Reciprocal role of GATA-1 and vitamin D receptor in human myeloid dendritic cell differentiation

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Two major pathways of human myeloid dendritic cell (DC) subset differentiation have previously been delineated. Langerhans cells (LCs) reside in epithelia in the steady state, whereas monocytes can provide dendritic cells (DCs) on demand in response to inflammatory signals. Both DC subset pathways arise from shared CD14+/CD16+ monocyte precursors, which in turn develop from myeloid committed progenitor cells. However, the underlying hematopoietic mechanisms still remain poorly defined. Here, we demonstrate that the vitamin D3 receptor (VDR) is induced by transforming growth factor β1 during LC lineage commitment and exerts a positive role during LC generation. In contrast, VDR is repressed during interleukin-4 (IL-4)–dependent monocyte-derived DC (moDC) differentiation. We identified GATA-1 as a repressor of VDR. GATA-1 is induced by IL-4 in moDCs. Forced inducible expression of GATA-1 mimics IL-4 in redirecting moDC differentiation and vice versa, GATA-1 knockdown arrests moDC differentiation at the monocyte stage. Moreover, ectopic GATA-1 expression stabilizes the moDC phenotype under monocyte-promoting conditions in the presence of vitamin D3 (VD3). In summary, human myeloid DC subset differentiation is inversely regulated by GATA-1 and VDR. GATA-1 mediates the repression of VDR and enables IL-4–dependent moDC differentiation. Conversely, VDR is induced downstream of transforming growth factor β1 and is functionally involved in promoting LC differentiation. (Blood. 2009;114:3813-3821)

Introduction

Two in vitro systems for the generation of human dendritic cells (DCs) are widely used for basic and clinically oriented research: monocyte-derived DCs (moDCs) and hematopoietic progenitor cell (HPC)–derived DCs. In the first system, granulocyte-macrophage colony-stimulating factor (GM-CSF) plus interleukin-4 (IL-4) are added to peripheral blood monocytes to generate moDCs. This system is most widely applied for generating human DCs. In the second model, CD34+ HPCs are stimulated in transforming growth factor β1 (TGF-β1)–supplemented serum-free cultures (containing basic DC cytokines GM-CSF, tumor necrosis factor α [TNFα], stem cell factor [SCF] with or without Flt3L) for generating Langerhans cells (LCs), a DC subtype that resides within epithelial tissue. Exclusion of TGF-β1 from this latter culture system abrogates LC differentiation and de-represses a program of monocyte generation from CD34+ cells. When these latter HPC cultures are initiated in the presence of serum without TGF-β1, 2 pathways of CD1a+ DCs are simultaneously generated, i.e., LCs and non-LC interstitial/dermal-type DCs. These populations arise from 2 separate precursor pathways at culture day 5. A CD1a+CD14+ precursor gives rise to LCs, whereas a CD1a−CD14+ precursor can give rise to non-LC DCs, which share many features with moDCs generated from peripheral blood monocytes. This CD1a+CD14+ intermediate population can be further subdivided into 2 functional subsets based on CD11b (CD11b−CD14+ versus CD11b+CD14+ monocyctic cells). CD11b+CD14+ cells can be induced by GM-CSF plus IL-4 to develop into intDC/moDCs or by M-CSF to macrophage differentiation. Conversely, CD11b−CD14+ intermediates, which represent early monocyctic cells, retain LC differentiation capacity in the presence of TGF-β1. Therefore, a CD14+ monocyctic precursor can be induced to develop along 2 separate pathways in vitro depending on IL-4 (moDCs) versus TGF-β1 (LCs) cytokine signals.

IL-4 versus TGF-β1 antagonize each other’s function to induce moDCs versus LCs. Neutralizing anti–TGF-β1 mAb represses LC generation in favor of CD14+ monocyte generation. Similarly, the addition of IL-4 to TGF-β1–supplemented LC generation cultures of CD34+ cells represses LC differentiation in favor of inducing non-LC DC differentiation. Conversely, the addition of TGF-β1 to GM-CSF plus IL-4–containing moDC cultures of peripheral blood monocytes polarizes these cells toward an LC-like phenotype. In line with this, TGF-β1 in the epidermal microenvironment is critical for LC differentiation in vivo. Functional differences between moDCs versus LCs are increasingly recognized. However, very little is known about the molecular mechanisms controlling the lineage commitment of these cells to develop into different DC subsets (i.e., LCs and non-LC moDCs).

We recently performed mechanistic studies on the involvement of family members of the nuclear hormone receptor system, i.e., retinoic X receptor-alpha (RXRα) and vitamin D3 receptor (VDR) in myelopoiesis. RXRα and VDR form heterodimeric complexes that, on ligand binding (ligand: calcitriol, 1,25-vitamin D3; VD3), can induce monocyte differentiation of myeloid progenitors. By using a functional screen we recently identified the hematopoietic master transcription factor GATA-1 as a dominant interfering


The online version of this article contains a data supplement.
molecule of VDR:RXRa-induced monocyte differentiation from U937 model cells (S.T. and H.S., unpublished data, February 2006). Because monocyte genes are consistently repressed concomitantly with LC2,3 or moDC1 subset differentiation, we studied whether the GATA-1/VDR:RXRa interdependence observed in our cell line model could possibly play a role in the development of monocytes to their resultant DC subsets. Moreover, a recent study has already implicated GATA-1 in murine DC development.14 We found that GATA-1 and VDR are inversely present in moDCs versus LCs and that moDCs require GATA-1 downstream of IL-4 for development. Furthermore, GATA-1 represses VDR in DC precursors. In sharp contrast, LC precursors up-regulate VDR in response to the lineage-instructive signal TGF-β1, and VDR plays a positive role in LC differentiation.

Methods

Cytokines and reagents

The following cytokines were used: SCF, thrombopoietin, IL-4, TNFα, IL-15, IFN-α (PeproTech); TGF-β1 (R&D Systems); Flt3L (Amgen); GM-CSF (Novartis); Ilu-2,25-dihydroxyvitamin D3 (1,25-VD3), 25-hydroxyvitamin D3 (25-VD3), doxycycline (DOX; Sigma-Aldrich); and VDR antagonist ZK159222 (Bayer Schering Pharma AG).

Cell isolation

Cord blood samples were collected during healthy full-term deliveries. Approval was obtained from the Medical University of Vienna Institutional Review Board for these studies. Informed consent was provided in accordance with the Declaration of Helsinki. CD34+ cells were isolated as previously described.15 CD14+ monocytes were isolated by magnetic-activated cell sorting (Miltenyi Biotec) from cord blood according to the manufacturer’s instructions. For experiments shown in Figure 2B, CD34+ cells were sorted by fluorescence-activated cell sorting (FACS; FACSAria; BD Biosciences) into greater than 95% pure CD34+ cells were sorted by fluorescence-activated cell sorting (FACS; FACSAria; BD Biosciences; CD45RA, clone MEM-56; Caltag). Enriched in myeloid progenitors (FITC: CD34, clone 581; BD Biosciences; PE: CD45RA, clone MEM-56; Caltag Laboratories; APC: CD19, clone F125-C1; Caltag).

DC generation cultures

X-VIVO 15 medium (BioWhittaker) was used for primary cell cultures supplemented with GlutaMAX (2.5 mM; Invitrogen), penicillin/streptomycin (P/S; 125 U/mL each; Sigma). In addition, 10% FBS (Invitrogen) were added for moDC generation. CD34+ progenitor cells were expanded in Flt3L, SCF, and thrombopoietin (each 50 ng/mL) for 1 to 3 days. LCs were generated in GM-CSF (100 ng/mL), SCF (20 ng/mL), Flt3-L (50 ng/mL), TNFa (2.5 ng/mL), and TGF-β1 (0.5 ng/mL). MoDCs from expanded CD34+ cells were obtained in GM-CSF (100 ng/mL), SCF (20 ng/mL), Flt3-L (50 ng/mL), TNFa (2.5 ng/mL) for 4 days, followed by GM-CSF (100 ng/mL), IL-4 (25 ng/mL) for 3 days. Monocytes were cultured with GM-CSF (50 ng/mL) and IL-4 (25 ng/mL) for 6 days. Absolute cell numbers for LCs and moDCs generated from 2 × 10^5 CD34+ cells were in the range of 1 to 2 × 10^6 and 2 to 4 × 10^6, respectively. Monocyte cultures were initiated with 10^5 cells, and cell numbers stayed equivalent or only slightly dropped until day 6. Cell viability usually exceeded 90%, as assessed routinely by morphologic inspection and trypan blue dye exclusion. Cell cultures containing less than 90% viable cells were excluded from further analyses.

Cell lines

Phoenix (Ph) cells were kindly provided by G.P. Nolan (Stanford University). 293T cells were purchased from ATCC, and both were maintained in DMEM (Sigma)/10% FBSA,-glutamine (Sigma) and P/S. U937Te cells are described elsewhere.15 K562 cells were obtained from ATCC. K562 and U937Te cells were cultured in RPMI with 10% FBS, t-glutamine, and P/S.

Human foreskin tissue preparation and immunofluorescence staining

Noninflamed human foreskin from routine circumcisions was obtained as discarded material after ethical committee approval of the Medical University of Vienna (code EK555/2005). Frozen sections of foreskin were fixed and incubated with mouse-αVDR mAb (D-6; Santa Cruz Biotechnology) followed by goat-αmouse F(ab’)2, Alexa Fluor 488 (Invitrogen), mouse-αhuman CD207-PE (DGGM4; Beckman Coulter), and mouse-αhuman CD45-APC (J33; Beckman Coulter). Immunostained sections were analyzed by a confocal laser scanning microscope (LSM 510; Carl Zeiss GmbH).

1,25-VD3 and 25-VD3 detection with radioimmunoassay

Skin pieces were homogenized in cold PBS by an Ultra Turrax (Janke-Kunkel GmbH). After centrifugation at 15,000 g for 10 minutes at 4°C, protein was precipitated with acetoniitril; 25-VD3 and 1,25-VD3 were analyzed with radioimmunoassay kits from DiaSorin according to the manufacturer.

Vector constructs and gene transduction

Murine stem cell virus–based retroviral vector MIG-R1 was obtained from G.P. Nolan (Stanford University). Human GATA-1 cDNA was amplified by polymerase chain reaction (PCR) by using the primers 5’-TCCCCAGAGGCTCCATGGAAG-3’ and 5’-GGTGTGGTCAGAATTCTGGCTAC-3’ from a human fetal liver cDNA library (Stratagene) and cloned into the EcoRI site of the MIG-R1. PINCO-VDR-GFP was kindly provided by A. Seshire (University of Frankfurt). The retroviral tetracycline-inducible system (Tet-on system) was described elsewhere.16 Briefly, the first vector encodes the Tet-activator, pT AlertsCDs. The second vector encodes human GATA-1 cDNA, which was cloned into the EcoRI site of the self-inactivating retroviral vector pHR-RES-NGFR and pHR-RES-GFP (kindly provided by F. Rossi, University of British Columbia, respectively). The packaging cell lines Ph-E, Ph-Gag-Pol (Ph-GP), and 293T were transfected as previously described.12,17 Retroviral infections of target cells was done as previously described.15 The silencing construct targeting GATA-1 (5’-ACGGCGCTCTATCACAAAGTAA-3’) in a hairpin structure (OligoID: V2HS_114063) was inserted into the Xhol-MluI sites of pGIPZ lentiviral vector (Open Biosystems). The nonsilencing pGIPZ (22-mer sequence 5’-ATCTGCTTGGGGCCGAGATAGAAG-3’) was obtained from the same company. The GIPZ vector backbone has a GFP marker molecule to track infected cells. Lentiviral infections were carried out according to standard procedures for silencing experiments. In brief, 293T cells were cotransfected with pMD2.G and pSAX2 (Addgene Inc) and the respective GIPZ-shRNA silencing construct by using 1 μg/mL Polybrevine (Sigma) to enhance lentiviral infections. Viral supernatants were used to infect monocytes, which were then differentiated to moDCs.

Flow cytometry

Flow cytometric analysis was performed as previously described.12,15,16 Sorting and FACS analyses were carried out on BD FACSAria, FACS Calibur, or LSRII cytometers, and data were analyzed with CellQuest Pro or FACS Diva software (all BD Biosciences).

Western blot analysis

Western blot analysis was performed as previously described.15 Detection was carried out with goat-αGATA-1 (C-20), rabbit-αPU.1 (T-21), rabbit-αVDR (H81) (Santa Cruz), rabbit-αActin (A-2066; Sigma), sheep-αI(1)OHase (Binding Site), followed by horseradish peroxide–conjugated donkey-αgoat or goat-αrabbit IgG (H+L) (Pierce) or rabbit-αsheep (Dako) antibodies. Cells (10^6) per lane were analyzed.

PCR

For reverse transcription (RT)–PCR, total RNA was isolated by using the RNeasy micro kit (QIAGEN) according to the manufacturer’s instructions. DNA-free total RNA was subjected to cDNA synthesis by using oligo-dT
primers and M-MuLV reverse transcriptase (Fermentas) according to the manufacturer’s instructions. Quantitative real-time RT-PCR was performed as described.19 The following primers were used: VDR, 5′-CCAGTTCTGG-TGGAATGATGG-3′ and 5′-AGATTGGAAGCTTGACGA-3′; HPRT, 5′-GACCAGTCAACAGGGGACAT-3′ and 5′-AACACTTGTGGGTGCC-TTTTC-3′; GAPDH, 5′-GAAATCCCATACACATCTCCAGG-3′ and 5′-CGCGGCCATCACGCACAGTTCC-3′.

Statistical analysis

Data are presented as mean plus or minus SEM. Statistical analysis was performed using the paired, 2-tailed Student t test. P values less than .05 were considered significant.

Results

Transcription factors GATA-1 and VDR are inversely expressed in human myeloid DC subsets

To study mechanisms of human myeloid DC differentiation, we established 2 in vitro differentiation culture models, initiated (1) by CD34+ umbilical cord blood cells or (2) by peripheral blood monocytes (Figure 1A). The first model allows for generation of either TGF-β1–dependent LCs3 or, by a 2-step culture system, IL-4–dependent moDCs.5 These IL-4–dependent moDCs share many phenotypic properties with moDCs generated from peripheral blood monocytes.13 CD11b has been previously identified as the marker molecule that best discriminates IL-4–dependent moDCs cells from LCs.5 Therefore, we used CD11b as a marker for moDCs (Figure 1A). A phenotypic comparison of in vitro–generated LCs and moDCs is shown in supplemental Figure 1 (available on the Blood website; see the Supplemental Materials link at the top of the online article). LCs and moDC subsets were identified as CD207+CD1a+ and CD11b+CD1a+ cells, respectively (see gate settings in supplemental Figure 1A) and analyzed for informative marker molecules. LCs but not moDCs express CD207 and E-cadherin. Conversely, LCs lack CD11b (supplemental Figure 1A). Furthermore, LCs and moDCs lack CD14. Moreover, we confirmed previous observations3 that moDCs derived from CD34+ cells but not LCs express CD209 (DC-SIGN) and that moDCs express higher levels of CD1c than do LCs (supplemental Figure 1B). Therefore, the here described in vitro generated DC subsets phenotypically resemble DC populations extensively characterized in previous studies.3,5,20 We recently identified the transcription factor GATA-1 as a repressor of VD3-induced monocytopoiesis in U937 mononuclear cells (S.T. and H.S., unpublished data, February 2006). Interestingly, ectopic GATA-1 expression decreases VDR mRNA expression levels compared with control transduced cells as shown from quantitative real-time RT-PCR analyses (Figure 1B). In line with this, a previous study has shown that monocytes down-regulate VDR expression during GM-CSF/IL-4–dependent moDC differentiation.21 These observations, together with a recent study that implicated GATA-1 in murine DC development,14 prompted us to perform a comparative analysis of GATA-1 and VDR in LCs versus moDCs. Western blot analyses showed that GATA-1 is detectable in GM-CSF/IL-4–dependent moDCs generated from CD34+ cells or peripheral blood monocytes, whereas LCs lack GATA-1 (Figure 1C). Conversely, VDR is expressed at higher levels in LCs compared with moDCs (Figure 1C). Therefore, GATA-1 and VDR are inversely expressed in these DC subsets. Time kinetics Western blot analyses of GM-CSF/IL-4–dependent moDC cultures have shown that GATA-1 expression is absent in monocytes at culture initiation (day 0). However, GATA-1 expression appears at day 2 and gradually increases thereafter, until day 6 (Figure 1D). Conversely, VDR is expressed in monocytes (day 0) and is gradually repressed by day 2 during GM-CSF/IL-4–dependent moDC differentiation (Figure 1D top panel). Phenotypic analyses have shown that at day 3, the monocyte marker CD14 was scarcely detectable and that approximately half of the population expressed CD1a, whereas at day 6, more than 90% of cells were CD1a+CD11b+CD14+ (Figure 1D bottom panel). These results indicate that GATA-1 up-regulation correlates with the differentiation of CD1a+CD11b+ moDCs between day 3 and day 6, as well as with the gradual repression of VDR during GM-CSF/IL-4–dependent moDC differentiation. In summary, GATA-1 and VDR are inversely expressed in DC subsets, suggesting a differential role during myeloid DC subset differentiation.

Functional involvement of VDR in TGF-β1–dependent LC differentiation

Next, we focused on the role of VDR in LCs. Analysis of human skin samples confirmed that CD45+CD207+ LCs express VDR in vivo. It can be seen from the high-power magnification insert in
For comparison, TGF-1 was omitted from parallel cultures. For H9252 were induced to differentiate into LCs in the presence of TGF-

VDR-transduced cells showed strongly increased gating in a separate FACS diagram (data not shown). In the presence of TGF-1 stimulation of myeloid progenitors (Figure H9252 RT-PCR analysis showed strong induction of VDR within 6 hours extending previous observations.22 Because LCs require TGF-1 for their generation from progenitors, 2 we next asked whether ectopic VDR might promote LC differentiation. Thus, myeloid progenitors (CD34 TGF-1 induces VDR concomitant with LC lineage differentiation. Therefore, gen-transduced progenitor cells were induced to develop into GM-CSF/IL-4–dependent CD1a CD207 CD45RA CD19 cells) CD1a versus CD207. Bars represent the mean of percent-

percentages of CD1a CD207 LCs relative to empty vector control (Figure 2C FACS and bar diagrams), indicating that VDR promotes LC differentiation. This effect was abrogated in the absence of TGF-1 (Figure 2C FACS and bar diagram), showing that VDR expression alone is not sufficient to induce LC differentiation. We confirmed that VDR-transduced LCs lack marker molecules associated with moDC differentiation or monocytes (CD11b– and CD14–; supplemental Figure 1C). Because VDR is repressed during GM-CSF/IL-4–dependent moDC generation, we next determined the role of ectopic VDR during this differentiation process. Therefore, gen-transduced progenitor cells were induced to develop into GM-CSF/IL-4–dependent CD1a CD11b moDCs according to Figure 1A. Ectopic VDR expression reduced the generation of GM-CSF/IL-4–dependent CD1a CD11b moDCs (Figure 2D), indicating an inhibitory effect of VDR expression on moDC differentiation. To address whether endogenous VDR is required for LC lineage differentiation, we generated LCs in the presence or absence of the VDR antagonist ZK159222 (10 \( \mu M \)). At day 7 cells were harvested and analyzed for CD207 versus CD1a. Data are representative of 3 independent experiments.
Endogenous GATA-1 mediates moDC differentiation downstream of IL-4

Because GATA-1 was specifically expressed in moDCs (Figure 1C), we next addressed the functional role of endogenous GATA-1 in these cells. MoDCs generated in response to GM-CSF plus IL-4 express GATA-1, whereas fresh monocytes lack detectable GATA-1 expression (Figure 3A). GATA-1 expression levels in moDCs are lower than in K562 leukemia cells, which typically serve as a reference cell line, given its abundant GATA-1 expression (Figure 3A). Importantly, omission of IL-4 from moDC generation cultures markedly reduces GATA-1 expression levels (Figure 3A), suggesting that IL-4 induces GATA-1 expression. This interpretation is supported by the time-kinetic experiment shown in Figure 1D, showing that GATA-1 protein levels gradually increase during IL-4–induced moDC differentiation from day 2 to day 6 and that GATA-1 expression positively correlates with the induction of a CD11b+/CD123+ moDC phenotype. We then asked whether endogenous GATA-1 is required for moDC differentiation downstream of IL-4. Therefore, we performed loss-of-function experiments of GATA-1 in moDC differentiation by using a lentiviral GATA-1 shRNA vector. Control experiments confirmed that this silencing vector reduces GATA-1 protein levels in K562 cells (Figure 3B top panel) and in moDCs (Figure 3B bottom panel) relative to nonsilencing shRNA control vector. Monocytes were transduced with shGATA1 or a nonsilencing control and were differentiated to nonsilencing shRNA control vector. Monocytes were transduced with a lentiviral vector encoding nonsilencing or GATA-1 shRNA sequences (ie, shGATA1-GFP or control). A simplified graphical representation of the vector backbone is shown. Infected monocytes from day 6–generated cells were analyzed for their phenotype (Figure 4A). The here used Tet-on system allowed tightly regulated DOX-dependent induction of gene expression as visualized by NGFR, because in the absence of DOX cells remained NGFR− (Figure 4C). DOX-induced GATA-1 expression promoted the generation of CD11b+/CD1a+ cells (FACS diagrams in Figure 4B and bar statistics in Figure 4D). Control experiments confirmed that GATA-1 expression in GM-CSF/IL-4–induced moDCs fails to inhibit CD1a expression (supplemental Figure 2). This observation is in line with above-described induction of endogenous GATA-1 during moDC differentiation (Figures 1D and 3A). Given the above-mentioned observations that ectopic GATA-1 inhibits VDR in U937 cells (Figure 1B), we then studied whether GATA-1 similarly inhibits VDR expression in LC precursors. Thus, we transduced cells under LC generation conditions with GATA-1–IRES-GFP or empty vector control, followed by cell sorting of GFP+ cells 48 hours after transduction. In line with our data in U937 cells (Figure 1B), ectopic GATA-1 clearly reduces VDR mRNA expression in LC precursors, as shown from quantitative real-time RT-PCR analysis (Figure 4E). Together these experiments show that ectopic GATA-1 mimics IL-4 stimulation in inducing CD11b+/CD123+ moDCs from day 5–generated LC precursors. Furthermore, ectopic GATA-1 in LC precursors diminishes VDR mRNA expression.
Ectopic GATA-1 allows CD1a<sup>+</sup>CD11b<sup>+</sup> moDC differentiation during VD3-induced monopoiesis

The addition of 1.25-VD3 to DC generation cultures is well described to inhibit CD1a<sup>+</sup> DCs in favor of monocyte generation. Consequently, GATA-1–mediated down-regulation of VDR may possibly inhibit the effects of the VDR ligand 1.25-VD3 on DC differentiation. Therefore, we hypothesized that low levels of ectopic GATA-1 expression as observed in moDCs (Figures 1D and 3A) may desensitize precursor cells to VD3-mediated inhibition of DC generation. Moreover, given the above-described ectopic expression experiments (Figure 4B), GATA-1 expression could possibly induce a CD1a<sup>+</sup>CD11b<sup>+</sup> moDC phenotype. Consequently, increased percentages of CD1a<sup>+</sup>CD11b<sup>+</sup> moDCs may occur among GATA-1–expressing cells in the presence of 1.25-VD3. In contrast, high GATA-1 expression levels typically found in erythroid cells might generally inhibit monocyte and moDC differentiation. To test these assumptions, we transduced CD34<sup>+</sup> progenitors with GATA-1–IRESGFP and cultured them in LC differentiation conditions in the presence of 1.25-VD3. As expected, GFP<sup>lo</sup> or GFP<sup>hi</sup> fractions sorted by FACS differed in CD34<sup>+</sup> cells when added at day 5 to LC generation cultures. CD34<sup>+</sup> cells were induced to differentiate to LCs. Day 5–generated cells were subcultured in LC cytokines in the absence or presence of IL-4 (25 ng/mL) for 4 days. At day 9 cells were harvested and analyzed for CD207 or CD11b versus CD1a. Data are representative of 3 independent experiments. (B-D) Tet-on-inducible system in primary LC precursors. CD34<sup>+</sup> cells were transduced with retroviral vectors encoding Tet-activator-IRES-lyt2, DOX-inducible GATA-1–IRESGFP, or empty vector control. Cells were stimulated with LC-promoting cytokines for 5 days. DOX (2 μg/mL) was added from day 5 to day 7. (B) FACS diagrams show day 5–generated cells (left) or day 7–generated NGFR<sup>+</sup> cells (right) analyzed for CD207 or CD11b versus CD1a. (C) FACS diagrams represent cells at day 7 analyzed for NGFR expression. (A-C) Each number in a quadrant represents the percentage of cells in the quadrant. (D) Bars represent mean percentages (SEM; n = 3) of CD1a<sup>+</sup>CD11b<sup>+</sup> cells from control vector or GATA-1–transduced cells at day 5 (total culture) and day 7 (gated NGFR<sup>+</sup> cells); *P < .05. (E) LC generation cultures were transduced at day 3 to day 4 with a retroviral vector encoding VDR–IRESGFP or empty control vector. Forty-eight hours after transduction, GFP<sup>+</sup> cells were isolated by using FACS. Equal numbers of GFP<sup>+</sup> cells were used for quantitative real-time RT-PCR analysis. Bars represent VDR mRNA levels in 1 representative experiment (cell numbers in PCR experiments: 3 × 10<sup>5</sup> to 2 × 10<sup>6</sup>; n = 4).

Figure 4. Constitutive and inducible GATA-1 expression in LC precursors. (A) IL-4 promotes CD11b<sup>+</sup>CD1a<sup>+</sup> cells at the expense of CD207<sup>+</sup> cells when added at day 5 to LC generation cultures. CD34<sup>+</sup> cells were induced to differentiate to LCs. Day 5–generated cells were subcultured in LC cytokines in the absence or presence of IL-4 (25 ng/mL) for 4 days. At day 9 cells were harvested and analyzed for CD207 or CD11b versus CD1a. (B-D) Tet-on-inducible system in primary LC precursors. CD34<sup>+</sup> cells were transduced with retroviral vectors encoding Tet-activator-IRES-lyt2, DOX-inducible GATA-1–IRESGFP, or empty vector control. Cells were stimulated with LC-promoting cytokines for 5 days. DOX (2 μg/mL) was added from day 5 to day 7. (B) FACS diagrams show day 5–generated cells (left) or day 7–generated NGFR<sup>+</sup> cells (right) analyzed for CD207 or CD11b versus CD1a. (C) FACS diagrams represent cells at day 7 analyzed for NGFR expression. (A-C) Each number in a quadrant represents the percentage of cells in the quadrant. (D) Bars represent mean percentages (SEM; n = 3) of CD1a<sup>+</sup>CD11b<sup>+</sup> cells from control vector or GATA-1–transduced cells at day 5 (total culture) and day 7 (gated NGFR<sup>+</sup> cells); *P < .05. (E) LC generation cultures were transduced at day 3 to day 4 with a retroviral vector encoding VDR–IRESGFP or empty control vector. Forty-eight hours after transduction, GFP<sup>+</sup> cells were isolated by using FACS. Equal numbers of GFP<sup>+</sup> cells were used for quantitative real-time RT-PCR analysis. Bars represent VDR mRNA levels in 1 representative experiment (cell numbers in PCR experiments: 3 × 10<sup>5</sup> to 2 × 10<sup>6</sup>; n = 4).
Discussion

Shared CD14+ monocytic precursors can give rise to LCs in response to TGF-β1 stimulation or to interstitial/inflammatory-type DCs in response to GM-CSF plus IL-4 independently of TGF-β1. Here, we studied the molecular mechanisms underlying separate lineage differentiation of monocytic cells to LCs or moDCs.

This study identified 2 key regulators of myeloid DC subset specification, VDR and GATA-1. We found that VDR is rapidly induced downstream of TGF-β1 during LC lineage specification. Conversely, GATA-1 is induced downstream of IL-4 during moDC differentiation. Gain- and loss-of-function analyses showed that VDR and GATA-1 are involved in LC and moDC differentiation. Moreover, we identified GATA-1 as a repressor of VDR in moDC precursors. Specifically, IL-4–induced moDC generation from blood monocytes is associated with diminished VDR expression. Similarly, ectopic GATA-1 represses VDR in moDC precursors. In this function, GATA-1 resembled a repressor of VDR signaling in desensitizing DC precursors to suppressive effects of 1.25-VD3. In line with these findings, ectopic VDR inhibited moDC differentiation.

A schematic summary of these findings is presented in Figure 6.

GM-CSF/IL-4–induced moDCs phenotypically resemble inflammatory dendritic epidermal cells (IDECs) present in lesions of the atopic eczema/dermatitis syndrome or in certain other inflammatory skin diseases. IDECs lack LC characteristics, appear at inflammatory sites, and share with GM-CSF/IL-4–induced moDCs the expression of CD206, CD11b, and CD1a.29 Furthermore, IDECs and moDCs share many phenotypic characteristics with dermal/interstitial DCs found at steady state.29 Therefore, reciprocal GATA-1 and VDR expression by LCs versus moDCs might be critical regulators of myeloid DC fates in steady-state versus inflammatory conditions (Figure 6). It will be interesting to study to which extent the here-described GATA-1– and VDR-dependent regulation could apply to other candidate inflammatory DC subsets.

Low GATA-1 levels in moDCs are reminiscent of low GATA-1 expression in previous studies of other leukocyte subsets, including mast cells, eosinophils31,32 and murine DC subsets.14 We showed that GATA-1 is induced in response to IL-4 stimulation during...
moDC differentiation. Importantly, shRNA knock-down experiments showed a functional requirement for endogenous GATA-1 in promoting moDC differentiation. Furthermore, ectopic GATA-1 diminished VDR mRNA expression, and low levels of GATA-1 were sufficient to desensitize DC precursors to inhibitory effects of 1,25-VD3. Repression of VDR by GATA-1 might desensitize DCs to possible inhibitory effects of VD3 metabolites and thereby may enable efficient antigen-presenting function. Taken together, GATA-1 represses VDR and is functionally involved in promoting moDC differentiation downstream of IL-4.

It was previously shown that DCs can produce IL-4 under certain conditions in an autocrine fashion. Moreover, exposure of DCs to IL-4 was shown to be necessary to induce IL-4 production in these cells. Most notably, GATA-1 binding sites were found in an IL-4 intronic enhancer element, and these sites were found to be essential for IL-4 expression in mast cells. Thus, GATA-1 in DCs might represent part of an IL-4–driven positive feedback loop. This possibility needs to be further elucidated.

We found that VDR mRNA is decreased within 48 hours after ectopic GATA-1 expression in primary cells and U937 cells; however, the underlying mechanism remains to be identified. One possibility is that GATA-1 acts as a direct transcriptional repressor of VDR, given the existence of several potential GATA-1 binding sites in the VDR promoter. Alternatively, but not mutually exclusive, GATA-1 might inhibit VDR mRNA expression by an indirect mechanism. Interestingly, the 5-kb VDR promoter region contains 3 potential PU.1 binding sites. Because GATA-1 binds and antagonizes PU.1, its negative effect on VDR expression might be secondarily mediated by PU.1 antagonism. Future studies should address this question.

We observed that overexpression of GATA-1 stabilizes the monocyte-like DC phenotype even in the presence of VDR agonist 1,25-VD3, therefore indicating that the actions of GATA-1 are dominant over those of vitamin D. Our data in U937 model cells seem to support that VDR down-regulation is critical for GATA-1–mediated induction of CD11b. Specifically, ectopic (nonregulated) VDR in the presence of VDR agonist partially impaired GATA-1–mediated induction of CD11b. Detailed mechanistic experiments in primary DCs are required to further study this point.

One key finding of our study was that VDR function intersects with TGF-β1 during LC lineage specification. Evidence for this is presented by the observations that purified myeloid progenitors rapidly up-regulate VDR in response to TGF-β1 signaling under LC lineage-instructive stimulation conditions and that ectopic VDR promotes TGF-β1–dependent LC differentiation. In the absence of TGF-β1, progenitors expressed low VDR levels and differentiated along a monocytic pathway. By using identical cytokine stimulation conditions as used in this study, we previously provided evidence that TGF-β1 redirects monocytes to LC differentiation in single-cell cultures of CD34+ cells. In support of these data, we have also shown that the addition of a VDR antagonist inhibited LC differentiation.

VDR immunohistology analyses of human skin samples showed a general bright staining pattern in epidermis. In particular, CD207+ epidermal LCs in situ showed a nuclear VDR expression pattern reminiscent of keratinocytes. Interestingly, unligated VDR was previously shown to induce keratinocyte differentiation. In support of this, deficiency of 1α-hydroxylase [1α(25)OHase, Cyp27B1], an enzyme converting 25-VD3 to the biologically active metabolite 1,25-VD3, in mice has no skin phenotype, whereas VDR-deficient mice show strongly increased keratinocyte proliferation and epidermal thickening. Several pieces of evidence indicate that unligated VDR might similarly promote LC differentiation in the absence of its ligand 1,25-VD3. First, the use of a serum-free culture model devoid of any VD3 showed that VDR strongly promotes LC differentiation. Second, the addition of a VDR inhibitor interfered with LC differentiation in these serum-free cultures devoid of VD3. Third, we detected only low levels of 25-VD3 in human skin (data not shown). Specifically, whereas serum levels of 1,25-VD3 and its precursor 25-VD3 are known to range from 40 to 150 pmol/L and 75 to 150 nmol/L, respectively, skin samples contained only low levels of 25-VD3 (9.0 and 9.5 nmol/L in 2 independent skin specimens) and 1,25-VD3 remained undetectable. However, LCs expressed similar amounts of 1α-hydroxylase [1α(25)OHase, Cyp27B1] as did CD34+ cells and moDCs (data not shown). Therefore, LCs might be constitutively exposed to low amounts of bioactive 1,25-VD3. In this respect it is interesting that 1,25-VD3 produced by DCs can instruct T cells to express CCR10 and to migrate toward the epithelial chemokine CCL27. In an attempt to mimic LC exposure to VD3 metabolites, we found that low concentrations of 25-VD3 (6.25 nmol/L, ie, in the range detected in skin) failed to inhibit LC differentiation, whereas 10-fold higher amounts (serum concentrations) abrogated LC differentiation in favor of monocyte induction (data not shown). In conclusion our data support a model whereby VDR is induced downstream of the LC-linage instructive TGF-β1 signal and promotes LC differentiation from monocytic cells in the epidermis, a process that could be independent of its ligand 1,25-VD3.

The here-presented findings are based on retroviral vector–mediated gain- and loss-of-function analyses of human primary DC subsets. Additional studies are required to address how GATA-1 and VDR alter DC subset-associated functions.

In conclusion, TGF-β1, the key cytokine for LC differentiation, induces VDR under LC lineage instructive conditions. Given that TGF-β1–dependent LC differentiation is an epithelia-specific process, this pathway seems to be operative in LC renewal in epithelia. In contrast, IL-4 induces GATA-1 during moDC differentiation and may thereby enable this DC subset to differentiate within inflammatory sites. We identified GATA-1 as a repressor of VDR in moDCs. Repression of VDR in DCs may desensitize them to VD3-mediated inhibition and hence allow a stable DC phenotype. Therefore, low VDR in these DCs may very well be critical for the induction of effective antigen-specific immunity to pathogens.

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Reciprocal role of GATA-1 and vitamin D receptor in human myeloid dendritic cell differentiation

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