a negative control, did not block binding even at 100x excess (Figure 4A-B). Overall, the competition results confirmed the specificity of the ELISAs as well as our general observation that complex antigen mixtures constitute higher affinity targets than relatively simple structures such as DNA or LPS.

Sequence analysis of human MALT lymphoma immunoglobulins

To be able to compare the sequence characteristics and specificity of the murine tumor immunoglobulins to their human counterparts, we cloned, sequenced, and recombinantly expressed the antibodies of 7 human low-grade MALT lymphomas.24 All 7 cases were clonal. Five of the 7 cases showed evidence of somatic hypermutation (Table 2, supplemental Figure 3). Interestingly, we found a bias toward use of the VH gene segment 1-69*01, which was used in 4 of the 7 cases (Table 2), and has been reported previously for MALT lymphomas of the parotid gland, stomach, and lung.13 No bias was detected in J and D segment use, but all matching light chains were of the \(\kappa\) isotype (Table 2), another seemingly common bias in MALT lymphoma antibodies.13,20 Three of 7 cases further showed evidence of intraclonal variation (Table 2, supplemental Figure 4). Interestingly, the CDR3 length of the heavy chains...
was higher than average (15.3 amino acids as opposed to the 12.7 amino acids usually found in human blood B-cell antibodies), confirming our observation from the murine model. Overall, the sequence analysis of our panel of 7 antibodies confirmed several observations reported previously for human gastric MALT lymphoma and the panel was therefore considered suitable for antigen binding studies.

Antigen reactivity of recombinant human MALT lymphoma–derived antibodies

To determine whether human MALT lymphoma Ig also follows a polyspecific binding pattern, matched heavy and light chains of all 7 human MALT lymphoma cases were screened for reactivity toward BSA, *H pylori* sonicate, LPS, IgG, ssDNA, and AGS cell extract (Figure 5A). All but 1 (hu Ab2) of the human MALT lymphoma antibodies exhibited a polyreactive binding profile very similar to the patterns observed with the murine tumor Igs. Interestingly, hu Ab2 was the only human tumor antibody on the panel to not have undergone either somatic hypermutation or ongoing mutation. As observed for the murine polyreactive MALT lymphoma antibodies, a trend of higher affinity binding toward more complex antigen compositions was evident (Figure 5A). To properly control for the validity of our ELISA readout in the human system, we chose 5 additional non-MALT lymphomas for amplification of their immunoglobulin genes, sequence analysis, and recombinant expression in 293T cells and subsequent ELISA analysis. We selected cases of “mutated” chronic lymphocytic leukemia

Figure 5. Recombinant human MALT lymphoma–derived antibodies are polyreactive. Three concentrations (0.1, 1, and 10 μg/mL) of 7 recombinant human MALT lymphoma antibodies (A) and 5 recombinant human CLL antibodies (B) were tested by ELISA for reactivity with the antigens indicated in the legends. All MALT lymphoma antibodies, with the exception of Ab2, but none of the CLL antibodies, exhibit a polyreactive binding pattern.
(CLL; ie, cases expressing somatically mutated Ig), as these are known to be monoreactive in contrast to their “unmutated” CLL counterparts. All 5 cases were clonal and showed on average 93% homology with the most closely related germline sequence. None of the antibodies displayed a polyreactive binding pattern similar to the one we observed for 6 of the 7 MALT lymphoma Igs. In fact, all but 1 of the 5 CLL antibodies failed to show any affinity for any of the targets of our ELISAs (one—Ab3—bound to ssDNA, but not to any of the other antigens). This result showed that MALT lymphoma antibodies have fundamentally different antigen-binding properties than monoreactive CLL antibodies, and confirms the validity of our ELISA.

Finally, the antigen specificity of the recombinant human MALT lymphoma antibodies was confirmed by a series of competitive inhibition ELISAs (Figure 6A-C); as observed for the murine antibodies, soluble-phase antigens were able to competitively block binding to the immobilized target in a dose-dependent manner, whereas BSA did not block binding even at 100× excess concentrations (Figure 6A-B right panels). Error bars represent SEM.

To assess the binding affinity of the various tumor Igs, we calculated $K_d$ values for a select group of 6 tumor antibody-antigen complexes using a curve-fitting approach as described. The $K_d$ values ranged from $1.1 \times 10^{-7}$ M (hu Ab5 reactivity toward LPS) to $6.3 \times 10^{-8}$ M (hu Ab4 toward $H. pylori$ extract). Murine tumor antibodies showed a similar range of affinity (eg, $K_d = 2 \times 10^{-8}$ M for mu Ab1 toward ssDNA; $K_d = 5 \times 10^{-8}$ M for mu Ab1 toward stomach extract). Overall, the binding affinity of polyclonal MALT lymphoma–derived antibodies is clearly lower than what is typically observed for monoreactive antibodies (which have $K_d$ of $10^{-8}$ to $10^{-11}$ M), but higher than the typical affinity of polyreactive, unmutated IgM expressed by immature B cells ($K_d$ of $10^{-2}$ to $10^{-3}$ M).

**Discussion** Persistent infection of BALB/c mice with various Helicobacter species results in the development of gastric MALT lymphoma in a majority of mice, with “clinical” features that are strikingly similar to the human form of the malignancy: late onset of disease, dependence on active infection with the organisms, and a low propensity for spreading. We show here that several molecular parameters of the murine tumors also mimic the human disease well: (1) the monoclonal status of more than half of all individually dissected tumors, (2) the antigen dependence of tumor cell proliferation in vitro, (3) the surface exposure of IgM and apparent lack of class switch recombination, (4) the somatic hypermutation of antibody sequences, accompanied in several cases by positive and/or negative antigen selection, and (5) the intraclonal variability that is a hallmark of continuing antigen exposure and, consequently, ongoing somatic mutation. We further demonstrate that both the murine tumor-derived antibodies as well as all but 1 of the antibodies derived from MALT lymphoma patient biopsies show a pattern of polyreactivity, displaying equally strong affinity toward a diverse panel of foreign but also self-antigens. This result is corroborated by our observation of a strong bias toward the use of $V_H$ gene segments that have previously been associated with autoantibodies or polyreactive antibodies in other B-cell malignancies or autoimmune pathologies.
In the murine setting, we often find that more than 1 tumor arises per stomach, that is, the lymphoma presents as a multifocal disease (as is the case in humans). In those cases in which we were able to dissect multiple monoclonal tumors from the same stomach, they were usually derived from independent clones, confirming that MALT lymphoma is a highly localized disease with little tendency to spread, even within the same organ. The monoclonal status of 55% of individually dissected tumors confirms the validity of our model; clonality rates of human MALT lymphomas range from 63% to 92% of histologically confirmed cases. The lower rates of clonality in the murine model may be due to the tiny size and multifocal nature of the murine tumors, which are difficult to dissect without contamination from surrounding normal tissue as well as neighboring tumor material.

Gastric MALT lymphomas are antigen dependent in their early stages, that is, they require the constant presence of or yet unknown antigen(s) for growth in vivo. Interestingly, this requirement for antigen is retained in ex vivo cultures of single-cell suspensions derived from individual murine tumors, which proliferate only in the presence of Helicobacter extract. Despite the apparent complexity of tumor cell suspensions, which contained a significant proportion of admixed T cells and other leukocytes, the proliferating population consisted predominantly of (tumor) B cells.

The immunoglobulin genes of murine as well as human gastric MALT lymphomas are subject to somatic hypermutation but fail to undergo class switch recombination. Indeed, expression of the enzyme responsible for both processes, activation-induced cytidine deaminase, could be detected by reverse-transcription–polymerase chain reaction in material generated from both murine tumor and human MALT lymphomas (data not shown). Although the 2 processes are known to be linked and usually happen simultaneously, antigen-driven malignancies. It has been described in detail for CLL, which, like MALT lymphoma, is believed to be an antigen-driven malignancy. A majority of unmutated, but not of mutated, CLL-derived antibodies tested with respect to their specificity have revealed polyreactive patterns in previous studies, binding to DNA, IgG, insulin, and LPS. In contrast, mutated CLL-derived antibodies are typically monoreactive. Indeed, none of the mutated 5 CLL antibodies we included as controls in our analysis showed evidence of polyreactivity. With respect to the link between mutational status and antigen reactivity, CLL-derived antibodies thus differ clearly from MALT lymphoma–derived antibodies. In fact, we found that the only human MALT lymphoma Ig that failed to exhibit polyreactivity was also the only unmutated antibody among those analyzed. It will be interesting to see whether the link between mutation status and polyreactive binding profile holds true in larger studies that include more of the rare unmutated cases.

Both polyreactive CLL and the MALT tumor immunoglobulins reported here share a strongly biased use of the IgV H1-69 gene segment; 4 of 7 of our panel of human MALT lymphoma antibodies use this segment and, of these, 3 combine the IgV H1-69 heavy chain with the same light chain V L segment IGKV3-20*01. This exact combination is typical of rheumatoid factors, with IgV H1-69 being preferentially used in autoimmune gammapathies reactive toward human IgG such as cryoglobulinemia and Waldenstrom macroglobulinemia. IgV H1-69 was also found to be used in a biased fashion in nongastric MALT lymphomas, including those of the parotid gland, tonsil, and lung. Overall, the strongly biased use of IgV H1-69 in MALT lymphoma and CLL is consistent with their shared polyreactive antibody specificity, as well as their shared dependence on antigenic stimulation.

Polyreactivity is thought to be caused by a special flexibility of the antigen-binding pocket, with multiple configurations or “isomers” of an antibody existing even before exposure to antigen, some evidence suggests that a longer CDR3 region of polyreactive antibodies might be underlying this flexibility. Indeed, we find that our human as well as our mouse MALT lymphoma antibodies possess longer than average CDR3 regions. In conclusion, many of the known molecular and structural hallmarks of polyreactive antibodies such as IgM class, CDR3 length, and preferential VH gene use are detected in MALT lymphoma antibodies and are consistent with our experimental finding of polyreactivity, suggesting a role for antigenic stimulation in MALT lymphoma pathogenesis.

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Authorship

Contribution: V.J.C. designed and performed research, analyzed data, and wrote the paper; I.A. and C.G. collected experimental animal material and performed fluorescence-activated cell sorting analyses; M.Q.H., T.W., A.N., and C.R. collected patient material and extracted mRNA; S.F. contributed vital tools and helped analyze data; and A.M. designed research, performed research, analyzed data, and wrote the paper.

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References


Gastric MALT lymphoma B cells express polyreactive, somatically mutated immunoglobulins

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