Adoptive transfer of allogeneic tumor-specific T cells mediates effective regression of large tumors across major histocompatibility barriers

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Graft-versus-tumor effects can be achieved after allogeneic bone marrow transplantation in patients with malignancies of the kidney or hematopoietic system but are often accompanied by severe graft-versus-host-disease (GVHD). We sought to maximize graft-versus-tumor while minimizing GVHD using tumor-specific allogeneic effector T cells rather than open-repertoire T cells. We transferred allogeneic CD8* pmel-1 or CD4* TRP-1 T cells specific for the melanoma-associated antigens, glycoprotein 100 (gp100) and tyrosinase-related protein-1 (TRP-1), respectively, into B16-melanoma-bearing mice. Mice receiving a preparative regimen of nonmyeloablating (5 Gy) total body irradiation experienced the rapid rejection of tumor-specific allogeneic lymphocytes with no impact on tumor growth. However, when mice were given more intense total body irradiation conditioning regimens combined with autologous bone marrow transplantation, adoptively transferred allogeneic tumor-specific T lymphocytes persisted at detectable levels for several weeks and mediated significant regression of large, vascularized tumors. We found that the risk of GVHD was low when tumor-specific T cells were transferred and significant toxicity was observed only when substantial numbers of open repertoire allogeneic naive T cells were mixed with the tumor-specific lymphocytes. Taken together, these data indicate that the use of tumor-specific allogeneic CD8+ T cells or CD4+ can result in significant antitumor effects in the absence of measurable GVHD. (Blood. 2008;112:4746-4754)

Introduction

The ability of T lymphocytes to recognize antigens with a high degree of discrimination and mediate tissue destruction makes them excellent candidates for treating malignancies. After successes obtained in treating hematologic malignancies with allogeneic bone marrow (BM) transplantation (BMT), many investigators have tried to use the same approach for solid tumors, relying on the capacity of allogeneic lymphocytes to mediate a graft-versus-tumor (GVT) effect. Unfortunately, most solid tumors have proven to be resistant to this therapy, and beneficial clinical responses were observed only for renal cell carcinoma1,2 with anecdotal successes to be resistant to this therapy, and beneficial clinical responses were obtained for renal cell carcinoma1,2 with anecdotal successes.
expression of MHC molecules, and the lack of highly immunogenic target antigens that are derived from foreign viral proteins.

One major concern is the survival of the effectors. The persistence of transferred cells might be crucial for a favorable clinical outcome. However, it is still not clear if persistence is a requirement for effective treatment or an epiphenomenon related to the other characteristics of therapeutically effective cells. Allogeneic cells with major MHC mismatches are rapidly rejected by the host immune system. In immunocompetent hosts, MHC-mismatched leukocytes present in nonirradiated blood products were shown to persist for not more than 5 to 6 days. Thus, although allogeneic cells specific for highly immunogenic viral antigens can be effective in the posttransplantation setting, it is not known whether allogeneic cells would be a viable option for adoptive immunotherapy of large, established solid tumors.

In the present paper, we sought to explore the feasibility of using tumor-specific allogeneic T cells in the treatment of the experimental mouse melanoma, B16, after a preparative regimen composed of total body irradiation (TBI) and autologous BMT. We use 2 different T-cell receptor (TCR) transgenic mouse models. The experimental mouse melanoma, B16, after a preparative regimen related protein-1 (TRP-1)–specific transgenic model. We also were previously described. Male Rag1−/− mice were bred at the National Institutes of Health and have been previously described. TRP-1 TCR-Tg mice were bred at the National Institutes of Health and have been previously described. Rag1−/−/H11001 mice (H-2b/b) were used as recipients in the present manuscript. We used the R&D Systems (Minneapolis, MN) antibody pair according to the manufacturer’s protocol for testing the release of interferon-γ (IFN-γ).

Adoptive cell transfer

Mice 6 to 12 weeks of age (n = 5-6 for all groups) were injected subcutaneously with 2 × 109 to 5 × 109 B16-F10 melanoma cells and treated 10 to 14 days later with intravenous adoptive transfer of in vitro–activated pmel-1 CD8+ T splenocytes or Rag1−/− B6 TRP-1 CD4+ cells derived from different F1 donors as indicated. Recombinant vaccinia virus expressing human gp100 (2 × 107 plaque-forming units) was administered intravenously, and interleukin-2 (IL-2) was given as 600 000 international units (IU) intraperitoneally twice daily for 6 doses. Lymphopenia was induced by nonmyeloablative (5 Gy) or myeloablative (9 Gy) TBI on the day before the cell transfer, and hematopoietic stem cells derived from bone marrow of donor animals were transferred on day 1 after treatment; 105 unsorted bone marrow cells were administered. Tumors were measured using calipers, and the tumor area was calculated as the products of the perpendicular diameters. All experiments were performed in a blinded, randomized fashion and performed independently at least twice, with similar results.

Enumeration of cells in vivo

At indicated time points, spleens were resected, and total cell numbers were enumerated by trypan blue exclusion. Samples were analyzed by flow cytometry for Thy1.1, CD8a, Vβ14, H-2 Dd, and Vβ13 (BD Biosciences, San Jose, CA). Cytofluorimetric analysis was done on a “lymphocytes gate” identified by physical parameters and staining with propidium iodide. Numbers of adoptively transferred cells were calculated by multiplying the total number of live splenocytes by the percentage of CD8+ Vβ13+ cells.

Histology

Eyes were enucleated 14 days after adoptive transfer, fixed in 10% formalin, embedded in methylacrylate, sectioned via papillary-optic nerve axis, and stained with hematoxylin and eosin. Images were obtained using a Nikon Eclipse E400 microscope (Tokyo, Japan) equipped with Nuance Multispectral Imaging System VIS and related software (CRI, Woburn, MA). Original magnification was ×100. Images were processed using Adobe Photoshop, version 7 (Adobe Systems, San Jose, CA).

Statistics

Tumor graphs were compared using analysis of variance. P values less than .05 were considered significant.

Results

The intensity of the conditioning regimen directly correlates with cell persistence after transfer

To address whether allogeneic tumor-specific T cells could be effectively used to treat established tumors, we created an allogeneic adoptive immunotherapy model based on the B16/pmel-1 system that we previously described. pmel-1 mice are Tg for a TCR that recognize the H-2 Dd-restricted epitope of the melanocyte differentiation antigen gp100 corresponding to amino acids 25 to 33 that is naturally expressed on the B16 melanoma tumor. To create a source of tumor-specific allogeneic effector cells, we crossed pmel-1 mice (H-2b/b) with wild-type BALB/c mice (H-2d/d) to generate F1-pmel-1 mice with a MHC H-2d/d genotype (pmel-1/1dd).

To assess the ability of allogeneic effector cells to persist in MHC-mismatched hosts, we transferred pmel-1/dd cells into either unmanipulated C57BL/6 mice or C57BL/6 mice that received 5 Gy....
TBI as lymphodepleting regimen before the transfer. The administration of 5 Gy TBI is a well-established conditioning regimen that induces a profound but transient lymphodepletion lasting for 7 to 10 days.\(^{30,31}\) This dose has previously been shown to augment the effectiveness of adoptive cell transfer therapy in the pmel-1 model through several mechanisms, including the removal of immunoregulatory elements, “cytokine sinks,” and activation of antigen-presenting cells.\(^{30,32}\) We wanted to investigate whether this conditioning regimen induced a sufficient degree of immunosuppression that would enable the transient engraftment of allogeneic cells. As a control, we transferred allogeneic pmel-1 cells into Rag1\(^{-/-}\) hosts, which genetically lack T and B lymphocytes. In both nonirradiated and 5 Gy TBI preconditioned mice, allogeneic cells were rapidly rejected with very little persistence observed at day 5 and no detectable cells at day 10 after transfer. In contrast, allogeneic cells that were transferred into immunodeficient Rag1\(^{-/-}\) recipients not only persisted but expanded over time (Figure S1, available on the Blood website; see the Supplemental Materials link at the top of the online article).

Because 5 Gy TBI did not confer any significant survival advantage to adoptively transferred allogeneic T cells, even at shorter time points, we sought to test whether a stronger lymphodepleting/immunosuppressive regimen would lead to a better engraftment. We treated tumor-bearing C57BL/6 mice preconditioned with 5 or 9 Gy TBI with pmel-1\(^{-bd}\) cells and added a specific vaccination and exogenous IL-2. We analyzed the animals at different time points for the presence of allogeneic cells expressing the MHC class I H-2 D\(^{d}\) molecule. Because 9 Gy is a myeloablative dose of TBI, all groups received an autologous BMT on the day after the transfer of allogeneic pmel-1 T cells. In mice that received 5 Gy TBI, allogeneic cells briefly persisted and were detectable in secondary lymphoid organs at day 5 but completely disappeared by day 10 (Figure 1A). In contrast, allogeneic donor cells were clearly measurable in 9 Gy irradiated hosts up to 24 days after transfer and were rejected only by day 36 (Figure 1A). Allogeneic CD8\(^{+}\) lymphocytes were also present at later time points in the draining lymph nodes and at the tumor site, indicating that they not only persisted but were capable of trafficking to peripheral sites (Figure S2).

We sought to have a more reliable quantitative comparison of adoptively transferred syngeneic and allogeneic T cells. Because pmel-1 T cells are CD8\(^{+}\) and express the V\(\beta\)13 rearrangement of the TCR, we analyzed the absolute numbers of CD8\(^{+}\)V\(\beta\)13\(^{+}\) cells present in the spleens. As shown in Figure 1B, pmel-1 cells reached a peak of expansion around day 5 in both the allogeneic and syngeneic groups and subsequently declined. However, in the 5-Gy setting, allogeneic cells did not persist after day 10. CD8\(^{+}\)V\(\beta\)13\(^{+}\) cells observed in the allogeneic group in Figure 1B past day 5 represented endogenous V\(\beta\)13\(^{+}\) cells because they did not express H-2 D\(^{d}\) (Figure 1A). In contrast, in the 9-Gy setting, allogeneic cells persisted for at least 24 days. These data indicated that, despite the MHC barrier, allogeneic effector cells can transiently engraft for relatively long periods after an increased intensity conditioning regimen.

**Effective tumor treatment with allogeneic effector cells is observed after administration of a highly immunosuppressive conditioning regimen**

Next, we wanted to investigate the ability of allogeneic effector cells to treat an established tumor. We used pmel-1\(^{-bd}\) cells to treat B16 melanoma-bearing C57BL/6 mice. This model eliminates the possibility of a GVHD-like reaction because F1 lymphocytes are tolerant to all antigens in the C57BL/6 background, but rejection of the donor cells remained efficient (Figure S1, Figure 1). This is because the endogenous lymphocytes present in the recipient recognize allogeneic MHC molecules expressed by the tumor-specific lymphocytes. Tumors were implanted subcutaneously and allowed to grow for 8 to 10 days before treatment. On the day of the transfer, recipient mice were preconditioned with 9 Gy TBI and received vaccination and adjuvant administration of exogenous recombinant IL-2. One day after treatment, all groups were injected with syngeneic (C57BL/6 H-2\(^{b}\)) bone marrow cells. This tripartite regimen was previously published by our group and proven highly effective in the eradication of well-established, large vascularized...
We sought to understand whether allogeneic effectors transferred after 9 Gy TBI were rejected by host lymphocytes that differentiate from the bone marrow stem cells or by surviving lymphocytes that repopulate the host by homeostatic expansion. We treated tumor-bearing C57BL/6 (H-2b/b) animals with a myeloablative dose of 9 Gy TBI and adoptive transfer of pmel-1-b/d cells along with vaccination and TBI and therapeutic efficacy, we exposed tumor-bearing mice to increasing doses of irradiation, starting at 5 Gy and escalating up to 11 Gy (Figure 3). Consistent with the previous findings, treatment outcome directly correlated with the dose of TBI. In particular, significant tumor treatment was evident when mice received at least 7 Gy TBI, whereas transfer of allogeneic antitumor cells did not show any effect in 5 Gy-irradiated hosts (5 Gy PVI vs 7 Gy PVI, \( P < .04 \)). Similar data were also obtained using a different pmel-1-F1 (B6-C3H F1 H-2\( ^{ab} \)) as a source of donor cells (data not shown). Taken together, these data suggest that an intense lymphodepleting/immunosuppressive regimen is critical for the therapeutic effectiveness of allogeneic tumor-specific lymphocytes, with obvious implications for the design of future clinical trials.

Adoptive transfer of allogeneic CD4\(^+\) T cells mediates effective tumor regression and ocular autoimmunity

We wanted to test whether our findings could be extended to adoptive immunotherapy based on allogeneic antitumor CD4\(^+\) T helper cells. We used CD4\(^+\) T lymphocytes expressing a Tg TCR-specific melanocyte differentiation antigen, TRP-1.\(^{29}\) These cells were recently described to be able to eradicate large tumors in an autologous setting.\(^{29}\) TRP-1-Tg mice were bred on a C57BL/6 Rag1\(^{--}\)/H-2\( ^{bb} \) background, and allogeneic effectors were generated by crossing them with BALB/c Rag1\(^{--}\)/H-2\( ^{da} \) mice to obtain TRP-1 H-2\( ^{bb} \)/H-2\( ^{da} \) Tg cells.

When given after a preparative regimen of 9 Gy TBI and autologous BMT, TRP-1-b/d cells were detectable for 2 weeks after transfer, as measured by VB14 (the TCR variable chain used by TRP-1) and D\( ^{a} \) (Figure 4A). As was observed with the pmel-1 model, allogeneic TRP-1 cells induced tumor regression (\( P < .02 \) NT vs AlloTI) and complete tumor cure was observed in 3 of 8 mice. Tumor regression using syngeneic T cells seemed more efficient than allogeneic T cells, but the difference did not achieve statistical significance (AlloTI vs SynTI, \( P = .04 \); AlloTI vs NT, \( P = .04 \)). Similar data were also obtained using a different pmel-1-F1 (B6-C3H F1 H-2\( ^{ab} \)) as a source of donor cells (data not shown). Taken together, these data suggest that an intense lymphodepleting/immunosuppressive regimen is critical for the therapeutic effectiveness of allogeneic tumor-specific lymphocytes, with obvious implications for the design of future clinical trials.

Rejection of adoptively transferred lymphocytes is mediated by endogenous T cells rather than those derived from transplanted bone marrow

We sought to understand whether allogeneic effectors transferred after 9 Gy TBI were rejected by host lymphocytes that differentiate from the bone marrow stem cells or by surviving lymphocytes that repopulate the host by homeostatic expansion. We treated tumor-bearing C57BL/6 (H-2\( ^{bb} \)) animals with a myeloablative dose of 9 Gy TBI and adoptive transfer of pmel-1-b/d cells along with vaccination and exogenous rhIL-2 administration. One day after ACT, we injected the animals with bone marrow derived either from wild-type C57BL/6 or Rag1\(^{--} \) mice. In animals transplanted with Rag1\(^{--} \) bone marrow, the host lymphocyte populations can only be replaced by surviving cells that homeostatically proliferate after TBI. Interestingly, we did not observe any statistically significant differences between the groups that received normal or Rag1\(^{--} \) bone marrow in terms of tumor treatment (Figure 5A). Furthermore, allogeneic cells were not detectable at day 30 in both groups (Figure 5B). These results indicate that allogeneic donor cells are
likely rejected by the host lymphocytes that survived the irradiation. Surviving host lymphocytes have been shown in previous studies to play a critical role in early immune-reconstitution of irradiated animals.\(^{37}\)

In the same experiment, we compared allogeneic cell treatment with syngeneic effector cells (Figure 5A). We injected syngeneic pmel-1\(^{\text{b/b}}\) into C57BL/6 mice transplanted either with wild-type or Rag1\(^{-/-}\) bone marrow. We did not observe any difference between the 2 groups that received syngeneic cells but different bone marrow. However, syngeneic cell treatment was superior to allogeneic adoptive transfer (alloPVI wild type [wt] BM vs synPVI wt BM, \(P < .008\); alloPVI Rag1\(^{-/-}\) BM vs synPVI Rag1\(^{-/-}\) BM, \(P < .03\)). Similar differences between allogeneic and syngeneic therapy were observed in other experiments (data not shown).

These data suggest that transient engraftment is sufficient to mediate antitumor activity, but the potency of the treatment is reduced compared with syngeneic treatment.

The risk of inducing a GVHD-like reaction is minimal when the TCR repertoire is confined

Allogeneic cell lines can induce GVHD-like reactions, especially considering the important requirement of a lymphodepleting regimen before cell transfer.\(^{15,30}\) For example, transfusion-associated GVHD could be a serious, even lethal, complication, and it might occur in immunosuppressed persons that receive nonirradiated blood components.\(^{38}\) We wanted to explore whether treatment with
allogeneic CD^8^ T lymphocytes bearing a restricted/monoclonal TCR repertoire had a potential for inducing a GVHD-like reaction. Theoretically, because the pmel-1 TCR is selected on the H-2^D^b MHC class I molecule, it could cross-react with self-antigens expressed on a different haplotype or even to react with a different MHC class I antigens in a peptide-independent manner. We first assessed the ability of pmel-1 cells to react against several different allogeneic MHC. C57BL/6–pmel-1 (H-2^b^/b) cells cocultured with irradiated splenocytes derived from different inbred mouse strains (SJL, H-2^s^/s and DBA, H-2^d^/d) displaying different allogeneic MHC haplotypes in the presence or the absence of the relevant peptide gp100_25-33 (+ pep). Syngeneic C57BL/6 H-2^s^/s irradiated splenocytes in the presence or the absence of the relevant gp100_25-33 peptide were used as positive and negative controls, respectively. Panels A and B are representative of 2 independent experiments. (C) pmel-1 cells were generated on a B6-C3H F1 background (H-2^b^/b) and used in combination with vaccinia virus encoding hgp100 and exogenous rHL-2 to treat B16 tumors established for 10 days in either B6-C3H F1 (SynPVI) or in B6-DBA F1 mice (H-2^d^/b) (AlloPVI). All groups were irradiated with 9 Gy TBI and given autologous BMT. Some groups also received different doses (10^4 or 10^5) of open repertoire B6-C3H F1 CD8^+^ naive T cells in conjunction with pmel-1 cells, vaccine, and rHL-2 (AlloPVI + 10^4 CD8^+^ and AlloPVI + 10^5 CD8^+^). Results of tumor area are the mean of measurements of at least 5 mice per group (± SEM). Data are representative of 2 independent experiments. (D) Percentage of initial weight of mouse groups is shown.

Discussion

In the present manuscript, we sought to explore the use of allogeneic MHC partially matched effector cells for adoptive immunotherapy of cancer. Our results indicate that this approach is a safe, feasible, and effective alternative to allogeneic BMT or autologous antitumor T cells in 2 TCR-Tg mouse models. We observed tumor regression in the absence of severe GVHD when a narrow repertoire of T cells was transferred. In vivo survival and persistence of allogeneic haploidentical cells were critically dependent on the administration of an immunosuppressive treatment before the transfer. With a strong lymphodepleting preparative regimen of 9 GyTBI, allogeneic lymphocytes could survive in vivo for several weeks before being eventually rejected. In addition, allogeneic T cells were able to efficiently migrate to secondary lymphoid organs as well as to the tumor site, where they were detectable in significant numbers.

Transient engraftment is minimal and sufficient to induce significant tumor regression. The persistence of adoptively transferred T cells has been previously linked to favorable outcomes in clinical trials of adoptive cell therapy, but it is uncertain whether this is a prerequisite for effective treatment or only an epiphenomenon correlated with successful immunotherapy but not causal. After the initial peak of activation and proliferation, T lymphocytes might become functionally exhausted or anergic. In the pmel-1 model, antitumor T lymphocytes recovered 2 weeks after transfer are no longer capable of readily
secreting cytokines and proliferating on antigen reencounter in vitro (data not shown). Moreover, our group has recently shown that elimination of effector cells using anti-CD8 antibody after adoptive cell transfer has no impact on tumor treatment as well as on the development of ocular autoimmunity when done 9 days after infusion.42 Long-term persistence might not be a crucial requirement for ACT. On the other hand, we observed diminished effectiveness of allogeneic compared with syngeneic effectors that is probable because of immune-mediated rejection. Interestingly, allogeneic CD4+ TRP-1-specific cells mediated a surprisingly strong antitumor effect in the absence of specific vaccination, even though they had a shorter survival time compared with allogeneic CD8+ pmel-1 cells. Furthermore, the difference between allogeneic and syngeneic CD4+ TRP-1 cells was much less pronounced than in the pmel-1 setting. In contrast to the CD8+ cell-based model, the differences between the 2 groups in the CD4+ cell-based therapy never reached statistical significance, although we did observe a trend toward a better treatment with the use of syngeneic TRP-1 cells. Although definitive conclusions are not easy to draw with the currently available data, these findings might be explained by the ability of T helper cells to recruit and activate endogenous effector cells from both the innate and adaptive branch of the immune system to the tumor site.34,43,44 In addition, CD4+ T cells facilitating the priming of specific antitumor CD8+ T cells in the tumor-draining lymph nodes.45-47 Thus, the requirement for long-term engraftment might be of lesser importance for antitumor CD4+ T cell–based therapies, making allogeneic T helper cells a particularly viable approach deserving a more detailed investigation.

In the trial published by Haque et al, patients received immunosuppression before ACT to prevent graft rejection.20 In that setting, immunosuppression was a mandatory component of a solid organ transplantation, which triggered posttransplantation lymphoproliferative disease, but it also enabled the allogeneic EBV-specific T cells to survive and function. Immunosuppression could be achieved in several ways, but our data indicate that intense lymphodepletion will be required for successful ACT.15,48 Lymphodepleting regimens offer allogeneic effector cells both the possibility to engraft and the chance to expand under the drive of homeostatic cytokines and the elimination of regulatory elements.10,33,49 In addition, TBI and chemotherapy can promote the generation of a proinflammatory environment that supports lymphocyte activation and function.32,50 It might be possible to take advantage of the myelotoxic/immunosuppressive properties of many chemotherapeutic agents by administering antitumor allogeneic cells as an adjunct to already existing regimens. Antimetabolite drugs such as fludarabine, which have already been used as preparatory regimen for adoptive cell transfer therapy,15 profoundly deplete host lymphocyte populations. Fludarabine has been linked to the development of transfusion-associated GVHD several months after its administration.51

Based on previous findings, hematopoietic stem cell transplantations might also be useful in promoting cell expansion.31 Immunosuppressive strategies aimed at decreasing host lymphocyte activation and function might be used to prolong in vivo survival and prevent the development of immunity against alloantigens. Steroids might delay the rejection of allogeneic cells without a negative impact on the tumor therapy.52 Selective in vivo depletions of host lymphocyte subpopulations with specific antibodies could also be used.53,54

Our data indicate that the risk of adverse reaction because of “off target” recognition may be low if a restricted or monoclonal TCR repertoire is used. We did not detect the induction of GVHD unless significant numbers of allogeneic open repertoire cells were admixed with tumor-specific cells. We observed a trend toward an improved tumor treatment in animals that died of GVHD, but this finding is difficult to interpret in the absence of further data. Many publications have suggested that a GVT reaction might effectively destroy solid tumors in the setting of allogeneic BMT,55,56 but we cannot exclude that slower tumor growth was not because of the severely compromised general health of the hosts in our experiments represented by Figure 6. Interestingly, even though allogeneic open repertoire cells were subjected to rejection, they were capable of causing fatal GVHD. A similar observation was made in a phase 1/2 clinical trial where patients were treated with autologous hematopoietic stem cell transplantation followed by the administration of low or high doses of allogeneic haploidentical donor lymphocytes.57 In this trial, patients treated with high-dose donor lymphocyte infusion developed severe adverse reactions and died of GVHD and bone marrow failure. Thus, it might be useful to screen allogeneic T-cell effectors and remove those cells that are alloreactive.58-60 However, because antitumor T cells generated from tumor-infiltrating lymphocytes have an effector/effecter memory phenotype and possess a limited TCR repertoire, the risk of GVHD in this setting should be considerably lower. Indeed, several reports have shown that naive T cells are much more efficient in initiating graft-versus-host reaction compared with memory T cells.51-54

Further applications might include the generation of genetically engineered cell lines, which might be modified to improve survival, proliferation, and effector functions or to deliver proinflammatory molecules to the tumor microenvironment.55-56 In this scenario, short-term engraftment might be an advantage, possibly preventing cells to cause excessive normal tissue damage42 or to undergo malignant transformation.70-74

Taken together, our findings show that allogeneic CD4+ and CD8+ T cells might have a future role in the cell-based immunotherapy of cancer. There are several reasons that make this strategy appealing. “Off-the-shelf” cell products would ensure improved selection, standardization, and quality and safety controls. The establishment of a bank of highly reactive tumor-specific T cells would allow the treatment of more patients and to spread this therapy where extensive laboratory expertise and equipment are not available. Multiple administrations, perhaps targeting different antigens, might be more feasible, but development of host immunity against allogeneic targets might thwart the effectiveness of repeat treatments.

In conclusion, a quickly available and reliable “off-the-shelf” cell source would reduce the time and labor required, speeding delivery of the treatment to the patient and reducing the associated costs.

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Authorship

Contribution: P.M. and A.B. wrote the paper and designed and performed experiments; L.C., C.W., C.M.P., D.C.P., L.G., C.S.H., and C.-C.C. helped with some of the experiments and wrote the paper; and S.A.R. and N.P.R. supervised the research and design and wrote the paper.

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