Brief report

A neopeptide generated by an FLT3 internal tandem duplication (FLT3-ITD) is recognized by leukemia-reactive autologous CD8+ T cells

Claudine Graf, Florian Heidel, Stefan Tenzer, Markus P. Radsak, Fian K. Solem, Cedrik M. Britten, Christoph Huber, Thomas Fischer, and Thomas Wölfel

The FLT3 receptor tyrosine kinase is expressed in more than 90% of acute myelogenous leukemias (AMLs), up to 30% of which carry an internal tandem duplication (ITD) within the FLT3 gene. Although varying duplication sites exist, most FLT3-ITDs affect a single protein domain. We analyzed the FLT3-ITD of an AML patient for encoding HLA class I–restricted immunogenic peptides. One of the tested peptides (YVDFREYEYY) induced in vitro autologous T-cell responses restricted by HLA-A*0101 that were also detectable ex vivo. These peptide-reactive T cells recognized targets transfected with the patient’s FLT3-ITD, but not wild-type FLT3, and recognized the patient’s AML cells. Our results demonstrate that AML leukemia blasts can in principle process and present immunogenic FLT3-ITD neoepitopes. Therefore, FLT3-ITD represents a potential candidate target antigen for the immunotherapy of AML. (Blood. 2007;109:2985-2988)

Introduction

The fibroblast-macrophage stimulating factor receptor (FMS)–like tyrosine kinase receptor 3 (FLT3) is involved in the proliferation, survival, and differentiation of early hematopoietic progenitor cells. More than 90% of acute myelogenous leukemias (AMLs) express FLT3. At the same time, FLT3 represents the most common mutated gene in AML. Two major types of mutation have been described. These are internal tandem duplications (FLT3-ITDs) within the juxtamembrane domain and point mutations within the kinase domain.1-3 FLT3-ITDs occur in 25% to 30% of AMLs and lead to leukemic transformation by constitutive phosphorylation and uncontrolled activation of the tyrosine kinase.4

Targeting FLT3 in AML by small molecules is a novel and promising therapeutic approach in AML.5,6 We reasoned that, in particular, FLT3-ITD–positive AML cells might be specifically targeted also by the T-cell system. Although FLT3-ITDs largely vary in length with duplications of 3 to more than 400 bp,7 about 60% of them occur between codons 591 and 601 and are in frame.1,8 It is likely that duplication regions encode immunogenic neoeptopes that can be presented by individual HLA alleles. Using a reverse-immunology approach, we demonstrated that an individual FLT3-ITD can indeed be immunogenic and can induce autologous antileukemia T-cell responses.

Patients, materials, and methods

Patients, lymphocytes, and cell lines

Sixty-nine-year-old patient J.C. (an anonymous designation) was diagnosed with an FLT3-ITD–positive AML (French-American-British [FAB] M5a). Standard chemotherapy with cytarabine and idarubicin induced a complete remission (CR), but the patient died from recurrent disease 4 months later. Leukapheresis cells (JC-AML cells) and bone marrow cells were obtained at diagnosis. In JC-AML cells, the white blood cell count was 224 \times 10^9/L (224 000/\mu L), 89% of which were monoblasts. In flow cytometry, 99% stained positive for CD33 and 73% for FLT3 (not shown). Peripheral blood mononuclear cells (JC-PBMCs) were collected in first CR after reconstitution. Sample collection was performed with informed consent. Approval was obtained from the Mainz University Hospital Institutional Review Board. The patient carried the HLA–class I (HLA-I) alleles HLA-A*0101, -A*0201, -B*0801, -B*4402, -Cw*0501, and -Cw*0702. K562/HLA cells were stably transfected with the aforementioned HLA-A and -B alleles as published.8 Epstein-Barr virus (EBV)–transformed B lymphocytes of healthy donor A.K. (AK-EBV-B) were cultivated as detailed before.9 Mature dendritic cells (EF-mDCs) were generated from PBMCs of HLA-I–matched healthy donor E.F. as described.10

FLT3-PCR and in vitro transcription of FLT3-mRNA

RNA was isolated from JC-AML cells for reverse transcriptase–polymerase chain reaction (RT-PCR). Full-length wild-type FLT3 (wFLT3) and JC-FLT3-ITD were amplified with primers 5'-atatcgacccacgctccggttg- gcggcgcgcac-3' and 5'-gcggatccggctcagtcgacct gcagctacctc-3', cloned into the vector pRESHyg3 (Clontech Laboratories, Mountain View, CA), and sequenced. The patient’s FLT3-ITD sequence, JC-FLT3-ITD, is shown in Figure 1. For in vitro transcription the plasmids were linearized with BamHI (New England Biolabs, Frankfurt am Main, Germany) permitting full-length transcription of the insert and subsequent polyadenylation of the mRNA as described.12 Polyadenylated mRNA was stored at −80°C.

Peptides

Peptides encoded by the JC-FLT3-ITD were predicted with public domain algorithms.13-15 They were synthesized in an automated peptide synthesizer EPS221 (Abimed, Langen, Germany) following the Fmoc/tBu strategy.

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Synthesis products were analyzed by high-performance liquid chromatography (HPLC) (Varian star, Darmstadt, Germany) and matrix-assisted laser desorption/ionization time-of-flight (MALDI-TOF) mass spectrometry (GSG-future, Bruchsal, Germany). Peptides of less than 80% purity were purified by HPLC.

**In vitro stimulation of JC-FLT3-ITD–specific T cells**

In vitro stimulation (IVS) was performed as published with slight modifications. JC-PBMCs were seeded in 96-well plates (5 × 10⁴ per well) in AIM-V supplemented with 5% human serum, IL-2 (20 U/mL), IL-7 (10 ng/mL), IL-4 (10 ng/mL), and synthetic JC-FLT3-ITD peptides (5 μg/mL) and restimulated on day 7. One of the IVS-responder populations was cloned by limiting dilution. Feeders were allogeneic AK-EBV-B lymphocytes (irradiated, 2 × 10⁶ per well). Stimulators were EF-DCs (irradiated, 1 × 10⁶ per well) transfected with JC-FLT3-ITD mRNA (1 μg per 0.2 × 10⁶ DCs) using TransMessenger Transfection Reagent (Qiagen, Hilden, Germany) according to the manufacturer’s instructions. Restimulations were performed in 7-day intervals.

**ELISPOT assays**

IFN-γ and granulocyte (Grb) enzyme-linked immunospot (ELISPOT) assays were performed as described. Spots were evaluated with computer-assisted video image analysis. Briefly, image acquisition was performed with an AxioCam MRc camera attached to a Zeiss Axio Imager M1 microscope via a 0.63× adapter. Spots were counted with a 3.15× magnification at a resolution of 1388 × 1040 pixels. The software was KS ELISPOT version 4.8. Targets were K562/HLA cells electroporated with mRNA at 300 μF and 250 V or loaded with synthetic peptides (10 μg/mL).

**Results and discussion**

The FLT3-ITD identified in JC-AML cells (JC-FLT3-ITD) encoded a duplication of amino acids 591 to 599 (Figure 1). The same duplication event has already been found in another AML patient, which is in line with the rather limited location of FLT3-ITD occurrence. Five peptides with high combined prediction scores were selected (Figure 1) and used for IVS with JC-PBMCs collected in CR. T-cell responses against peptide YVDFREYEEEY (YVD/A1), predicted to bind to HLA-A*0101, could be repeatedly generated. The peptide was only recognized in association with HLA-A*0101 but not with any other HLA-A or -B allele of patient J.C. (not shown). YVDF/A1-reactive IVS responders recognized K562/A1 transfected with full-length JC-FLT3-ITD mRNA but not K562/A1 transfected with wtFLT3 mRNA (Figure 2A). This indicated that the peptide YVDF/A1 was endogenously processed and proved the existence of JC-FLT3-ITD–specific T cells.

IVS-responder populations containing YVDF/A1-reactive T cells also recognized JC-AML cells (Figure 2A). We cloned IVS JC#1 using HLA-1–matched EF-DCs transfected with JC-FLT3-ITD as stimulators. Ten of 19 T-cell clones recognized YVDF/A1 and JC-AML equally well (Figure 2B), which demonstrated that JC-FLT3-ITD was expressed and processed by JC-AML cells leading to the presentation of YVDF/A1 on their cell surface. When YVDF/A1 was titrated, half-maximal recognition was observed at 20 nM (Figure S1, available on the Blood website; see the Supplemental Figure link at the top of the online article), which is in the range of moderate avidity T cells.

YVDF/A1–specific T cells were detectable ex vivo at similar frequencies in the patient’s peripheral blood lymphocytes (Figure 2C) and in bone marrow lymphocytes (not shown) both drawn at diagnosis. This indicated an in vivo expansion of these T cells. In addition, ex vivo CD8+ T cells secreted Grb in response to YVDF/A1, which demonstrated their cytotoxic ability. It is widely accepted that progressive malignant disease in the presence of antitumor T cells is not an oxymoron considering the complexity of resistance mechanisms already recognized.

Recently, Scholl and colleagues showed the potency of FLT3-ITD–encoded peptides and related mimotopes to bind to HLA-I molecules. However, whether these peptides were processed and recognized by T cells was not explored. Analogous to JC-FLT3-ITD, fusion proteins occurring in leukemia subsets have been found to induce T-cell responses (eg, ETV6/AML1 and TEL/AML1). As newly synthesized FLT3-ITD is retained in the endoplasmic reticulum because of inefficient folding and chaperoning, it appears likely that FLT3-ITD molecules are preferentially degraded after synthesis and introduced into the HLA-I–presenting pathway. This would favor T-cell recognition in case peptides are immunogenic.

It has been shown that FLT3-ITD are present in leukemia stem cells. Targeting of leukemogenic molecules with T cells might improve the chances to eliminate also the quiescent fraction of leukemia stem cells as compared with small inhibitory molecules.

To fully recognize the potential of FLT3-ITD as an immunotherapeutic target, it will be necessary to study in a next step a cohort of FLT3-ITD–positive patients for anti–FLT3-ITD T-cell responses and to include CD8+ as well as CD4+ T-cell responses via all individual HLA-I and -II alleles.

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Authorship

Contribution: C.G. performed the experiments and drafted the manuscript as part of her PhD thesis; F.H. performed FLT3-ITD sequencing and cloning; S.T. and M.P.R. performed peptide predictions; F.K.S. performed PCR-based FLT3-ITD screening and prepared clinical samples; C.M.B. generated stable K562/HLA transfectants; C.H. contributed to the design of the project; T.F. contributed to the design of the project and to the writing of the manuscript; and T.W., C.G.’s thesis supervisor, designed the project and finalized the manuscript.

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Correspondence: Thomas Wolfgel, III Medizinische Klinik, Johannes Gutenberg-Universität, D-55101 Mainz, Germany; e-mail: t.woelfel@3-med.klinik.uni-mainz.de.
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