A reliable estimate of peripheral blood stem cell (PBSC) mobilization response to granulocyte colony-stimulating factor (G-CSF) may identify donors at risk for poor mobilization and help optimize transplant approaches. We studied 639 allogeneic PBSC collections performed in 412 white, 75 black, 116 Hispanic, and 36 Asian/Pacific adult donors who were prescribed G-CSF dosed at either 10 or 16 μg/kg per day for 5 days followed by large-volume leukapheresis (LVL). Additional LVL (mean, 11 L) to collect lymphocytes for donor lymphocyte infusion (DLI) and other therapies was performed before G-CSF administration in 299 of these donors. Day 5 preapheresis blood CD34+ cell counts after mobilization were significantly lower in whites compared with blacks, Hispanics, and Asian/Pacific donors (79 vs 104, 94, and 101 cells/μL, P < .001). In addition, donors who underwent lymphapheresis before mobilization had higher CD34+ cell counts than donors who did not (94 vs 79 cells/μL, P < .001). In multivariate analysis, higher post–G-CSF CD34+ cell counts were most strongly associated with the total amount of G-CSF received, followed by the pre–G-CSF platelet count, pre–G-CSF mononuclear count, and performance of prior LVL for DLI collection. Age, white ethnicity, and female gender were associated with significantly lower post–G-CSF CD34+ cell counts.

**Methods**

**Donors**

Study subjects comprised 639 consecutive allogeneic donors who underwent PBSC collection by large-volume leukapheresis (LVL) after G-CSF mobilization between January 1999 and March 2006. All subjects were healthy sibling donors at least 18 years of age, who were undergoing their first PBSC mobilization and gave informed consent in accordance with the Declaration of Helsinki and National Institutes of Health Institutional Review Board–approved transplantation protocols. According to individual protocol design, 299 of these donors also underwent an unstimulated (non–G-CSF mobilized) lymphapheresis collection by LVL before initiation of G-CSF to collect cells for use in posttransplantation strategies designed to enhance transplant outcomes, including donor lymphocyte infusions,14 scheduled T-cell “add-backs,”6,15 Th2/Tc2 cell generation,16,17 and other ex vivo graft manipulation therapies. At each visit, donor demographic information, including age, sex, height, and weight, was...
collected. Ethnicity was self-categorized as white, black, Hispanic, or Asian/Pacific and is reported throughout this study as ethnicity rather than race for consistency.

**Mobilization and collection of PBSC**

All donors received 5 days of G-CSF (filgrastim; Amgen, Thousand Oaks, CA) with leukapheresis initiated on the morning of day 5, at least 2 hours after the last G-CSF dose. All data in this report for PBSC CD34+ apheresis yields were obtained from a single apheresis procedure per donor, performed on day 5. The G-CSF dose was prescribed as either 10 µg/kg per day or 16 µg/kg per day; the actual dose administered was obtained in all cases from review of pharmacy and nursing records. LVL procedures were performed with use of a model CS-3000 Plus continuous-flow apheresis device (Fenwal Division, Baxter, Deerfield, IL) using prophyllactic intravenous calcium and magnesium as previously described. Volume processed per procedure ranged from 15 to 25 L for PBSC collections (mean ± SD, 19 ± 4 L), depending on the preapheresis CD34+ cell count, and from 7 to 15 L (11 ± 2 L) for LVL conducted to collect lymphocytes for DLI. Apheresis procedures were well tolerated, with no serious adverse events reported.

**Laboratory analyses**

Complete blood counts were measured at baseline before G-CSF administration (pre–G-CSF), before and after PBSC collection after G-CSF mobilization (post–G-CSF and postapheresis, respectively), and before and after lymphapheresis for donors who underwent LVL for collection of cells for DLI (pre-DLI and post-DLI, respectively). The complete blood count was assayed using an electronic cell counter, and quantification of CD34+ cells before and after PBSC collection was performed by flow cytometry as previously described.

**Statistical analysis**

Graphics and standard data analysis were performed with a spreadsheet application (Excel, Microsoft, Redmond, WA). Body mass index (BMI) was calculated as BMI = (weight in kilograms)/(height (in meters)²). The percentage body fat was calculated as [(body weight – lean body weight)/body weight] × 100, where lean body weight = [(1.07 × body weight (kg)) – 148 × (body weight (kg)/100 × height (m)²)] for women and [(1.10 × body weight (kg)) – 128 × (body weight (kg)/100 × height (m)²)]. The total mononuclear cell count was calculated as the sum of total lymphocyte and total monocyte count reported on the white blood cell differential from the complete blood cell count. Actual administered values, which were within 10% of the prescribed dose, were used for comparisons of G-CSF doses at 10 versus 16 µg/kg per day. Significance tests for comparisons between 2 groups were conducted with 2-tailed, nonpaired t tests. Values between more than 2 groups were compared using analysis of variance. Multivariate analyses were performed using stepwise forward logistic regression, based on parameters having significance in univariate analysis, using a commercial statistics program (StatView; Abacus Concepts, Berkeley, CA). Proportions between 2 groups were compared using a 2-tailed Fisher exact test, and comparisons of proportions between multiple groups were made using χ² analysis. Results are provided as the mean plus or minus SD, unless otherwise stated.

**Table 2. Post–G-CSF peripheral blood CD34+ cell counts and CD34+ apheresis yields according to G-CSF dose (µg/kg) and ethnicity**

<table>
<thead>
<tr>
<th>Administered G-CSF dose, µg/kg</th>
<th>All donors</th>
<th>White</th>
<th>Black</th>
<th>Hispanic</th>
<th>Asian/Pacific</th>
</tr>
</thead>
<tbody>
<tr>
<td>10 µg/kg</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>N</td>
<td>561</td>
<td>350</td>
<td>67</td>
<td>108</td>
<td>36</td>
</tr>
<tr>
<td>Male, %</td>
<td>51%</td>
<td>51%</td>
<td>50%</td>
<td>50%</td>
<td>52%</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>78 ± 17</td>
<td>79 ± 17</td>
<td>81 ± 16</td>
<td>76 ± 17</td>
<td>68 ± 16</td>
</tr>
<tr>
<td>Total G-CSF received, µg/d</td>
<td>788 ± 169</td>
<td>801 ± 169</td>
<td>815 ± 152</td>
<td>766 ± 169</td>
<td>688 ± 160</td>
</tr>
<tr>
<td>CD34+ response</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Peripheral blood, µL</td>
<td>84 ± 51†</td>
<td>77 ± 45</td>
<td>102 ± 59</td>
<td>93 ± 49</td>
<td>101 ± 74</td>
</tr>
<tr>
<td>Yield, 10^5/µL processed</td>
<td>32 ± 20‡</td>
<td>30 ± 19</td>
<td>40 ± 26</td>
<td>33 ± 20</td>
<td>36 ± 24</td>
</tr>
<tr>
<td>16 µg/kg</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>N</td>
<td>27</td>
<td>21</td>
<td>2</td>
<td>4</td>
<td>0</td>
</tr>
<tr>
<td>Male, %</td>
<td>37%</td>
<td>38%</td>
<td>0%</td>
<td>50%</td>
<td>—</td>
</tr>
<tr>
<td>Weight, kg</td>
<td>78 ± 15</td>
<td>78 ± 16</td>
<td>82 ± 10</td>
<td>74 ± 10</td>
<td>—</td>
</tr>
<tr>
<td>Total G-CSF received, µg/d</td>
<td>1247 ± 236</td>
<td>1253 ± 255</td>
<td>1335 ± 191</td>
<td>1170 ± 159</td>
<td>—</td>
</tr>
<tr>
<td>CD34+ response</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>Peripheral blood, µL</td>
<td>119 ± 49†</td>
<td>114 ± 52</td>
<td>144 ± 26</td>
<td>130 ± 40</td>
<td>—</td>
</tr>
<tr>
<td>Yield, 10^5/µL processed</td>
<td>39 ± 17‡</td>
<td>38 ± 18</td>
<td>46 ± 15</td>
<td>41 ± 17</td>
<td>—</td>
</tr>
</tbody>
</table>

*Includes donors with actual G-CSF received within 10% of prescribed dose of 10 or 16 µg/kg per day.
†P < .001, 16 versus 10 µg/kg per day G-CSF for peripheral blood CD34+ cell count.
‡P = .02, 16 versus 10 µg/kg per day G-CSF for CD34+ apheresis yields per L processed.

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NS, not significant.

*For ANOVA comparison among all groups.
Results

Donor demographics, apheresis parameters, and CD34+ cell counts

Donor demographics, peripheral blood CD34+ cell counts after G-CSF mobilization (post-G-CSF), and CD34+ apheresis yields per liter processed are shown according to donor ethnicity in Table 1. Of the 639 subjects, 66% (n = 412) were white, 11% (n = 75) were black, 18% (n = 116) were Hispanic, and 5% (n = 36) were Asian/Pacific Islander. Compared with other groups, whites were older, whereas Asian/Pacific donors had lower body weight and lower BMI. The mean weight for the entire study population was 79 kg (± 18 kg). Asians had the lowest mean weight at 68 kg (± 16 kg), followed by Hispanics, 75 kg (± 17 kg), whites, 80 kg (± 19 kg) and blacks, 82 kg (± 16 kg). There was no difference in sex distribution; 51% of study subjects overall were male. As illustrated in Table 1, peripheral blood post–G-CSF CD34+ cell counts and CD34+ apheresis yields were significantly lower in white donors than in the other ethnic groups.

Association of CD34+ cell counts with G-CSF dose, and demographic and laboratory variables

In the study group of 639 donors, 561 received an administered G-CSF dose that was within 10% of 10 μg/kg per day (10.1 ± 0.4; range, 9.0-11.0 μg/kg per day) and 27 received an administered G-CSF dose that was within 10% of 16 μg/kg per day (16.0 ± 0.4; range, 15.0-16.7 μg/kg per day), as listed in Table 2. The remaining 51 donors received a mean administered G-CSF dose outside of these ranges (mean 9.5 ± 1.7; range, 6.4-13.9 μg/kg per day).

As shown in Figure 1, donors with greater weight had significantly higher post–G-CSF CD34+ cell counts (Figure 1A), whereas donors who were older had lower CD34+ cell counts (Figure 1B). In both instances, CD34+ responses were lowest in white donors (Figure 1A,B). As shown in Figure 2A and Table 2, donors who received G-CSF at 16 μg/kg per day (8 μg/kg twice a day) had significantly higher CD34+ mobilization responses than those who received G-CSF at 10 μg/kg per day (P < .001). However, responses were similar when post–G-CSF CD34+ cell counts were plotted against the total amount of G-CSF received per day rather than as a function of dose per kilogram of donor weight.
than the dose per kilogram (Figure 2B), suggesting that the total amount of G-CSF received, rather than the dose per kilogram, exerted a larger effect on CD34⁺ mobilization in this study population.

Donors with higher platelet and total mononuclear cell count before administration of G-CSF (pre--G-CSF) also had significantly higher peripheral blood post--G-CSF CD34⁺ cell counts, as shown in Figure 3A,B. The strongest response was observed in Asian/Pacific donors. The lowest values for CD34⁺ mobilization response were observed in whites, with intermediate responses in blacks and Hispanics.

**Effect of prior lymphapheresis on CD34⁺ mobilization**

In accord with individual protocol designs, 299 of the 639 donors in this study underwent an unstimulated lymphapheresis for collection of cells for subsequent DLI, before receiving G-CSF stimulation and PBSC collection. Comparison of prelymphapheresis and postlymphapheresis complete blood counts in these donors demonstrated that the procedure significantly lowered blood hemoglobin concentration, platelet and total white blood cell count, and lymphocyte counts (Table 3). As shown in Table 4, peripheral blood hemoglobin and platelet levels, but not absolute lymphocyte levels, remained low after subsequent G-CSF administration, both before and after PBSC collection (post--G-CSF and postapheresis), and were significantly lower in these 299 donors than in the other donors who did not undergo lymphapheresis. Interestingly, the donors who underwent a prior lymphapheresis procedure had significantly higher post--G-CSF CD34⁺ cell counts than those who did not (mean 94 vs 79/μL, P < .001), an effect that was most marked in the 164 of 299 donors who underwent lymphapheresis within 2 days of beginning G-CSF mobilization (mean post--G-CSF CD34⁺ count, 98/μL, P = .005, Figure 4). Donors who underwent a prior lymphapheresis procedure also had higher CD34⁺ apheresis yields per liter processed (34 vs 30 × 10⁶ CD34⁺ cells/L processed, P = .003; 36 × 10⁶ CD34⁺ cells/L processed when lymphapheresis was performed within 2 days of beginning G-CSF, P = .02) than donors who did not undergo this procedure.

**Univariate and multivariate stepwise regression analysis of factors affecting CD34⁺ mobilization**

In univariate logistic regression analysis, the strongest association with higher post--G-CSF CD34⁺ cell counts was observed with total G-CSF dose administered (P < .001), followed by weight (P < .001), pre--G-CSF donor platelet count (P < .001), pre--G-CSF donor mononuclear cell count (P < .001), prior lymphapheresis for DLI collection (P = .008), and G-CSF dose per kg (P = .006). In contrast, white ethnicity (P < .001) and female sex (P = .006) were associated with lower CD34⁺ cell counts in univariate analysis, whereas age (P = .06) exhibited a trend toward association with lower CD34⁺ cell counts. In stepwise regression after adjustment for total G-CSF dose received, higher post--G-CSF CD34⁺ cell counts remained significantly associated with the pre--G-CSF platelet count (P < .001), pre--G-CSF mononuclear cell count (P < .001), and prior lymphapheresis for DLI collection (P = .02), and were negatively associated with white ethnicity (P < .001) and age (P < .004); after this adjustment (total amount of G-CSF received), the associations of CD34⁺ cell counts with sex (P = .45), weight (P = .598), and G-CSF dose per kilogram (P = .85) no longer retained significance. Interestingly, donor height, which was not significantly associated with CD34⁺ cell counts in univariate analysis (P = .54), exhibited a strong negative association with this parameter (P = .001) after adjustment for the total amount of G-CSF received.

| Table 3. Hematologic parameters before and after donor lymphocyte collection by apheresis before PBSC mobilization in 299 allogeneic donors |
|---------------|---------------|---------------|---------------|
|               | Platelets, 10⁹/L | Hemoglobin, g/L | White blood cells, 10⁹/L | Lymphocytes, 10⁹/L |
| Pre-DLI       | 259 ± 55       | 137 ± 15.3     | 6.36 ± 1.6     | 1.93 ± 0.6 |
| Post-DLI      | 178 ± 50       | 129 ± 17       | 5.72 ± 1.7     | 1.51 ± 0.4 |
| P             | < .001         | < .001         | < .001         | < .001      |
Table 4. Parameters at the time of PBSC collection after G-CSF mobilization in donors who did, compared with those who did not, undergo a prior lymphapheresis procedure

<table>
<thead>
<tr>
<th>Prior donor lymphocyte collection</th>
<th>Post–G-CSF peripheral blood CD34⁺ per µL</th>
<th>CD34⁺ yield, 10⁹/L processed</th>
<th>Hemoglobin, g/L</th>
<th>Platelets, x10⁹/L</th>
<th>Lymphocytes, x10⁹/L</th>
</tr>
</thead>
<tbody>
<tr>
<td>Before PBSC collection</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>79 ± 46</td>
<td>—</td>
<td>135 ± 13</td>
<td>235 ± 53</td>
<td>3.91</td>
</tr>
<tr>
<td>Yes</td>
<td>94 ± 59</td>
<td>—</td>
<td>132 ± 16</td>
<td>214 ± 52</td>
<td>3.78</td>
</tr>
<tr>
<td>P</td>
<td>&lt;.001</td>
<td>—</td>
<td>.02</td>
<td>&lt;.001</td>
<td>.4</td>
</tr>
<tr>
<td>After PBSC collection</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
</tr>
<tr>
<td>No</td>
<td>—</td>
<td>30 ± 21</td>
<td>127 ± 14</td>
<td>160 ± 42</td>
<td>2.21 ± 0.6</td>
</tr>
<tr>
<td>Yes</td>
<td>—</td>
<td>34 ± 25</td>
<td>122 ± 16</td>
<td>141 ± 47</td>
<td>2.11 ± 0.6</td>
</tr>
<tr>
<td>P</td>
<td>—</td>
<td>.003</td>
<td>.002</td>
<td>.002</td>
<td>.09</td>
</tr>
</tbody>
</table>

— indicates not applicable.

In the final multivariate stepwise regression, higher post–G-CSF CD34⁺ cell counts were positively associated with total G-CSF dose (P < .001), pre–G-CSF platelet count (P < .001), pre–G-CSF absolute mononuclear cell count (P = .005), and prior lymphapheresis for DLI collection (P = .007), and were negatively associated with height (P < .001), female gender (P < .001), white ethnicity (P = .004), and age (P = .003); no significant association was present with weight (P = .5) or G-CSF dose per kilogram (P = .6).

In this study, female donors overall had lower post–G-CSF CD34⁺ cell counts than men (79 vs 93 cells/µL, P < .001), and female sex was significantly associated with lower post–G-CSF CD34⁺ cell counts in univariate analysis. The overall impact of sex on post–G-CSF CD34⁺ cell counts was complicated by the fact that female donors weighed less than male donors (71 vs 86 kg, respectively, P < .001), received lower total amounts of G-CSF (740 vs 877 µg, respectively, P < .001), were shorter (163 vs 177 cm, respectively, P < .001), and had higher baseline pre–G-CSF platelet counts (266 vs 246 x 10⁹/µL, respectively). Thus, when post–G-CSF CD34⁺ cell counts were adjusted for the total amount of G-CSF received, responses in women were overall similar to men, as illustrated in Figure 5 for white donors.

Demographic and laboratory parameters in donors with a poor mobilization response

Table 5 lists clinical and laboratory data for donors who experienced a poor mobilization response to G-CSF, defined as either a post–G-CSF CD34⁺ cell count less than 20 (n = 29) or less than 30 (n = 73) cells per microliter. In both groups, donors were significantly more likely to be female or white, have lower weight, lower BMI, lower estimated percentage body fat, lower pre–G-CSF platelet, and total mononuclear cell counts, and to have received lower total amounts of G-CSF compared with donors who did not have a poor mobilization. Furthermore, in each instance, the proportion of donors who were female or white was higher, and mean values for weight, BMI, estimated percentage body fat, pre–G-CSF platelet and mononuclear cell counts, and total G-CSF dose received tended to be lower in donors with a post–G-CSF CD34⁺ cell count of 20 cells/µL versus 30 cells/µL. There was no significant association with mean age or height, nor with the fraction of donors who received G-CSF at 16 versus 10 µg/kg per day. However, donors with poor mobilization responses tended to be older than those who did not mobilize poorly, and none of the donors who received G-CSF dosed at 16 µg/kg per day exhibited a poor mobilization. Finally, there was no significant difference in the proportion of donors who underwent a prior lymphapheresis for DLI collection in those with versus those without a poor mobilization response, reflecting the fact that the impact of this procedure was less evident on donors with low body weight, as shown in Figure 4.

Discussion

This study represents the largest published analysis of PBSC yields in allogeneic donors. It is also the only series to examine the effect of donor ethnicity on CD34⁺ mobilization responses. We found that the single strongest factor affecting CD34⁺ apheresis yields was the total amount of G-CSF administered to the donor and that donor age, gender, and weight had reduced impact after accounting for this parameter. Furthermore, donor ethnicity significantly affected CD34⁺ mobilization response, a novel finding that remained strongly significant in multivariate analysis. White donors had the lowest CD34⁺ cell counts and black donors had the highest CD34⁺ cell counts after mobilization, whereas Asian/Pacific donors had the highest CD34⁺ cell counts when adjusted for BMI and body weight. Although a wide variation in individual responses to G-CSF has been reported, the reason for a blunted CD34⁺ response to G-CSF in white donors is unknown. Our data indicate that donor ethnicity may play a significant role in the mobilization...
response to G-CSF and should be taken into account in designing mobilization regimens.

Our findings for mobilized PBSC counts in black donors are in contrast with those reported in cord blood collections, wherein blacks have been noted to have lower CD34+ cell counts. Similarly, reference values for unstimulated total white cell and neutrophil counts are also known to be lower in blacks than other races. Several small reports have also shown that blacks did not increase their total leukocyte counts as much as whites after one dose of hydrocortisone or after participating in athletic activity. A trend toward more robust CD34+ mobilization in black subjects with sickle cell trait versus those without trait was noted in a small number of healthy donors undergoing PBSC collection; however, we did not evaluate the presence of sickle cell trait among our donors.

The significant variation in PBSC mobilization response between ethnic groups suggests that there may be genetic regulation of mobilization response. Quantitative variations in PBSC mobilization response to G-CSF have been noted in inbred strains of mice. Genetic analysis of high-responder (DBA/2) and poor-responder (C57BL/6) mice revealed that progenitor cell release in response to G-CSF was linked to loci on chromosomes 2 and 11. Genetic variations could exist in putative stem cell pool size, intrinsic migration properties of hematopoietic stem cells, or interaction of stem cells with stromal and endothelial cell types in the bone marrow niche. A trend toward more robust CD34+ mobilization in black subjects with sickle cell trait versus those without trait was noted in a small number of healthy donors undergoing PBSC collection; however, we did not evaluate the presence of sickle cell trait among our donors.

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### Table 5. Clinical and laboratory data in donors with a poor mobilization response to G-CSF

<table>
<thead>
<tr>
<th>Prior lymphapheresis, years (%)</th>
<th>Pre–G-CSF</th>
<th>Post–G-CSF</th>
<th>CD34+ cell count</th>
<th>MNC, %</th>
<th>Pre–G-CSF G-CSF dose, mg/kg</th>
<th>Post–G-CSF</th>
<th>Estimated body fat, %</th>
<th>Height, cm</th>
<th>Weight, kg</th>
<th>BMI, kg/m²</th>
<th>Total G-CSF, mg/d</th>
<th>LN</th>
<th>Pre–G-CSF lymphapheresis yields</th>
<th>Post–G-CSF lymphapheresis yields</th>
</tr>
</thead>
<tbody>
<tr>
<td>All donors</td>
<td>639</td>
<td>49/51</td>
<td>64/12/18/6</td>
<td>96/4</td>
<td>6.4/7/6/9</td>
<td>596/1000</td>
<td>48/54/2/8</td>
<td>69/170/27.1</td>
<td>98/4</td>
<td>68/41.7</td>
<td>26/40.4</td>
<td>2.43</td>
<td>80/8</td>
<td>80/8</td>
</tr>
<tr>
<td>Less than 30</td>
<td>29</td>
<td>74/26</td>
<td>81/4/15/0</td>
<td>100/0</td>
<td>6.4/7/6/9</td>
<td>596/1000</td>
<td>48/54/2/8</td>
<td>69/170/27.1</td>
<td>98/4</td>
<td>68/41.7</td>
<td>26/40.4</td>
<td>2.43</td>
<td>80/8</td>
<td>80/8</td>
</tr>
<tr>
<td>More than or equal to 30</td>
<td>610</td>
<td>49/51</td>
<td>64/12/18/6</td>
<td>96/4</td>
<td>6.4/7/6/9</td>
<td>596/1000</td>
<td>48/54/2/8</td>
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significantly associated with subsequent CD34+ cell counts, an effect that was independent of the association with platelet counts. Interestingly, the CD34+ cell count associations with baseline levels of both platelets and mononuclear cells in this study were strongest in donors of Asian/Pacific ethnicity. Our data thus confirm and extend the findings of Suzuya et al., in a larger study of donors of diverse backgrounds.

The association of higher baseline platelet counts with improved CD34+ mobilization may be related to common pathways of thrombopoiesis and progenitor cell mobility. Increased plasma levels of SDF-1 have been shown to enhance human thrombopoiesis and mobilize human colony forming cells in nonobese diabetic/severe combined immunodeficiency mice.48 In addition, CD34+ cells may exhibit changes similar to platelet-derived microparticles during G-CSF mobilization.49 Chemokine-mediated interactions of hematopoietic progenitors with the bone marrow vascular niche may also allow progenitors to relocate to a microenvironment that is permissive for megakaryocyte maturation and thrombopoiesis.40

Our study indicates that donors with a poor mobilization response to G-CSF are more likely to have received a low total amount of G-CSF and to be white, female, and older, and to have lower BMI and lower baseline platelet and mononuclear cell counts. The data allow an educated assessment of risk for either a poor or an excessively robust mobilization response and allow a priori intervention with a higher or lower total G-CSF dose to mitigate the effect of otherwise fixed demographic factors. Such strategies could be used in cases where lighter-weight donors are matched with heavier-weight recipients or in situations where additional CD34+ cells are required for ex vivo graft manipulation. In unrelated donor programs where a fixed dose of G-CSF is prescribed, selection of younger, male donors might be considered to optimize responses when higher cell doses are desired. Furthermore, with the development of newer mobilizing agents, such as CXCR4 antagonists, knowledge of predictive factors for mobilization to G-CSF may potentially be used to determine the best mobilizing agent for a donor. Use of this information in clinical mobilization protocols could also result in overall benefit to donors, avoidance of multipleapheresis procedures, maximization of cell yields by apheresis, and improved transplantation outcomes.

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Authorship

Contribution: S.V. wrote the paper and interpreted data; S.F.L. designed research and analyzed data; J.F.T., M.M.H., R.W.C., A.J.B., D.H.F., M.R.B., E.M.K., H.L.M., H.M.K., and C.E.D. performed research and provided clinical data; Y.Y. designed research, performed research, collected data, and analyzed data; R.W. provided statistical expertise; and C.D.B. designed research and analyzed data.

Conflict-of-interest disclosure: The authors declare no competing financial interests.

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