Plasmacytoid dendritic cells regulate B-cell growth and differentiation via CD70

Joanne Shaw, Yui-Hsi Wang, Tomoki Ito, Kazuhiko Arima, and Yong-Jun Liu

1Department of Immunology, University of Texas, M. D. Anderson Cancer Center, Houston; and 2University of Texas Graduate School of Biomedical Sciences at Houston

The ability of plasmacytoid dendritic cells (pDCs) to promote plasma cell differentiation and immunoglobulin (Ig) secretion is dependent on the production of type I interferon (IFN)–producing cells, which include type I IFN–producing cells, are one of the two main populations of dendritic cells in human peripheral blood. They are known as interferon and interleukin-6 (IFN-α, IFN-γ, IL-6) producing cells, and are well-established memory B-cell markers. Engagement of CD70 with its receptor CD27 expressed on memory B cells promotes plasma cell differentiation and Ig secretion. Using a pDC/B-cell coculture system, we found that CpG-stimulated pDCs can induce the proliferation of CD40L-activated human peripheral B cells and Ig secretion. This occurs independently of interferon and residual CpG, and requires physical contact between pDCs and B cells. CpG-stimulated pDCs can induce the proliferation of both naive and memory B cells, although Ig secretion is restricted to the memory subset. Blockage of the interaction of CD70 with CD27 using an antagonist anti-CD70 antibody reduces the induction of B-cell proliferation and IgG secretion by CpG-stimulated pDCs. This factor has therefore identified CD70 as a dominant factor in the regulation of B-cell growth and differentiation by pDCs. (Blood. 2010; 115(15):3051-3057)

Introduction

Dendritic cells are a heterogeneous population of cells that play an important role in the initiation and regulation of both innate and adaptive immune responses.1,2 Plasmacytoid dendritic cells (pDCs), also known as type I interferon (IFN)–producing cells, are one of the two main populations of dendritic cells in human peripheral blood. They selectively express Toll-like receptor 7 (TLR7), which allows them to respond to RNA viruses, and TLR9, which allows them to respond to DNA viruses and CpG oligonucleotides.3,6 On exposure to virus, pDCs produce vast amounts of IFN,7 directly inhibiting viral replication and contributing to the activation of B cells.8,10

Several recent studies have indicated an important role for pDCs in the regulation of B-cell differentiation.5-10 Influenza virus-stimulated pDCs are capable of inducing the differentiation of human B cells into plasma cells secreting virus-specific antibodies.8 IFN secretion by pDCs mediates the differentiation of B cells into plasmablasts and pDC-derived interleukin-6 (IL-6) promotes the subsequent development of plasmablasts into immunoglobulin (Ig)–secreting plasma cells. Blocking the function of IFN and IL-6 significantly reduces the production of IgG by B cells stimulated with virus-activated pDCs; however, CD40L-activated B cells cultured with IL-2 and IL-6 produce levels of IgG that are lower than those observed in pDC/B-cell coculture experiments potentially indicating a requirement for additional factors. Indeed, in a recent study comparing the ability of pDCs and myeloid dendritic cells to promote B-cell proliferation and differentiation,10 it was observed that pDCs, but not myeloid dendritic cells, could enhance the plasma cell differentiation of memory, but not naive B cells stimulated with TLR7/8 ligands, via an IFN-dependent mechanism. It has also been observed that pDCs enhance the activation, plasma cell differentiation, and Ig secretion of B cells activated by B-cell receptor cross-linking and CpG-C.9 Separating the pDCs and B cells in a transwell system or blocking the interaction of IFN reduced but did not eliminate the activation of B cells observed in the presence of pDCs, again indicating a role for additional factors, including direct cell-to-cell contact.

CD27 is a member of the tumor necrosis factor receptor family and is a well-established memory B-cell marker.16-18 Engagement of CD27 with its ligand CD70 promotes the production of IgG, IgM, and IgA by human peripheral B cells cultured with IL-2 and IL-10.19,20 It also enhances the differentiation of CD40L-activated B cells into plasma cells.21 CD40L stimulation strongly enhances B-cell proliferation induced by Staphylococcus aureus Cowan I strain; however, engagement of CD27 has little effect.21 Instead, engagement of CD27 promotes plasma cell differentiation and IgG production from B cells cultured with S. aureus Cowan I strain and IL-2.21 B cells cultured with agonist anti-CD40 antibody and IL-4 produce a significant amount of IgE but do not undergo plasma cell differentiation. Engagement of CD27 significantly enhances the production of IgE from B cells cultured with agonist anti-CD40 antibody and IL-4 and also promotes plasma cell differentiation and expression of the plasma cell-specific gene BLIMP1.22 Thus, CD27 plays an important role in the regulation of plasma cell differentiation and Ig secretion from human memory B cells. Here we examine the expression of CD70 in pDCs and investigate the role of CD70 in the regulation of B-cell growth and differentiation by pDCs to better understand the mechanism of interaction between pDCs and B cells.
Methods

pDC and B-cell isolation

pDCs and B cells were isolated from buffy coat of healthy adult donors obtained from the Gulf Coast Regional Blood Center. This study was approved by the Institutional Review Board of the University of Texas M. D. Anderson Cancer Center, and informed consent was provided in accordance with the Declaration of Helsinki. A pDC-enriched population was isolated from peripheral blood mononuclear cell by positive selection using CD304 (BDCA-4/neutrophil-1) microbead kit (Miltenyi Biotec), and pDCs were isolated by sorting CD3+ (HIT3a) CD14+ (MOP9) CD16− (3G8) CD56− (B159) CD19− (HIB19) CD11c− (Bly6) CD4+ (RPA-T4) CD123+ (6H6; BioLegend) cells. A B cell–enriched population was isolated from peripheral blood mononuclear cell by negative selection using the human B-cell isolation kit II (Miltenyi Biotec) and sorting CD3− CD14− CD16− CD56− BDCA-2+ (AC144; Miltenyi Biotec) CD19+ cells. For some experiments, naive and memory B cells were isolated by sorting total B cells on the basis of CD27 (M-T271) expression. Unless indicated, all antibodies were purchased from BD Biosciences. Purity of sorted cells is greater than 98%.

pDC/B-cell coculture

pDCs and B cells were cultured in RPMI 1640 (Invitrogen) containing 10% fetal bovine serum (Atlanta Biologicals), 2mM l-glutamine, 100 U/mL penicillin, 100 U/mL streptomycin, 1mM sodium pyruvate, and 10mM N-2-hydroxyethylpiperazine-N'-2-ethanesulfonic acid. pDCs were cultured for 24 hours at a density of 10^6/mL in medium alone or with 1μM CpG (B type, 2006; Alpha DNA) and washed 3 times with culture medium to remove residual CpG before use in subsequent culture. B cells were cultured over CD40L-transfected L cells irradiated 6500 to 7500 cGy at an L-cell/B-cell ratio of 1:5. In some experiments, cells were cultured with recombinant CD40L (Alexis) at a concentration of 50 to 200 ng/mL. Similar results were obtained in both culture systems (data not shown). For analysis of B-cell proliferation, 2 × 10^6 B cells were cultured in 96-well round-bottom plates with 10^4 activated pDCs for a total of 4 days. A total of 1 μCi of [3H]-thymidine was added to each well for the last 18 hours, and cellular incorporation was measured. Alternatively, to analyze B-cell proliferation by carboxyfluorescein succinimidyl ester (CFSE) dilution, 5 × 10^6 B cells were labeled with CFSE and cultured with 2.5 × 10^4 activated pDCs. Cells were harvested after 5 days of culture, stained with allophycocyanin-labeled anti-CD19 to allow residual pDCs to be excluded, and CFSE dilution analyzed by flow cytometry. For analysis of immunoglobulin secretion, 2 × 10^9 B cells were cultured with 10^5 activated pDCs, IL-2 (50 U/mL) and IL-10 (10 ng/mL) for 14 days, at which time supernatant was collected and IgG and IgM levels determined by enzyme-linked immunosorbent assay (ELISA; Bethesda Laboratories). In some experiments, neutralizing rabbit polyclonal anti–IFN-α (2000 neutralizing U/mL; PBL Biomedical Laboratories), and anti–IFN-β antibodies (1000 neutralizing U/mL; PBL Biomedical Laboratories), and mouse anti–IFN-α/β receptor monoclonal antibody (10 μg/mL, MMHAR-2; PBL Biomedical Laboratories) were used.

Flow cytometry

To analyze CD70 expression, freshly isolated pDCs or pDCs cultured with CpG-A (1μM; Alpha DNA), CpG-B (1μM; Alpha DNA), irradiated herpes simplex virus (HSV)-1 (10 pfu/cell), influenza A virus (10 pfu/cell), or IL-3 (10 ng/mL; R&D Systems) were stained with purified anti-CD70 (clone KI-24; BD Biosciences) or purified isotype control (mouse IgG1; BD Biosciences) followed by phycoerythrin-labeled goat anti–mouse Ig (BD Biosciences). To analyze CD70 expression, freshly isolated B cells or B cells cultured with CD40L with or without IL-2 (50 U/mL) and IL-10 (10 ng/mL) were stained with phycoerythrin-labeled anti-CD27 (BD Biosciences).

Generation of CD70 transfectants

Full-length CD70 cDNA was amplified from cDNA derived from CpG-B-stimulated pDCs and cloned into the lentiviral vector FG9. The plasmid was cotransfected into HEK293T cells with the packaging constructs pMDLg/pRRE, RSV-Rev and CMV-VSVg.23 After 2 days, the culture supernatant containing the virus was collected and used to infect parental L cells. Expression of CD70 was confirmed by flow cytometry, and cells with a high level of CD70 expression were sorted and used in the generation of the antagonist anti-CD70 antibody.

Generation of antagonist anti-CD70 antibody

A BALB/c mouse less than 6 months of age was immunized with CD70-transfected L cells. Hybridomas secreting monoclonal antibodies recognizing CD70 were identified by ELISA and flow cytometry, and antagonist function was identified by screening clones for the ability to block the induction of proliferation of B cells cultured with IL-2 and IL-10 by CD70. Clone 113-16, which is IgG1 isotype, was used in all experiments.

Microarray analysis

Microarray analysis was carried out as described previously.15,24

Results

Expression of CD70 and CD27 in human immune cells

Microarray analysis (Figure 1A) indicates that CD70 was not expressed in resting pDCs, although expression of CD70 by pDCs was observed after stimulation with certain TLR7 and TLR9 ligands, including influenza virus (fluv) and CpG oligonucleotides but not HSV or IL-3. CD70 expression in pDCs was confirmed using flow cytometry. Strong and stable up-regulation of CD70 expression in pDCs was observed within 12 hours of CpG-B stimulation, reaching peak levels after 48 hours of stimulation (Figure 1B). Expression of CD70 in pDCs was observed after stimulation for 48 hours with influenza virus and CpG-A, but not HSV or IL-3 (Figure 1C). CD27, the receptor for CD70, is specifically expressed on memory B cells. Microarray analysis indicates that CD27 was expressed by B cells, CD8+ T cells, and CD4+ T-cell subsets (Figure 1A). Total B cells were used for microarray analysis, which can account for the relatively low level of CD27 expression that was observed. Expression of CD27 on human peripheral B cells, both resting and on stimulation with CD40L or CD40L plus IL-2 and IL-10, was confirmed using flow cytometry (Figure 1D). CD27 expression was observed on a subset of resting B cells and decreases on activation. Because CD70 is expressed on activated pDCs, it could potentially play an important role in the interaction between pDCs and CD27+ memory B cells.

pDCs stimulated with CpG promote the proliferation and Ig secretion of human peripheral B cells

To examine the role of pDCs in the induction of B-cell proliferation and Ig secretion, an in vitro pDC/B-cell coculture system was established. CpG-B was selected to stimulate pDCs because it has a strong capacity to activate pDCs and induce the expression of CD70 but does not lead to significant IFN production.25 B cells were labeled with CFSE and cultured over irradiated CD40L L cells alone or with IL-2, IL-10, or IL-2 plus IL-10. Resting pDCs or pDCs, which had been stimulated for 24 hours with CpG-B (CpG-B-pDCs), were added to B cells, and proliferation was examined after 5 days of coculture (Figure 2A). The data indicate
that CpG-B-pDCs had a strong capacity to promote the proliferation of CD40L-activated B cells. Addition of IL-2 had little effect on the induction of B-cell proliferation by CpG-B-pDCs. Addition of IL-10 or IL-2 plus IL-10 enhanced B-cell proliferation in response to CD40L stimulation but did not enhance the ability of CpG-B-pDCs to promote B-cell proliferation. Similar results were observed when proliferation was measured using [3H]-thymidine incorporation (data not shown) and when B cells were activated with recombinant CD40L instead of CD40L-transfected cells (data not shown). A maximum 2-fold increase in actual B-cell numbers was observed in pDC/B-cell coculture experiments (data not shown). CpG-B alone was able to enhance the proliferation of B cells cultured over irradiated CD40L-transfected L cells alone or with IL-2, IL-10, or IL-2 plus IL-10 (Figure 2A). Using a similar pDC/B-cell coculture system, the ability of CpG-B-pDCs to promote the secretion of IgM and IgG from B cells was examined (Figure 2B). The addition of CpG-B-pDCs to B cells cultured with CD40L alone could not promote secretion of IgM or IgG. A moderate increase in IgG secretion was observed when CpG-B-pDCs were added to B cells cultured with CD40L and IL-2 compared with resting pDCs, although the increase in IgM secretion observed was minimal. Addition of CpG-B-pDCs to B cells cultured with CD40L...
Data indicate that the induction of B-cell Ig secretion by pDCs is contact-dependent and independent of IFN-α and residual CpG-B. (A) pDC/B-cell coculture was established with pDCs sequestered in the upper transwell chamber and B cells in the lower chamber. B cells were labeled with CFSE before culture, and proliferation was assessed by measuring CFSE dilution on day 5. The shaded histogram represents B cells cultured with CD40L; dashed histogram, B cells cultured with resting pDCs; and solid histogram, B cells cultured with CpG-B-pDCs. Data shown indicate 1 donor representative of 3. Supematant was collected on day 14, and IgG and IgM concentrations were determined by ELISA. The mean plus or minus SEM of duplicate results from 3 individual donors is shown. (B) pDC/B-cell coculture was carried out as described previously, except that a cocktail of antagonist antibodies against IFN-α, IFN-β, and IFN-α/β receptor was added. Proliferation was assessed by measuring [3H] thymidine incorporation on day 5, and IgG and IgM levels in supernatant were measured by ELISA on day 14. Mean results ± SEM from 1 experiment representative of 3 are shown. (C) B cells were labeled with CFSE and cultured with CD40L and 1 μM CpG-B, resting pDCs, or CpG-B-pDCs. Chloroquine was added to the B cells at the indicated concentrations 30 minutes before the addition of CD40 or pDCs. Proliferation was assessed by measuring CFSE dilution on day 5. Control experiments: Results are shown in the top panel; shaded histogram represents B cells cultured with CD40L; dotted histogram, B cells cultured with CD40L plus 1 μM CpG-B, pDC/B-cell coculture experiments; Results are shown in the bottom panel; shaded histogram represents B cells cultured with CD40L; dashed histogram, B cells cultured with resting pDCs; and solid histogram, B cells cultured with CpG-B-pDCs. Supernatant was collected on day 14, and IgG and IgM concentrations were determined by ELISA. Chloroquine was titrated to determine the optimal concentration. Fixed numbers of B cells and pDCs were added. Results of 1 experiment representative of 3 are shown. Data were analyzed by unpaired t test: *P < .05 and **P < .01.

Induction of B-cell proliferation and Ig secretion by CpG-stimulated pDCs is contact dependent and independent of type I interferon and residual CpG-B

Previous studies have indicated that the interaction between pDCs and B cells is largely mediated by soluble factors, including IFN-α and IL-6,8-10. We have shown that CD70 is expressed by CpG-B-pDCs (Figure 1A-C) and that CpG-B-pDCs are capable of inducing B-cell proliferation and Ig secretion. If the induction of B-cell proliferation and Ig secretion by CpG-B-pDCs is mediated by CD70, then it should occur in a contact-dependent manner. To address whether the induction of B-cell proliferation and Ig secretion by CpG-B-pDCs is mediated by cell-cell interactions or soluble molecules, we conducted a transwell experiment by culturing pDCs in the upper chamber and B cells in the lower chamber. Both wells were coated with CD40L-L cells, and proliferation was measured by CFSE dilution. Blocking the interaction between pDCs and B cells completely abrogated the induction of proliferation and Ig secretion by CpG-B-pDCs (Figure 3A). Using a flow cytometry-based assay, conjugate formation between pDCs and B cells was observed, further supporting the physical interactions between pDCs and B cells (data not shown). Published data indicate that the induction of B-cell Ig secretion by pDCs is largely mediated through IFN-α, 8-10 To exclude a role for IFN-α in the induction of B-cell proliferation and Ig secretion by CpG-B-pDCs, pDC/B-cell coculture was carried out in the presence of a cocktail of antagonistic antibodies against IFN-α, IFN-β, and IFN-α/β receptor. In the presence of the antagonistic antibody cocktail, the induction of B-cell proliferation by pDCs was significantly enhanced, although the induction of IgM and IgG secretion was not significantly affected (Figure 3B). Because CpG-B alone was able to induce B-cell proliferation (Figure 2A) and Ig secretion (Figure 2B), experiments were conducted to ensure that the induction of proliferation and Ig secretion observed on the addition of pDCs were not the result of carryover of residual CpG-B. To rule out the possibility that the B-cell proliferation and Ig secretion observed after coculture with CpG-B-pDCs was the result of residual CpG-B carried over on the pDCs after stimulation, chloroquine was added to culture wells. At a chloroquine concentration of 1 μg/mL, the induction of proliferation of CD40L-activated B cells by CpG-B was completely blocked, although the induction of B-cell proliferation by CpG-B-pDCs was only slightly reduced (Figure 3C). The ability of CpG-B to induce IgG and IgM secretion from B cells activated by CD40L, IL-2, and IL-10 was blocked by 0.5 μg/mL of chloroquine (Figure 3C). At this concentration, the induction of IgM secretion by CpG-B-pDCs was reduced to a level similar to that observed with resting pDCs, although the induction of IgG secretion by CpG-B-pDCs was unaffected (Figure 3C). These data show that pDCs have the ability to promote B-cell proliferation and Ig secretion in a contact-dependent, interferon-independent manner, and that the induction of B-cell proliferation and the secretion of IgG occur independent of residual CpG-B.
pDCs stimulated with CpG-B preferentially promote the proliferation and Ig secretion of CD27+ memory B cells

If CpG-B-pDCs promote B-cell proliferation and Ig secretion via engagement of CD27 on B cells with CD70, then it would be anticipated that CpG-B-pDCs would preferentially promote the induction of proliferation and Ig secretion from CD27+ memory B cells. Naive and memory B cells were sorted from total CD19+ B cells based on CD27 expression and used in pDC/B-cell coculture with resting pDCs or CpG-B-pDCs. Proliferation was measured by CFSE dilution on day 5. CpG-B-pDCs induced proliferation of both naive and memory B cells stimulated by CD40L, although a higher level of proliferation was observed for memory B cells compared with naive B cells (Figure 4A). Supernatant was collected on day 14, and IgG and IgM levels were determined by ELISA. CpG-B-pDCs were able to promote a slight but significant increase in IgM secretion from naive B cells but did not have a significant effect on IgG secretion (Figure 4B). CpG-B-pDCs were able to promote a significant increase in the induction of both IgG and IgM from CD27+ memory B cells (Figure 4B). The data indicate that, although CpG-B-pDCs are able to induce the proliferation of both naive and memory B cells, the induction of Ig secretion is restricted to the memory subset.

Antagonist anti-CD70 antibody limits the induction of B-cell proliferation and Ig secretion by pDC

To specifically investigate the role of CD70 in the induction of B-cell proliferation and Ig secretion by CpG-B-pDCs, an antagonist anti-CD70 antibody was generated that could completely block the function of CD70 at a concentration of 10 µg/mL (data not shown). Resting or CpG-B-pDCs were preincubated with antagonist anti-CD70 antibody or purified mouse IgG1 isotype before the addition of B cells to the culture. In the presence of antagonist anti-CD70 antibody, the ability of CpG-B-pDCs to induce the proliferation of total B cells was significantly reduced (Figure 5A). In the presence of antagonist anti-CD70 antibody, IgG secretion was significantly reduced to a level comparable with that induced by resting pDCs (Figure 5B). The induction of IgM secretion was reduced in some but not all donors, but the result is not statistically significant. This is consistent with earlier data indicating that the induction of IgM secretion by CpG-B-pDCs was largely the result of residual CpG-B (Figure 3C). These data indicate an important role for CD70 in the regulation of B-cell proliferation and IgG secretion by CpG-B-pDCs.

Discussion

The ability of pDCs to promote plasma cell differentiation and Ig secretion through the production of soluble factors, including IFN and IL-6, has been well documented,8-10 although the role of additional factors, including tumor necrosis factor receptor-ligand interactions, has not been addressed. We have shown that CD70 is highly expressed on pDCs in response to stimulation with the TLR9 ligand CpG-B. Using a pDC/B-cell coculture system, the interaction between pDCs and B cells was further characterized and CD70 was identified as an important factor in the regulation of B-cell proliferation and Ig secretion by pDCs.

CpG-B-pDCs can promote the proliferation of CD40L-activated peripheral B cells in the absence of additional cytokines. However, the addition of IL-2 and IL-10 is required for CpG-B-pDCs to enhance the secretion of IgG and IgM from CD40L-activated B cells. The induction of B-cell proliferation and Ig secretion by CpG-B-pDCs was shown to be contact-dependent, and independent of IFN. These data conflict with published studies that
indicate an important role for IFN in the induction of IgG secretion from CD40L-activated B cells by virus stimulated pDCs, and a marginal role if any for cell-cell contact.8-10 On stimulation with CpG-B, pDCs produce very little IFN but exhibit robust expression of CD70. In contrast, pDCs stimulated with influenza virus produce a large amount of IFN but express a relatively low level of CD70 on only a subset of cells. The level of CD70 expressed by pDCs in response to flu stimulation may be too low to influence B-cell proliferation or differentiation. In addition, IFN is produced very rapidly on stimulation with flu, usually within the first 12 hours,15 whereas peak levels of CD70 expression are observed after 48 hours of stimulation. It is possible that the B cell would be exposed to IFN before CD27 ligation, which might alter the response to CD70 or that there are 2 distinct phases of interaction between pDCs and B cells, a rapid IFN-dependent phase, followed by a later IFN-independent CD70-dependent phase. It is also possible that IFN stimulation might affect the expression level of CD27 on B cells or the level of CD70 expressed by pDCs. Initial studies indicated that pDCs promoted mostly IgG secretion from B cells activated by CD40L stimulation or B-cell receptor cross-linking in the presence of CpG-C and led to speculation that pDCs may target the memory B-cell subset.8-10 It was shown recently that pDCs could specifically promote the proliferation of memory B cells in response to CpG-B stimulation.10 CpG-B-pDCs were shown to promote the proliferation of both naive and memory B cells; however, Ig secretion was restricted to the CD27+ memory B cell subset. In addition, CD70 was shown to have an important role in the induction of B-cell proliferation and Ig secretion by pDCs. This provides a mechanism to explain the ability of pDCs to specifically influence the growth and differentiation of CD27+ memory B cells.

Systemic lupus erythematosus (SLE) is an autoimmune disease characterized by tissue damage resulting from immune complex deposition, and high titers of autoantibodies specific for DNA and nucleosomes.20-28 Low numbers of pDCs are found in the blood of SLE patients as they have migrated to sites of inflammation. However, increased levels of IFN-α, probably produced by pDCs, are observed in the serum of approximately 20% to 30% of SLE patients. IFN-α enhances the differentiation of monocytes into dendritic cells that can take up and express antigens from apoptotic cells leading to autoimmunity. IFN-α also promotes the development of autoreactive plasmablasts. Abnormalities in the composition of the peripheral B-cell compartment have been observed in SLE patients. Although decreased numbers of naive and CD27+ memory B cells are found in SLE patients, increased numbers of CD27+ high plasma cells29 and CD27+ high plasma cell precursors30 have been observed and levels of these cells have been found to correlate with disease severity.31 SLE patients treated with rituximab, an antibody that depletes CD20+ B cells, who exhibit a delayed reconstitution of CD27+ memory B cells after treatment, have a better response to the therapy than other patients with better relief from symptoms and reduced levels of autoantibodies in the serum.12 There is considerable evidence that pDCs are activated in SLE and that CD27+ B cells have an important role in the disease. It is possible that activated pDCs expressing CD70 may drive the expansion of these cells and promote their differentiation into antibody-secreting plasma cells, thereby promoting autoantibody production and exacerbation of symptoms.

Acknowledgments

The authors thank David He, Karen Ramirez, and Amy Cortes for assistance with cell sorting, and Long Vien for assistance with the generation of the antagonist anti-CD70 antibody. This work was supported by the M. D. Anderson Cancer Foundation and in part by the National Institutes of Health through M. D. Anderson Cancer Center Support Grant CA016672.

Authorship

Contribution: J.S. designed and performed research, analyzed data, and wrote the manuscript; Y.-H.W. and T.I. performed research and analyzed data; K.A. designed and performed research; and Y.-J.L. designed research and reviewed and approved the manuscript.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

Correspondence: Yong-Jun Liu, University of Texas, M. D. Anderson Cancer Center, Department of Immunology, Unit 901, PO Box 130402, Houston, TX 77030-1903; e-mail: yjliu@mdanderson.org.


Plasmacytoid dendritic cells regulate B-cell growth and differentiation via CD70

Joanne Shaw, Yui-Hsi Wang, Tomoki Ito, Kazuhiko Arima and Yong-Jun Liu